

# Oncogene Overdose: Too Much of a Bad Thing for Oncogene-Addicted Cancer Cells

## Supplementary Issue: Signaling Pathways as Biomarkers

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**ABSTRACT:** Acquired resistance to targeted inhibitors remains a major, and inevitable, obstacle in the treatment of oncogene-addicted cancers. Newer-generation inhibitors may help overcome resistance mutations, and inhibitor combinations can target parallel pathways, but durable benefit to patients remains elusive in most clinical scenarios. Now, recent studies suggest a third approach may be available in some cases—exploitation of oncogene overexpression that may arise to promote resistance. Here, we discuss the importance of maintaining oncogenic signaling at “just-right” levels in cells, with too much signaling, or oncogene overdose, being potentially as detrimental as too little. This is highlighted in particular by recent studies of mutant-BRAF in melanoma and the fusion kinase nucleophosmin–anaplastic lymphoma kinase (NPM–ALK) in anaplastic large cell lymphoma. Oncogene overdose may be exploitable to prolong tumor control through intermittent dosing in some cases, and studies of acute lymphoid leukemias suggest that it may be specifically pharmacologically inducible.

**KEYWORDS:** oncogene addiction, oncogene overdose, intermittent dosing, the Goldilocks principle, ALK, BRAF

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## Introduction

For most cancers, the hope that single-molecule targeted therapy would revolutionize treatment has been thwarted by acquired resistance. Mechanisms may be target-dependent, typically mutations or overexpression, or target-independent, such as the activation of parallel pathways.<sup>1</sup> Although rational strategies exist to overcome parallel signaling and resistance-conferring mutations (reviewed elsewhere<sup>2–6</sup>), the intriguing possibility of exploiting target over-expression is now also entering discussion. Here, we review the feasibility of this approach, where target upregulation can cause an overdose of oncogenic signaling that is detrimental to cancer cell survival. By exploiting the “Goldilocks principle”—the idea that even for oncogenes “just-right” levels are required—strategies such as intermittent dosing may permit prolonged tumor control in some patients.

## Oncogene Addiction

Though cancer cells accumulate numerous genetic changes, deactivation of individual oncogenes often cause pronounced tumor regressions in mouse-model systems. Bernard Weinstein

first used the term “oncogene addiction” in 2002 to describe these findings.<sup>7</sup> Strong clinical support came with the remarkable and durable efficacy of breakpoint cluster region–Abl kinase (BCR–ABL) tyrosine kinase inhibitors (TKIs) in treating chronic myeloid leukemia (CML) without significant toxicity.<sup>8,9</sup> Such findings revolutionized cancer drug development toward targeted therapies to exploit every tumor's perceived Achilles' heel and ultimately driving a new paradigm of personalization of cancer care through precision medicine.<sup>6,10–12</sup> Unfortunately, however, CML still stands more or less alone in the success of this strategy. Inhibitors targeting other driver kinases produce progression-free survival (PFS) times of typically a few months and have failed to replace frontline chemotherapy in most diseases.<sup>13–16</sup> Even when other inhibitors have moved to the frontline, as the epidermal growth factor receptor (EGFR) inhibitor erlotinib and the anaplastic lymphoma kinase (ALK) inhibitor crizotinib have for appropriate advanced lung-cancer patients, median PFS is ~9–10 months.<sup>17,18</sup> In TKI-treated CML by contrast, median PFS is still unknown because it has not been reached after a decade or more of follow-up.<sup>9,19–22</sup>



## Newer Understandings of Oncogene Addiction

Difficulties in recapitulating the success of BCR-ABL kinase inhibitors vs. CML in other cancers have highlighted the deep gaps in our comprehension of oncogene addiction's complexities. Through recent work, however, it is now well understood that the inhibition of a single target can lead to the rewiring of signaling pathways that may rescue the activation of key downstream processes. For example, in triple-negative breast cancer, Duncan et al<sup>23</sup> showed how MEK/ERK inhibition promotes the degradation of MYC, leading to the expression and activation of various receptor tyrosine kinases (RTKs). These RTKs overcome inhibition of MEK2 (but not MEK1), reactivating ERK and culminating in drug resistance. Knock-down of ERK, MYC, AKT, or mTOR recapitulates this reprogramming of the kinome, while preventing MYC's proteasomal degradation blocks it.<sup>23–25</sup>

**Oncogenic shock.** The “oncogenic shock” model provides a framework for conceptualizing such results. Briefly, while oncogenes promote proliferation and survival, they paradoxically activate signals to promote apoptosis or cell cycle arrest.<sup>26,27</sup> This may be because of feedback repression of normal survival pathways (as above) and/or through stress imposed by increased cellular growth rates, similar to oncogene-induced senescence in nontransformed cells.<sup>28</sup> Either way, when an inhibitor of a driver oncogene is introduced to the system, both the growth/survival and the proapoptotic/growth-arrest signals are inhibited. In truly oncogene-addicted cells, abrupt loss of the survival signal destroys the cells before the loss of the apoptotic signal can allow them to survive and vice versa for the cells that survive.<sup>6,26,27</sup>

The oncogenic shock model, which was originally proposed nearly a decade ago, has received strong support from recent results with key clinical targeted inhibitors. In CML, the poster-child disease for oncogene addiction, for example, Asmussen et al<sup>29</sup> found that BCR-ABL represses normal myeloid survival pathways through feedback inhibition mediated by activated MEK, and these pathways come rushing back several hours after BCR-ABL inhibition. However, drug sensitivity is preserved because cells commit to apoptosis before rescue can occur. The story is different for BRAF<sup>V600E</sup>-driven melanoma (Fig. 1A), where inhibitors typically delay progression for only a few months. Here, the time taken for MAPK pathway rebound is a lot quicker than that for CML (2–4 hours vs. 8–24 hours).<sup>29,30</sup> Therefore, resistance arises in mutant BRAF melanomas, as growth factor pathway restoration is established before a significant proportion of cells enter apoptosis.

