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ORIGINAL RESEARCH

LncRNA LINC00665 Promotes Prostate Cancer Progression via miR-1224-5p/SND1 Axis

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Background: Increasing researches have revealed a critical role of long noncoding RNAs (lncRNAs) in tumor progression. LINC00665 is a poorly investigated lncRNA. In this research, we sought to determine the potential role of LINC00665 in prostate cancer (PC) progression.

Methods: LINC00665 expression was analyzed by bioinformatics method and qRT-PCR. Proliferation was determined via CCK8 and colony formation assays. Transwell assay was conducted to analyze migration and invasion. Xenograft assay was used to test the roles of LINC00665 in vivo. Luciferase reporter assay, pulldown assay and RIP assay were utilized to confirm the interaction between LINC00665 and miR-1224-5p.

Results: LINC00665 expression was increased in PC samples in contrast to control tissues, according to bioinformatics analysis and qRT-PCR validation. LINC00665 high expression was related to a poor prognosis. LINC00665 knockdown markedly attenuated growth and metastasis of PC cells and impaired tumor propagation in vivo. Mechanistic investigation revealed that LINC00665 was the sponge for miR-1224-5p. By inhibiting miR-1224-5p level, LINC00665 dramatically promoted the expression of SND1 in PC cells. Ectopic expression of SND1 significantly rescued the effects of LINC00665 silencing.

Conclusion: LINC00665 is a novel oncogenic gene in PC by targeting miR-1224-5p/SND1 pathway and may be a therapeutic target.

Keywords: LINC00665, miR-1224-5p, SND1, prostate cancer, progression

Introduction

Prostate cancer (PC) is a very malignant and common cancer in men worldwide.¹ This cancer has gradually become one of the leading causes, threatening men's health.² Although some improvement achieved on PC diagnosis, about 10% of new cases show advanced stages and about 5% ones display distant metastasis.³ Nowadays, the prognosis of PC patients is still unsatisfactory.⁴ Thus, understanding the molecular mechanism of PC pathogenesis is urgently required.

Long noncoding RNAs (lncRNAs) are characterized by no or limited proteincoding potential and have over 200 nucleotides in length.⁵ As the development of RNA sequencing, over 60,000 lncRNAs have been identified in human tissues.⁶ Although some reports have demonstrated the diverse functions of lncRNAs in biological processes, including tumor growth, metastasis and differentiation. Roles of most lncRNAs are still unclear.^{7,8} For example, LINC00460 upregulation in bladder urothelial cancer indicates poor prognosis.⁹ LncRNA H19 overexpression promotes epithelial–mesenchymal transition of papillary thyroid cancer.¹⁰ Aberrant downregulation of lncRNA STXBP5-AS1 in gastric cancer leads to accelerated growth and enhanced metastasis of tumor cells through activation of PI3K/AKT

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© 2020 Chen et al. This work is published and licensed by Dove Medical Press Limited. The full terms of this license are available at https://www.dovepress.com/terms.php you hereby accept the Terms. Non-commercial uses of the work are permitted without any further permission from Dove Medical Press Limited, provided the work is properly attributed. For permission for commercial use of this work, please see paragraphs 4.2 and 5 of our Terms (http://www.dovepress.com/terms.php). signal.¹¹ Due to their important roles, several lncRNAs may be effective biomarkers for diagnosis or prognosis in cancer.^{12,13}

LINC00665 has been reported to promote progression of several cancers, including lung cancer, gastric cancer and breast cancer.14-16 Nevertheless, the functions and mechanism of LINC00665 in PC are unclear. We found that LINC00665 was highly expressed in prostate cancer tissues through bioinformatics analysis and qRT-PCR method. Thus, we sought to determine the potential function and mechanism of LINC00665 in the regulation of PC progression. LINC00665 levels were elevated in PC tissues. LINC00665 knockdown suppressed PC cell proliferation, migration and invasion. Furthermore, LINC00665 was identified to be an effective sponge for miR-1224-5p, which further directly targeted SND1. Through regulating miR-1224-5p/SND1 axis, LINC00665 was demonstrated to act as oncogenic roles. Our results suggested that LINC00665 may serve as a novel target for PC therapy.

Materials and Methods

Clinical Tissue Specimens

Forty-one pairs of PC samples and normal controls were collected from First Affiliated Hospital of Wenzhou Medical University. All samples were histopathologically confirmed by two independent pathologists. This study was approved by the Ethics Committee of First Affiliated Hospital of Wenzhou Medical University. Written informed consents were achieved from patients. This study was conducted in accordance with the Declaration of Helsinki.

