1 **TITLE: A protective and broadly binding antibody class engages the influenza virus**
2 **hemagglutinin head at its stem interface**
3

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21 **Abstract:**
22 Influenza infection and vaccination impart strain-specific immunity that protects against neither
23 seasonal antigenic variants nor the next pandemic. However, antibodies directed to conserved sites
24 can confer broad protection. Here we identify and characterize a class of human antibodies that engage
25 a previously undescribed, conserved epitope on the influenza hemagglutinin (HA) protein. Prototype
26 antibody S8V1-157 binds at the normally occluded interface between the HA head and stem.
27 Antibodies to this HA head-stem interface epitope are non-neutralizing *in vitro* but protect against lethal
28 influenza infection in mice. Antibody isotypes that direct clearance of infected cells enhance this
29 protection. Head-stem interface antibodies bind to most influenza A serotypes and seasonal human
30 variants, and are present at low frequencies in the memory B cell populations of multiple human
31 donors. Vaccines designed to elicit these antibodies might contribute to "universal" influenza immunity.
32

33 **INTRODUCTION:**
34 Influenza pandemics arise from antigenically novel zoonotic influenza A viruses transmitted to humans
35 from animals. Historically, pandemic viruses generally have become endemic and have continued to
36 circulate as seasonal viruses. Sustained viral circulation is enabled by on-going antigenic evolution that
37 leads to escape from population-level immunity elicited by previous exposures [1]. Although antibodies
38 provide the strongest protection against infection, they also drive the antigenic evolution of the viral
39 surface proteins [1]. Therefore, neither infection nor the seasonal flu vaccine confers enduring immunity
40 against future seasonal variants or new pandemic viruses. Nevertheless, broadly protective monoclonal
41 antibodies that engage conserved influenza sites have been isolated from human donors [2]. Passive
42 transfer of these antibodies to animal models imparts broadly protective immunity. A next generation
43 influenza vaccine that elicits similar antibodies would likely confer more durable protection than that
44 offered by current seasonal vaccines [3-5].
45

46 The influenza hemagglutinin (HA) protein is the major target of protective antibodies [6]. HA facilitates
47 cell entry by attaching to cells via an interaction with its receptor, sialic acid, and by acting as a virus-
48 cell membrane fusogen. HA is synthesized as a polyprotein, HA0, which forms homotrimers that are
49 incapable of undergoing the full series of conformational rearrangements required for membrane fusion.
50 Cellular proteases (often resident on the target cell) cleave the HA0 into HA1 and HA2 domains,
51 resulting in a fusion-competent trimer [7-9]. HA1 includes the globular HA head that contains the receptor
52 binding site (RBS), while HA2 contains the helical stem regions that rearrange during endosomal
53 acidification to drive fusion of viral and cellular membranes. The requirement for HA to transit through
54 multiple conformations during fusion likely accounts for the intrinsic propensity of HA0 or HA1-HA2 to

55 transiently explore states that deviate from its prefusion conformation, which reproducibly forms
56 ordered protein crystal lattices that yield high-resolution structures determined by X-ray crystallography
57 [10-13].

59 Large genetic and antigenic differences separate influenza A HA serotypes, which are classified into
60 group 1 (H1, H2, H5, H6, H8, H9, H11, H12, H13, H16) and group 2 (H3, H4, H7, H10, H14, H15).
61 Serotypes of a second glycoprotein, neuraminidase (NA), are denoted by a similar convention;
62 consequently, influenza viruses are named by their HA and NA content (e.g., H2N2, H10N8). Currently,
63 divergent H1N1 and H3N2 viruses circulate as seasonal human influenza viruses. Humans can
64 produce antibodies that engage surfaces conserved between serotypes and thereby provide cross-
65 serotype protection. These epitopic regions include the HA RBS, stem, anchor, head interface, the so-
66 called "long alpha-helix", and the HA2 β-hairpin [2,14-19].

68 Here, we report a class of human antibodies directed to a previously unreported, widely conserved site
69 at the base, or neck, of the HA head. In the "resting state" prefusion structure of the HA trimer, the
70 epitope for prototype antibody S8V1-157 is occluded at the interface of the head and stem.
71 Biochemical, cellular, and *in vivo* passive transfer experiments indicate that this HA head-stem interface
72 epitope is sufficiently exposed to allow antibodies to bind and confer strong protection against lethal
73 influenza virus infection in murine challenge models. Many humans harbor antibodies recognizing this
74 epitope; some of these antibodies bind HAs from divergent influenza serotypes, groups, and >50 years
75 of human seasonal virus antigenic variation. Immunogens that elicit such antibodies might be included
76 in influenza vaccines intended to confer broader protection.

78 **RESULTS:**
79 **A class of antibodies engages the interface of the HA head and stem regions**
80 By culturing individual human memory B (Bmem) cells and then screening the culture supernatants
81 containing secreted IgGs, we identified (as reported previously [20]) 449 HA-reactive antibodies from four
82 donors (S1, S5, S8 and S9). These antibodies represent the Bmem cells circulating in the blood at the
83 time the donors were immunized with the TIV 2015-2016 seasonal influenza vaccine (visit 1; V1), or 7
84 days later (visit 2; V2). From donors S1 and S8, we found four IgGs (S1V2-17, S1V2-60, S1V2-65 and
85 S8V1-157) that had a novel pattern of HA reactivity (Figure 1A). All bound seasonal H1s and H3s and
86 an H5 HA; they also bound recombinant HA constructs comprising only the HA head, indicating the
87 IgGs did not target stem epitopes (Figure 1A). These four antibodies competed with each other for HA-
88 binding, implying overlapping epitopes; however, the four antibodies did not compete with other well-
89 characterized antibodies that engage conserved, structure-verified, broadly protective epitopes
90 spanning the HA molecule (Figure 1B).

92 All four new antibodies have light chains encoded by IGKV2-28*01 or IGKV2D-28*01 (Figure 1C), with
93 identical germline coding sequences. The heavy chains are encoded by either IGHV1-24*01 or IGHV5-
94 51*01 and have short (11-13 amino acid) third complementarity-determining regions (HCDR3s). S1V2-
95 60 and S1V2-65 are clonally related, while S1V2-17 and S8V1-157 share a common IGHV1-24*01-
96 IGKV2-28*01 pairing, despite their derivation from different donors.

98 We determined the structure of S8V1-157 complexed with a monomeric HA head construct. Crystals
99 were only obtained using an A/American black duck/New Brunswick/00464/2010(H4N6) (H4-NB-2010)
100 HA (Figure 2A and Table S1). S8V1-157 engages an epitope at the base of the head, just where it
101 faces the stem [21]. In the context of trimeric, full-length, prefusion HA, this epitope is occluded by the
102 association of HA1 with HA2 (Figure 2A). Therefore, displacement of the HA1 head from the HA2
103 helical stem would be required for B cells or antibodies to recognize this site.

105 S8V1-157 contacts residues mediating the intra-protomer interaction, which are conserved across
106 divergent HAs (Figures 2B and S1). Evolutionary constraints imposed by the requirement that HA1 and
107 HA2 stably associate at neutral pH likely account for the conservation of the head-stem interface
108 epitope, and by extension, the binding breadth of the corresponding antibodies.

109
110 The compact HCDR3 of S8V1-157 packs against the heavy chain CDR and framework regions (FR),
111 HCDR1-FR1 and FR2-HCDR2, to produce a cleft between the heavy and light chains that
112 accommodates the head-stem interface epitope (Figure 2C). Light chain CDR1 residues N33, Y35 and
113 Y37 make extensive contacts with a conserved HA surface. The δ-amide of N33 donates and receives
114 hydrogen bonds from the main chain of HA residue HA-C305. Its main chain amide also donates a
115 hydrogen bond to the carboxyl group of HA-D304. Y35 donates a hydrogen bond to the main chain
116 carbonyl of HA-T291 and participates in van der Waals interactions by stacking upon HA-P293. Y37
117 receives a hydrogen bond from the main chain residue HA-K307, donates a hydrogen bond to HA-
118 P293, and participates in van der Waals contacts with the aryl group of HA-F294 and the aliphatic side
119 chain of HA-K307.
120
121 The structure of the S8V1-157-HA complex explains the genetic features common to this class of head-
122 stem interface antibodies. The LCDR1 NXYXY motif is germline-encoded by a small subset of IGKV-
123 genes. Of these, only IGKV2-28/IGKV2D-28a also encode a hydrophobic residue at position 55. L55
124 participates in van der Waals contacts with HA-P293 (Figure 2C) and contributes to a hydrophobic
125 surface on S8V1-157 that complements a hydrophobic patch within its epitope on the HA head (Figure
126 S2). Polar residues at position 55 in otherwise similar light chains are predicted to be less favorable in
127 this local environment. Similar sequence features are not present in the IGLV locus. Additionally, the
128 short HCDR3 (common among these antibodies) creates shallow cavity that accommodates the
129 hydrophobic surfaces that would normally pack against HA2 in the prefusion HA trimer.
130
131 **Antibodies to the HA head-stem interface epitope are broadly binding.**
132 We determined the binding of head-stem interface antibodies to divergent HA serotypes using
133 recombinantly expressed soluble HA ectodomains and recombinantly expressed antibodies in enzyme-
134 linked immunosorbent assays (ELISA). All HAs except H5 and H7 were expressed as HA0, which is
135 incapable of transitioning to the postfusion conformation without first being processed by a trypsin-like
136 protease[8,22]. H5 and H7 HAs contain polybasic cleavage sites that are processed by endogenous furin-
137 like proteases in the expressing cell line, resulting in HA1-HA2[23-28].
138
139 The HA head-stem interface antibodies had very similar breadth: each bound all seasonal H1s (1977-
140 2019) and H3s (1968-2020) assayed, and also bound several other HA subtypes within groups 1 and 2
141 (Figures 3A and S3). Typically, if an HA was bound by one head-stem epitope antibody, it was also
142 bound by the other three (Figure 3). Affinities and breadth of binding were generally higher for group 2
143 HAs but extended to group 1 HAs, including seasonal H1, pandemic H2, and pre-pandemic H5 HAs.
144 Overall, these antibodies engaged HAs from 11 of the 16 non-bat influenza A HA serotypes. Failure to
145 engage specific HAs was not due to epitope inaccessibility, since the antibodies also did not bind
146 matched, soluble, HA head that present the HA head-stem epitope without steric hindrance (Figures S4
147 and S5). None of the head-stem interface antibodies bound a truncated HA head lacking the S8V1-157
148 epitope. All four antibodies likely engage a common, discrete epitope comprising the terminus of the HA
149 head.
150
151 In a complementary flow cytometry assay (in the absence of exogenous trypsin[19,29]), HA head-stem
152 epitope antibodies bound divergent HAs stably expressed in their native, transmembrane form on the
153 cell surface (Figure 4). The binding pattern was consistent with our ELISA data. We used a curated
154 panel of antibodies (largely overlapping with those in Figure 1), including K03.12[30] and H5.3[31] which
155 recognize the fully exposed receptor binding site. HA head-stem interface antibodies generally labeled
156 cells less brightly than did control antibodies directed to solvent-exposed epitopes on the HA head
157 (e.g., K03.12, H5.3, HC19); however, head-stem interface antibodies labeled cells comparably to the
158 HA head-interface antibody S5V2-29 [20]. Together, these observations suggest that HA interface
159 epitopes are not occupied as fully by antibody as are fully exposed epitopes. Epitope accessibility
160 and/or differences in the number of IgG molecules bound per HA trimer may account for these
161 differences.
162

163   To determine whether head-stem interface antibodies can engage native HA0 on the cell surface, 293F
164   cells were transiently transfected with an expression vector encoding full-length, membrane-anchored
165   A/Aichi/02/1968(H3N2)(X-31) HA. Western blotting analysis using an antibody against a linear epitope
166   contained within HA1 confirmed that the transfected cells expressed only unprocessed, 83 kD HA0
167   (Fig. S5B). Processed HA1, which under reducing and denaturing conditions would be observed as a
168   55 kD band, was not detected. Flow cytometry analysis showed that S8V1-157 labeled the transfected
169   cells (Fig. S5C), as did antibodies recognizing the RBS (HC19) and head interface epitope (S5V2-29).
170   The head-stem interface epitope is therefore exposed, if only transiently, on native HA0 expressed on
171   the cell surface.

172

173   **Antibodies to the HA head-stem epitope protect against lethal influenza virus infection.**
174   S8V1-157 failed to neutralize influenza virus *in vitro*, in an assay in which virus and antibody were co-
175   incubated, added to cells, and neutralization scored four days post-infection (Figure S6). To determine
176   if head-stem interface antibodies confer protection *in vivo*, we produced the antibodies as recombinant
177   mouse IgG1 or IgG2c, then passively transferred the antibodies to mice and challenged them with a
178   lethal dose of influenza virus. In mice, the IgG2c isotype potently directs protective, Fc-dependent,
179   effector functions, including antibody-dependent cellular cytotoxicity (ADCC) and complement
180   deposition (ADCD). In contrast, the IgG1 isotype does not appreciably stimulate these effector
181   functions. Each mouse received 150 μg (~7.5 mg/kg) of the head-stem interface antibodies S8V1-157
182   or S5V2-65, HC19[32] (which potently neutralizes A/Aichi/02/1968(H3N2)(X-31)), S5V2-29[20] (an HA-head
183   interface antibody that protects against lethal infection by Fc-dependent mechanisms), or CR3022[33]
184   (SARS-CoV antibody). The ~7.5 mg/kg antibody dose is comparable or below commonly administered
185   doses of antiviral monoclonal antibody therapeutics[33-38].

186

187   HC19 protected mice from infection-induced weight loss, while the irrelevant antibody CR3022 offered
188   no protection, requiring recipient mice to be ethically euthanized by day 10 post-infection (Figure 5A).
189   All animals administered HA head-stem interface antibodies S8V1-157 or S1V2-65 recovered after
190   experiencing mild to moderate weight loss; by day 10 post-infection, their body weights were similar to
191   the HC19-treated group. IgG2c versions of S8V1-157 and S1V2-65 conferred slightly better protection
192   than the corresponding IgG1s, indicating that Fc effector functions had a role in controlling the infection
193   (Figure 5B-D). Similarly, head interface antibody S5V2-29 protected better against infection-induced
194   weight loss when infused as an IgG2c. Thus, despite failing to neutralize the initial infection, HA head-
195   stem interface antibodies protected influenza-infected mice against severe disease and mortality.