While apoptosis precedes the restoration of oncogenic signaling through network rewiring in wild-type BCR-ABL cells,<sup>29</sup> complications arise in the context of resistance. For example, the BCR-ABL inhibitors imatinib, dasatinib, and nilotinib have weak off-target activity against RAF that drives RAS-dependent paradoxical BRAF and CRAF activation.<sup>31</sup> Because BCR-ABL is upstream of RAS

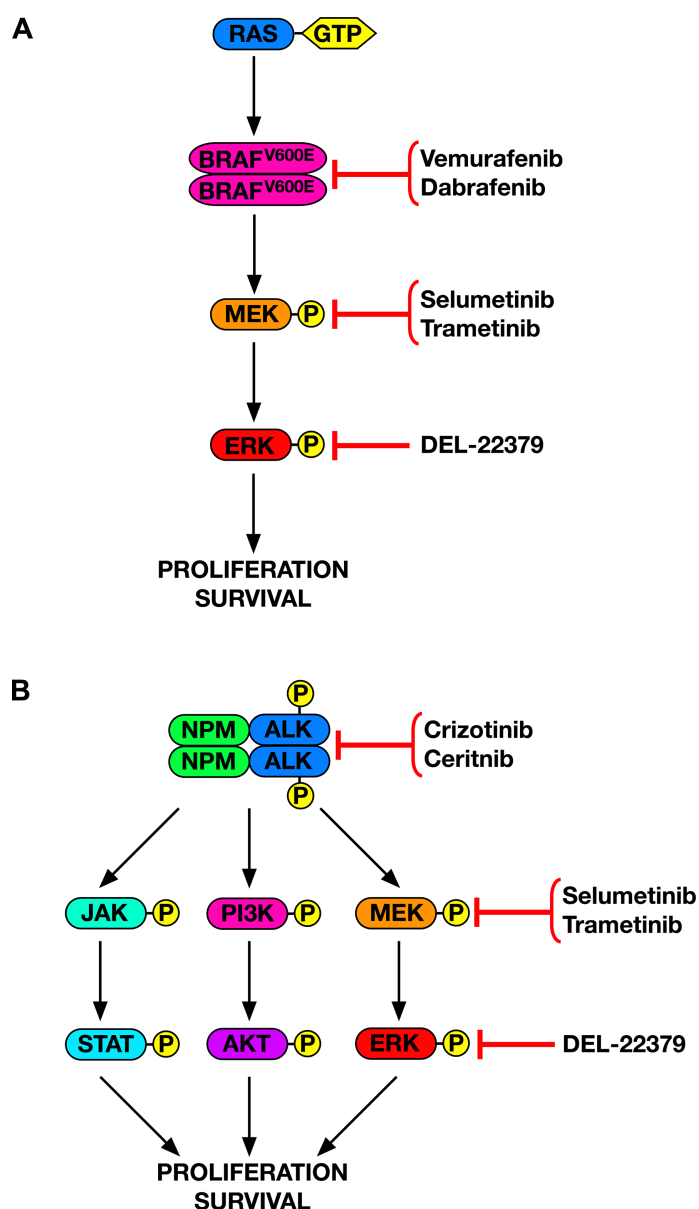
activation in sensitive cells, inhibition with these agents also suppresses RAS. In drug-resistant cells with the T315I gatekeeper BCR-ABL mutation, however, RAS activity permits the drugs' paradoxical BRAF/CRAF and downstream MEK/ERK activation (Fig. 1A). This results in dependency on this pathway, such that combining BCR-ABL inhibitors with a MEK inhibitor synergistically inhibited resistant cell growth in vitro and in vivo.<sup>31</sup> However, prolonged MEK inhibition can cause autocrine activation of STAT3 via fibroblast growth factor receptors and Janus kinases (JAKs),<sup>32</sup> supporting even further combination therapy, such as adding JAK/STAT inhibitors to the drug cocktail. Combined drug toxicities to patients, however, become a rising concern as more and more drugs are put together. Instead, what may be more practical clinically would be scheduling drug dosing intermittently, carefully pulsed to ensure target inhibition followed by withdrawal to prevent pro-survival pathway reactivation. Important work is being carried out to understand the specific timings of these antagonistic processes and their responses in different contexts based on factors like protein turnover.<sup>6</sup> In addition, a new approach to target the MAPK pathway—the direct inhibition of ERK with the dimerization inhibitor DEL-22379—was recently reported.<sup>33</sup> This drug showed ability to overcome many resistance mechanisms that thwart MEK inhibitors but has not yet been tested in BCR-ABL-driven models.

## Oncogene Overdose

While extensive efforts are underway targeting signals to which cancer cells are addicted, the prospect of using these very signals to overwhelm the system is just now entering exploration. Similar to drug addicts overdosing on the very thing they require to avoid withdrawal, cancer cells may be susceptible to the induction of overdoses in oncogenic signaling. Recent studies indicate that this may be feasible in BRAF<sup>V600E</sup>-addicted melanoma, ALK-addicted T-cell lymphoma, and BCR-ABL+ B-cell acute lymphoblastic leukemia (B-ALL).

### Oncogene overdose in BRAF<sup>V600E</sup> melanoma.

While BRAF<sup>V600E</sup>-addicted melanomas rely on continual BRAF<sup>V600E</sup> → MEK → ERK signaling<sup>34</sup> (Fig. 1A), Das Thakur et al<sup>35</sup> found that the continual administration of vemurafenib caused resistant tumors to actually become dependent on the continued inhibition of this very pathway. Resistance arose through elevated BRAF<sup>V600E</sup> expression, which permitted survival during drug exposure but proved toxic when the drug was withdrawn. MEK1/2 inhibition with AZD6244 (selumetinib) also rescued from the BRAF<sup>V600E</sup> overdose signaling. These observations were made in patient-derived xenograft models and confirmed similar findings reported previously in melanoma cell lines.<sup>36</sup> The fitness benefit provided by increased BRAF<sup>V600E</sup> expression during drug exposure therefore becomes a fitness liability when the inhibitor is withdrawn<sup>35</sup> (Fig. 2). Resistant tumors showed impaired