Cell Lines and Culture

Human PC cell lines (LNCaP, PC-3, DU-145 and 22RV1) and normal prostate epithelial cell line RWPE-1 were obtained from the American Type Culture Collection (ATCC, Manassas, VA, USA). Cells were cultured as pre-viously described.¹⁷

qRT-PCR

RNA was isolated with TRIzol (Beyotime, Shanghai, China) and then reversely transcribed into complementary DNA (cDNA) using RETROscriptTM Reverse Transcription Kit (Invitrogen). qPCR was carried out by using VeriQuest SYBR Green qPCR Master Mix (Thermo Fisher Scientific, Waltham, MA). U6 was the normalized control for miRNA and 18S was the normalized control for lncRNA. Expression was calculated according to the $2^{-\Delta\Delta Ct}$ method.

Cell Transfection

To knock down LINC00665, siLINC00665 (#1: 5'-TACAA AGAAACAGGTGAAATT-3'; #2: 5'-GGGATTACAGGTCC AGATTTT-3') or control siRNA (5'-AATTCTCCGAACGT GTCACGT-3') was transfected into DU-145 or LNCaP cells. After 48 hrs, the transfection efficiency was validated by qRT-PCR. siRNAs were purchased from GenePharma (Shanghai, China). For SND1 overexpression, the coding sequence of SND1 was constructed into the pcDNA3 vector.

CCK8 Assay

CCK8 assay (Dojindo, Kumamoto, Japan) was used to detect cell proliferation. In brief, 2000 cells per well were seeded into 96-well plates. After cultured for 0 day, 1 day, 2 days and 3 days, 10 μ L CCK8 solution per well was added and incubated for 2 hrs. Then, absorbance at 450 nm was determined using a VarioskanTM LUX microplate reader (Thermo Fisher Scientific).

Colony Formation Assay

For colony formation assay, 500 cells per well were seeded into the 6-well plates. After cultured for 14 days, the clones were fixed and stained using crystal violet. And the colony number was calculated.

EdU Assay

EdU assay was performed as described before.¹⁸ Briefly, tumor cells were incubated with 10 μ mol/L EdU for 4 h, followed by Hoechst staining. EdU positive cells were then analyzed by FACS.

Transwell Assay

Transwell assays were performed as reported.¹⁹

Xenograft Assay

Four-week Balb/c nude female mice were from Shanghai Laboratory Animal Company (Shanghai, China). DU-145 cells were injected subcutaneously into the right flank of nude mice (n=5 for each group). And the tumor volumes were measured every week. After 5 weeks, the tumor tissues were removed and weighed. The animal experiments were approved by the Ethics Committee of First Affiliated Hospital of Wenzhou Medical University. All animal operations were performed in accordance with the Animal Policy and Welfare Committee of First Affiliated Hospital of Wenzhou Medical University.

Luciferase Reporter Assay

Luciferase reporters were designed and reporter assay was performed as previously described.²⁰ The association between LINC00665 and miR-1224-5p or between miR-1224-5p and SND1 was tested by using the Dual-Luciferase Reporter Assay System (Promega, Madison, WI). In brief, 5000 DU-145 cells per well were seeded into 24-well plates and transfected with pmirGLO reporter (Promega) containing wild-type (wt) or mutant (mut) LINC00665 or SND1-3'-UTR region and miR-1224-5p mimics using Lipofectamine 3000 reagent (Invitrogen). After 48h, luciferase activity was determined according to the manufacturer's protocols.

Statistical Analysis

GraphPad Prism Software was used for statistical analyses. Student's *t*-test and one-way ANOVA were used for analysis of statistical significance as appropriately. Kaplan-Meier analysis and log-rank test were used for analysis of survival rate. P < 0.05 was considered as statistically significant.

Results

Expression Patterns of LINC00665 in PC Tissues and Cell Lines

To analyze important lncRNAs in PC progression, we used bioinformatics approach and identified LINC00665 as a significantly upregulated lncRNA in PC tissues (Figure 1A), which was further confirmed via TCGA data (Figure 1B). Through qRT-PCR analysis, we validated LINC00665 upregulation in 41 PC tissues compared to adjacent normal tissues (Figure 1C). Kaplan-Meier analysis show overexpression of LINC00665 predicted decreased survival rate in PC patients (Figure 1D). Then, LINC00665 level in PC cell lines was examined. We found that LINC00665 levels were also upregulated in PC cell lines (Figure 1E).