196

197   **The HA head-stem epitope is immunogenic in humans.**
198   We screened additional human Bmem cell cultures for S8V1-157-competing antibodies (Figure 6). The
199   samples were taken from seven donors, including the same subjects as before (S1, S5, S8, S9), but
200   vaccinated and sampled during subsequent flu seasons (see Materials and Methods for details);
201   subject S12, who was immunized and sampled at the same times; and subjects KEL01 and KEL03,
202   who were sampled after receiving TIV in 2014-2015. From 528 clonal cultures that produced HA-
203   binding IgG, we identified eight additional supernatants that inhibited S8V1-157 binding to HA by >90%.
204   (Figure 6A). In a screen of antibody reactivity, five of the eight competing antibodies reacted with both
205   group 1 and group 2 HAs.

206

207   Paired heavy and light chain sequences were recovered from seven of the eight S8V1-157-competing
208   antibodies (Figure S7). Two antibodies, S1V4-P4-C5 and S12V6-P7-H4, use an IGKV2-28 light chain
209   paired with IGHV1-24*01, like S8V1-157 and S1V2-17. All four antibodies have short, 11-12 amino acid
210   HCDR3s. These features, shared with S8V1-157, likely define a public antibody class present in at
211   least three human donors. The other newly identified S8V1-157 competitors use other *IGHV* and *IGKV*
212   genes and have varying HCDR3 lengths (10-19 amino acids). These antibodies could either have
213   footprints that overlap the head-stem interface epitope or that prevent its exposure.

214

215   **DISCUSSION:**

216   We identified and characterized a human antibody response directed to a previously unrecognized,
217   widely conserved, cryptic epitope on the influenza HA head. These antibodies bind broadly to divergent
218   HA serotypes found in animals and seasonal antigenic variants of human viruses. Prophylactic passive
219   transfer of these antibodies to mice protects against lethal influenza virus disease. Approximately 1% of
220   circulating, HA-reactive Bmem recognize the head-stem interface and are present in multiple donors
221   both before and after seasonal influenza vaccination, though most such Bmem were isolated following
222   vaccination. Head-stem interface antibodies have a bias, but not a restriction, for IGKV2-28 usage.
223   Across the 12 known examples we find no additional constraints. Other humans are therefore likely to
224   produce similar antibodies and to have the capacity to mount polyclonal, broadly protective antibody
225   responses.
226
227   That HA head-stem interface antibodies confer robust protection against lethal influenza challenge
228   demonstrates that the epitope is exposed on infected cells and/or virions, if only transiently. Exposure
229   of the HA head-stem interface would be predicted to require a large-scale displacement of the HA head
230   from HA2, and additional conformational rearrangements within the HA trimer might accompany such
231   events. Nevertheless, our *in vitro* experiments, which used recombinant HA0 (either as soluble
232   ectodomain or native, membrane-anchored HA on the surface of transiently-transfected cells),
233   confirmed that head-stem interface antibodies must be able to recognize their epitope on some
234   conformation of prefusion HA, because HA0 cannot transition to a stable postfusion structure[8].
235   Structural, biophysical, and computational approaches indicate that HA trimers transiently adopt
236   conformations that expose epitopes normally occluded in the defined prefusion state[10-13]. Such
237   transient fluctuations might explain how B cells and antibodies recognize ordinarily occluded
238   epitopes[16,17,20,39-41]. In addition to binding HA0, head-stem interface antibodies might also directly
239   recognize postfusion HA on the surface of infected cells, as has been proposed for some HA
240   antibodies, including LAH31[17], S1V2-72[19] and m826[41]. It is plausible that, *in vivo*, HA0 is cleaved into
241   HA1/HA2 by proteases and that some fraction of postfusion HA containing the exposed head-stem
242   interface epitope accumulates on infected cell-surfaces and/or among the HAs presented to B cells in
243   germinal centers.
244
245   Humans and mice mount antibody responses to several apparently occluded HA epitopes [14-17,19,20,39-43].
246   Prevalence of such antibodies in immune repertoires indicates that these epitopes are immunogenic.
247   That several of the human head-stem interface antibodies reported here had 5-11% IGHV gene
248   mutation frequencies implies that the epitope was encountered repeatedly over recurrent influenza
249   exposures (whether by infection or vaccination). The immunogenicity of the HA head-stem epitope
250   poses a critical question for vaccinology: how does the antigen presented to a B cell in a germinal
251   center reaction relate to its form on infectious virions or to what has been defined in the laboratory in
252   considerable biochemical and structural detail? The existence of antibodies to interface/buried epitopes
253   on other, unrelated viral glycoproteins demonstrates that our lack of understanding extends beyond HA
254   [44].
255
256   Several of the most conserved epitopes on HA are occluded. Adjacent protomers and/or membranes
257   hinder access to some stem/anchor epitopes, and interface epitopes are normally hidden [2,14,15,19,45].
258   Nevertheless, antibodies recognizing these epitopes protect against lethal influenza virus disease in
259   small animal models. These antibodies are typically less potently neutralizing than antibodies targeting
260   fully exposed sites, and their protection is often enhanced by or fully dependent on Fc-mediated
261   effector functions [16,17,19,20,40,43,46,47]. Their resistance to human influenza antigenic evolution and ability to
262   engage emerging, pre-pandemic, and other animal viruses make their selective elicitation a potential
263   strategy to improve current influenza vaccines. Given the characteristics of head-stem interface
264   antibodies, a rigorous understanding of their potential to enhance human protective immunity and/or
265   ameliorate disease is needed.
266
267

268 **METHODS:**
269 *Human subjects.*
270 Peripheral blood mononuclear cells (PBMCs) were obtained from human donors KEL01 (male, age 39)
271 and KEL03 (female, age 39), under Duke Institutional Review Board committee guidelines. Written
272 informed consent was obtained from both subjects. KEL01 and KEL03 received the trivalent inactivated
273 seasonal influenza vaccine (TIV) 2014-2015 Fluvirin, which contained A/Christchurch/16/2010, NIB-74
274 (H1N1), A/Texas/50/2012, NYMC X-223 (H3N2), and B/Massachusetts/2/2012, NYMC BX-51B. Blood
275 was drawn on day 14 post-vaccination, and PBMCs isolated by centrifugation over Ficoll density
276 gradients (SepMate-50 tubes, StemCell Tech) were frozen and kept in liquid nitrogen until use.
277
278 PBMCs were also obtained from human donors S1 (female, age 51-55), S5 (male, age 21-25), S8
279 (female, age 26-30), S9 (female, age 51-55), and S12 (male, age 35-40), under Boston University
280 Institutional Review Board committee guidelines. Written informed consent was obtained from all five
281 subjects. Donors met all of the following inclusion criteria: between 18 and 65 years of age; in good
282 health, as determined by vital signs [heart rate (<100 bpm), blood pressure (systolic ≤ 140 mm Hg and
283 ≥ 90 mm Hg, diastolic ≤ 90 mm Hg), oral temperature (<100.0 °F)] and medical history to ensure
284 existing medical diagnoses/conditions are not clinically significant; can understand and comply with
285 study procedures, and; provided written informed consent prior to initiation of the study. Exclusion
286 criteria included: 1) life-threating allergies, including an allergy to eggs; 2) have ever had a severe
287 reaction after influenza vaccination; 3) a history of Guillain-Barre Syndrome; 4) a history of receiving
288 immunoglobulin or other blood product within the 3 months prior to vaccination in this study; 5) received
289 an experimental agent (vaccine, drug, biologic, device, blood product, or medication) within 1 month
290 prior to vaccination in this study or expect to receive an experimental agent during this study; 6) have
291 received any live licensed vaccines within 4 weeks or inactivated licensed vaccines within 2 weeks prior
292 to the vaccination in this study or plan receipt of such vaccines within 2 weeks following the vaccination;
293 7) have an acute or chronic medical condition that might render vaccination unsafe, or interfere with the
294 evaluation of humoral responses (includes, but is not limited to, known cardiac disease, chronic liver
295 disease, significant renal disease, unstable or progressive neurological disorders, diabetes mellitus,
296 autoimmune disorders and transplant recipients); 8) have an acute illness, including an oral
297 temperature greater than 99.9°F, within 1 week of vaccination; 9) active HIV, hepatitis B, or hepatitis C
298 infection; 10) a history of alcohol or drug abuse in the last 5 years; 11) a history of a coagulation
299 disorder or receiving medications that affect coagulation. Subjects S1, S5, S8, S9, and S12 received
300 seasonal influenza vaccination during three consecutive North American flu seasons (2015-2016, 2016-
301 2017, 2017-2018), and had blood drawn on day 0 (pre-vaccination; visits 1, 3, and 5) and day 7 (post-
302 vaccination, visits 2, 4, and 6) each year. During the 2015-2016 season (visits 1 and 2), the subjects
303 received the TIV Fluvirin, which contained A/reassortant/NYMC X-181 (California/07/2009 x NYMC X-
304 157) (H1N1), A/South Australia/55/2014 IVR-175 (H3N2), and B/Phuket/3073/2013. During the 2016-
305 2017 season (visits 3 and 4), the subjects received the quadrivalent inactivated vaccine Flucelvax,
306 containing A/Brisbane/10/2010 (H1N1), A/Hong Kong /4801/2014 (H3N2), B/Utah/9/2014, and B/Hong
307 Kong/259/2010. During the 2017-2018 season (visits 5 and 6), the subjects received the quadrivalent
308 inactivated vaccine Flucelvax, containing A/Singapore/GP1908/2015 IVR-180 (H1N1),
309 A/Singapore/GP2050/2015 (H3N2), B/Utah/9/2014, and B/Hong Kong/259/2010.
310
311 *Cell lines.*
312 Human 293F cells were maintained at 37°C with 5-8% CO_2 in FreeStyle 293 Expression Medium
313 (ThermoFisher) supplemented with penicillin and streptomycin. HA-expressing K530 cell lines [29] (*Homo*
314 *sapiens*) were cultured at 37°C with 5% CO_2 in RPMI-1640 medium plus 10% FBS (Cytiva), 2-
315 mercaptoethanol (55 μM; Gibco), penicillin, streptomycin, HEPES (10 mM; Gibco), sodium pyruvate (1
316 mM; Gibco), and MEM nonessential amino acids (Gibco). Madin-Darby canine kidney (MDCK) were
317 maintained in Minimum Essential medium supplemented with 10% fetal bovine serum, 5 mM L-
318 glutamine and 5 mM penicillin/streptomycin.
319 *Recombinant Fab expression and purification*.
320 The heavy and light chain variable domain genes for Fabs were cloned into a modified pVRC8400
321 expression vector, as previously described[48-50]. Fab fragments used in crystallization were produced

322 with a noncleavable 6xhistidine (6xHis) tag on the heavy chain C-terminus. Fab fragments were
323 produced by polyethylenimine (PEI) facilitated, transient transfection of 293F cells. Transfection
324 complexes were prepared in Opti-MEM (Gibco) and added to cells. 5 days post transfection, cell
325 supernatants were harvested and clarified by low-speed centrifugation. Fabs were purified by passage
326 over TALON Metal Affinity Resin (Takara) followed by gel filtration chromatography on Superdex 200
327 (GE Healthcare) in 10 mM tris(hydroxymethyl)aminomethane (tris), 150 mM NaCl at pH 7.5 (buffer A).
328
329 *Single B cell Nojima cultures.*
330 Nojima cultures were previously performed[18-20]. Briefly, peripheral blood mononuclear cells (PBMCs)
331 were obtained from four human subjects S1 (female, age range 51-55), S5 (male, age 21-25), S8
332 (female, age 26-30), and S9 (female, age 51-55). Single human Bmem cells were directly sorted into
333 each well of 96-well plates and cultured with MS40L-low feeder cells in RPMI1640 (Invitrogen)
334 containing 10% HyClone FBS (Thermo scientific), 2-mercaptoethanol (55 μM), penicillin (100 units/ml),
335 streptomycin (100 μg/ml), HEPES (10 mM), sodium pyruvate (1 mM), and MEM nonessential amino
336 acid (1X; all Invitrogen). Exogenous recombinant human IL-2 (50 ng/ml), IL-4 (10 ng/ml), IL-21 (10
337 ng/ml) and BAFF (10 ng/ml; all Peprotech) were added to cultures. Cultures were maintained at 37°
338 degrees Celsius with 5% CO_2. Half of the culture medium was replaced twice weekly with fresh medium
339 (with fresh cytokines). Rearranged *V(D)J* gene sequences for human Bmem cells from single-cell
340 cultures were obtained as described[20,48,51]. Specificity of clonal IgG antibodies in culture supernatants
341 and of rIgG antibodies were determined in a multiplex bead Luminex assay (Luminex Corp.). Culture
342 supernatants and rIgGs were serially diluted in 1 × PBS containing 1% BSA, 0.05% NaN_3 and 0.05%
343 Tween20 (assay buffer) with 1% milk and incubated for 2 hours at room temperature with the mixture of
344 antigen-coupled microsphere beads in 96-well filter bottom plates (Millipore). After washing three times
345 with assay buffer, beads were incubated for 1 hour at room temperature with Phycoerythrin-conjugated
346 goat anti-human IgG antibodies (Southern Biotech). After three washes, the beads were re-suspended
347 in assay buffer and the plates read on a Bio-Plex 3D Suspension Array System (Bio-Rad).
348
349 *Recombinant HA expression and purification.*
350 Recombinant HA head constructs and Full-length HA ectodomains (FLsE) were expressed by
351 polyethylenimine (PEI) facilitated, transient transfection of 293F cells. To clone HA heads, synthetic
352 DNA for the region was subcloned into a pVRC8400 vector encoding a C-terminal rhinovirus 3C
353 protease site and a 6xHis tag. To produce FLsE constructs, synthetic DNA was subcloned into a
354 pVRC8400 vector encoding a T4 fibritin (foldon) trimerization tag and a 6xHis tag. Transfection
355 complexes were prepared in Opti-MEM (Gibco) and added to cells. 5 days post transfection, cell
356 supernatants were harvested and clarified by low-speed centrifugation. HA was purified by passage
357 over TALON Metal Affinity Resin (Takara) followed by gel filtration chromatography on Superdex 200
358 (GE Healthcare) in buffer A. HA heads used for crystallography underwent the following additional
359 purification steps. HA heads were cleaved using the Pierce 3C HRV Protease Solution Kit (Ref 88947)
360 and passed over TALON Metal Affinity Resin to capture cleaved tags. Cleaved HA heads were then
361 further purified by gel filtration chromatography on Superdex 200 (GE Healthcare) in buffer A.
362
363 *ELISA*
364 Five hundred nanograms of rHA FLsE or HA head were adhered to high-capacity binding, 96 well-
365 plates (Corning 9018) overnight in PBS pH 7.4 at 4°C. HA coated plates were washed with a PBS-
366 Tween-20 (0.05%v/v) buffer (PBS-T) and then blocked with PBS-T containing 2% bovine serum
367 albumin (BSA) for 1 hour at room temperature. Blocking solution was then removed, and 5-fold dilutions
368 of IgGs (in blocking solution) were added to wells. Plates were then incubated for 1 hour at room
369 temperature. Primary IgG solution was removed and plates were washed three times with PBS-T.
370 Secondary antibody, anti-human IgG-HRP (Abcam ab97225) diluted 1:10,000 in blocking solution, was
371 added to wells and incubated for 30 minutes at room temperature. Plates were then washed three
372 times with PBS-T. Plates were developed using 150μl 1-Step ABTS substrate (ThermoFisher,
373 Prod#37615). Following a brief incubation at room temperature, HRP reactions were stopped by the
374 addition of 100μl of 1% sodium dodecyl sulfate (SDS) solution. Plates were read on a Molecular
375 Devices SpectraMax 340PC384 Microplate Reader at 405 nm. KD values for ELISA were obtained as

