

## The clinical and immunological features of the post-extracorporeal shock wave lithotripsy anti-glomerular basement membrane disease

Beining Wang<sup>a,b,c,d,e</sup>, Jia Xiaoyu<sup>a,b,c,d,e</sup>, Xiaojuan Yu<sup>a,b,c,d,e</sup>, Zhao Cui<sup>a,b,c,d,e</sup> and Minghui Zhao<sup>a,b,c,d,e,f</sup>

<sup>a</sup>Renal Division, Peking University First Hospital, Beijing, China; <sup>b</sup>Institute of Nephrology, Peking University, Beijing, China; <sup>c</sup>Key Laboratory of Renal Disease, Ministry of Health of China, Beijing, China; <sup>d</sup>Key Laboratory of CKD Prevention and Treatment, Ministry of Education of China, Beijing, China; <sup>e</sup>Research Units of Diagnosis and Treatment of Immune-mediated Kidney Diseases, Chinese Academy of Medical Sciences, Beijing, China; <sup>f</sup>Peking-Tsinghua Center for Life Sciences, Beijing, China

### ABSTRACT

**Introduction:** Extracorporeal shock wave lithotripsy (ESWL) is a noninvasive modality to treat urolithiasis, with complications including tissue damage and hematoma of kidney parenchyma. Anti-glomerular basement membrane (GBM) disease is suggested to be a rare complication of ESWL since it was reported in several cases to occur after ESWL. However, the clinical and immunological features of the ESWL-associated anti-GBM disease have not been fully investigated so far.

**Case Presentation:** Here, we present the clinical, pathological, and immunological characteristics of three patients with the post-ESWL anti-GBM disease in our hospital. Anti-GBM disease occurred within a median of 22 months after ESWL treatment. It presented with similar clinical features to the classic anti-GBM disease, including fever, gross hematuria, and rapidly progressive glomerulonephritis (RPGN) with poor renal prognosis. Sera from all patients recognized the  $\alpha 3(\text{IV})\text{NC1}$  in GBM, but with IgG2 and IgG4 as the dominant IgG subclasses.

**Conclusion:** Although further exploration is required to prove the causal relationship in this rare condition, our study reminds physicians that patients developing acute renal insufficiency after ESWL should lead to the suspicion of anti-GBM disease and in-time diagnosis and treatment.

### ARTICLE HISTORY

Received 28 September 2020  
Revised 1 December 2020  
Accepted 16 December 2020

### KEYWORDS



ESWL; lithotripsy; anti-glomerular basement membrane disease; antigen


### Introduction

Anti-glomerular basement membrane (GBM) disease or Goodpasture (GP) disease is a rare but fulminant and fatal disorder. It is widely accepted as a prototypical autoimmune disease induced by autoantibodies targeting the glomerular and alveolar basement membrane [1,2]. The well-defined autoantigen is localized to the non-collagen domain (NC1) of the  $\alpha 3$  chain of type IV collagen [ $\alpha 3(\text{IV})\text{NC1}$ ] in GBM [3]. In addition, the autoantibodies have been proven to recognize the other 4  $\alpha$  chains ( $\alpha 1$ , 2, 4, and 5) within type IV collagen [4,5]. Two conformational epitopes, EA and EB, were mapped inside  $\alpha 3(\text{IV})\text{NC1}$  [6] and sequestered within the GBM in healthy individuals [7]. It is still unknown how the immune tolerance would be breached and epitopes exposed. One of the accepted mechanisms is that factors such as smoking [8], hydrocarbon exposure [9], infection [10], or preceding glomerular diseases [11–13]

would damage the basement membrane and unmask the epitopes [14]. Rare cases have been reported about anti-GBM diseases associated with urinary obstruction [15–18] or nephrectomy [19].

Extracorporeal shock wave lithotripsy (ESWL) is a common and noninvasive modality to treat urolithiasis, disintegrating stones by shock waves. Although generally safe [20], it could cause short-term complications as renal colic or obstruction by stone fragments, tissue damage or hematoma, and even temporary reduction of glomerular filtration rate [21]. Renal histopathological findings of human and animal models shortly after ESWL included focal damage to vascular endothelium, nephron, renal tubules, and interstitia [20,22–24]. In 1990, Guerin et al. first reported a 67-year-old male with rapidly progressive renal dysfunction 7 months after ESWL. His serum anti-GBM antibody was positive, while the serum before ESWL was not [25]. Since then,

**CONTACT** Minghui Zhao  [mhzhao@bjmu.edu.cn](mailto:mhzhao@bjmu.edu.cn)  Renal Division, Peking University First Hospital, No. 8 Xishiku Street, Xicheng District, 100034 Beijing, China

 Supplemental data for this article can be accessed [here](#).