Effects of LINC00665 Knockdown on PC Cell Proliferation

Then, we chose DU-145 and LNCaP cells for experiments. We knocked down LINC00665 expression in DU-145 and LNCaP cells by two independent siRNAs (Figure 2A). CCK8 assay was then performed. LINC00665 knockdown significantly inhibited proliferation of DU-145 and LNCaP cells (Figure 2B). Similarly, LINC00665 silencing led to reduced colony number and EdU incorporation in DU-145 and LNCaP cells (Figure 2C and D), suggesting LINC00665 regulates PC cell proliferation. Moreover, we performed

xenograft experiments. At indicated time points, we measured tumor volumes. And 5 weeks later post-injection, we analyzed tumor weights. The results showed that LINC00665 knockdown suppressed tumor volumes and weights significantly (Figure 2E and F). Thus, LINC00665 promotes PC cell growth in vitro and in vivo.

LINC00665 Knockdown Suppressed PC Cell Migration and Invasion

Afterwards, Transwell assay was carried out. We found that LINC00665 knockdown markedly reduced numbers of migrated and invaded DU-145 and LNCaP cells (Figure 3A and B), indicating that LINC00665 regulates PC cell metastasis.

LINC00665 Was a Sponge for miR-1224-5p

To further explore the mechanism, we analyzed the possible targets of LINC00665 through bioinformatics analysis (miRDB tool). We chose the ten most potential target miRNAs for validation. Through luciferase reporter assay, we found that only miR-1224-5p mimics inhibited the luciferase activity of LINC00665 reporter (Figure 4A). Then, we further constructed LINC00665-wt and mut luciferase reporter vectors (Figure 4B). Luciferase reporter assays demonstrated miR-1224-5p mimics suppressed the activity of LINC00665-wt vector but not the mutant one (Figure 4C). RNA pulldown assay also showed that LINC00665 was enriched by biotin-labeled miR-1224-5p-wt but not the mutant one or control (Figure 4D), suggesting a direct interaction between miR-1224-5p and LINC00665. RIP assay using anti-Ago2 also confirmed the direct interaction between LINC00665 and miR-1224-5p (Figure 4E). Importantly, LINC00665 knockdown also increased the level of miR-1224-5p and vice versa (Figure 4F and G). We also found a negative correlation between miR-1224-5p and LINC00665 in PC tissues (Figure 4H). Thus, LINC00665 was a sponge for miR-1224-5p.

SND1 Was Targeted by miR-1224-5p

We also analyzed the targets of miR-1224-5p through TargetScan7 tool. Many potential candidates were identified. And SND1 ranked top among all candidates. Thus, we chose it to validate. Firstly, the wt and mut SND1 luciferase reporter vectors were cloned (Figure 5A). Luciferase reporter assay showed that SND1-WT reporter activity was decreased as miR-1224-5p overexpression (Figure 5B). MiR-1224-5p mimics also inhibited SND1 level (Figure 5C). Moreover,

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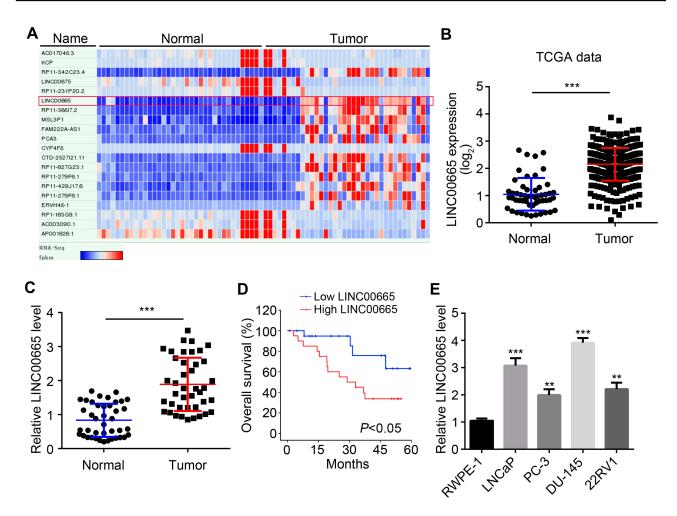