376 follows. All measurements were performed in technical triplicate. The average background signal (no
377 primary antibody) was subtracted from all absorbance values. Values from multiple plates were
378 normalized to the FI6v3[47] standard (FluA20 was used for the HA head assays) that was present on
379 each ELISA plate. The average of the three measurements were then graphed using GraphPad Prism
380 (v9.0). KD values were determined by applying a nonlinear fit (One site binding, hyperbola) to these
381 data points. The constraint that Bmax must be greater than 0.1 absorbance units was applied to all KD
382 analysis parameters.
383
384 *Recombinant IgG expression and purification.*
385 The heavy and light chain variable domains of selected antibodies were cloned into modified
386 pVRC8400 expression vectors to produce full length human IgG1 heavy chains and human lambda or
387 kappa light chains. IgGs were produced by transient transfection of 293F cells as specified above. Five
388 days post-transfection supernatants were harvested, clarified by low-speed centrifugation, and
389 incubated overnight with Protein A Agarose Resin (GoldBio) at 4°C. The resin was collected in a
390 chromatography column and washed with one column volume of buffer A. IgGs were eluted in 0.1M
391 Glycine (pH 2.5) which was immediately neutralized by 1M tris (pH 8.5). Antibodies were then dialyzed
392 against phosphate buffered saline (PBS) pH 7.4.
393
394 *Recombinant mIgG expression and endotoxin free purification.*
395 The heavy and light chain variable domains of selected antibodies were cloned into respective modified
396 pVRC8400 expression vector to produce full length murine IgG1 and IgG2C heavy chains and murine
397 lambda or kappa light chains. IgGs were produced by transient transfection of 293F cells as specified
398 above. Five days post-transfection supernatants were harvested, clarified by low-speed centrifugation.
399 20% volume of 1M 2-morpholin-4-ylethanesulfonic acid (MES) pH 5 and Immobilized Protein G resin
400 (Thermo Scientific Prod#20397) were added to supernatant and sample was incubated overnight at
401 4°C. The resin was collected in a chromatography column and washed with one column volume of
402 10mM MES, 150 mM NaCl at pH 5. mIgGs were eluted in 0.1M Glycine (pH 2.5), which was
403 immediately neutralized by 1M tris (pH 8.5). Antibodies were then dialyzed against phosphate buffered
404 saline (PBS) pH 7.4. All mIgGs administered to mice were purified using buffers made with endotoxin-
405 free water (HyPure Cell Culture Grade Water, Cytiva SH30529.03) and were dialyzed into endotoxin-
406 free Dulbecco's PBS (EMD Millipore, TMS-012-A). Following dialysis, endotoxins were removed using
407 Pierce High Capacity Endotoxin Removal Spin Columns (Ref 88274).
408
409 *Virus microneutralization assays.*
410 Two-fold serial dilutions of 50 ug/mL of HC19, S8V1-157 or CR3022 were incubated with $10^{3.3}$ $TCID_{50}$ of
411 A/Aichi/02/1968 H3N2 (X-31) influenza virus for 1 hour at room temperature with continuous rocking.
412 Media with TPCK was added to 96-well plates with confluent MDCK cells before the virus:antibody
413 mixture was added and left on the cells for the duration of the experiment. After 4 days, cytopathic effect
414 was determined and the neutralizing antibody titer was expressed as the reciprocal of the highest dilution
415 of serum required to completely neutralize the infectivity each virus on MDCK cells. The concentration of
416 antibody required to neutralize 100 $TCID_{50}$ of virus was calculated based on the neutralizing titer dilution
417 divided by the initial dilution factor, multiplied by the antibody concentration.
418
419 *Protection:*
420 C57BL/6 female and male mice were obtained from the Jackson Laboratory. All mice were housed
421 under pathogen-free conditions at Duke University Animal Care Facility. Eight to 10-week old mice
422 were injected *i.p.* with 150 μg of recombinant antibody diluted to 200 μl in PBS. Three hours later, mice
423 were anesthetized by *i.p.* injection of ketamine (85 mg/kg) and xylazine (10 mg/kg) and infected
424 intranasally with $3 \times LD_{50}$ (1.5×10^4 PFU) of A/Aichi/2/1968 X-31 (H3N2) in 40 μL total volume (20
425 μL/nostril). Mice were monitored daily for survival and body weight loss until 14 days post-challenge.
426 The humane endpoint was set at 20% body weight loss relative to the initial body weight at the time of
427 infection. All animal experiments were approved by the standards and guidance set forth by Duke
428 University IACUC.
429

430   Female C57BL/6J mice at 8 weeks old, purchased from Japan SLC, were i.p. injected with 150 µg of
431   the antibodies in 150 µL PBS. Three hours later, the mice were intranasally infected with $5LD_{50}$ of
432   A/Aichi/02/1968(H3N2)(X31), kindly gifted from Dr. Takeshi Tsubata (Tokyo Medical and Dental
433   University), under anesthesia with medetomidine-midazolam-butorphanol. Survival and body weight
434   were daily assessed for 14 days with a humane endpoint set as 25% weight loss from the initial body
435   weight. The experimental procedures were approved by the Animal Ethics Committee of the National
436   Institute of Infectious Diseases, Japan, and performed in accordance with the guidelines of the
437   Institutional Animal Care and Use Committee. Statistical significance of body weight change and
438   survival of mice after lethal influenza challenge was calculated by Two-way ANOVA test and Mantel-
439   Cox test, respectively, using GraphPad Prism (v10.0) software

441   *Competitive inhibition assay*
442   Competitive binding inhibition was determined by a Luminex assay, essentially as described[20,30].
443   Briefly, serially diluted human rIgGs or diluted (1:10) Bmem cell culture supernatants were incubated
444   with HA-conjugated Luminex microspheres for 2 h at room temperature or overnight at 4°C. S8V1-157
445   mouse IgG1 was then added at a fixed concentration (100 ng/ml final for competition with rIgGs, or 10
446   ng/ml final for competition with culture supernatants) to each well, and incubated with the competitor
447   Abs and HA-microspheres for 2 h at room temperature. After washing, bound S8V1-157 was detected
448   by incubating the microspheres with 2 µg/ml PE-conjugated rat anti-mouse IgG1 (SB77e,
449   SouthernBiotech) for 1 hr at room temperature, followed by washing and data collection. Irrelevant rIgG
450   or culture supernatants containing HA-nonbinding IgG were used as non-inhibiting controls.

452   *Flow cytometry*
453   Flow cytometry analysis of rIgG binding to HA-expressing K530 cell lines was performed essentially as
454   described [29]. Briefly, K530 cell lines were thawed from cryopreserved aliquots and expanded in culture
455   for ≥3 days. Pooled K530 cells were incubated at 4°C for 30 min with 0.4 µg/ml recombinant human
456   IgGs diluted in PBS plus 2% fetal bovine serum. After washing, cells were labeled with 2 µg/ml PE-
457   conjugated goat anti-human IgG (Southern Biotech) for 30 min at 4°C. Cells were then washed and
458   analyzed with a BD FACSymphony A5 flow cytometer. Flow cytometry data were analyzed with FlowJo
459   software (BD).

461   Flow cytometry analysis of rIgG binding to HA-expressing 293F cells was performed as follows. 293F
462   cells were transfected using PEI with either plasmid encoding full-length HA from
463   A/Aichi/02/1968(H3N2)(X31) or with empty vector. 36 hours post-transfection, cells were incubated at
464   4°C for one hour with 0.4 µg/ml recombinant human IgGs diluted in PBS plus 2% fetal bovine serum.
465   After washing, cells were labeled with BB515 mouse anti-human IgG (BD Biosciences) for 30 min at
466   4°C at the manufacturer's recommended concentration, followed by washing and fixation in 2%
467   paraformaldehyde. Cells were analyzed with a BD LSRFortessa flow cytometer. Flow cytometry data
468   were analyzed with FlowJo software (BD).

470   *Western blots*
471   293F cells were transfected using PEI with either plasmid encoding full-length HA from
472   A/Aichi/02/1968(H3N2)(X31) or with empty vector. 24 hours post-transfection, cells were lysed in RIPA
473   buffer (25mM Tris pH7.6, 150mM NaCl, 1% NP40 alternative, 1% sodium deoxycholate, 0.1% SDS).
474   Cell lysates were clarified of debris and boiled with Laemmli buffer with 2-mercaptoethanol. Samples
475   were run on 4-20% acrylamide gels and transferred to nitrocellulose membranes. Membranes were
476   blocked in 5% milk in PBS with 0.05% Tween 20 and probed with anti-HA tag antibody (Thermo-Fisher
477   A01244-100; 0.3 µg/ml), followed by IR800-conjugated goat anti-mouse immunoglobulin (LiCor 926-
478   32210; 0.1 µg/ml) and DyLight680-conjugated mouse anti-rabbit GAPDH antibody (BioRad
479   MCA4739D680; 0.3 µg/ml). Membranes were imaged using a LiCor Odyssey CLx imager.

481   *Crystallization*

482 S1V2-157 Fab fragments were co-concentrated with the HA-head of A/American black duck/New
483 Brunswick/00464/2010(H4N6) at a molar ratio of ~1:1.3 (Fab to HA-head) to a final concentration of
484 ~20 mg/ml. Crystals of Fab-head complexes were grown at 18°C in hanging drops over a reservoir
485 solutions containing 0.1 M Lithium sulfate, 0.1 M Sodium chloride, 0.1 M 2-(N-
486 morpholino)ethanesulfonic acid (MES) pH 6.5 and 30% (v/v) poly(ethylene glycol) (PEG) 400. Crystals
487 were cryprotected with 30% (v/v) PEG 400, 0.12 M Lithium sulfate, 0.3 M Sodium chloride, and 0.06 M
488 MES pH 6.5. Cryprotectant was added directly to the drop, crystals were harvested, and flash cooled in
489 liquid nitrogen.
490
491 *Structure determination and refinement*
492 We recorded diffraction data from a single moderate to poorly diffracting crystal at the Advanced
493 Photon Source on beamline 24-ID-C. The dataset was strongly anisotropic, variable mosaicity (average
494 1.4°) and comprised contributions from multiple lattices. Efforts to improve these crystals were
495 unsuccessful. Data were processed and scaled (XSCALE) with XDS[52]. Molecular replacement was
496 carried out with PHASER[53], dividing each complex into four search models (HA-head, Vh, Vl and
497 constant domain). Search models were 5XL2, 6EIK , 5BK5 and 6E56.We carried out refinement
498 calculations with PHENIX[54] and model modifications, with COOT[55]. Refinement of atomic positions and
499 B factors was followed by translation-liberation-screw (TLS) parameterization. All placed residues were
500 supported by electron density maps and subsequent rounds of refinement. Final coordinates were
501 validated with the MolProbity server[56]. Data collection and refinement statistics are in Table S1. Figures
502 were made with PyMOL (Schrödinger, New York, NY.
503
504 **DATA AND SOFTWARE AVAILABILITY**:
505 Coordinates and diffraction data have been deposited at the PDB, accession number 8US0. Antibody
506 sequences have been deposited in NCBI GenBank, accession numbers OR825693-OR825700.
507
517
518 **FIGURE LEGENDS:**
519
520 **Figure 1: Identification of a novel, broadly binding, HA head-directed antibody class.** A. Luminex
521 screening of Bmem-cell Nojima culture supernatants identified four antibodies that broadly react with
522 influenza A HA FLsEs and heads. The mean fluorescence intensity (MFI) values are colored according
523 to the key. B. In a Luminex competitive binding assay, the four antibodies from (A) that share a pattern
524 of reactivity did not compete with antibodies that engage known HA epitopes, but compete with each
525 other for HA binding. Structures of Fab-HA complexes were aligned on an HA trimer from A/American
526 black duck/New Brunswick/00464/2010(H4N6) (PDB: 5XL2)[21]. Fab structures include HC19[32] (PDB
527 2VIR), S5V2-29[20] (PDB 6E4X), HC45[57] (PDB 1QFU), CR9114[58] (PDB 4FQY), CR8020[59] (PDB 3SDY)
528 and FI6v3[47] (PDB 3ZTJ). SARS-CoV antibody CR3022[33] was used as an HA non-binding control. C.
529 The cross-competing HA antibodies share genetic signatures.
530
531 **Figure 2: Human antibodies engage a recessed surface at the head-stem interface of the**
532 **influenza HA molecule.** A. Structure of antibody S8V1-157 complexed with the HA head of
533 A/American black duck/New Brunswick/00464/2010(H4N6) colored in gray. The heavy chain is colored
534 darker blue and the light chain is lighter blue. Engagement of this site is incompatible with the defined
535 prefusion H4 HA trimer[21], colored in white (PDB: 5XL2) or with individual HA monomers. B. A surface