© 2021 The Author(s). Published by Informa UK Limited, trading as Taylor & Francis Group.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

there have been several case reports about the post-ESWL anti-GBM disease, proposing anti-GBM disease as a rare complication of ESWL [26–29]. However, the causal relationship between the two has not been elucidated yet, nor the antigen spectrum or IgG subclasses of the autoantibodies in this rare entity.

In this study, we identified three patients with the post-ESWL anti-GBM disease after retrospectively reviewing 166 consecutive patients diagnosed with the anti-GBM disease in Peking University First Hospital from January 1, 2010 to January 31, 2020. We analyzed their clinical, pathological, and immunological features and reviewed five previously reported cases, aiming to draw a more comprehensive picture and bring insight into possible etiologies of this rare condition.

## Case report

### Case 1

A 36-year-old male was otherwise healthy except for undergoing one session of ESWL for kidney stones 24 months ago. Kidney function (represented as serum creatinine) was normal. Twenty-four months later, he presented with fever, cough, and hemoptysis for 5 days. He was anemic with hemoglobin 75 g/L and serum creatinine was 691  $\mu\text{mol/L}$  (eGFR, 8 mL/min/1.73m<sup>2</sup>). Two days later, he developed gross hematuria and serum creatinine rose to 884  $\mu\text{mol/L}$  (eGFR, 6 mL/min/1.73m<sup>2</sup>), at which time he was referred to our hospital. He was a long-term cigarette smoker and had gasoline exposure 15 days ago.

On admission, he was pale and febrile with no lymphadenopathy. Breath sounds were weak in both lungs on auscultation. Serum creatinine rapidly increased to 1279  $\mu\text{mol/L}$  (eGFR, 4 mL/min/1.73m<sup>2</sup>) with oliguria and hemoglobin decreased to 49 g/L. Chest radiography suggested bilateral pulmonary hemorrhage. The circulating anti-GBM antibody was positive, with the titer >200 RU/mL (Euroimmune ELISA kit, normal range <20 RU/mL), while the serum anti-neutrophil cytoplasmic antibody (ANCA) was undetected (Table 1).

Anti-GBM disease was diagnosed based on the renal-pulmonary involvement and positive serum anti-GBM antibody. The patient was prompted to plasma exchange, intravenous methylprednisolone pulse treatment, cyclophosphamide, and hemodialysis. After 17 sessions of plasma exchange, he became afebrile without hemoptysis. His serum anti-GBM antibody decreased to 33 RU/mL and serum creatinine to 425  $\mu\text{mol/L}$  (eGFR, 14 mL/min/1.73m<sup>2</sup>). Repeated chest radiography indicated the absorption of his hemorrhage. He

was dialysis-independent on discharge and a 3-month follow-up.

Sera from this patient were tested for antigen spectrum using recombinant human  $\alpha$ 1–5(IV)NC1 chains and chimeric proteins containing epitopes EA and EB on  $\alpha$ 3(IV)NC1 (Supplementary materials and methods). The patient's serum recognized  $\alpha$ 1 and  $\alpha$ 3(IV)NC1 domains, and both epitopes EA and EB of  $\alpha$ 3(IV)NC1. The immunoglobulin G (IgG) subclass distribution against  $\alpha$ 3(IV)NC1 was also tested for the sera. All 4 subclasses of IgG were detected, with the dominance of IgG2 and IgG4 (Table 1; Figure S1).

### Case 2

A 50-year-old male undertook one session of ESWL for ureter stones with normal kidney function 22 months ago. He was a 20-year cigarette smoker but had quit smoking for 10 years. Twenty-two months later he was referred to our hospital for fever, gross hematuria, and rapidly progressive renal dysfunction. He had suffered from intermittent fever for 2 months without other indicating symptoms and it could not be resolved by antibiotics. Fifteen days prior to admission, he developed gross hematuria and the serum creatinine progressed from 160  $\mu\text{mol/L}$  (eGFR, 40 mL/min/1.73m<sup>2</sup>) to 504  $\mu\text{mol/L}$  (eGFR, 11 mL/min/1.73m<sup>2</sup>). No stones were found by renal ultrasound.