Figure I Expression patterns of LINC00665 in PC tissues and cell lines. (A) Differentially expressed lncRNAs in PC tissues and normal tissues according to TCGA database were analyzed through bioinformatics method (http://incrnator.ewha.ac.kr/index.html). The heatmap data were directly available from the lncRNAtor database. TCGA: the cancer genome atlas. (B) Expression patterns of LINC00665 in PC tissues and normal controls based on TCGA data according to the TANRIC tool (https://ibl.mdanderson.org/tanric/_design/basic/query.html). (C) 41 pairs of PC tissues and normal tissues were used for the analysis of LINC00665 expression by qRT-PCR. 185 was as the normalized control. (D) Survival rate analysis of PC patients based on LINC00665 expression levels. The 41 patients were divided into LINC00665 high or low expression groups. (E) LINC00665 expression was determined in PC cell lines by qRT-PCR. **P<0.01 and ***P<0.001.

qRT-PCR analysis and TCGA data indicated that SND1 expression was upregulated in PC tissues (Figure 5D and E), suggesting it may be an oncogene. We also observed that miR-1224-5p expression was negatively correlated with SND1 in PC tissues (Figure 5F).

Ectopic Expression of SND1 Reversed the Effects of LINC00665 Knockdown

Through qRT-PCR analysis, we found that LINC00665 silencing suppressed SND1 expression (Figure 6A). However, inhibition of miR-1224-5p reversed it (Figure 6A). Thus, LINC00665 promoted SND1 expression by inhibiting miR-1224-5p in PC. To determine whether LINC00665 regulates PC progression through SND1, we overexpressed SND1 and confirmed its upregulation by qRT-PCR (Figure 6B). Then, we performed rescue assays. CCK8, colony formation and EdU incorporation

assays indicated that ectopic expression of SND1 significantly promoted proliferation of DU-145 and LNCaP cells after LINC00665 knockdown (Figure 6C–E). Similarly, the abilities of migration and invasion were also rescued by SND1 overexpression in LINC00665-depleted DU-145 and LNCaP cells (Figure 6F and G). More importantly, we found that SND1 overexpression further promoted PC progression compared to the control group (Figure 6C–G). Thus, LINC00665 promotes PC progression through upregulating SND1 expression.

Discussion

As increasing numbers of lncRNAs are identified by RNAsequencing, more and more attention are attracted to their functions in tumorigenesis.²¹ In our work, we revealed LINC00665 expression was upregulated in PC tissues. Furthermore, LINC00665 upregulation indicated unsatisfactory prognosis.

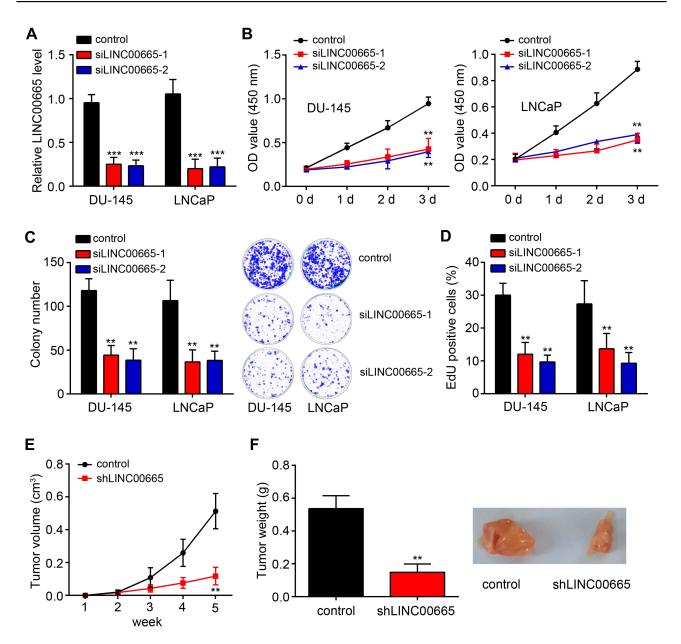


Figure 2 Effects of LINC00665 knockdown on PC cell proliferation. (A) DU-145 and LNCaP cells were transfected with siRNAs against LINC00665 or control. LINC00665 expression was then analyzed. (B–D) Proliferation of PC cells was analyzed by CCK8, colony formation and EdU incorporation assays after siRNA transfection. (E) Tumor volumes were measured at indicated time points. (F) Tumor weights were analyzed 5 weeks later. **P<0.01 and ***P<0.001.