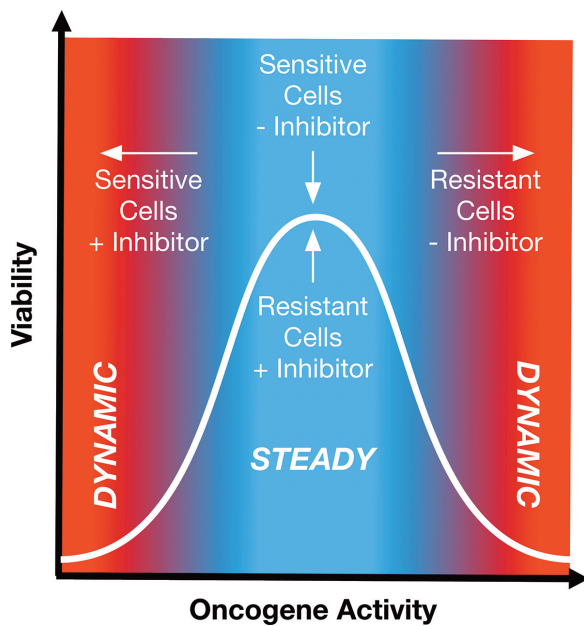
**Figure 1.** Downstream signaling and targeted pathway inhibition for mutant BRAF in melanoma (A) NPM-ALK in ALCL (B).

**Notes:** (A) The dimerization of mutant BRAF in melanoma, activated by RAS, turns on several pro-survival and pro-proliferation pathways driving tumor growth in melanoma, which can be targeted using small molecule inhibitors. (B) The same pathways are activated through dimerization and subsequent trans-autophosphorylation of several ALK-fusion kinases (NPM-ALK shown above).

engraftment to additional host animals treated with vehicle vs. those treated with vemurafenib. In addition, drug withdrawal in vemurafenib-treated animals produced tumor regressions concomitant with spikes in MEK/ERK activity.

To combat resistance to *BRAF*<sup>V600E</sup> inhibitors that reestablishes continual BRAF<sup>V600E</sup> → MEK → ERK signaling despite target inhibition, clinical trials have assessed the efficacy of dual BRAF<sup>V600E</sup> and MEK inhibitor therapy.<sup>37,38</sup> However, resistance to this dual approach is already reported, arising due to augmentation and/or combination of single-agent resistance mechanisms. Moriceau et al<sup>39</sup> established several cell lines from melanomas that had acquired resistance to dual BRAF<sup>V600E</sup> and MEK inhibition. One showed

ultra-amplification of *BRAF*<sup>V600E</sup> (>160 copies), which leads to *CRAF* activation. Another showed low copy number gain (20 copies) together with an *MEK1* mutation that increases BRAF<sup>V600E</sup>-mutant-MEK interactions. Both events led to ERK activation, resulting in growth and survival signals (Fig. 1A). However, both cell lines were also strongly addicted to both inhibitors, as dual inhibitor withdrawal led to a loss of viability via pERK hyperactivation.<sup>39</sup> In fact, a stronger drug addiction phenotype was observed in resistant lines grown ex vivo when patients had been administered dual inhibitors as opposed to single BRAF<sup>V600E</sup> inhibition. Unfortunately, while looking over the cases of old patients, although the authors were able to identify tumor regression after dual drug



**Figure 2.** Oncogene-addicted cells can require “just the right” amount of signaling for survival. Oncogene-addicted cells constitutively express the amount of activated oncogene required for growth and proliferation. Targeted inhibition of the oncogene shuts off this signaling and shifts the cells into a dynamic state leading to death. Resistance can be achieved by increased expression and/or activation of the oncogene in question, which allows resistant cells to grow in the presence of the inhibitor originally designed to kill them. Resistance by this means is often accompanied by dependence, such that inhibitor withdrawal causes an overdose of oncogenic signaling that overwhelms the cells and also results in death.

cessation, they did not observe disease stabilization or uniform tumor regression culminating in clinical remission. They only observed decelerated tumor growth in melanomas where single BRAF<sup>V600E</sup> inhibitor therapy was stopped.

**Oncogene overdose in ALK+ ALCL.** The anaplastic lymphoma kinase (ALK) is an important new therapeutic target activated through chromosomal translocations that fuse its C-terminal kinase domain to the N-terminus of various constitutively expressed proteins (Fig. 1B). Examples include t(2;5)(p23;q35) creating *NPM* (nucleophosmin)-*ALK*, found in ~70% of anaplastic large cell lymphomas (ALCL), and inv(2)(p21;p23) creating *EML4* (echinoderm microtubule associated protein like 4)-*ALK*, found in 3–5% of non-small cell lung cancers (NSCLC).<sup>40–42</sup> Two U.S. Food and Drug Administration approved ALK TKIs, crizotinib and ceritinib, show activity in both ALK+ NSCLC and ALCL.<sup>16,18,43,44</sup> While resistance in ALK+ NSCLC mainly occurs via the activation of alternative signaling pathways, such as EGFR and c-KIT, as well as the acquisition of second-site mutations, the major mechanism of resistance in ALK+ ALCL reported thus far has been ALK-domain mutations.<sup>45–55</sup>

We recently established, however, that over-expression of *NPM-ALK* reliably arises during resistance selections and was in fact the predominant resistance mechanism initially

observed.<sup>56</sup> Through serially plating patient-derived ALK+ ALCL cell lines in increasing concentrations of crizotinib or ceritinib, we generated resistant subclones. Each line acquired resistance by overexpressing NPM-ALK at the levels of both transcription and translation. Surprisingly, however, inhibitor withdrawal induced apoptosis, associated with massive amounts of activated ALK signaling, suggesting that these cells were overdosing on this heightened ALK activity (Fig. 2). This was confirmed by the fact that multiple different ALK TKIs could rescue the toxicity caused by signaling overdose, showing that this is a kinase-dependent consequence of NPM-ALK overexpression. Therefore, similar to the above-mentioned study of mutant BRAF melanoma, these ALK+ ALCL cells had grown to not only become resistant to, but also dependent on, the very inhibitors intended to kill them: a phenomenon we termed “resistance/dependence.” Importantly, while three of the five resistant lines established from this study harbored kinase domain mutations, the “resistance/dependence” phenotype, leading to ALK overdose upon drug washout, was seen regardless. This stresses that the upregulation of ALK activity was the major mechanism of resistance acquired by each of these cell lines. The mechanism by which the overdose of ALK activity promoted toxicity is unknown, as individual inhibition of downstream ALK targets (Fig. 1B), including MEK/ERK, failed to rescue from the overdose effects (unpublished observations). Therefore, unbiased approaches are currently underway to ascertain the basis of oncogene overdose in this disease.