After admission, he was diagnosed with anti-GBM disease due to positive serum anti-GBM antibody (183 RU/mL). His serum ANCA was tested negative. He received 7 sessions of plasma exchange, intravenous methylprednisolone pulse treatment, and hemodialysis. His condition improved and hematuria disappeared afterward. Serum creatinine decreased to 353  $\mu\text{mol/L}$  (eGFR, 16 mL/min/1.73m<sup>2</sup>) and anti-GBM antibody titer to 51 RU/mL on discharge. The patient was dialysis-dependent during a 60-month follow-up (Table 1).

The patient's sera reacted with all five  $\alpha$  chains and both EA and EB epitopes of  $\alpha$ 3(IV)NC1. All 4 IgG subclasses to  $\alpha$ 3(IV)NC1 were detected with the dominance of IgG2 (Table 1; Figure S1).

### Case 3

A 74-year-old female received one session of ESWL for kidney stones 10 months ago when the renal function was normal. Ten months after ESWL, she complained of fever and gross hematuria with urinary irritation symptoms for 2 weeks. No leukocytosis was found, but her urinalysis revealed white and red blood cells. Initial workups showed elevated C-reactive protein (96 mg/L)

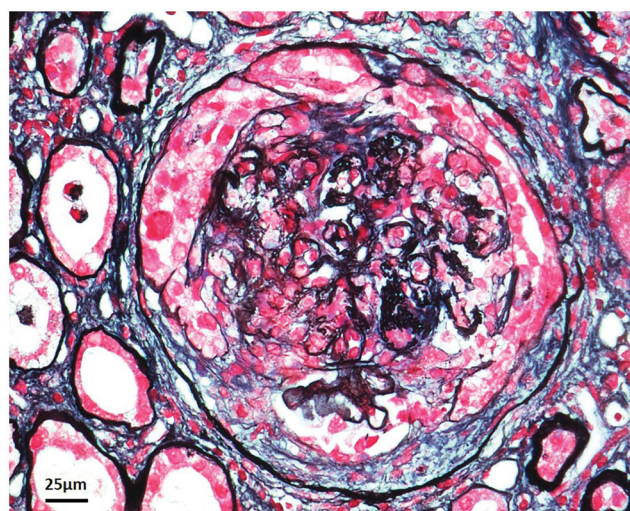
**Table 1.** Clinical, pathological, and immunological features of patients with post-ESWL anti-GBM disease in our hospital.

	Patient 1	Patient 2	Patient 3
Sex/age, y	M/36	M/50	F/74
Hydrocarbon exposure (Y/N)	Y	N	N
Smoking (Y/N)	Y	Y	N
Prodromal infection (Y/N)	Y	N	Y
Fever (Y/N)	Y	Y	Y
Pulmonary hemorrhage (Y/N)	Y	N	N
Gross hematuria (Y/N)	Y	Y	Y
Oliguria/Anuria (Y/N)	Y	N	Y
Hemoglobin on diagnosis, g/L	49.0	102.0	63.0
Serum albumin, g/L	30.0	27.1	29.0
Urinary protein, g/24h	2.0	1.6	0.6
Nephrotic syndrome (Y/N)	N	N	N
Serum creatinine on diagnosis, $\mu\text{mol/L}$	1279	364	1066
eGFR on diagnosis, $\text{mL/min}/1.73\text{m}^2$	4.0	15.0	3.0
Anti-GBM antibodies on diagnosis, RU/mL	>200	183	196
Positive ANCA (Y/N)	N	N	N
Treatment	PE/MP/Pred/CTX	PE/MP/Pred	MP/Pred/CTX
Anti-GBM antibody on discharge, RU/mL	33	51	102
Serum creatinine on discharge, $\mu\text{mol/L}$	425	353	618
eGFR on discharge, $\text{mL/min}/1.73\text{m}^2$	14.0	16.3	5.3
Dialysis-dependent on discharge (Y/N)	N	Y	Y
Duration of follow-up, m	3	60	48
Dialysis-dependent at last follow-up (Y/N)	N	Y (PD)	Y (HD)
Death at last follow-up (Y/N) (cause of death)	N	N	Y (Esophagus cancer)
Stone location	kidney	ureter	kidney
Total ESWL number	1	1	1
ESWL-to-onset interval, m	24	22	10
Anti- $\alpha 1$ (IV)NC1 antibody (ref range, <0.04)	0.06	0.89	0.33
Anti- $\alpha 2$ (IV)NC1 antibody (ref range, <0.05)	0.01	0.83	0.12
Anti- $\alpha 3$ (IV)NC1 antibody (ref range, <0.06)	2.11	2.08	2.15
Anti- $\alpha 4$ (IV)NC1 antibody (ref range, <0.58)	0.27	1.43	0.70
Anti- $\alpha 5$ (IV)NC1 antibody (ref range, <0.02)	0.00	0.48	0.10
Anti- $\alpha 3$ EA antibody (ref range, <0.07)	1.39	1.67	1.61
Anti- $\alpha 3$ EB antibody (ref range, <0.15)	0.59	1.54	1.35
Anti- $\alpha 3$ (IV)NC1 IgG1 (ref range, <0.02)	0.68	0.57	0.73
Anti- $\alpha 3$ (IV)NC1 IgG2 (ref range, <0.07)	1.51	1.32	1.25
Anti- $\alpha 3$ (IV)NC1 IgG3 (ref range, <0.01)	0.53	0.49	0.63
Anti- $\alpha 3$ (IV)NC1 IgG4 (ref range, <0.01)	2.34	0.56	0.43