536 projection showing the degree of amino acid conservation among HAs engaged by this antibody class
537 (see Figure 3). The head-stem epitope is circumscribed in blue in the rightmost panel. Conservation
538 scores were produced using ConSurf [60,61]. C. Key S8V1-157 contacts. The orientation relative to panel
539 A is indicated.
540
541 **Figure 3: Breadth of HA binding by HA head-stem epitope antibodies.** A. Equilibrium dissociation
542 constants (K_d), determined by ELISA. Broadly binding influenza A HA antibody FI6v3 [47] and influenza B
543 HA antibody CR8071 [58] served as binding controls. B. Phylogenetic relationships of HAs used in our
544 panel. HAs bound by HA head-stem epitope antibodies are indicated. Binding data from Figure S4 are
545 incorporated into panel B.
546
547 **Figure 4: HA head-stem epitope antibodies bind cell surface-anchored HA.** Flow cytometry
548 histograms depict the fluorescence intensities of recombinant IgG binding to K530 cell lines expressing
549 recombinant, native HA on the cell surface. K530 cells were labeled with 400 ng/ml of the four head-
550 stem epitope antibodies or control antibodies targeting the HA receptor binding site (HC19[32], K03.12[30]
551 and H5.3[31]), the head interface (S5V2-29[20]), a lateral head epitope (HC45)[57], stem (FI6v3[47]), or SARS-
552 CoV spike protein (CR3022[33]). HA abbreviations correspond to: H1/SI06: A/Solomon
553 Islands/3/2006(H1N1), H1/CA09: A/California/04/2009(H1N1), H2/JP57: A/Japan/305/1957(H2N2),
554 H5/VN04: A/Viet Nam/1203/2004(H5N1), H6/TW13: A/Taiwan/2/2013(H6N1), H8/CA07: A/northern
555 shoveler/California/HKWF1204/2007(H8N4),H3/HK68: A/Aichi/02/1968(H3N2)(X31), H3/TX12:
556 A/Texas/50/2012(H3N2), H4/NB10:A/American black duck/New Brunswick/00464/2010, H7/TW13:
557 A/Taiwan/1/2017(H7N9), H10/JX13: A/Jiangxi/IPB13/2013(H10N8),H14/WI10:
558 A/mallard/Wisconsin/10OS3941/2010(H14N6).
559
560 **Figure 5: HA head-stem epitope antibodies protect against lethal influenza virus infection and**
561 **severe disease.** C57BL/6 mice (n = 7 per group) were intraperitoneally injected with 150 μg of
562 recombinant antibody via intraperitoneal injection three hours prior to intranasal challenge with 5xLD50
563 of A/Aichi/02/1968(H3N2)(X31). Mice were weighed daily and euthanized at a humane endpoint of 25%
564 loss of body weight. Antibodies passively transferred included musinized IgG1 and IgG2c versions of
565 HA head-stem epitope antibodies S8V1-157 and S1V2-65, neutralizing antibody HC19[32], head interface
566 antibody S5V2-29[20] and SARS-CoV antibody CR3022[33]. Mice injected with PBS were included as an
567 additional control. A. Post-infection survival rate. B-D. Body weight curves for infected mice
568 administered S8V1-157 (B), S1V2-65 (C), or S5V2-29 (D) antibodies, compared with controls. *p <
569 0.05 and ***p < 0.001 compared with isotype control CR3022. Not significant (n.s.), p 0.05; † p < 0.05,
570 †† p < 0.01, and †††† p < 0.0001 IgG2c compared with IgG1.
571
572 **Figure 6: The HA head-stem epitope epitope is immunogenic in humans.** A. An additional 528
573 Nojima culture supernatants from donors K01, K03, S1, S5, S8, S9 and S12 were screened for
574 competition with a recombinant musinized S8V1-157 IgG1 for HA binding. Culture supernatants that
575 inhibited S8V1-157 binding by >90% are colored and specified. B. HA reactivity of S8V1-157-competing
576 Nojima culture supernatants, as determined by multiplex Luminex assay. N.D.: not determined.
577
578 **Supporting Figures:**
579
580 **Supporting Table 1: Data collection and refinement statistics.**
581
582 **Figure S1: S8V1-157 Contacts conserved sites.** An alignment of HA head regions for HAs used in
583 this study. S8V1-157 contacting residues in A/American black duck/New Brunswick/00464/2010(H4N6)
584 are colored red. HAs not bound by S8V1-157 are in denoted in gray. Sites identical to the A/American
585 black duck/New Brunswick/00464/2010(H4N6) sequence are indicated by a ".". Amino acid numbering
586 corresponds to standard H3N2 numbering.
587
588 **Figure S2: Surface hydrophobicity on the HA molecule.** Surface hydrophobicity and the head-stem
589 epitope are shown on the HA trimer of A/American black duck/New Brunswick/00464/2010(H4N6)

590   (PDB: 5XL2) [21]. Coloring is based on the PyMOL Color h script that utilizes a normalized consensus
591   hydrophobicity scale[62]. Views and orientations match those in Figure 2. The head-stem epitope is
592   circled in the far right panel.
593
594   **Figure S3: ELISA titrations of antibodies on HA coated plates.** The broadly binding stem antibody
595   FI6v3[47] was used as a positive control and an influenza B specific head antibody, CR8071[58], as a
596   negative control for influenza A isolates. Data points represent the average of three technical replicates.
597   The standard error of the mean is shown for each point. KDs were calculated from the curves fit to
598   these data points.
599
600   **Figure S4: Head-stem epitope antibody binding to HA head-only constructs.** A. Structures of HA
601   head used in these experiments the S8V1-157-HA head complex is shown for reference. A compact
602   head[49] , is shown for reference (PDB 7TRH). B. HA heads for HAs not bound or not expressed in
603   Figure 3 were produced alongside a positive control A/California/07/2009(H1N1)(X-181) and a compact
604   head version. Sequences of the HA head regions are aligned to the A/Aichi/02/1968(H3N2)(X-31)
605   reference sequence and numbered by H3 convention. C. Dissociation constants from ELISA
606   measurements of head-stem epitope antibodies to HA heads. Head interface antibody FluA-20[40] was
607   used as a positive control and influenza B head antibody CR8071[58] as a negative control.
608
609   **Figure S5: ELISA titrations of antibodies on HA head coated plates and characterization of cell**
610   **expressed HA.** A. Head interface antibody FluA-20[40] was used as a positive control and influenza B
611   head antibody CR8071[58] as a negative control. Data points represent the average of three technical
612   replicates. The standard error of the mean is shown for each point. KDs were calculated from the
613   curves fit to these data points. B. Lysates from 293F transfected with either HA from
614   A/Aichi/02/1968(H3N2)(X31) or empty vector were subjected to western blotting under reducing and
615   denaturing conditions with either anti-HA tag antibody, which recognizes an endogenous sequence in
616   the H3 HA head (HA1), or anti-GAPDH antibody. Molecular weights of unprocessed HA0 and
617   processed HA1 are indicated with open or closed arrowheads, respectively. C. 293F transfected with
618   either HA from A/Aichi/02/1968(H3N2)(X31) or empty vector were stained with the indicated antibodies
619   and analyzed by flow cytometry. Gates denoting cells bound by antibody are shown, with the percent of
620   the total population indicated above.
621
622   **Figure S6: S8V1-157 is a non-neutralizing antibody.** Neutralization IC50 values for S8V1-157,
623   neutralizing antibody HC19[32] (positive control) and SARS-CoV antibody CR3022[33] (negative control).
624
625   **Figure S7: Antibody genetics of S8V1-157 competing antibodies.** Antibody names, gene usage
626   and HCDR3 sequences for S8V1-157 competing antibodies identified in Figure 6.
627
628
629
630
631

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794
795

A.

Antibody	H1 2009 X	H1 2009	H1 2006	H3 1968	H3 1968 H	H3 1994 H	H3 2005	H3 2014	H5 2004	BYam 2013	BVic 2004	IgG	Igκ	Igλ
S1V2-17	45,566	3,699	1,801	20,292	193,960	89,944	9,237	18,673	64,650			85,284	66,873	
S1V2-60	58,140	6,418	6,148	25,742	198,166	139,909	15,213	27,634	78,010			89,707	73,143	
S1V2-65	47,698	4,273	3,224	24,328	182,993	129,196	12,551	23,051	74,100			89,661	66,611	
S8V1-157	17,600	386	53	2,558	162,331	44,285	25,049	6,644	12,032			38,425	23,142	

MFI				
<100	100-1,000	1,000-10,000	10,000-100,000	>100,000

B.

C.

Antibody	IGHV	IGHJ	IGHJ	HCDR3	Length	IGVL	JL	LCDR3	Length
S1V2-17	IGHV1-24*01	IGHJ4*02	IGHD2-21*02	C VTWLVGTTVAAY W	12	IGKV2-28*01[a]	IGKJ1*01	C MQALQAPRT F	9
S1V2-60	IGHV5-51*01	IGHJ4*02	IGHD5-12*01	C VRLGYGDSPPFDS W	13	IGKV2-28*01[a]	IGKJ2*01	C MQALQDPYT F	9
S1V2-65	IGHV5-51*01	IGHJ4*02	IGHD5-12*01	C ARLGYGDSPPFDF W	13	IGKV2-28*01[a]	IGKJ2*01	C MQALETPYT F	9
S8V1-157	IGHV1-24*01	IGHJ4*02	IGHD1-26*01	C ATLMGANPFDY W	11	IGKV2-28*01[a]	IGKJ2*01	C KQALQTLYT F	9

[a]or IGKV2D-28*01

Figure 1: Identification of a novel, broadly binding, HA head-directed antibody class. A. Luminex screening of Bmem-cell Nojima culture supernatants identified four antibodies that broadly react with influenza A HA FLsEs and heads. The mean fluorescence intensity (MFI) values are colored according to the key. B. In a Luminex competitive binding assay, the four antibodies from (A) that share a pattern of reactivity did not compete with antibodies that engage known HA epitopes, but compete with each other for HA binding. Structures of Fab-HA complexes were aligned on an HA trimer from A/American black duck/New Brunswick/00464/2010(H4N6) (PDB: 5XL2)[21]. Fab structures include HC19[32] (PDB 2VIR), S5V2-29[20] (PDB 6E4X), HC45[57] (PDB 1QFU), CR9114[58] (PDB 4FQY), CR8020[59] (PDB 3SDY) and FI6v3[47] (PDB 3ZTJ). SARS-CoV antibody CR3022[33] was used as an HA non-binding control. C. The cross-competing HA antibodies share genetic signatures.
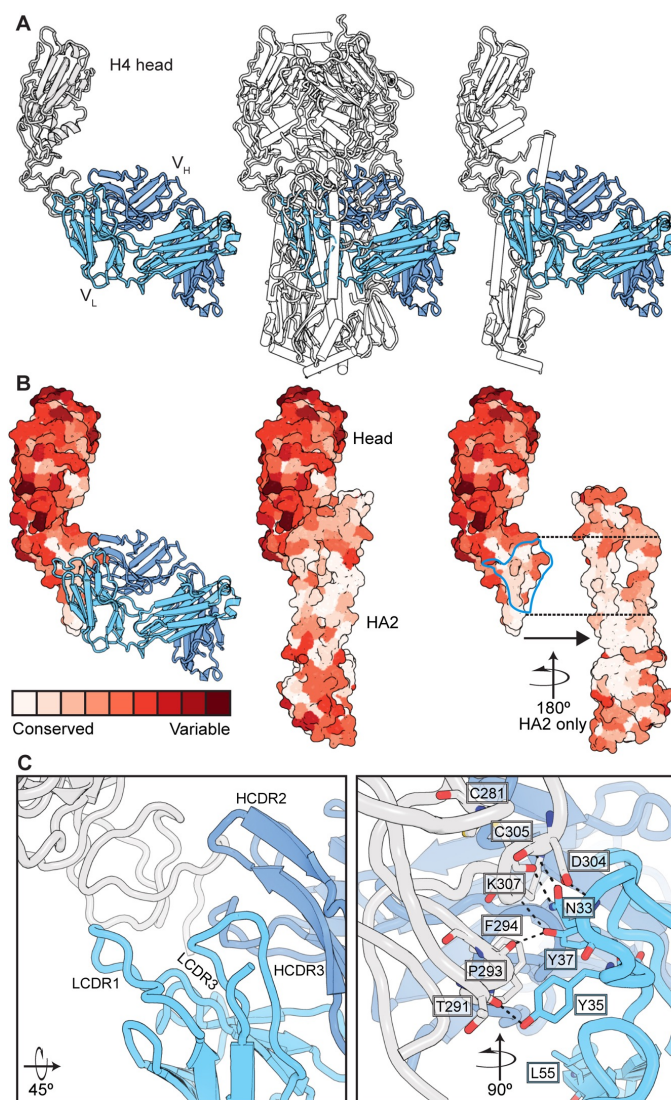
810
Figure 2: Human antibodies engage a recessed surface at the head-stem interface of the influenza HA molecule. A. Structure of antibody S8V1-157 complexed with the HA head of A/American black duck/New Brunswick/00464/2010(H4N6) colored in gray. The heavy chain is colored darker blue and the light chain is lighter blue. Engagement of this site is incompatible with the defined prefusion H4 HA trimer[21], colored in white (PDB: 5XL2) or with individual HA monomers. B. A surface projection showing the degree of amino acid conservation among HAs engaged by this antibody class (see Figure 3). The head-stem epitope is circumscribed in blue in the rightmost panel. Conservation scores were produced using ConSurf [60,61]. C. Key S8V1-157 contacts. The orientation relative to panel A is indicated.