In vivo, even stronger than the described observations in melanoma, we found that tumor engraftment of resistant cells was only seen in mice dosed with an ALK TKI (ceritinib).<sup>56</sup> Complete absence of engraftment in vehicle-treated mice underlines the drug dependent phenotype that these resistant cells acquired. Careful passaging of resistant cells in vitro at high confluence, however, allowed us to re-establish lines able to grow in the absence of inhibitor. ALK activity in these lines returned to baseline, and the cells were resensitized to the ALK TKIs. This is similar to observations from the above-mentioned study of melanoma, which showed resensitization through forced knockdown of BRAF<sup>V600E</sup>.<sup>35</sup>

These studies of mutant BRAF melanoma and ALK+ ALCL highlight the importance of having “just the right” amount of signaling with respect to tumor survival (Fig. 2).

### Goldilocks Principle

These findings fit well into the “Goldilocks principle,” the idea that certain biologic factors require precise levels to promote fitness, with either too much or too little being toxic. For example, the restoration of calcium release in heart cells,<sup>57</sup> the levels of oxygen administered by postcardiac arrest,<sup>58</sup> the amount of vitamin D in the body,<sup>59</sup> the redox environment of the cell with respect to oxidative stress,<sup>60</sup> and the levels of MeCP2 in causing Rett syndrome immune defects<sup>61</sup> must fall within the “Goldilocks zone.” Even a person’s body mass



index, which should be within the “normal” range, can be thought of in terms of the Goldilocks principle.

**MAPK cascade and the Goldilocks principle.** Although not usually described in these terms, RAF signaling also follows a Goldilocks paradigm, having dual roles in cell cycle progression or arrest depending on level.<sup>62,63</sup> Studies mostly in mouse fibroblasts, for instance, showed decreases in Raf activity can promote passage through the cell cycle through activation of Cyclin D1/Cdk4 and Cyclin E-Cdk2. Too much Raf, however, can cause cell cycle arrest through the induction of the Cdk inhibitors p21<sup>CIP1</sup> and p16<sup>INK4A</sup>.<sup>62,63</sup> Interestingly, Raf appears to suppress its own activity to maintain levels in the Goldilocks zone.<sup>64</sup> In addition, further highlighting the paradoxical nature of RAS → RAF → MEK → ERK signaling, the overactivation of any step in the pathway can cause oncogene-induced senescence via replicative stress leading to the activation of p16<sup>INK4A</sup> and p19<sup>ARF</sup> tumor suppressor pathways.<sup>28,65–67</sup> In an inducible mouse model with titratable levels of Ras, increased signaling led to senescence, while lower levels induced tumor formation.<sup>68</sup> Therefore, optimum tumor development also requires “just-right” levels of RAS, and indeed, this can be achieved by way of balancing autophagy.<sup>69,70</sup> Furthermore, autophagy itself appears to follow the Goldilocks principle in tumor development. On the one hand, it allows cells to scavenge their own reserves during nutritional and ischemic stress. On the other hand, too much autophagy opposes tumor establishment and development, through nascent tumor death or the prevention of further mutations that favor progression caused by reactive oxygen species that are generated by the buildup of damaged organelles.<sup>71</sup>

### Intermittent dosing

The requirement that at least some oncogenes maintain signaling at “just-right” levels may be exploitable using intermittent dosing to delay the onset of fatal resistance (Table 1). As discussed above, pulsed TKI dosing may allow BCR-ABL and its downstream signaling cascade to be effectively turned off on-drug, without permitting the reactivation of growth factor signaling when off-drug.<sup>6,29</sup> This can provide a window where there are only proapoptotic signals and no pro-survival signals present, enhancing cell death. Furthermore, pulsed BCR-ABL inhibition elicits the activation of the proapoptotic BIM protein with the same kinetics seen with continual exposure.<sup>72</sup> This approach, however, is highly specific depending on the oncogene in question. For example, while the inhibition of FLT3-ITD fusion in acute myeloid leukemia turns off oncogenic signaling, growth factor receptor signaling is restored at a faster rate upon inhibitor discontinuation, which means that there is not enough time for the induction of apoptosis before the reactivation of pro-survival signaling.<sup>6,29</sup> It will require meticulous tweaking of dosing schedules to exploit each oncogene to bring about death before the restoration of pro-survival signaling.

Systems in which target overexpression drives both resistance and dependence may be the most amenable to intermittent-dosing strategies. Using patient-derived BRAF<sup>V600E</sup> melanoma xenografts, Das Thakur et al<sup>35</sup> showed that intermittent vemurafenib dosing prolonged tumor control compared to continuous dosing. Both individualized drug interruptions based on tumor burden and up-front scheduled intermittent dosing were superior to continuous dosing in this report. Considering strategies to combat targeted-drug

**Table 1.** Studies that favor intermittent dosing to prolong tumor control.

ONCOGENE	INHIBITOR	DISEASE	STUDY	REFERENCE
BCR-ABL	Dasatinib Imatinib	Chronic Myeloid Leukemia	Pulsed dosing favors clinical remission in patients	72
BCR-ABL	Dasatinib Imatinib	Chronic Myeloid Leukemia	Pulsed inhibition prevents downstream growth factor rewiring	29
EGFR	Erlotinib	Chronic Myeloid Leukemia	Pulsed dosing prevents cytotoxicity	72
Mutant BRAF	Vemurafenib	Melanoma	Intermittent dosing prolongs tumor control	35
Mutant BRAF	Dabrafenib Trametinib	Melanoma	Ongoing joint NCI-SWOG randomized trial assessing the feasibility of intermittent dosing in melanoma using dual BRAF and MEK inhibitors	NCT02196181
Mutant BRAF	Vemurafenib Selumetinib	Melanoma	Cells with acquired resistance to dual BRAF and MEK inhibition are sensitive to drug withdrawal	39
Mutant BRAF	Vemurafenib Dabrafenib	Melanoma	Successfully rechallenged 2 mutant BRAF melanoma patients following a drug holiday	73
NPM-ALK	Crizotinib Ceritinib	ALK+ Anaplastic Large Cell Lymphoma	Intermittent dosing forestalls the fatal onset of resistance	56
EML4-ALK	Crizotinib	ALK+ Non-Small Cell Lung Cancer	Patients were successfully rechallenged with crizotinib after a drug holiday	74,75
EGFR	Erlotinib Gefitinib	Non-Small Cell Lung Cancer	Rechallenging with EGFR inhibitors decreased tumor volume	85