M: male; F: female; eGFR: estimated glomerular filtration rate; GBM: glomerular basement membrane; ANCA: anti-neutrophil cytoplasmic antibody; PE: plasma exchange; MP: methylprednisolone pulse; Pred: prednisone; CTX: cyclophosphamide; PD: peritoneal dialysis; HD: hemodialysis; ESWL: extracorporeal shock wave lithotripsy; IgG: immunoglobulin G; ref: reference; Y: yes; N: no.

and normal serum creatinine ( $87\ \mu\text{mol/L}$ ; eGFR,  $55\ \text{mL/min}/1.73\text{m}^2$ ). Renal ultrasound showed no stones. She was diagnosed with urinary tract infection and treated by intravenous antibiotics, after which she reported no symptom improvement but developed oliguria and bilateral edema of lower extremities. Her serum creatinine deteriorated to  $1066\ \mu\text{mol/L}$  (eGFR,  $3\ \text{mL/min}/1.73\text{m}^2$ ) within 9 days. Then she was referred to our hospital.

On admission, physical examination revealed a sick and pale woman with no rales on auscultation. Serum anti-GBM antibody was positive ( $196\ \text{RU/mL}$ ) while the serum ANCA was not. Renal biopsy revealed severe necrosis of glomerular capillary walls and cellular/cellular-fibrous crescents in all 32 glomeruli on light microscopy (Figure 1), with no electron-dense deposits on electron microscopy. The immunofluorescence microscopy was negative on a sclerotic glomerulus. She was treated by methylprednisolone pulse and cyclophosphamide. Plasma exchange was not performed, given



**Figure 1.** Renal pathology of patient 3 with post-extracorporeal shock wave lithotripsy (ESWL) anti-GBM disease showed cellular crescent formation in a glomerulus by periodic acid-silver methenamine and Masson trichrome stain on light microscopy ( $400\times$ ).

**Table 2.** Clinical and pathological data of previously reported post-ESWL anti-GBM disease cases.

	1	2	3	4	5
Authors	Cranfield et al.	Sellin et al.	Xenocostas et al.	Iwamoto et al.	Guerin et al.
Publication year	2015	2005	1999	1998	1990
Sex/age, y	F/67	M/32	M/72	F/37	M/67
Hydrocarbon exposure (Y/N)	NA	NA	NA	NA	NA
Smoking (Y/N)	NA	NA	NA	NA	Y
Prodromal infection (Y/N)	N	N	Y	N	N
Fever (Y/N)	Y	Y	Y	Y	Y
Pulmonary hemorrhage(Y/N)	Y	N	N	N	N
Gross hematuria (Y/N)	NA	Y	N	Y	NA
Serum creatinine on diagnosis, $\mu\text{mol/L}$	1179	919	NA	1114	1074
Positive anti-GBM antibody(Y/N)	Y	Y	Y	Y	Y
Positive ANCA (Y/N)	N	NA	N	N	N
HLA phenotype	DR4, DQ6	DRB1*11 & 13	DR15	DR2	DR2
Treatment	PE/MP/Pred/CTX	Pred/CTX	NA	PE/MP/Pred/CTX	NA
Dialysis-dependent (Y/N)	Y	Y	Y	Y	Y
Renal biopsy (Y/N)	NA	Y	Y	Y	Y
Immunofluorescence	NA	Linear IgG deposits along GBM	Linear IgG deposits along GBM	Linear IgG and C3 deposits along GBM	Linear IgG and C3 deposits along GBM
Light microscopy	NA	Crescent formation in 23/25 glomeruli	Crescent formation in nearly all glomeruli	Cellular crescents in all glomeruli	Cellular crescents in all glomeruli
Electron-dense deposits on electron microscopy	NA	NA	N	N	NA
Stone location	NA	Infundibulum	Left renal pelvis	Right kidney	Left kidney
Stone component	NA	Calcium oxalate	NA	NA	NA
Total ESWL number	2 within 4 weeks	3 within 4 months	1	NA	2 within 10 days
ESWL-to-onset interval	1 week	5 months	3 months	3 months	7 months
Shock number	3200	NA	4000	1000	NA
Energy data	75 kPa	NA	NA	17 kV	NA
Renal function at last ESWL	NA	NA	Normal	Normal	Normal
Anti-GBM antibody before ESWL	NA	NA	NA	Negative	Negative