A

		S1V2-17	S1V2-60	S1V2-65	S8V1-157	CR8071	FI6
H1N1	A/USSR/90/1977	7.0	1.2	2.2	2.7	>100	0.32
	A/Chile/01/1983	13	3.7	5.3	8.1	>100	0.33
	A/Texas/36/1991	2.9	0.73	1.0	1.2	>100	0.28
	A/Solomon Islands/3/2006	14	3.5	4.1	8.3	>100	0.67
	A/California/04/2009	6.9	0.86	1.0	3.8	>100	0.42
	A/Victoria/2570/2019(IVR-215)	2.8	0.39	0.40	2.1	<100	0.35
H2N2	A/Japan/305/1957	6.5	8.8	10	2.5	>100	0.36
H3N2	A/Aichi/02/1968(X31)	0.34	0.16	0.16	0.18	>100	0.31
	A/Philippines/02/1982	0.58	0.25	0.21	0.29	>100	1.2
	A/Sydney/05/1997	0.50	0.27	0.25	0.21	>100	3.0
	A/Wisconsin/67/2005	0.99	0.26	0.21	0.42	>100	3.5
	A/Hong Kong/4801/2014	0.85	0.26	0.21	0.49	>100	0.88
	A/Tasmania/503/2020(IVR-221)	0.62	0.28	0.21	0.22	>100	2.8
H4N6	A/American black duck/New Brunswick/00464/2010	0.55	0.17	0.20	0.32	>100	1.4
H5N1	A/Viet Nam/1203/2004	0.14	0.10	0.10	<0.1	>100	0.44
H6N1	A/Taiwan/2/2013	>100	>100	>100	>100	>100	0.47
H7N9	A/Taiwan/1/2017	0.11	0.13	0.12	<0.1	>100	0.24
H8N4	A/northern shoveler/California/HKWF1204/2007	9.2	0.77	1.7	4.5	>100	0.29
H9N2	A/turkey/Wisconsin/1/1966	>100	3.2	>100	>100	>100	0.37
	A/swine/Hong Kong/9/98	13	0.48	8.0	26	>100	0.40
H11N9	A/duck/Memphis/546/1974	>100	>100	>100	>100	>100	0.74
H12N5	A/duck/Alberta/60/1976	>100	>100	>100	>100	>100	0.44
H14N6	A/mallard/Wisconsin/10OS3941/2010	1.9	0.26	0.30	1.2	>100	1.4
H15N9	A/wedge-tailed shearwater/Western Australia/2576/1979	61	0.65	8.0	6.0	>100	2.5
H16N3	A/black-headed gull/Sweden/2/99	>100	20	>100	>100	>100	0.55
Influenza B	B/Brisbane/60/2008	>100	>100	>100	>100	0.35	>100

Kd in nM				
<1	1-5	5-25	25-100	>100

B

Figure 3: Breadth of HA binding by HA head-stem epitope antibodies. A. Equilibrium dissociation constants (K_d), determined by ELISA. Broadly binding influenza A HA antibody FI6v3 [47] and influenza B HA antibody CR8071 [58] served as binding controls. B. Phylogenetic relationships of HAs used in our panel. HAs bound by HA head-stem epitope antibodies are indicated. Binding data from Figure S4 are incorporated into panel B.
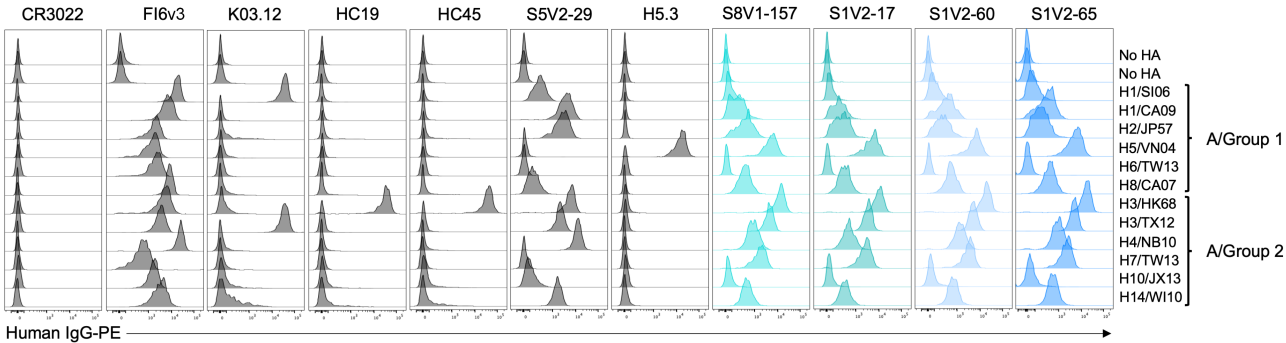
Figure 4: HA head-stem epitope antibodies bind cell surface-anchored HA. Flow cytometry histograms depict the fluorescence intensities of recombinant IgG binding to K530 cell lines expressing recombinant, native HA on the cell surface. K530 cells were labeled with 400 ng/ml of the four head-stem epitope antibodies or control antibodies targeting the HA receptor binding site (HC19[32], K03.12[30] and H5.3[31]), the head interface (S5V2-29[20]), a lateral head epitope (HC45)[57], stem (FI6v3[47]), or SARS-CoV spike protein (CR3022[33]).
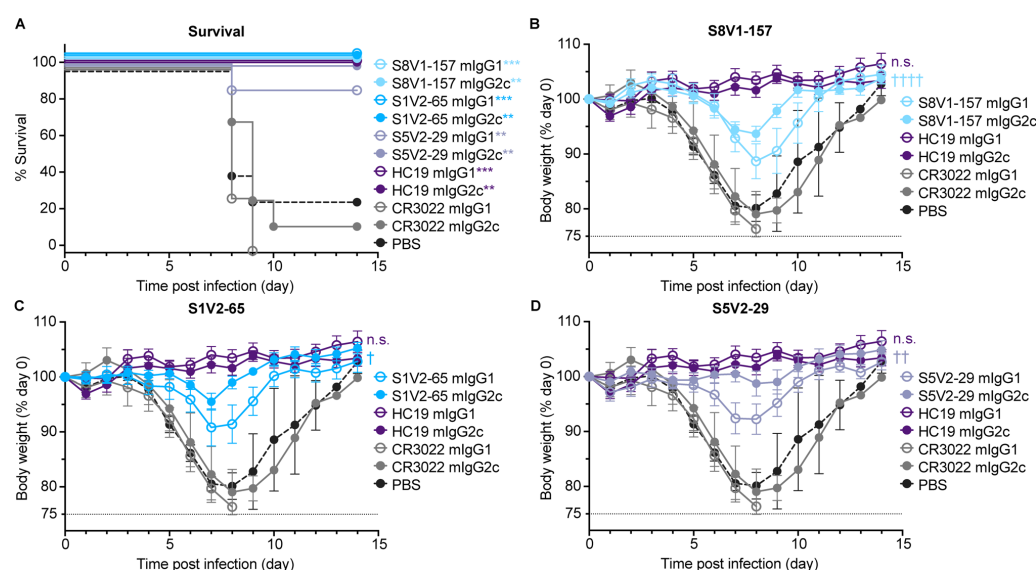
Figure 5: HA head-stem epitope antibodies protect against lethal influenza virus infection and severe disease. C57BL/6 mice (n = 7 per group) were intraperitoneally injected with 150 μg of recombinant antibody via intraperitoneal injection three hours prior to intranasal challenge with 5xLD50 of A/Aichi/02/1968(H3N2)(X31). Mice were weighed daily and euthanized at a humane endpoint of 25% loss of body weight. Antibodies passively transferred included musinized IgG1 and IgG2c versions of HA head-stem epitope antibodies S8V1-157 and S1V2-65, neutralizing antibody HC19[32], head interface antibody S5V2-29[20] and SARS-CoV antibody CR3022[33]. Mice injected with PBS were included as an additional control. A. Post-infection survival rate. B-D. Body weight curves for infected mice administered S8V1-157 (B), S1V2-65 (C), or S5V2-29 (D) antibodies, compared with controls. *p < 0.05 and ***p < 0.001 compared with isotype control CR3022. Not significant (n.s.), p 0.05; † p < 0.05, †† p < 0.01, and †††† p < 0.0001 IgG2c compared with IgG1.

Antibody	H1 2009 X	H1 2009	H1 2006	H3 1968	H3 2005	H3 2014ᵃ	H3 2014ᵇ	H5 2004	BYam2013	BVic 2004	IgG	Igκ	Igλ
S8V5 P5-C6	211,868	N.D.	108,575					N.D.			85,808	80,971	
S8V6 P7-H9		N.D.		57,232	41,063	17,839	35,872	N.D.			100,560	104,082	
S8V6 P10-G1	183,143	N.D.	27,673					N.D.			80,617		65,397
S1V4 P4-C5	11,528	2,664		26,677	1,387	13,996	7,592	2,315	200		64,575	78,170	
S5V4 P1-A2		1,555		53,091	67,485	29,348	111,273				64,266	74,232	
S5V4 P2-D2		2,711	200	93,375	105,075	79,837	166,328				82,873	91,355	
S12V4 P1-F1	42,514	19,363	600	9,458	3,555	22,409	5,793	16,648			59,983	73,887	
S12V6 P7-H4		1,225		600	18,737	11,681	1,041				66,471	68,493	

MFI				
<100	100-1,000	1,000-10,000	10,000-100,000	>100,000

Figure 6: The HA head-stem epitope epitope is immunogenic in humans. A. An additional 528 Nojima culture supernatants from donors K01, K03, S1, S5, S8, S9 and S12 were screened for competition with a recombinant musinized S8V1-157 IgG1 for HA binding. Culture supernatants that inhibited S8V1-157 binding by >90% are colored and specified. B. HA reactivity of S8V1-157-competing Nojima culture supernatants, as determined by multiplex Luminex assay. N.D.: not determined.

862 **Supporting Figures:**

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865 **Supporting Table 1: Data collection and refinement statistics.**

	S8V1-157-A/American black duck/New Brunswick/00464/2010(H4N6) Fab-HA head complex
PDB ID	8US0
Data Collection	APS 24-ID-C
Number of datasets	1
Resolution, Å	49.17 - 3.704 (3.836 - 3.704)
Wavelength (Å)	0.9792
Space Group	P 1 21 1
Unit cell dimensions (a, b, c), Å	127.92 241.54 144.13
Unit cell angles (α, β, γ) °	90.00 101.62 90.00
I/σ	7.00 (1.20)
Rmeas	0.15 (1.12)
Rpim	0.10 (0.80)
Rmerge, %	0.10 (0.80)
CC*	1.00 (0.79)
CC½	0.99 (0.45)
Completeness, %	98.77 (98.49)
Number of reflections	175949 (17445)
Redundancy	2.0 (2.0)
Refinement	
Number of reflections:	
Working	89804 (8919)
Free	4506 (491)
Rwork, %	24.45 (38.84)
Rfree, %	29.14 (41.97)
Ramachandran plot, % (favored, disallowed)	94.83 (0.14)
Rmsd bond lengths, Å	0.002
Rmsd bond angles, °	0.49
Average B-factor	144.76

Rmerge, $\Sigma_{h,k,l}\Sigma_i|I_i(hkl) - <I(hkl)>|/\Sigma_{h,k,l}\Sigma_i I_i(hkl)$, where I is an intensity that is observed i times; I/σ, signal-to-noise ratio (average observed intensity divided by average standard deviation of the observed intensity); Rwork, $\Sigma_{h,k,l}||F_{obs}| - |F_{calc}||/\Sigma_{h,k,l}|F_{obs}|$, where h, k, l covers the "working set" of observed structure factor amplitude (F_{obs}) reflections used in refinement (total reflections minus the test set) and F_{calc} is the calculated structure factor amplitude; Rfree, calculated as for Rwork but on 5% of data excluded prior to refinement. Values in parentheses refer to highest-resolution shell. Related to Experimental Procedures.