resistance, intermittent dosing emerges as one carrying the least amount of both expense and toxicity to patients. Therefore, the melanoma report both established the preclinical proof of principle and highlighted its potential flexibility. A small case series, meanwhile, reported successful BRAF-inhibitor rechallenge in two patients who had previously developed resistance and were then off-drug for several months, but the cause of resistance was not investigated.<sup>73</sup> A joint NCI-SWOG randomized trial specifically comparing intermittent vs. continuous dosing of dabrafenib and trametinib for treating *BRAF*<sup>V600E</sup>-mutant melanomas is currently enrolling patients (NCT02196181).

Although we found that ALK+ ALCL lines selected for TKI resistance reliably developed resistance-dependence, we have only begun exploring intermittent dosing as a therapeutic strategy. The improvement of ALK+ ALCL mouse models using genome editing with CRISPR/Cas9 should allow this possibility to be more rigorously investigated. However, two case reports have shown that patients may respond positively to rechallenge with crizotinib, lending support to the clinical investigation of discontinuous dosing with ALK TKIs.<sup>74,75</sup>

### Pharmacologically Inducing Oncogene Overdose

Another potential way to exploit therapeutically oncogene overdose may be through its forced pharmacological induction. In order to investigate this possibility, the mechanism(s) underlying oncogene overdose must be well understood. In ALK+ ALCL, our knowledge is incomplete, and, as mentioned, the inhibition of downstream targets of ALK failed to rescue cell viability (unpublished observations). We are undertaking several unbiased approaches to elucidate the specific mechanisms underlying overdose in the hope that forced induction may be translatable into a therapeutic strategy.

In another lymphocyte-derived malignancy, however, both the basis for and means to induce an overdose of oncogenic signaling were recently established. Briefly, the Goldilocks principle can be thought of as applying to the positive and negative selection of developing lymphocytes. The positive selection of immature B and T cells requires their newly created B-cell receptor (BCR) or T-cell receptor (TCR) to bind test antigens with a minimum affinity to weed out weak clones not likely to be useful in immune defense. However, negative selection eliminates clones with too strong of an affinity to self-antigens, eliminating clones likely to precipitate autoimmune phenomena. In both the cases, the degree of affinity is translated into the strength of downstream signaling, with too little or too much both being potentially fatal. Therefore, lymphocytes carry a propensity in their life history to die due to the overactivation of their core survival pathways.<sup>76-79</sup>

Approximately 25% of B-ALL cases contain the chromosomal fusion BCR-ABL discussed above, which in this context mimics the constitutive activation of pre-BCR signaling.<sup>80,81</sup> A comprehensive study by Chen et al<sup>82</sup> showed that BCR-ABL ALL cells express increased amounts of several immunoreceptor tyrosine-based inhibitory motif

(ITIM)-containing proteins, such as PECAM1, CD300A, and LAIR1. These ITIMs recruit several phosphatases, such as PTPN6 (also known as SHP1) and INPP5D, which down-regulate the activation of SYK, an early step in BCR signaling whose activation is normally markedly reduced in BCR-ABL ALL cells. Therefore, the authors used a small molecule inhibitor that targets the inhibitory phosphatase INPP5D to induce SYK activation. This led to the selective death of BCR-ABL+ B-ALL cells, both in vitro and in vivo. Furthermore, this was achieved regardless of the mutational status of BCR-ABL, as SYK induction also caused BCR-ABL<sup>T3151</sup> ALL cell death. This study highlights the potential of pharmacologically inducing overdose as a form of therapy.

This paradigm may also be applicable to T-ALL. A study of *TEL-JAK2* T-leukemogenesis in transgenic mice crossed these animals with the transgenic Marilyn strain, which expresses an MHC class II-restricted TCR $\alpha\beta$  recognizing specifically the H-Y male antigen.<sup>83</sup> Male primary mice, and even male mice engrafted with tumor cells from females, rarely developed tumors, in contrast with females, whose disease onset was the same as baseline *TEL-JAK2* animals. The cause was hypothesized to be self-antigen-induced negative selection but was not specifically established. Nor has a pharmacologic means of inducing such an overdose of signaling in T cells yet been reported.

### Conclusions

More work is warranted to assess the impact of oncogene overdose as a general mechanism leading to cell death in the context of acquired resistance, and intermittent dosing to exploit overdose may be fruitful in prolonging tumor control in selected patients. The in vitro and in vivo work discussed above in mutant BRAF melanoma and ALK+ ALCL shows great promise for potentially implementing this treatment paradigm in a patient setting, where intermittent dosing forestalled the onset of resistance.<sup>35,56</sup> The findings of these studies are further corroborated by clinical cases where discontinuous dosing has been highly efficacious (discussed above and summarized in Table 1). However, care must be taken when administering this regimen, as drug withdrawal prior to the onset of resistance, as well as the dose of drug initially administered, could accelerate acquired resistance.<sup>6,84</sup> Therefore, up-front intermittent schedules must be carefully studied and established. Otherwise, individualized approaches may be better, particularly for the short term. Precise characterization of the mechanisms of oncogene overdose will shed clearer light on both the therapeutic potential and appropriate strategies of intermittent dosing and is required before any efforts to pharmacologically induce overdose can be undertaken. It is also important to note that while intermittent dosing is less toxic and expensive than most other treatment strategies, it is not a cure. The in vivo experiments in mutant BRAF melanoma show that although the forestalling of resistance is achieved via intermittent dosing, eventually the cell finds a way and



resistance refractory to intermittent dosing sets in.<sup>35</sup> When feasible, regular tumor screening and genotyping to ascertain the mechanisms at play when tumor burden increases at each drug cycle will help clinicians stay one step ahead of tumor evolution and prevent the fatal onset of resistance.