F: female; M: male; GBM: glomerular basement membrane; ANCA: anti-neutrophil cytoplasmic antibody; HLA: human leukocyte antigen; PE: plasma exchange; MP: methylprednisolone pulse; Pred: prednisone; CTX: cyclophosphamide; IgG: immunoglobulin G; C3: complement 3; ESWL: extracorporeal shock wave lithotripsy; ref: reference; Y: yes; N: no; NA: not available.

the poor prognosis based on renal biopsy. Her renal function did not recover and she remained dialysis-dependent. The patient died of esophagus cancer 2 years after discharge.

The patient's sera recognized all five  $\alpha$  chains and both EA and EB epitopes of  $\alpha 3(\text{IV})\text{NC1}$ . As for circulating IgG to  $\alpha 3(\text{IV})\text{NC1}$ , all four subclasses were detected with the dominance of IgG2 (Table 1; Figure S1).

## Discussion

In this study, we described three patients with anti-GBM disease occurring within 2 years after ESWL. The incidence of such cases among all anti-GBM patients from our hospital in the past 10 years was 1.81% (3/166). Their clinical manifestations, pathological features, and antigen spectrum were similar to the classic anti-GBM disease. All three patients presented with fever, gross hematuria, and rapidly progressive glomerulonephritis (RPGN) with poor renal prognosis. One of them suffered from pulmonary hemorrhage. The one available renal biopsy revealed 100% crescent formation in the glomeruli. All three patients recognized

$\alpha 3(\text{IV})\text{NC1}$  and EA/EB epitopes. None were positive for anti-neutrophil cytoplasmic antibody (ANCA).

Post-ESWL anti-GBM disease has been rarely reported. After searching in PubMed using keywords 'anti-GBM', 'Goodpasture', 'ESWL', and 'lithotripsy', we found five previous cases of the post-ESWL anti-GBM disease with full text in English, reviewed in Table 2. Among the five patients, three of them were male and two were female, with a median age of 67 years old (range, 32–72 years). Human leukocyte antigen (HLA) phenotyping was performed in all five patients, with four of them expressing susceptible serotypes for anti-GBM disease [30]. All previous cases presented with shorter ESWL-to-onset intervals (1 week to 7 months) than ours, possibly due to more ESWL treatment numbers. Renal damage by ESWL appears to be cumulative in animal experiments and human, proportional to the application frequency and the number of treatments [31–33]. Their clinical manifestations, renal pathologies, and prognosis were similar to ours. Specifically, fever occurred in all previous reports and ours. However, only a few of them (3/8, 37.5%) were reported to bear prodromal infections and none in either group were positive for ANCA. Fever was a common feature in

anti-GBM disease patients, as shown in our previous study in a 140-patient cohort, and 78.7% of febrile patients had infections [34]. The infection rate in post-ESWL anti-GBM disease patients seemed to be lower, which might indicate distinct disease triggers other than prodromal infections in these patients.

Further studies are still needed to elucidate the effect of ESWL on the initiation of anti-GBM disease. However, there has been evidence about the collagen cleaving effect of ultrasound since early 1980s when researchers tried to isolate and define the components of GBM by physical methods. Glomeruli were sequestered and sonicated to separate the GBM, which then released split products of collagen with antigenicity [35], indicating ultrasound could cause local damage of the GBM. There was also evidence of transient nephrotic-range proteinuria immediately after ESWL in a patient cohort [36] and mesangial proliferative glomerulopathy after ESWL in experimental animal models of pig [37]. Moreover, shock wave lithotripsy is utilized in pancreatic and large common bile duct stones or sialolithiasis, but no cases of anti-GBM disease have been reported in either of these conditions yet, suggesting ESWL for urinary stones may affect kidneys more directly. Therefore, ESWL could possibly damage and expose the antigen inside the GBM *via* direct damage by shock waves or secondary damage by immune-complex induced by ESWL debris [37].