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                                                                 37                                                                                                      147
A/American black duck/New Brunswick/00464/2010(H4N6):          -----------VAAQELVESQHLPELCPS-PLRLVDGQTCDIVNGALGSPGCDHLNG-AEWDIFIERPTAV-DTCYPFDVPDYQSLRSILANNGKFEFI---AEE----FQWSTV-KQNGKSGACKRA-NVNDFFN
A/Aichi/02/1968(X-31)(H3N2):                                   -----------TN.T...Q.SSTGKI.NN-.H.IL..ID.TLIDAL..D.H..VFQN-ET..L.V..SK.F-SN...Y.....A....LV.SS.TL..---T.G----.T.TG.-T...G.N....G-PGSG..S
A/Philippines/2/1982(H3N2):                                    -----------TN.T...Q.SSTGRI.D.-.H.IL..KN.TLIDAL..D.H..GFQN-EK..L.V..SK.F-SN...Y.....A....LV.SS.TL..---N.TG.-T.S.G.ST...G-SN.S..S
A/Sydney/5/97(H3N2):                                           -----------TN.T...Q.SSTGRI.D.-.H.IL..EN.TLIDAL..D.H..GFQN-K...L.V..SK.Y-SN...Y.....A....LV.SS.TL..N---N.S----.N.TG.-A...T.Y...S-SIKS..S
A/Wisconsin/67/2005(H3N2):                                     -----------TN.T...Q.SSTGGI.D.-.HQIL..EN.TLIDAL..D.Q..GFQN-KK..L.V..SK.Y-SN...Y.....A....LV.SS.TL..N---N.S----.N.TG.-T...T.SS...R-SN.S..S
A/Hong Kong/4801/2014(H3N2):                                   -----------TN.T...QNSSIG.I.D.-.HQIL..EN.TLIDAL..D.Q..GFQN-KK..L.V..SK.Y-SN...Y.....A....LV.SS.TL..N---N.S----.N.TG.-T...T.SS..I.R-SN.S..S
A/Tasmania/503/2020(IVR-221)(H3N2):                            -----------TN.T...QNSSIG.I.D.-.HQIL..EN.TLIDAL..D.Q..GFQN-K...L.V..SK.N-SN...Y.....A....LV.SS.TL..K---N.S----.N.TG.-...T.S..I.G-SSSS..S
A/mallard/Wisconsin/10OS3941/2010(H14N6):                      -----------.S.K....TN.IN.....-.K.....D.LI...........QD-TT..V......M---------.....SS.SL..---..Q----.T.NG.-TVD.S..S..LL-G-GR.G..T
A/Taiwan/01/2017(H7N9):                                        -----------.N.T.T..RTNT.RI.SK-GK.T..LGQ.GLLGTIT.P.Q..QFLE-FSA.LI...REGS-.V...GKFVNEEA...Q..RES.GIDKE---PMG-----TYNGT-RT..VTS..R.--SGSS.YA
A/chicken/Germany/n/1949(H10N7):                               -----------TN.T.T...TN.NK..MK-GRSYK.LGN.HP.GMLI.T.V..PHLT-GT..TL...EN.I-AH...GATINEEA..QKIMES.GISKM---STG-----TYGSSINSA.TTK..M.N-GGDS.YA
A/wedge-tailed shearwater/Western Australia/2576/1979(H15N9):  -----------.N.T.T..ITGIDKV.TK-GKKA..LGS.G.LGTII.P.Q..LHLE-FKA.LI...RNSS-.I...GRFTNEEA...Q.IRES.GIDKE---SMG-----RY.GI-RTD.ATS....-T.SS.YS
A/USSR/90/1977(H1N1):                                          -----------THSVN.L.DS.NGK..RLKGIAPLQLGK.N.AGWI..N.E.ES.FSKKS.SYIA.T.NSENG....GYFA..EE..EQ.SSVSS..RFEIFPK.----RS.PKHNVTR.VTAS.SHK-GKSS.YR
A/Chile/1/1983(H1N1):                                          -----------THSVN.L.DN.NGK..RLKGIAPLQLGK.N.AGWI..N.E.ES.FSKKS.SYIA.T.NSENG....GYFA..EE..EQ.SSVSS..RFEIFPK.----RS.PKHNVTK.VTA..SHK-GKSS.YR
A/Texas/36/1991(H1N1):                                         -----------THSVN.L.DS.NGK..RLKGIAPLQLGK.N.SVAGWI..N.K.ES.FSKES.SYIA.T.NPENG....GYFA..EE..EQ.SSVSS..RFEIFPK.----RS.PNHTVTK.VTTS.SHN-GKSS.YR
A/Solomon Islands/03/2006(H1N1):                               -----------THSVN.L.DS.NGK..LLKGIAPLQLGN.SVAGWI..N.E.EL.ISRES.SYIV.K.NPENG....GHFA..EE..EQ.SSVSS..RFEIFPK.----SS.PNHTT-T.V.AS.SHN-GESS.YK
A/California/04/2009(H1N1):                                     -----------THSVN.L.DK.NGK..KLRGVAPLHLGK.N.AGWI..N.E.ES.STASS.SYIV.T.SSDNG....G.FI..EE..EQ.SSVSS..RFEIFPKT----SS.PNHDSNK.VTA..PH.-GAKS.YK
A/Victoria/2570/2019(IVR-215)(H1N1):                           -----------THSVN.L.DK.NGK..KLRGVAPLHLGK.N.AGWI..N.E.ES.STARS.SYIV.TSNSDNG....G.FIN.EE..EQ.SSVSS..RFEIFPKT----SS.PNHDSD..VTA..PH.-GAKS.YK
A/Japan/305/1957(H2N2):                                        -----------TH.KDIL.KT.NGK..KLNGIPPLELGD.S.AGWL..N.E..R.LSVP..SYIM.KENPRDGL...GSFN..EE.KHL.SSVKH..KVKILPK-----DR.TQHTTT-..G.R..AVS-GNPS..R
A/Viet Nam/1203/2004(H5N1):                                    -----------TH..DIL.KK.NGK..DLDGVKPLILRD.SVAGWL..N.M..FIHVP..SYIV.KANP.N.L...G.FN..EE.KHL.SRINH..K.QIIPK-----SS..SHEASL.V.S..PYQ-GKSS..R
A/Taiwan/2/2013(H6N1):                                         -----------THSV..L.N.KEKRF.KIMNKAPL.LKD.T.EGWI..N.K..L.L.DQS.SYIV...N.QNGI...GVLNELEE.KAFIGSGERV.RFEMFPK-----ST.AG.DTSR.VTN..PSYTIDSS.YR
A/northern shoveler/California/HKWF1204/2007(H8N4):            -----------TQTM....TEKH.AY.NTDLGTPLELRD.K.EAVIY.N.K..THLKDQG.SYIV...S.PEGM...GS.ENLEE..FVFS.AASYKR.RLFDY-----SR.NVT--SS.T.K..NASTGGQS.YR
A/swine/Hong Kong/9/98(H9N2):                                  -----------TH.K..LHTE.NGM..ATNLGHPLILD..T.EGLIY.N.S..L.L.GR..SYIV...S..NGM...GN.ENLEE..LFSSASSYQR.QIFPD-----TI.NVS--YS.T.K..SD-----S.YR
A/turkey/Wisconsin/1/1966(H9N2):                               -----------THTK..LHTE.NGM..ATDLGHPLILD..T.EGLIY.N.S..I.L.GK..SYIV..SS..NGM...GN.ENLEE..LFSSAKSYKR.QIFPD-----KT.NVT--YS.T.R..SN-----S.YR
A/duck/Alberta/60/1976(H12N5):                                 -----------TQVE...HGGID.I..GTELGSPLVLDD.SLEGLI..N.K..LYLNGR..SYIV...KEMEGV...GSIENQEE..LFSSIK.Y.RVKMFDF-----TK.NVT--YT.T.K..NNTS.QGS.YR
A/duck/Memphis/546/1974(H11N9):                                -----------TSSV....NEYTGSF.SIDGKAPISLGD.SFAGWI..N.M..D.I.KTS.SYIV.K.NPENG....GTLENEEE..LKFSGVLE.NKFEAFTS-----NG.GS.NSGA.VTA...FG-SS.S..R
A/duck/Hokkaido/W345/2012(H13N2):                              -----------TSSVD...TN.TGTY.ALGGISP.HLGD.SFEGWIV.N.A.ASNL.IR..SYL..D.S.PHGL...GELDNNGE..HLFSGIRS.SRTELI.P-----TS.GA.--ND.V.A..TDR-GASS.YR
A/black-headed gull/Sweden/2/99(H16N3):                        -----------TSSID...TN.TGTY.SLNGVSPIHLGD.SFEGWIV.N.S.ASNINIR..SYL..D.N.PHKL.F.GE.DNNGE..HLFSGVNS.SRTELIPP-----SS.GDI--LE.TTAS.QNR-GA.S.YR
B/Brisbane/60/2008:                                            TGVIPLTTTPTKSHFAN.KGTETRGK...K-------CLN.TDLDV...R.K.TGKIPS.RVS.LH.V-RP.TSG.F.IM-H.RTKI.QLPNLLRGY.H.RLSTHNVINAENAPGGPYKI.T..S.PNIT.G.G..A


                                                                 150                                                                                               265
A/American black duck/New Brunswick/00464/2010(H4N6):         RLNWLTK-SDGN----AYPLQNLTKINNGDYARLYIWGVHHPSTDTEQTNLYKNNPGRVT-VSTKTSQTSVVPNIGSRPWV-----RGQSGRISFYWTIVEPGDLIVFNTIGNLIAPRGHYKLNSQKKS--------
A/Aichi/02/1968(X-31)(H3N2):                                  .......-.G-S----T..VL.V.MP..DNFDK.....I....NQ...S..VQAS....-.RR..QTII................-----L.S...I....K...VL.I.SN........YF.MRTG-..--------
A/Philippines/2/1982(H3N2):                                   .....YE-.E-S----K..VL.V.MP..KFDK.....I....K.....IRAS....-.R..QT.I................------L.S...I....K...ILLI.ST........YF.IRTG-..--------
A/Sydney/5/97(H3N2):                                          .....HQ-LA-Y----K..VL.V.MP..DKFDK...........SD..SI.AQAS....-.R..QT.I................-----.I.S...IH....K...ILLI.ST........YF.IR.G-..--------
A/Wisconsin/67/2005(H3N2):                                    ......Q-LK-F----K..AL.V.MP..EKFDK..........V..ND.IF..AQAS..I.----.R..QT.I......RI------NIPS...I....K...ILLI.ST........YF.IR.G-..--------
A/Hong Kong/4801/2014(H3N2):                                  ......H-LN-Y----T..AL.V.MP.+EQFDK........G..KD.IF..AQSS..I.-...R..QA.I......RI------DIPS..I...K...ILLI.ST........YF.IR.G-..--------
A/Tasmania/503/2020(IVR-221)(H3N2):                           ......H-LN-Y----K..AL.V.MP.KEQFDK........Y.....KD.IS.FAQPS..I.-...R..QA.I......RI------.IPS...I....K...ILLI.ST........YF.IR.G-..--------
A/mallard/Wisconsin/10OS3941/2010(H14N6):                     ......R-VKNG-----N.GPI.V..E.T.S.V...L.....S....D...VAT......-.RSD.I.II......R.------N....T...L.N...S.I..S...........I.KST.G--------
A/Taiwan/01/2017(H7N9):                                       EMK..LSNT.NA----.F.QMTKSYK.TRESPAIIV..I..SVSTA...K..GSGNKL..-.GSSNY.Q.F..SP.A..Q.------N.....D.H.L.IN.N.TVT.SFN.AF..DRASF.R---------GKS
A/chicken/Germany/n/1949(H10N7):                              E.K..VSKTK.Q----NF.QTTN.YR.TDTAEH.I...I....STQ.KND..QTQSLSIS-.ESS.Y.NNF..VV.A..Q.------N....D.H..L.Q...N.T.SHN.G...SRVS..T---------GRG
A/wedge-tailed shearwater/Western Australia/2576/1979(H15N9): EMK..SSSMNNQ----VF.QL.Q.YR.TRKEPA.IV--.S.SLD..NK..GTGNKLI-..GSSKY.Q.FS.SP.A..K.------N..A..D.H.MLLD...TVT.TFN.AF..DRATF.R.NAP.GIEYNGKS
A/USSR/90/1977(H1N1):                                         N.L....E-K-NG----S..NLSKSYV..KEKEV.VL.......NIED.KTI.RKENAY.S-.VSSNYNRRFT.E.AE..K.------..A...NY...LL....T.I.EAN.....WHAFA..RG--------FGS
A/Chile/1/1983(H1N1):                                         N.L....E-K-NG----S..NLSKSYV..KEKEV.VL.......NIED.KTI.RKENAY.S-.VSSHYNRRFT.E.AK..K.------N.E...NY...LL....T.I.EAN.....WYAFA.SRG--------FGS
A/Texas/36/1991(H1N1):                                        N.L....E-K-NG----L..NLSKSYV..KEKEV.VL.......NIRD.RAI.HTENAY.S-.VSSHYSRRFT.E.AK..K.------..E...NY...LL....T.I.EAN.....WYAFA.SRG--------FGS
A/Solomon Islands/03/2006(H1N1):                              N.L...G-K-NG----L..NLSKSYA..KEKEV.VL.......NIGD.RA..HTENAY.S-.VSSHYSRRFT.E.AK..K.------N.L...K...T.EAT...VV..YAFA.SNG-------AGS
A/California/04/2009(H1N1):                                    N.I..V.-K-.----S..KLSKSY..DKGEKV.VL..I.....SAD.QS..Q.ADTY.F-.GSSRYSKKFK.E.AI..K.------N.K...K.T.EAT...VV..YAFAMERN--------AGS
A/Victoria/2570/2019(IVR-215)(H1N1):                          N.I..V-K-.S----.KI.Q.V..DKGKEV.VL..I...P.IAD.QS..Q.EDAY.F-.G.SRYSKKFK.E.AT..K.------.DRE..MNY...L.....K.T.EAT...V..YAFTMERD--------AGS
A/Japan/305/1957(H2N2):                                       NMV....-K-.S----N..VAKGSYN.TSGEQM.I......NDE...RT..Q.VGTY.S-.G.S.LNKRST.E.AT..K.------N.LG..ME.S..LLDMW.T.N.EST....EYGF.ISKR--------GSS
A/Viet Nam/1203/2004(H5N1):                                   NVV...I.-K-NS----T..TIKRSYN.TNQEDL.VL..I...NDAA...K..Q.PTTYIS-.G.S.LNQRL..R.AT.SK.------N....ME.F...LK.N.A.N.ESN..F...EYA..IVKK--------GDS
A/Taiwan/2/2013(H6N1):                                        N.V.IV.-T.SA----T..VIKG.YN.T.TQPI..F.....LDT.V.D...GSGDKY.R-MG.ESMNFAKS.E.AA..A.------N...RS..DY..SVLR..ETLNVESN.....WYA..FV.T--------NKKG
A/northern shoveler/California/HKWF1204/2007(H8N4):           SI.....-KKPD-----T.DFNEGSYV..E.GDIIFL..I...PDTK...T....ANTLSS-.T.N.INR.FQ....P..L.------...Q..MDY..G.LKR.ETLKIR.N.....EFG.L.KGE--------SHG
A/swine/Hong Kong/9/98(H9N2):                                 SMR....Q-K-N----S..F..AHYT..ERENI.FM..I....P....D...KADTTTS-.T.EDINRTFK.V..P..L.------N..Q...DY..SVLK..QTLRIGSN.....WYGHV.TGE--------SHG
A/turkey/Wisconsin/1/1966(H9N2):                              SMR....L-K-SG----QF.V.TDEYK.TR.SDIVFT.AI...P.SD..VK....PDTLSS-.T.DEINR.FK..P..L.------..Q..MDY..AVLK..QTVKIQ.N.....EPGHLITGK--------GNG
A/duck/Alberta/60/1976(H12N5):                                NMV..IH-Q-SG----T..VIRR.FN.TKGRDV.MV......A.LK.HQD...KDNSY.A-.GSESYNRRFT.E.ST..K.------N..A..MT.....K.EEA.T.ESN.AFL...YAFE.V.L--------GNG
A/duck/Memphis/546/1974(H11N9):                               N.V.FVN-R-.----N..VIRGAYN.TTGRDV.V..I..VSTD.VRQ..AKDNPYTL-..RSWSRRYNLET.V..GY------N..RSWMKI..YLLH..ES.S.ESN.G.L..KYG.IIEEY--------GKG
A/duck/Hokkaido/W345/2012(H13N2):                             N.I..VN-K-L.----K..VVKGEYN.TTGRDV.VL..I...D.EATANK...V.KNPYTL-...EWSRRYELE..T.IG------D..RSWMKI..HLMH..ER.T.ESS.G.L...YG.IIEKY--------GTG
A/black-headed gull/Sweden/2/99(H16N3):                       N.I..VN-K.L.----K..VVKGEYN.TTGRDV.VL..I...D.EATANK...V.KNPYTL-...EWSRRYELE..T.IG------D..RSWMKI..HLMH..ER.T.ESS.G.L...YG.IIEKY--------GTG
B/Brisbane/60/2008:                                           TMA.AVPKN.K.KTATNPLTIEVPY.CTEGEDQITV..F.SDD-E.GMKL.PKQFTS.ANGVT.HY.SQ..GF.NQTEDGGLP....VVDYMVQKS.KTGTITYQRGILL.QKVWCAS----------GRS