### Author Contributions

Wrote the first draft of the manuscript: ADA and JHS. Contributed to the writing of the manuscript: ADA, SSR, MJG, PP and JHS. Jointly developed the structure and arguments for the paper: ADA and JHS. Made critical revisions and approved final version: ADA, SSR, MJG, PP and JHS. All authors reviewed and approved of the final manuscript.

### REFERENCES

- van der Kuip H, Wohlbold L, Oetzel C, Schwab M, Aulitzky WE. Mechanisms of clinical resistance to small molecule tyrosine kinase inhibitors targeting oncogenic tyrosine kinases. *Am J Pharmacogenomics*. 2005;5(2):101–112.
- Camidge DR, Pao W, Sequist LV. Acquired resistance to TKIs in solid tumours: learning from lung cancer. *Nat Rev Clin Oncol*. 2014;11(8):473–481.
- Friedman R. Drug resistance missense mutations in cancer are subject to evolutionary constraints. *PLoS One*. 2013;8(12):e82059.
- Gazdar AF. Activating and resistance mutations of EGFR in non-small-cell lung cancer: role in clinical response to EGFR tyrosine kinase inhibitors. *Oncogene*. 2009;28(suppl 1):S24–S31.
- Logue JS, Morrison DK. Complexity in the signaling network: insights from the use of targeted inhibitors in cancer therapy. *Genes Dev*. 2012;26(7):641–650.
- Pagliarini R, Shao W, Sellers WR. Oncogene addiction: pathways of therapeutic response, resistance, and road maps toward a cure. *EMBO Rep*. 2015;16(3):280–296.
- Weinstein IB. Cancer. Addiction to oncogenes—the Achilles heel of cancer. *Science*. 2002;297(5578):63–64.
- Druker BJ, Tamura S, Buchdunger E, et al. Effects of a selective inhibitor of the Abl tyrosine kinase on the growth of Bcr-Abl positive cells. *Nat Med*. 1996;2(5):561–566.
- Druker BJ, Guilhot F, O'Brien SG, et al; IRIS Investigators. Five-year follow-up of patients receiving imatinib for chronic myeloid leukemia. *N Engl J Med*. 2006;355(23):2408–2417.
- Sharma SV, Settleman J. Oncogene addiction: setting the stage for molecularly targeted cancer therapy. *Genes Dev*. 2007;21(24):3214–3231.
- Torti D, Trusolino L. Oncogene addiction as a foundational rationale for targeted anti-cancer therapy: promises and perils. *EMBO Mol Med*. 2011;3(11):623–636.
- Weinstein IB, Joe A. Oncogene addiction. *Cancer Res*. 2008;68(9):3077–3080.
- Arbiser JL. Why targeted therapy hasn't worked in advanced cancer. *J Clin Invest*. 2007;117(10):2762–2765.
- Pearson MA, Fabbro D. Targeting protein kinases in cancer therapy: a success? *Expert Rev Anticancer Ther*. 2004;4(6):1113–1124.
- Shaw AT, Engelman JA. ALK in lung cancer: past, present, and future. *J Clin Oncol*. 2013;31(8):1105–1111.
- Shaw AT, Kim DW, Mehra R, et al. Ceritinib in ALK-rearranged non-small-cell lung cancer. *N Engl J Med*. 2014;370(13):1189–1197.
- Rosell R, Carcereny E, Gervais R, et al; Spanish Lung Cancer Group in collaboration with Groupe Français de Pneumo-Cancérologie and Associazione Italiana Oncologia Toracica. Erlotinib versus standard chemotherapy as first-line treatment for European patients with advanced EGFR mutation-positive non-small-cell lung cancer (EURTAC): a multicentre, open-label, randomised phase 3 trial. *Lancet Oncol*. 2012;13(3):239–246.
- Shaw AT, Kim DW, Nakagawa K, et al. Crizotinib versus chemotherapy in advanced ALK-positive lung cancer. *N Engl J Med*. 2013;368(25):2385–2394.
- de Lavallade H, Apperley JF, Khorashad JS, et al. Imatinib for newly diagnosed patients with chronic myeloid leukemia: incidence of sustained responses in an intention-to-treat analysis. *J Clin Oncol*. 2008;26(20):3358–3363.
- Gugliotta G, Castagnetti F, Breccia M, et al; GIMEMA CML Working Party. Long-term outcome of a phase 2 trial with nilotinib 400 mg twice daily in first-line treatment of chronic myeloid leukemia. *Haematologica*. 2015;100(9):1146–1150.
- Hochhaus A, O'Brien SG, Guilhot F, et al; IRIS Investigators. Six-year follow-up of patients receiving imatinib for the first-line treatment of chronic myeloid leukemia. *Leukemia*. 2009;23(6):1054–1061.
- Kalmanti L, Saussele S, Laussek M, et al. Safety and efficacy of imatinib in CML over a period of 10 years: data from the randomized CML-study IV. *Leukemia*. 2015;29(5):1123–1132.
- Duncan JS, Whittle MC, Nakamura K, et al. Dynamic reprogramming of the kinome in response to targeted MEK inhibition in triple-negative breast cancer. *Cell*. 2012;149(2):307–321.
- Chandarlapaty S, Sawai A, Scaltriti M, et al. AKT inhibition relieves feedback suppression of receptor tyrosine kinase expression and activity. *Cancer Cell*. 2011;19(1):58–71.