Anti-GBM IgG subclass distribution is associated with disease severity [38]. The IgG1 and IgG3 dominated the IgG subclasses in patients with severe renal impairment, while IgG2 and IgG4 were associated with milder renal damage [38]. In post-ESWL anti-GBM disease patients from this study, the IgG2 and IgG4 were the dominant subclasses against  $\alpha 3(\text{IV})\text{NC1}$ . However, these patients presented with severe renal damage and poor prognosis. The IgG subclass switching after B cell activation follows the sequence of IgG3→IgG1→IgG2→IgG4. It has been suggested that IgG4 production results from chronic or repetitive antigenic stimulation [39,40]. Moreover, low-level natural anti-GBM autoantibodies existed in healthy human sera, predominantly of IgG2 and IgG4 [41]. We speculated that ESWL might expose GBM autoantigens and induce IgG autoantibodies chronically during the ESWL-to-onset interval, allowing them to complete subclass switching. The autoantibodies may accumulate beyond a threshold to disturb the immune tolerance in healthy individuals and provoke the pathogenic autoimmunity.

Anti-GBM diseases were also reported scarcely to associate with obstructive uropathy due to urinary malignancy, neurogenic bladder, or ureteral stenosis

[15–18]. The anti-GBM antibody level and renal function seemed to be parallel with treatment efficacy of hydro-nephrosis [18]. Intact NC1 hexamer could be detected in the serum and be secreted in the urine of healthy individuals [17,42]. Rabbits immunized with their own urinary concentrate developed anti-GBM glomerulonephritis [43]. It was suggested that urinary  $\alpha 3(\text{IV})\text{NC1}$  under urinary obstruction might enter the renal interstitia, dissociate under acidic pH changed by inflammatory infiltrate (such as infection) and act as an immunogen [17]. These studies provide another theory for the induction of anti-GBM disease associated with urinary stones, including patients treated by ESWL.

The limitations of our study lay in at least two points. Firstly, the number of cases is too small to draw the causal relationship between ESWL and anti-GBM disease. Moreover, we cannot rule out the possibility that individuals with anti-GBM disease or with susceptible HLA phenotypes may bear a higher risk of urinary obstructions, which lead to the production of anti-GBM antibodies. A prospective cohort study in patients undergoing ESWL might be required to further elucidate the relationship between the procedure and anti-GBM disease. Secondly, not all patients had detailed ESWL information (stone component, shock wave frequency, and energy), kidney biopsy, and HLA phenotypes for us to draw a full picture.

In summary, the anti-GBM disease could happen within weeks to months after ESWL treatment and present with similar clinical features, antigen spectrum, and prognosis to classic anti-GBM disease. IgG2 and IgG4 were the two dominant subclasses against  $\alpha 3(\text{IV})\text{NC1}$ . Although the causal relationship between ESWL and anti-GBM disease still needs further exploration, our study here may act as a reminder for physicians that patients developing acute renal insufficiency after ESWL should lead to the suspicion of anti-GBM disease and in-time diagnosis and treatment.

### Geolocation information

City: Beijing, Latitude: 39.9289, Longitude: 116.3883.

### Ethical policy and institutional review board statement

Written informed consents were obtained from the patients for publication of the three cases and any accompanying images. The experiments in this study complied with the Declaration of Helsinki and were approved by the ethics committee of Peking University First Hospital.

## Disclosure statement

No potential conflict of interest was reported by the author(s).

## Funding

This work was supported by the National Natural Science Foundation of China [grant number 81870482] and CAMS Innovation Fund for Medical Sciences [grant number 2019-I2M-5-046].