We revealed that LINC00665 could activate SND1 expression via sponging miR-1224-5p. Then, upregulated SND1 expression led to enhanced proliferation, migration and invasion of PC cells.

Previously, several lncRNAs were reported to modulate various biological processes of PC cells.^{22,23} For example, lncRNA FOXC2-AS1 promotes growth and metastasis of PC cells through regulating miR-1253/EZH2 axis.²⁴ LncRNA LOXL1-AS1 promotes CCND1 expression to facilitate the growth of PC cells via sponging miR-541-3p.²⁵ PVT1 promotes PC cell proliferation and migration through modulating P38 expression.²⁶ As a poorly researched lncRNA,

LINC00665 was reported to promote lung cancer progression and tumor cell resistance to Gefitinib.^{14,27} Whether LINC00665 participates in PC progression remains largely unclear. We firstly identified LINC00665 as a significantly upregulated lncRNA in PC tissues by bioinformatics analysis. We also validated it by qRT-PCR. Moreover, we revealed that LINC00665 expression was closely linked to patients' prognosis. Thus, LINC00665 might be a biomarker for prognosis in PC patients. Then, through loss-of-function assay, we demonstrated that LINC00665 knockdown suppressed growth and metastasis of PC cells.

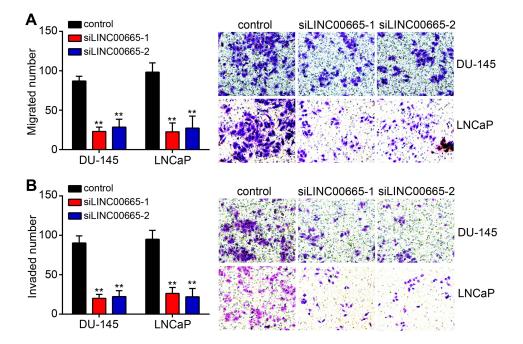


Figure 3 LINC00665 knockdown suppressed PC cell migration and invasion. (A) After transfection with indicated siRNAs, migration of DU-145 and LNCaP cells was analyzed by Transwell assay. (B) LINC00665 silencing suppressed the invasion of DU-145 and LNCaP cells. **P<0.01.

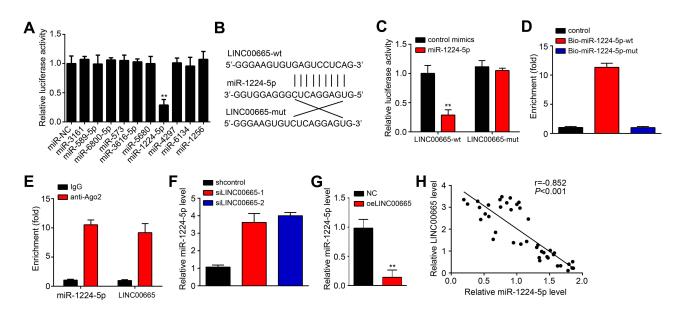


Figure 4 LINC00665 was a sponge for miR-1224-5p. (A) Luciferase reporter assay was performed using LINC00665 reporter and the ten most potential target miRNAs. (B) Diagram of the putative binding site in LINC00665 with miR-1224-5p and strategy for construction of LINC00665 mutation. wt: wild-type; mut: mutant. (C) Activity of LINC00665-wt reporter was inhibited by miR-1224-5p mimics. (D) LINC00665 was precipitated by bio-miR-1224-5p-wt, but not bio-miR-1224-5p-mut or control. (E) MS2based RIP assay indicated that LINC00665 directly interacted with miR-1224-5p in DU-145 cells. (F) LINC00665 knockdown led to increased expression of miR-1224-5p in DU-145 cells. U6 was as miR-1224-5p control. (G) LINC00665 overexpression inhibited the expression of miR-1224-5p in DU-145 cells. (H) There was a reverse correlation between miR-1224-5p and LINC00665 in PC tissues. U6 was as miR-1224-5p control and 18S was as LINC00665 control. **P<0.01.