- Rodrik-Outmezguine VS, Chandarlapaty S, Pagano NC, et al. mTOR kinase inhibition causes feedback-dependent biphasic regulation of AKT signaling. *Cancer Discov*. 2011;1(3):248–259.
- Sharma SV, Gajowniczek P, Way IP, et al. A common signaling cascade may underlie "addiction" to the Src, BCR-ABL, and EGF receptor oncogenes. *Cancer Cell*. 2006;10(5):425–435.
- Sharma SV, Settleman J. Exploiting the balance between life and death: targeted cancer therapy and "oncogenic shock". *Biochem Pharmacol*. 2010;80(5):666–673.
- Serrano M, Lin AW, McCurrach ME, Beach D, Lowe SW. Oncogenic ras provokes premature cell senescence associated with accumulation of p53 and p16INK4a. *Cell*. 1997;88(5):593–602.
- Asmussen J, Lasater EA, Tajon C, et al. MEK-dependent negative feedback underlies BCR-ABL-mediated oncogene addiction. *Cancer Discov*. 2014;4(2):200–215.
- Lito P, Pratilas CA, Joseph EW, et al. Relief of profound feedback inhibition of mitogenic signaling by RAF inhibitors attenuates their activity in BRAFV600E melanomas. *Cancer Cell*. 2012;22(5):668–682.
- Packer LM, Rana S, Hayward R, et al. Nilotinib and MEK inhibitors induce synthetic lethality through paradoxical activation of RAF in drug-resistant chronic myeloid leukemia. *Cancer Cell*. 2011;20(6):715–727.
- Lee HJ, Zhuang G, Cao Y, Du P, Kim HJ, Settleman J. Drug resistance via feedback activation of Stat3 in oncogene-addicted cancer cells. *Cancer Cell*. 2014;26(2):207–221.
- Herrero A, Pinto A, Colón-Bolea P, et al. Small molecule inhibition of ERK dimerization prevents tumorigenesis by RAS-ERK pathway oncogenes. *Cancer Cell*. 2015;28(2):170–182.
- Fecher LA, Amaravadi RK, Flaherty KT. The MAPK pathway in melanoma. *Curr Opin Oncol*. 2008;20(2):183–189.
- Das Thakur M, Salangsang F, Landman AS, et al. Modelling vemurafenib resistance in melanoma reveals a strategy to forestall drug resistance. *Nature*. 2013;494(7436):251–255.
- Petti C, Molla A, Vegetti C, Ferrone S, Anichini A, Sensi M. Coexpression of NRASQ61R and BRAFV600E in human melanoma cells activates senescence and increases susceptibility to cell-mediated cytotoxicity. *Cancer Res*. 2006;66(13):6503–6511.
- Flaherty KT, Infante JR, Daud A, et al. Combined BRAF and MEK inhibition in melanoma with BRAF V600 mutations. *N Engl J Med*. 2012;367(18):1694–1703.
- Flaherty KT, Robert C, Hersey P, et al; METRIC Study Group. Improved survival with MEK inhibition in BRAF-mutated melanoma. *N Engl J Med*. 2012;367(2):107–114.
- Moriceau G, Hugo W, Hong A, et al. Tunable-combinatorial mechanisms of acquired resistance limit the efficacy of BRAF/MEK cotargeting but result in melanoma drug addiction. *Cancer Cell*. 2015;27(2):240–256.
- Morris SW, Kirstein MN, Valentine MB, et al. Fusion of a kinase gene, ALK, to a nucleolar protein gene, NPM, in non-Hodgkin's lymphoma. *Science*. 1994;263(5151):1281–1284.
- Rikova K, Guo A, Zeng Q, et al. Global survey of phosphotyrosine signaling identifies oncogenic kinases in lung cancer. *Cell*. 2007;131(6):1190–1203.
- Soda M, Choi YL, Enomoto M, et al. Identification of the transforming EML4-ALK fusion gene in non-small-cell lung cancer. *Nature*. 2007;448(7153):561–566.
- Gambacorti-Passerini C, Messa C, Pogliani EM. Crizotinib in anaplastic large-cell lymphoma. *N Engl J Med*. 2011;364(8):775–776.
- Gambacorti Passerini C, Farina F, Stasia A, et al. Crizotinib in advanced, chemoresistant anaplastic lymphoma kinase-positive lymphoma patients. *J Natl Cancer Inst*. 2014;106(2):djt378.
- Ceccon M, Mogni L, Bisson W, Scapozza L, Gambacorti-Passerini C. Crizotinib-resistant NPM-ALK mutants confer differential sensitivity to unrelated Alk inhibitors. *Mol Cancer Res*. 2013;11(2):122–132.
- Ceccon M, Mogni L, Giudici G, et al. Treatment efficacy and resistance mechanisms using the second-generation Alk inhibitor Ap26113 in human Npm-Alk-positive anaplastic large cell lymphoma. *Mol Cancer Res*. 2014;13(4):775–783.
- Doebele RC, Pilling AB, Aisner DL, et al. Mechanisms of resistance to crizotinib in patients with ALK gene rearranged non-small cell lung cancer. *Clin Cancer Res*. 2012;18(5):1472–1482.
- Fontana D, Ceccon M, Gambacorti-Passerini C, Mogni L. Activity of second-generation ALK inhibitors against crizotinib-resistant mutants in an NPM-ALK model compared to EML4-ALK. *Cancer Med*. 2015;4(7):953–965.
- Friboulet L, Li N, Katayama R, et al. The ALK inhibitor ceritinib overcomes crizotinib resistance in non-small cell lung cancer. *Cancer Discov*. 2014;4(6):662–673.