## References

- [1] McAdoo SP, Pusey CD. Anti-glomerular basement membrane disease. *Clin J Am Soc Nephrol*. 2017;12(7):1162–1172.
- [2] Cashman SJ, Pusey CD, Evans DJ. Extraglomerular distribution of immunoreactive Goodpasture antigen. *J Pathol*. 1988;155(1):61–70.
- [3] Saus J, Wieslander J, Langeveld JPM, et al. Identification of the Goodpasture antigen as the  $\alpha 3$ (IV) chain of collagen IV. *J Biol Chem*. 1988;263(26):13374–13380.
- [4] Hellmark T, Johansson C, Wieslander J. Characterization of anti-GBM antibodies involved in Goodpasture's syndrome. *Kidney Int*. 1994;46(3):823–829.
- [5] Yang R, Hellmark T, Zhao J, et al. Antigen and epitope specificity of anti-glomerular basement membrane antibodies in patients with goodpasture disease with or without anti-neutrophil cytoplasmic antibodies. *JASN*. 2007;18(4):1338–1343.
- [6] Netzer KO, Leinonen A, Boutaud A, et al. The goodpasture autoantigen. Mapping the major conformational epitope(s) of alpha3(IV) collagen to residues 17-31 and 127-141 of the NC1 domain. *J Biol Chem*. 1999;274(16):11267–11274.
- [7] Hudson BG, Tryggvason K, Sundaramoorthy M, et al. Alport's syndrome, Goodpasture's syndrome, and type IV collagen. *N Engl J Med*. 2003;348(25):2543–2556.
- [8] Donaghy M, Rees A. Cigarette smoking and lung haemorrhage in glomerulonephritis caused by autoantibodies to glomerular basement membrane. *Lancet*. 1983;322(8364):1390–1393.
- [9] Stevenson A, Yaqoob M, Mason H, et al. Biochemical markers of basement membrane disturbances and occupational exposure to hydrocarbons and mixed solvents. *QJM*. 1995;88(1):23–28.
- [10] Couser WG, Johnson RJ. The etiology of glomerulonephritis: roles of infection and autoimmunity. *Kidney Int*. 2014;86(5):905–914.
- [11] Olson SW, Arbogast CB, Baker TP, et al. Asymptomatic autoantibodies associate with future anti-glomerular basement membrane disease. *JASN*. 2011;22(10):1946–1952.
- [12] Basford AW, Lewis J, Dwyer JP, et al. Membranous nephropathy with crescents. *J Am Soc Nephrol*. 2011;22(10):1804–1808.
- [13] Jia XY, Hu SY, Chen JL, et al. The clinical and immunological features of patients with combined anti-glomerular basement membrane disease and membranous nephropathy. *Kidney Int*. 2014;85(4):945–952.
- [14] Pedchenko V, Bondar O, Fogo AB, et al. Molecular architecture of the Goodpasture autoantigen in anti-GBM nephritis. *N Engl J Med*. 2010;363(4):343–354.
- [15] Bazari H, Mauiyyedi S. Case records of the Massachusetts General Hospital. Weekly clinicopathological exercises. Case 4-2002. A 75-year-old man with acute renal failure five months after cystoprostatectomy and urethrectomy for carcinoma. *N Engl J Med*. 2002;346(5):353–360.
- [16] Blas A, 3rd, McGill R, Sandroni S. Case 4-2002: cancer-associated obstruction and glomerular damage. *N Engl J Med*. 2002;346(22):1751–1752.
- [17] Weber M, Pullig O, Boesken WH. Anti-glomerular basement membrane disease after renal obstruction. *The Lancet*. 1990;336(8713):512–513.
- [18] Takeuchi Y, Takeuchi E, Kamata K. A possible clue for the production of anti-glomerular basement membrane antibody associated with ureteral obstruction and hydronephrosis. *Case Rep Nephrol Dial*. 2015;5(1):87–95.
- [19] O'Hagan E, Mallett T, Convery M, et al. Anti-GBM disease after nephrectomy for xanthogranulomatous pyelonephritis in a patient expressing HLA DR15 major histocompatibility antigens: a case report. *Clin Nephrol Case Stud*. 2015;3:25–30.
- [20] Türk C, Petřík A, Sarica K, et al. EAU guidelines on interventional treatment for urolithiasis. *Eur Urol*. 2016;69(3):475–482.
- [21] Skolarikos A, Alivizatos G, de la Rosette J. Extracorporeal shock wave lithotripsy 25 years later: complications and their prevention. *Eur Urol*. 2006;50(5):981–990; discussion 90.
- [22] Karlsen SJ, Smevik B, Hovig T. Acute morphological changes in canine kidneys after exposure to extracorporeal shock waves. A light and electron microscopic study. *Urol Res*. 1991;19(2):105–115.
- [23] Recker F, Hofmann W, Bex A, et al. Quantitative determination of urinary marker proteins: a model to detect intrarenal bioeffects after extracorporeal lithotripsy. *Journal of Urology*. 1992;148(3 Part 2):1000–1006.
- [24] Delvecchio F, Auge BK, Munver R, et al. Shock wave lithotripsy causes ipsilateral renal injury remote from the focal point: the role of regional vasoconstriction. *J Urol*. 2003;169(4):1526–1529.
- [25] Guerin V, Rabian C, Droz D, et al. Anti-glomerular-basement-membrane disease after lithotripsy. *Lancet*. 1990;335(8693):856–857.
- [26] Iwamoto I, Yonekawa S, Takeda T, et al. Anti-glomerular basement membrane nephritis after extracorporeal shock wave lithotripsy. *Am J Nephrol*. 1998;18(6):534–537.
- [27] Xenocostas A, Jothy S, Collins B, et al. Anti-glomerular basement membrane glomerulonephritis after extracorporeal shock wave lithotripsy. *Am J Kidney Dis*. 1999;33(1):128–132.