                                                                 266                              317
A/American black duck/New Brunswick/00464/2010(H4N6):         TILN-TAVPI-GSCVSKCH-TDRGSITTTKPF-QNISRISIGDCPKYVKQGSLKLATG
A/Aichi/02/1968(X-31)(H3N2):                                  S.MR-SDA..-DT.I.E.I-.PN...PND...-..VNK.TY.A........NT......
A/Philippines/2/1982(H3N2):                                   S.MR-SDA..-.T.S.E.I-.PN...PND...-..VNK.TY.A.R....NT......
A/Sydney/5/97(H3N2):                                          S.MR-SDA..-.K.N.E.I-.PN...PND...-..VN..TY.A..R....NT......
A/Wisconsin/67/2005(H3N2):                                    S.MR-SDA..-.K.E.I-.PN...PND...-..VN..TY.A..R....NT......
A/Hong Kong/4801/2014(H3N2):                                  S.MR-SDA..-.K.K.E.I-.PN...PND...-..VN..TY.A..R....HST......
A/Tasmania/503/2020(IVR-221)(H3N2):                           S.MR-SDA..-.K.K.E.I-.PN...PND...-..VN..TY.A..R....ST......
A/mallard/Wisconsin/10OS3941/2010(H14N6):                     .V.K-SDKK.-..T.P.L-..QSD...-.V...A..N........M....
A/Taiwan/01/2017(H7N9):                                       MGIQ-SG.QVDAN.EGD.Y-HSG.T.ISNL..-...DSRTV.K..R....R..L.....
A/chicken/Germany/n/1949(H10N7):                              LGIQ-SEAL.DN..E...F-WRG....N.KL..-..L.PRTV.Q......N.R..L....
A/wedge-tailed shearwater/Western Australia/2576/1979(H15N9): LGIQ-SDAQ.DE..EGE.F-YSG.T.NSPL..-...DSRAV.K..R....S..P..L.
A/USSR/90/1977(H1N1):                                         G.IT-SNASM-DE.DT..Q-.PQ.A.NSSL..-..HPVT...E.....RSTK.RMV..
A/Chile/1/1983(H1N1):                                         G.IT-SNASM-DE.DA..Q-.PQ.A.NSSL..-..VHPVT...E.....RSTK.RMV..
A/Texas/36/1991(H1N1):                                        G.IT-SNASM-DE.DA..Q-.PQ.A.NSSL..-..VHPVT...E.....RSTK.RMV..
A/Solomon Islands/03/2006(H1N1):                              G.I.-SNA.M-DE.DA..Q-.PQ.A.NSSL..-..VHPVT...E.....RSAK.RMV..
A/California/04/2009(H1N1):                                    G.II-SDT.V-HD.NTT.Q-.PK.A.N.SL..-..HP.T..K....STK.R.....
A/Victoria/2570/2019(IVR-215)(H1N1):                          G.II-SDT.V-HD.NTT.Q-.PE.A.N.SL..-..VHP.T..E.....STK.R.....
A/Japan/305/1957(H2N2):                                       G.MK-.EGTL-EN.ET..Q-.PL.A.N..L..-H.VHPLT..E.....SEK.V.....
A/Viet Nam/1203/2004(H5N1):                                   ..MK-SELEY-.N.NT..Q-.PM.A.NSSM..-H..HPLT..E.....SNR.V....
A/Taiwan/2/2013(H6N1):                                        AVFK-SDL..-EN.DAT.Q-.IT.VLR.N.T-..V.PLW..E.....SE..R.....
A/northern shoveler/California/HKWF1204/2007(H8N4):           R.IQNEDI..-.N.HT..Q-.YA.A.NSS...-..A..HYM.E.....KA..R..V.
A/swine/Hong Kong/9/98(H9N2):                                 R..K-.DLNS-.N..VQ.Q-.E..GLN..L..-H.V.KYAF.N.....GVK......V.
A/turkey/Wisconsin/1/1966(H9N2):                              R..K-.DLNN-.N..VQ.Q-.EK.GLN..L..-H...KYAF.N.....GVK......V.
A/duck/Alberta/60/1976(H12N5):                                R..K-NNL.M-.Q..TE.Q-LNE.VMN.S...-..T.KHY..K.....IPS.......I.
A/duck/Memphis/546/1974(H11N9):                               KLFR-SDLN.-E..ST..Q-SEI.W.N.NRS.-HSVH.NT........NVK.......
A/duck/Hokkaido/W345/2012(H13N2):                             R.FQ-SRIR-.AK.NA..Q-.SV.G.N.N.T-...E.NAL......I.S.Q......
A/black-headed gull/Sweden/2/99(H16N3):                       R.FQ-SG.RL-AK.NT..Q-.PM.A.N.N.T-...E.NAL......I.S.Q......
B/Brisbane/60/2008:                                           KVIK-GSL.LI.EA--D.LHEKY.GLNKS..YYTGEHAKA..N..IW..T-P....N.
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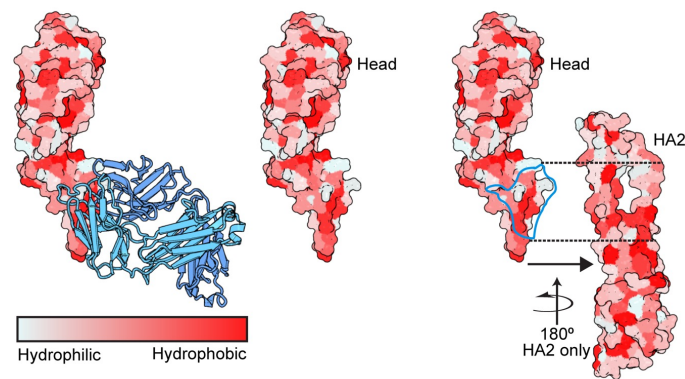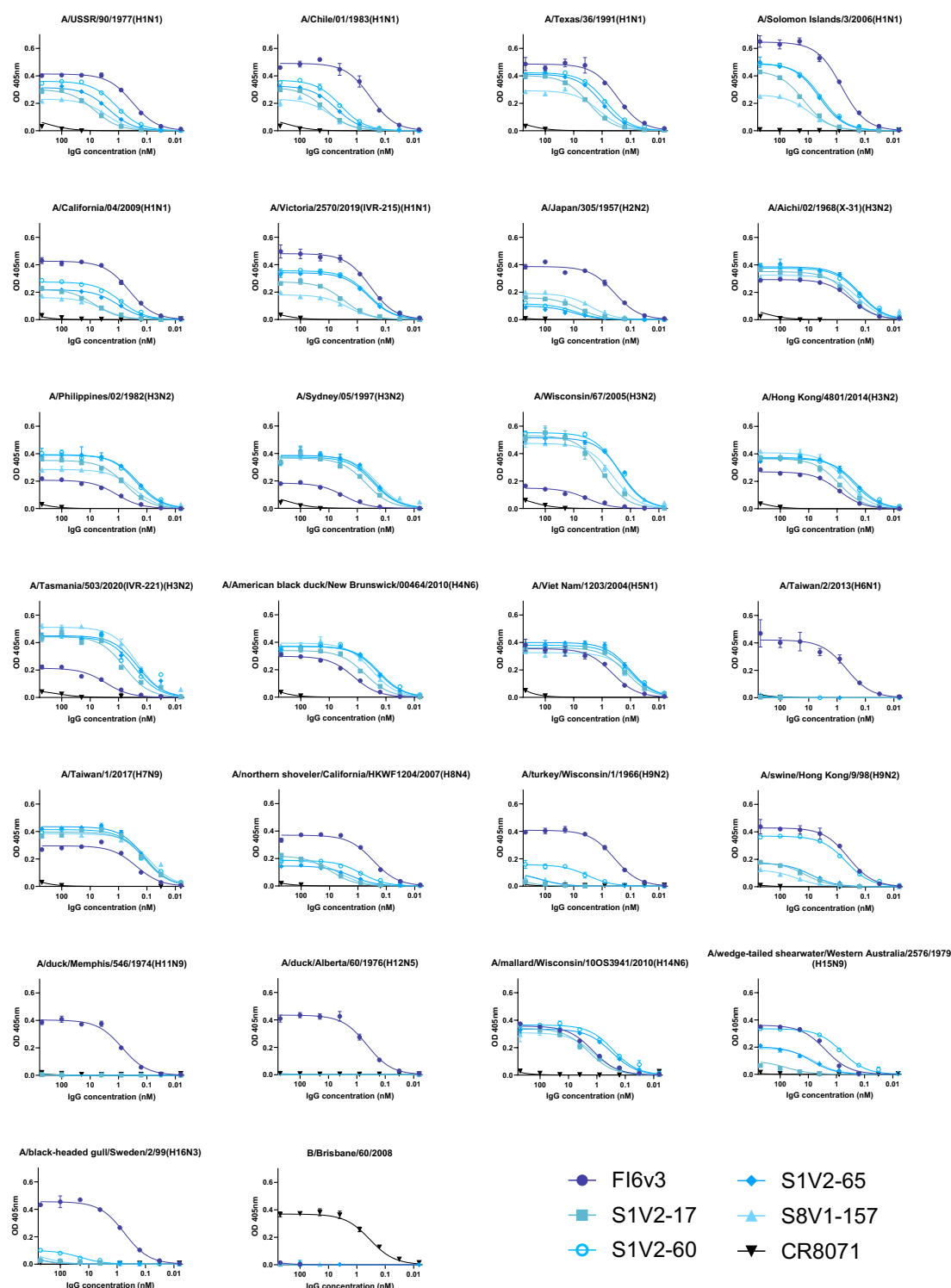
869
870
871 **Figure S1: S8V1-157 Contacts conserved sites.** An alignment of HA head regions for HAs used in
872 this study. S8V1-157 contacting residues in A/American black duck/New Brunswick/00464/2010(H4N6)
873 are colored red. HAs not bound by S8V1-157 are in denoted in gray. Sites identical to the A/American
874 black duck/New Brunswick/00464/2010(H4N6) sequence are indicated by a ".". Amino acid numbering
875 corresponds to standard H3N2 numbering.
876

877

878
879 **Figure S2: Surface hydrophobicity on the HA molecule.** Surface hydrophobicity and the head-stem
880 epitope are shown on the HA trimer of A/American black duck/New Brunswick/00464/2010(H4N6)
881 (PDB: 5XL2) [21]. Coloring is based on the PyMOL Color h script that utilizes a normalized consensus
882 hydrophobicity scale[62]. Views and orientations match those in Figure 2. The head-stem epitope is
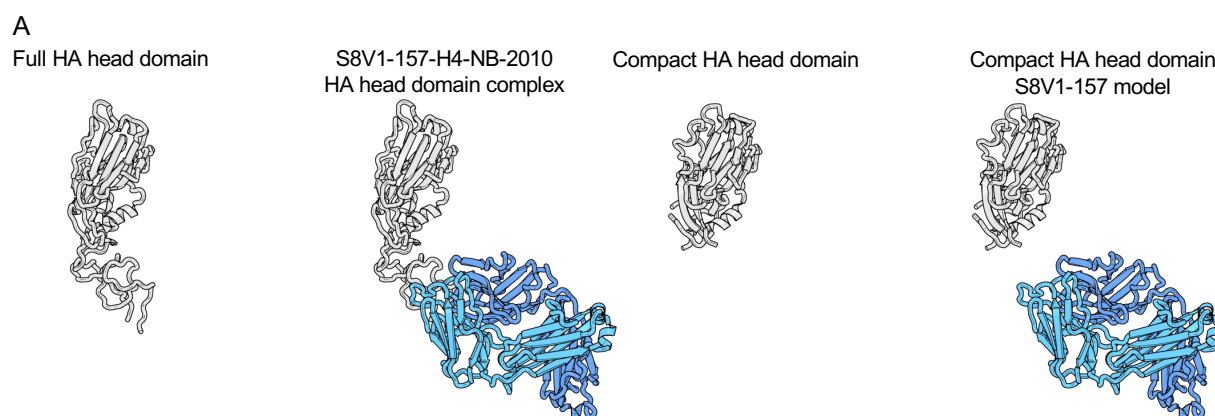883 circled in the far right panel.
884
885

886

887
888 **Figure S3: ELISA titrations of antibodies on HA coated plates.** The broadly binding stem antibody
889 FI6v3[47] was used as a positive control and an influenza B specific head antibody, CR8071[58], as a
890 negative control for influenza A isolates. Data points represent the average of three technical replicates.
891 The standard error of the mean is shown for each point. KDs were calculated from the curves fit to
892 these data points.
893
894

A

Full HA head domain   S8V1-157-H4-NB-2010   Compact HA head domain   Compact HA head domain
                       HA head domain complex   S8V1-157 model

B

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                                                  37                                                              114
A/Aichi/02/1968(X31)(H3N2)              :TNATELVQSSSTGKICN-NPHRILDGIDCTLIDALLGDPHCDVFQNET-WDLFVERSKAFSN-CYPYDVPDYASLRSLVASSG
A/California/7/2009(X181)(H1N1)(compact):---------------------AP.HLGK.NIAGWI..N.E.ESLSTASS.SYI..TPSSDNGT...G.FI..EE..EQLS.VS
A/California/7/2009(X181)(H1N1)         :.HSVN.LEDKHN..L.KLRGVAP.HLGK.NIAGWI..N.E.ESLSTASS.SYI..TPSSDNGT...G.FI..EE..EQLS.VS
A/Taiwan/2/2013(H6N1)                   :.HSV..LENQKEKRF.KIMNKAP..LK...IEGWI..N.K..LLLGDQS.SYI...PN.QNGI...GVLNELEE.KAFIG.GE
A/turkey/Wisconsin/1/1966(H9N2)         :.HTK...LHTEHN.ML.ATDLGHP.ILDT..IEGLIY.N.S..ILLGGKE.SYI....S.VNGM...GN.ENLEE....FS.AK
A/chicken/Germany/n/1949(H10N7)         :.....T.E.TNLN.L.-MKGRSYK.LGN.HPVGM.I.T.V..PHLTG.-..TLI..EN.IAH-...GATINEEA..QKIME..
A/duck/Memphis/546/1974(H11N9)          :.SSV...ENEY.SF.SIDGKAPISLG..SFAGWI..N.M..DLIGK.S.SYI..KPNPINGI...GTLENEEE..LKFSGVL
A/duck/Alberta/60/1976(H12N5)           :.QVE...HGGIDPIL.GTELGSP.VLD..S.EGLI..N.K..LYL.GRE.SYI...P.EMEGV...GSIENQEE....FS.IK
A/duck/Hokkaido/W345/2012(H13N2)        :.SSVD..ETNH..TY.ALGGISPVHLG..SFEGWIV.N.A.ASNLGIRE.SYLI.DPS.PHGL...GELDNNGE..H.FSGIR
A/black-headed gull/Sweden/2/99(H16N3)  :.SSID..ETNH..TY.SL.GVSPIHLG..SFEGWIV.N.S.ASNI.IRE.SYLI.DPN.PHKL.F.GE.DNNGE..H.FSGVN