50. Katayama R, Khan TM, Benes C, et al. Therapeutic strategies to overcome crizotinib resistance in non-small cell lung cancers harboring the fusion oncogene EML4-ALK. *Proc Natl Acad Sci U S A*. 2011;108(18):7535–7540.
51. Katayama R, Shaw AT, Khan TM, et al. Mechanisms of acquired crizotinib resistance in ALK-rearranged lung cancers. *Sci Transl Med*. 2012;4(120):120ra117.
52. Katayama R, Friboulet L, Koike S, et al. Two novel ALK mutations mediate acquired resistance to the next-generation ALK inhibitor alectinib. *Clin Cancer Res*. 2014;20(22):5686–5696.
53. Kodama T, Tsukaguchi T, Yoshida M, Kondoh O, Sakamoto H. Selective ALK inhibitor alectinib with potent antitumor activity in models of crizotinib resistance. *Cancer Lett*. 2014;351(2):215–221.
54. Mologni L, Cecon M, Pirola A, et al. NPM/ALK mutants resistant to ASP3026 display variable sensitivity to alternative ALK inhibitors but succumb to the novel compound PF-06463922. *Oncotarget*. 2015;6(8):5720–5734.
55. Sakamoto H, Tsukaguchi T, Hiroshima S, et al. CH5424802, a selective ALK inhibitor capable of blocking the resistant gatekeeper mutant. *Cancer Cell*. 2011;19(5):679–690.
56. Amin AD, Rajan SS, Liang WS, et al. Evidence suggesting that discontinuous dosing of ALK kinase inhibitors may prolong control of ALK+ tumors. *Cancer Res*. 2015;75(14):2916–2927.
57. Liu OZ, Lederer WJ, Sobie EA. Does the goldilocks principle apply to calcium release restitution in heart cells? *J Mol Cell Cardiol*. 2012;52(1):3–6.
58. Hommers C. Oxygen therapy post-cardiac arrest: the 'Goldilocks' principle? *Resuscitation*. 2010;81(12):1605–1606.
59. Drake MT. Vitamin D and the goldilocks principle: too little, too much, or just right? *J Clin Endocrinol Metab*. 2014;99(4):1164–1166.
60. Alleman RJ, Katunga LA, Nelson MA, Brown DA, Anderson EJ. The "goldilocks zone" from a redox perspective—adaptive vs. deleterious responses to oxidative stress in striated muscle. *Front Physiol*. 2014;5:358.
61. Boothby M, Williams CL. The goldilocks effect. *Sci Transl Med*. 2012;4(163):163fs142.
62. Woods D, Parry D, Cherwinski H, Bosch E, Lees E, McMahon M. Raf-induced proliferation or cell cycle arrest is determined by the level of Raf activity with arrest mediated by p21Cip1. *Mol Cell Biol*. 1997;17(9):5598–5611.
63. Zhu J, Woods D, McMahon M, Bishop JM. Senescence of human fibroblasts induced by oncogenic Raf. *Genes Dev*. 1998;12(19):2997–3007.
64. Hall-Jackson CA, Eyers PA, Cohen P, et al. Paradoxical activation of Raf by a novel Raf inhibitor. *Chem Biol*. 1999;6(8):559–568.
65. Kilbey A, Terry A, Cameron ER, Neil JC. Oncogene-induced senescence: an essential role for Runx. *Cell Cycle*. 2008;7(15):2333–2340.
66. Palmero I, Pantoja C, Serrano M. p19ARF links the tumour suppressor p53 to Ras. *Nature*. 1998;395(6698):125–126.
67. Yaswen P, Campisi J. Oncogene-induced senescence pathways weave an intricate tapestry. *Cell*. 2007;128(2):233–234.
68. Sarkisian CJ, Keister BA, Stairs DB, Boxer RB, Moody SE, Chodosh LA. Dose-dependent oncogene-induced senescence in vivo and its evasion during mammary tumorigenesis. *Nat Cell Biol*. 2007;9(5):493–505.
69. Guo JY, Chen HY, Mathew R, et al. Activated Ras requires autophagy to maintain oxidative metabolism and tumorigenesis. *Genes Dev*. 2011;25(5):460–470.
70. Schmukler E, Kloog Y, Pinkas-Kramarski R. Ras and autophagy in cancer development and therapy. *Oncotarget*. 2014;5(3):577–586.
71. Martin SJ. Oncogene-induced autophagy and the goldilocks principle. *Autophagy*. 2011;7(8):922–923.
72. Shah NP, Kasap C, Weier C, et al. Transient potent BCR-ABL inhibition is sufficient to commit chronic myeloid leukemia cells irreversibly to apoptosis. *Cancer Cell*. 2008;14(6):485–493.
73. Seghers AC, Wilgenhof S, Lebbe C, Neyns B. Successful rechallenge in two patients with BRAF-V600-mutant melanoma who experienced previous progression during treatment with a selective BRAF inhibitor. *Melanoma Res*. 2012;22(6):466–472.
74. Browning ET, Weickhardt AJ, Camidge DR. Response to crizotinib rechallenge after initial progression and intervening chemotherapy in ALK lung cancer. *J Thorac Oncol*. 2013;8(3):e21.
75. Schrödl K, von Schilling C, Tufman A, Huber RM, Gamarra F. Response to chemotherapy, reexposure to crizotinib and treatment with a novel ALK inhibitor in a patient with acquired crizotinib resistance. *Respiration*. 2014;88(3):262–264.
76. Burrows PD, Suematsu S, Watanabe T. Activation of self-reactive B cells and autoimmune diseases. *Rev Immunogenet*. 2000;2(1):38–51.
77. Keenan RA, De Riva A, Corleis B, et al. Censoring of autoreactive B cell development by the pre-B cell receptor. *Science*. 2008;321(5889):696–699.
78. Wang B, Primeau TM, Myers N, et al. A single peptide-MHC complex positively selects a diverse and specific CD8 T cell repertoire. *Science*. 2009;326(5954):871–874.
79. Yun TJ, Bevan MJ. The goldilocks conditions applied to T cell development. *Nat Immunol*. 2001;2(1):13–14.
80. Feldhahn N, Klein F, Mooster JL, et al. Mimicry of a constitutively active pre-B cell receptor in acute lymphoblastic leukemia cells. *J Exp Med*. 2005;201(11):1837–1852.
81. Klein F, Feldhahn N, Harder L, et al. The BCR-ABL1 kinase bypasses selection for the expression of a pre-B cell receptor in pre-B acute lymphoblastic leukemia cells. *J Exp Med*. 2004;199(5):673–685.
82. Chen Z, Shojaee S, Buchner M, et al. Signalling thresholds and negative B-cell selection in acute lymphoblastic leukaemia. *Nature*. 2015;521(7552):357–361.
83. dos Santos NR, Rickman DS, de Reynies A, et al. Pre-TCR expression cooperates with TEL-JAK2 to transform immature thymocytes and induce T-cell leukemia. *Blood*. 2007;109(9):3972–3981.
84. Foo J, Chmielecki J, Pao W, Michor F. Effects of pharmacokinetic processes and varied dosing schedules on the dynamics of acquired resistance to erlotinib in EGFR-mutant lung cancer. *J Thorac Oncol*. 2012;7(10):1583–1593.
85. Riely GJ, Kris MG, Zhao B, et al. Prospective assessment of discontinuation and reinitiation of erlotinib or gefitinib in patients with acquired resistance to erlotinib or gefitinib followed by the addition of everolimus. *Clin Cancer Res*. 2007;13(17):5150–5155.