- [28] Sellin L, Quack I, Weiner SM, et al. Nephrolithiasis and hematuria-sometimes a stony road to diagnosis. *Clin Nephrol.* 2005;64(2):151–154.
- [29] Cranfield A, Mathavakkannan S. Goodpasture's disease following extracorporeal shock wave lithotripsy: a case report & literature review. *Clin Case Rep.* 2015;3(3):160–164.
- [30] Fisher M, Pusey CD, Vaughan RW, et al. Susceptibility to anti-glomerular basement membrane disease is strongly associated with HLA-DRB1 genes. *Kidney Int.* 1997;51(1):222–229.
- [31] Koga H, Matsuoka K, Noda S, et al. Cumulative renal damage in dogs by repeated treatment with extracorporeal shock waves. *Int J Urol.* 1996;3(2):134–140.
- [32] Kang DH, Cho KS, Ham WS, et al. Comparison of high, intermediate, and low frequency shock wave lithotripsy for urinary tract stone disease: systematic review and network meta-analysis. *PLoS One.* 2016;11(7):e0158661
- [33] Chung JM, Park BK, Kim JH, et al. Impact of repeated extracorporeal shock wave lithotripsy on prepubertal rat kidney. *Urolithiasis.* 2018;46(6):549–558.
- [34] Gu QH, Xie LJ, Jia XY, et al. Fever and prodromal infections in anti-glomerular basement membrane disease. *Nephrology (Carlton).* 2018;23(5):476–482.
- [35] Lubec G. Anti-glomerular basement membrane disease after lithotripsy. *Lancet.* 1990;335(8702):1405.
- [36] Gilbert BR, Riehle RA, Vaughan ED. Extracorporeal shock wave lithotripsy and its effect on renal function. *J Urol.* 1988;139(3):482–485.
- [37] Banner B, Ziesmer D, Collins LA. Proliferative glomerulopathy following extracorporeal shock wave lithotripsy in the pig. *Journal of Urology.* 1991;146(5):1425–1428.
- [38] Zhao J, Yan Y, Cui Z, et al. The immunoglobulin G subclass distribution of anti-GBM autoantibodies against rAlpha3(IV)NC1 is associated with disease severity. *Hum Immunol.* 2009;70(6):425–429.
- [39] Magnusson CG, Cesbron JY, Djurup R, et al. Raised serum IgG4 levels in patients with atopy and filariasis: application of an automated particle-counting immunoassay using monoclonal antibody. *Int Arch Allergy Appl Immunol.* 1986;81(3):238–244.
- [40] Ljungstrom I, Hammarstrom L, Kociecka W, et al. The sequential appearance of IgG subclasses and IgE during the course of *Trichinella spiralis* infection. *Clin Exp Immunol.* 1988;74(2):230–235.
- [41] Cui Z, Wang HY, Zhao MH. Natural autoantibodies against glomerular basement membrane exist in normal human sera. *Kidney Int.* 2006;69(5):894–899.
- [42] Schuppan D, Besser M, Schwarting R, et al. Radioimmunoassay for the carboxy-terminal cross-linking domain of type IV (basement membrane) procollagen in body fluids. Characterization and application to collagen type IV metabolism in fibrotic liver disease. *J Clin Invest.* 1986;78(1):241–248.
- [43] Lerner RA, Dixon FJ. The induction of acute glomerulonephritis in rabbits with soluble antigens isolated from normal homologous and autologous urine. *J Immunol.* 1968;100(6):1277–1287.