LncRNA was shown to function as ceRNA for miRNAs.^{28,29} For instance, lncRNA LINC00460 sponges miR-939-5p to promote colorectal cancer invasion.³⁰ LncRNA SLCO4A1-AS1 contributes to bladder cancer progression by inhibiting miR-335-5p.³¹ LncRNA PVT1

facilitates bladder cancer development by sponging miR-143.³² Thus, we analyzed the potential targets of LINC00665. We identified miR-1224-5p and demonstrated miR-1224-5p bound to LINC00665. MiR-1224-5p was found to suppress the development of several

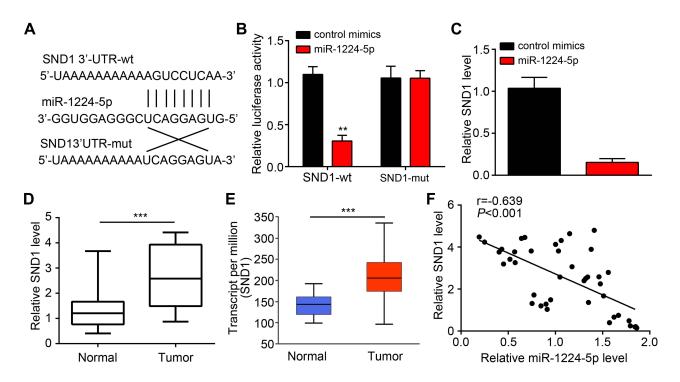


Figure 5 SND1 was targeted by miR-1224-5p. (**A**) Diagram of the putative binding site in SND1 with miR-1224-5p and strategy for construction of SND1 mutation. wt: wild-type; mut: mutant. (**B**) Activity of SND1-wt reporter was inhibited by miR-1224-5p mimics. (**C**) SND1 expression was inhibited by miR-1224-5p in DU-145 cells. miR-1224-5p mimics were transfected into cells for miR-1224-5p overexpression. Then, qRT-PCR was performed to detect SND1 expression. I8S was as the normalized control of SND1. (**D**) SND1 expression was upregulated in the 41 PC tissues compared to corresponding control normal tissues. (**E**) In TCGA data, SND1 level was increased in PC tissues compared to normal controls. (**F**) An inverse correlation between SND1 and miR-1224-5p was observed. **P<0.01 and **P<0.001.

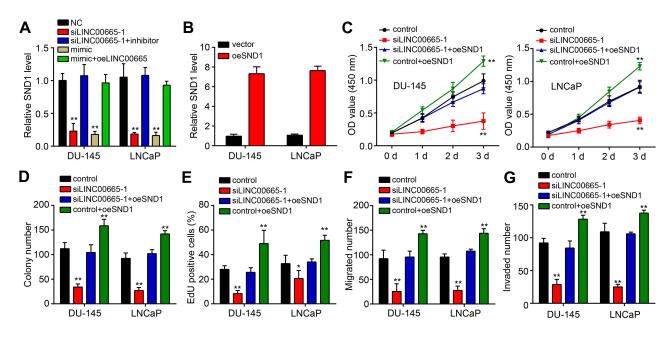


Figure 6 Ectopic expression of SND1 reversed the effects of LINC00665 knockdown. (A) qRT-PCR was used to analyze SND1 expression in DU-145 and LNCaP cells transfected with indicated vectors. GAPDH was used for normalized control. Inhibitor indicates miR-1224-5p inhibitors. (B) The upregulation of SND1 was confirmed after transfection by qRT-PCR. GAPDH was as the normalized control. (C–E) CCK8, colony formation and EdU incorporation assays were utilized to test cell proliferation. (F and G) Cell migration and invasion were determined by Transwell assay. *P<0.05 and **P<0.01.

cancers, including gliomas and melanoma.^{33,34} Its roles in PC have not been reported. In our research, our data indicated that miR-1224-5p may be a tumor suppressor

in PC. Furthermore, we also found the direct target of miR-1224-5p by bioinformatics analysis and identified SND1 (Staphylococcal nuclease domain-containing 1).

Importantly, we found that LINC00665 promotes SND1 expression by inhibiting miR-1224-5p in PC cells.

SND1 has been shown to be overexpressed in several human tumor tissues. A recent study reported that SND1 overexpression promotes growth and metastasis of glioma cells.³⁵ Besides, other works also indicated that SND1 upregulation contributes to tumorigenesis, such as osteo-sarcoma, hepatocellular carcinoma and breast cancer.^{36–38} Our data indicated that SND1 expression was upregulated in PC tissues. Moreover, we found that SND1 significantly enhanced proliferation, migration and invasion of PC cells transfected with LINC00665 siRNA, indicating the oncogenic roles of SND1 in PC.

Summarily, we demonstrated LINC00665 promoted PC progression through targeting miR-1224-5p/SND1.

Data Sharing Statement

All data were included in the manuscript.

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Disclosure

The authors report no conflicts of interest in this work.

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