                                                  117                                                             190
A/Aichi/02/1968(X31)(H3N2)              :T---LEFITEGFTWTGVTQN-GGSNACKRGPG-SGFFSRLNWLTKS--GSTYPVLNVTMPNNDNFDKLYIWGIHHPSTNQEQT
A/California/7/2009(X181)(H1N1)(compact):SFERF.IFPKTSS.PNHDSDK.VTA..PHAGA-KS.YKN.I..V.K--.NS..K.SKSYI.DKGKEV.VL........SAD.Q
A/California/7/2009(X181)(H1N1)         :SFERF.IFPKTSS.PNHDSDK.VTA..PHAGA-KS.YKN.I..V.K--.NS..K.SKSYI.DKGKEV.VL........SAD.Q
A/Taiwan/2/2013(H6N1)                   :RVERF.MFP-KS..A..DTSR.VT...PSYTID.S.YRN.V.IV.TD-SA....IKG.YN.TGTQPI..F..V...LDTTV.D
A/turkey/Wisconsin/1/1966(H9N2)         :SYKRIQIFP-DK..NVTYS--..T.R..S-----NS.YRSMR...HKS-N-S..FQ.AHYT..ERENI.FM......P.DT...
A/chicken/Germany/n/1949(H10N7)         :GISKMST---..YGSSINSA.TTK..M.NG.-DS.YAE.K..VSKTK.QNF.QTTN.YR.T.TAEH.I.....ST..KN
A/duck/Memphis/546/1974(H11N9)          :EFNKF.AF.-SNG.GS.NSGA.VTA...F.SS-NS..RNMV..IHQS-.-....IRR.FN.TKGR.V.MV..V...A.LK.HQ
A/duck/Alberta/60/1976(H12N5)           :KYERVKMFD-FTK.NVTYT--.T.K..NNTSNQGS.YRSMR...LKS-.-QF..QTDEYK.TRDS.IVFT.A....P.SD..V
A/duck/Hokkaido/W345/2012(H13N2)        :SFSRT.L.A-PTS.GA.ND--..V.A..TDRGA-.S.YRN.V.FVNRG-N-N...IRGAYN.TTGR.V.V......VSTD.VR
A/black-headed gull/Sweden/2/99(H16N3)  :SFSRT.L.P-PSK.GDILE--..TTAS.QNRGA-NS.YRN.I..VNK--LNK...VKGEYN.TTGR.V.VL......D.EATAN

                                                  193                                                             271
A/Aichi/02/1968(X31)(H3N2)              :SLYVQASGRVTVSTRRSQQTIIPNIGSRPWVRGLSSRISIYWTIVKPGDVLVINSNGNLIAPRGYFKM--RTGKSSIMRSDAP
A/California/7/2009(X181)(H1N1)(compact):...QN.DAY.F.GSS.YSK.FK.E.AI..K..DREG.MNY...L.E...KITFEAT...VV..YA.A.ERNA.S---------
A/California/7/2009(X181)(H1N1)         :...QN.DAY.F.GSS.YSK.FK.E.AI..K..DREG.MNY...L.E...KITFEAT...VV..YA.A.ERNA.SG-.II..T.
A/Taiwan/2/2013(H6N1)                   :.N..GSGDKY.RMG.ESMNFAKS.E.AA..A.N.QR...DY..SVLR..ET.NVE.......WYAY.FVSTNK.GAVFK..L.
A/turkey/Wisconsin/1/1966(H9N2)         :D..KK.DTTTS.T.EDINR.FK.V..P.N.QQG..DY..SVL...QT.R.G.......WYGHVLTGESH-GR.LKT.LN
A/chicken/Germany/n/1949(H10N7)         :D..GTQ.LSIS.ESSTY.NNFV.VV.A..Q.N.Q.G..DFH..L.Q...NITFSH..G....SRVS.LTG.GLG--.QSEALI
A/duck/Memphis/546/1974(H11N9)          :D..KKDNSY.A.GSESYNRRFT.E.ST..K.N.QAG.MTF.......EEAITFE...AFL....YA.ELVSLGN-GKLF..LN
A/duck/Alberta/60/1976(H12N5)           :K..KNPDTLSS.T.DEINRSFK....P..L...QQG.MDY..AVL...QTVK.QT......EYGHLITGKSH-GR.LKNNL.
A/duck/Hokkaido/W345/2012(H13N2)        :Q..AKDNPYTL....SWSRKYNLET.V..GYN.QK.WMK...YLLH..ESISFE...G.L..KYGYIIEEYGK-GR.FQ.RIR
A/black-headed gull/Sweden/2/99(H16N3)  :K...NKNPYTL....KEWSRRYELE..T.IG-D.QR.WMK...HLMH..ERITFE.S.G.L...YGYIIEKYGT-GR.FQ.GVR

                                                  274                                     317
A/Aichi/02/1968(X31)(H3N2)              :IDTCISECITPNGSIPNDKPFQNVNKITYGACPKYVKQNTLKLATG
A/California/7/2009(X181)(H1N1)(compact):----------------------------------------------
A/California/7/2009(X181)(H1N1)         :VHD.NTT.Q..K.A.NTSL....IHP..I.K.....STK.R....
A/Taiwan/2/2013(H6N1)                   :.EN.DAT.Q.IT.VLRTN.T....SPLWI.E......SES.R....
A/turkey/Wisconsin/1/1966(H9N2)         :NGN.VVQ.Q.EK.GLNTTL..H.IS.YAF.N.....GVKS....V.
A/chicken/Germany/n/1949(H10N7)         :DNS.E.K.FWRG...NTKL....LSPR.V.Q.....N.RS.L....
A/duck/Memphis/546/1974(H11N9)          :.ES.STK.QSEI.W.NTNRS.HS.HRN.I.D.....NVKS......
A/duck/Alberta/60/1976(H12N5).          :MGQ.VT..QLNE.VMNTS.....TS.HYI.K....IPSGS....I.
A/duck/Hokkaido/W345/2012(H13N2)        :.AK.NAK.Q.SV.G.NTN.T...IERNAL.D....I.SGQ......
A/black-headed gull/Sweden/2/99(H16N3)  :LAK.NTK.Q.SM.G.NTN.T...IERNAL.D....I.SGQ......
```

C

			S1V2-17	S1V2-60	S1V2-65	S8V1-157	CR8071	FluA20
Compact HA head domain	H1N1	A/California/7/2009(X181)	>100	>100	>100	>100	>100	14
Full HA head domain	H1N1	A/California/7/2009(X181)	0.24	0.15	0.11	<0.1	>100	0.18
	H3N2	A/Aichi/02/1968(X31)	1.4	1.4	1.1	0.35	>100	0.57
	H6N1	A/Taiwan/2/2013	>100	>100	>100	>100	>100	0.60
	H9N2	A/turkey/Wisconsin/1/1966	>100	>100	>100	>100	>100	0.18
	H10N7	A/chicken/Germany/n/1949	>100	>100	>100	>100	>100	0.24
	H11N9	A/duck/Memphis/546/1974	>100	>100	>100	>100	>100	0.13
	H12N5	A/duck/Alberta/60/1976	>100	>100	>100	>100	>100	0.18
	H13N2	A/duck/Hokkaido/W345/2012	>100	>100	>100	>100	>100	62
	H16N3	A/black-headed gull/Sweden/2/99	80	4.3	76	28	>100	>100

| Kd in nM				
<1	1-5	5-25	25-100	>100

Figure S4: Head-stem epitope antibody binding to HA head-only constructs. A. Structures of HA head used in these experiments the S8V1-157-HA head complex is shown for reference. A compact head[49], is shown for reference (PDB 7TRH). B. HA heads for HAs not bound or not expressed in Figure 3 were produced alongside a positive control A/California/07/2009(H1N1)(X-181) and a compact head version. Sequences of the HA head regions are aligned to the A/Aichi/02/1968(H3N2)(X-31)

902 reference sequence and numbered by H3 convention. C. Dissociation constants from ELISA
903 measurements of head-stem epitope antibodies to HA heads. Head interface antibody FluA-20[40] was
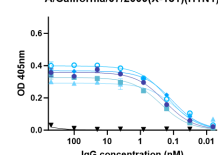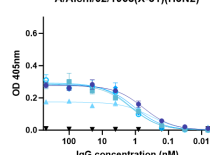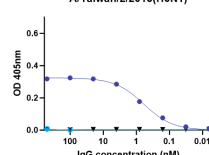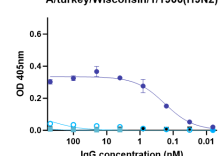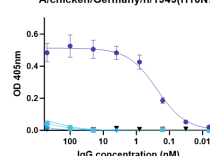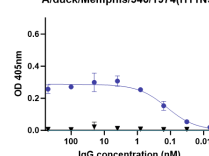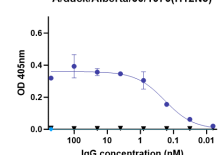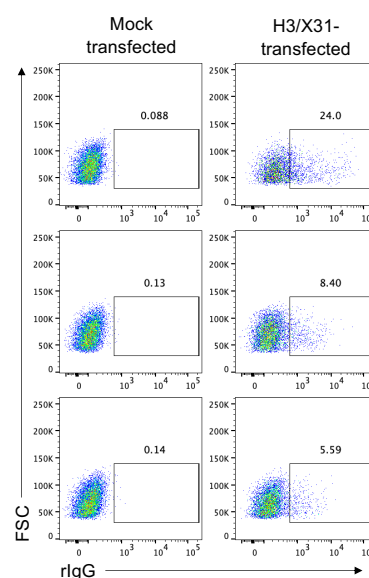904 used as a positive control and influenza B head antibody CR8071[58] as a negative control.
905
906
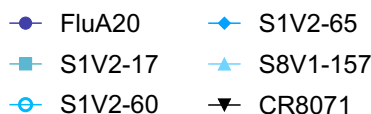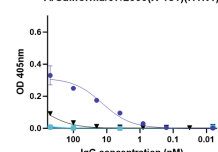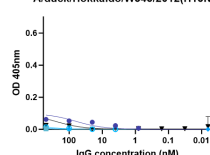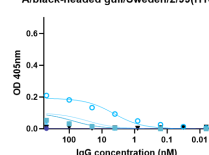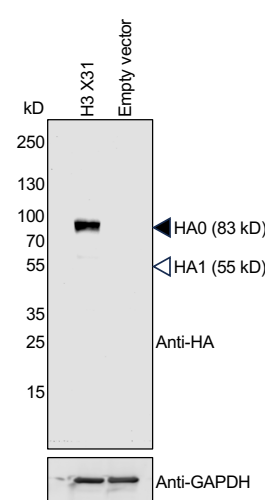
Figure S5: ELISA titrations of antibodies on HA head coated plates and characterization of cell expressed HA. A. Head interface antibody FluA-20[40] was used as a positive control and influenza B head antibody CR8071[58] as a negative control. Data points represent the average of three technical replicates. The standard error of the mean is shown for each point. KDs were calculated from the curves fit to these data points. B. Lysates from 293F transfected with either HA from A/Aichi/02/1968(H3N2)(X31) or empty vector were subjected to western blotting under reducing and denaturing conditions with either anti-HA tag antibody, which recognizes an endogenous sequence in the H3 HA head (HA1), or anti-GAPDH antibody. Molecular weights of unprocessed HA0 and processed HA1 are indicated with open or closed arrowheads, respectively. C. 293F transfected with either HA from A/Aichi/02/1968(H3N2)(X31) or empty vector were stained with the indicated antibodies

918　and analyzed by flow cytometry. Gates denoting cells bound by antibody are shown, with the percent of
919　the total population indicated above.
920
921

	A/Aichi/02/1968(H3N2)(X-31)		
Antibody	HC19	S8V1-157	CR3022
IC50 µg/mL	<3.5	>3535.5	>3535.5

922
923 **Figure S6: S8V1-157 is a non-neutralizing antibody.** Neutralization IC50 values for S8V1-157,
924 neutralizing antibody HC19[32] (positive control) and SARS-CoV antibody CR3022[33] (negative control).
925
926

Antibody	Vaccination	IGHV	% VH SHM	IGHJ	IGHJ	HCDR3	Length	IGVL	% VL SHM	JL	LCDR3	Length
S1V2-17	Post	IGHV1-24*01	6.94	IGHJ4*02	IGHD2-21*02	C VTWLVGTTVAAY W	12	IGKV2-28*01ᵃ	4.21	IGKJ1*01	C MQALQAPRT F	9
S1V2-60	Post	IGHV5-51*01	9.72	IGHJ4*02	IGHD5-12*01	C VRLGYGDSPPFDS W	13	IGKV2-28*01ᵃ	4.21	IGKJ2*01	C MQALQDPYT F	9
S1V2-65	Post	IGHV5-51*01	7.99	IGHJ4*02	IGHD5-12*01	C ARLGYGDSPPFDF W	13	IGKV2-28*01ᵃ	6.67	IGKJ2*01	C MQALETPYT F	9
S8V1-157	Pre	IGHV1-24*01	5.90	IGHJ4*02	IGHD1-26*01	C ATLMGANPFDY W	11	IGKV2-28*01ᵃ	2.46	IGKJ2*01	C KQALQTLYT F	9

Antibody	Vaccination	IGHV	% VH SHM	IGHJ	IGHJ	HCDR3	Length	IGVL	% VL SHM	JL	LCDR3	Length
S8V5_P5-C6	Pre	IGHV4-61*08	6.32	IGHJ6*02	IGHD3-10*01	C ARVSMVPGEGWFFGVDV W	17	IGKV3-20*01	4.61	IGKJ4*01	C QHYGGSLT F	8
S8V6_P7-H9	Post	IGHV1-8*01	2.43	IGHJ4*02	IGHD6-13*01	C VRGDXAAAXY W	10	IGKV4-1*01	0.34	IGKJ2*01	C QQYYSTPMYT F	10
S8V6_P10-G1	Post	unrecoverable	-	-	-	–	-	unrecoverable	-	-	–	-
S1V4_P4-C5	Post	IGHV1-24*01	6.94	IGHJ6*03	IGHD4-23*01	C AITLSYYYMNV W	11	IGKV2-28*01ᵃ	5.10	IGKJ1*01	C MQALQTPPWT F	10
S5V4_P1-A2	Post	IGHV1-18*01	6.25	IGHJ3*02	IGHD3-3*01	C ARSKFGLVGRDVFDI W	15	IGKV4-1*01	1.01	IGKJ4*01	C QQYYDTPPS F	9
S5V4_P2-D2	Post	IGHV1-18*01	10.76	IGHJ3*02	IGHD4-23*01	C ARSKLGLVGRDVFDI W	15	IGKV4-1*01	2.02	IGKJ4*01	C QQYYDTPPS F	9
S12V4_P1-F1	Post	IGHV4-31*04	9.28	IGHJ6*02	IGHD1-7*01	C ARETGIISGSNFFYYGMDV W	19	IGKV3-15*01	2.15	IGKJ1*01	C QQYNNWPLRT F	10
S12V6_P7-H4	Post	IGHV1-24*01	6.25	IGHJ4*02	IGHD3-3*02	C AINLGGVPTTIL W	12	IGKV2-28*01ᵃ	4.42	IGKJ5*01	C MQALQTPIT F	9

ᵃ or IGKV2D-28*01

927

Figure S7: Antibody genetics of S8V1-157 competing antibodies. Antibody names, gene usage and HCDR3 sequences for S8V1-157 competing antibodies.