

## Review Article

# Canonical Transient Receptor Potential (TRPC) Channels in Nociception and Pathological Pain

Zhi-Chuan Sun,<sup>1,2</sup> Sui-Bin Ma,<sup>2</sup> Wen-Guang Chu,<sup>2</sup> Dong Jia,<sup>1</sup> and Ceng Luo<sup>2</sup> 

<sup>1</sup>Department of Neurosurgery, Tangdu Hospital, Fourth Military Medical University, Xi'an 710038, China

<sup>2</sup>Department of Neurobiology, School of Basic Medicine, Fourth Military Medical University, Xi'an 710032, China

Correspondence should be addressed to Ceng Luo; [luoceng@fmmu.edu.cn](mailto:luoceng@fmmu.edu.cn)

Received 27 September 2019; Revised 26 February 2020; Accepted 7 March 2020; Published 21 March 2020

Academic Editor: Gernot Riedel

Copyright © 2020 Zhi-Chuan Sun et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Chronic pathological pain is one of the most intractable clinical problems faced by clinicians and can be devastating for patients. Despite much progress we have made in understanding chronic pain in the last decades, its underlying mechanisms remain elusive. It is assumed that abnormal increase of calcium levels in the cells is a key determinant in the transition from acute to chronic pain. Exploring molecular players mediating  $\text{Ca}^{2+}$  entry into cells and molecular mechanisms underlying activity-dependent changes in  $\text{Ca}^{2+}$  signaling in the somatosensory pain pathway is therefore helpful towards understanding the development of chronic, pathological pain. Canonical transient receptor potential (TRPC) channels form a subfamily of nonselective cation channels, which permit the permeability of  $\text{Ca}^{2+}$  and  $\text{Na}^+$  into the cells. Initiation of  $\text{Ca}^{2+}$  entry pathways by these channels triggers the development of many physiological and pathological functions. In this review, we will focus on the functional implication of TRPC channels in nociception with the elucidation of their role in the detection of external stimuli and nociceptive hypersensitivity.

## 1. Introduction

Chronic pathological pain represents a major challenge to clinical practice and basic science. Activity-dependent neural plasticity is assumed to be a prime mechanism underlying various physiological and pathological processes including clinical transitions from acute, physiological pain to chronic, pathological pain [1, 2]. Accumulating evidence has revealed that the second messenger  $\text{Ca}^{2+}$  and  $\text{Ca}^{2+}$ -dependent pathways play a crucial role in the neural plasticity, i.e., peripheral and central sensitization associated with pathological pain. Mobilization of intracellular  $\text{Ca}^{2+}$  upon neuronal activation is the main trigger for activation of a variety of signaling mediators, such as CamKII- $\alpha$ , Protein Kinase A, and extracellular receptor-activated kinases (ERK1/2); these, in turn, regulate the expression and functions of downstream proteins determining the excitability of neurons, which are involved in pain processing [1, 2]. Exploring molecular players mediating  $\text{Ca}^{2+}$  entry into cells and molecular mechanisms underlying activity-dependent changes in  $\text{Ca}^{2+}$  signaling in the somatosensory pain pathway is therefore

helpful towards understanding the development of chronic, pathological pain.

The discovery of transient receptor potential (TRP) channels during the last 5 decades has increased impressively our knowledge of the molecular players mediating  $\text{Ca}^{2+}$  mobilization in the cells. TRP proteins comprise nonselective cation channels that permit the permeability of  $\text{Ca}^{2+}$  and  $\text{Na}^+$  into the cells [3]. TRP channels contribute to changes in cytosolic-free  $\text{Ca}^{2+}$  concentration either by inducing  $\text{Ca}^{2+}$  influx across plasma membrane or by driving  $\text{Ca}^{2+}$  release from several organelles. Given the unique importance of  $\text{Ca}^{2+}$  and  $\text{Ca}^{2+}$ -dependent signaling in the cells, it is not surprising that TRP channels and its dysfunctions are closely linked with many physiological and pathological processes, including pain and sensitization. On the basis of amino acid homology, TRP superfamily is divided into six subfamilies, TRP canonical or classical (TRPC), TRP vanilloid (TRPV), TRP melastatin (TRPM), TRP ankyrin (TRPA), TRP polycystin (TRPP), and TRP mucolipin (TRPML) [4–9]. Among which, TRPV1, TRPM8, and TRPA1 have been extensively investigated and considered to be molecular detectors for

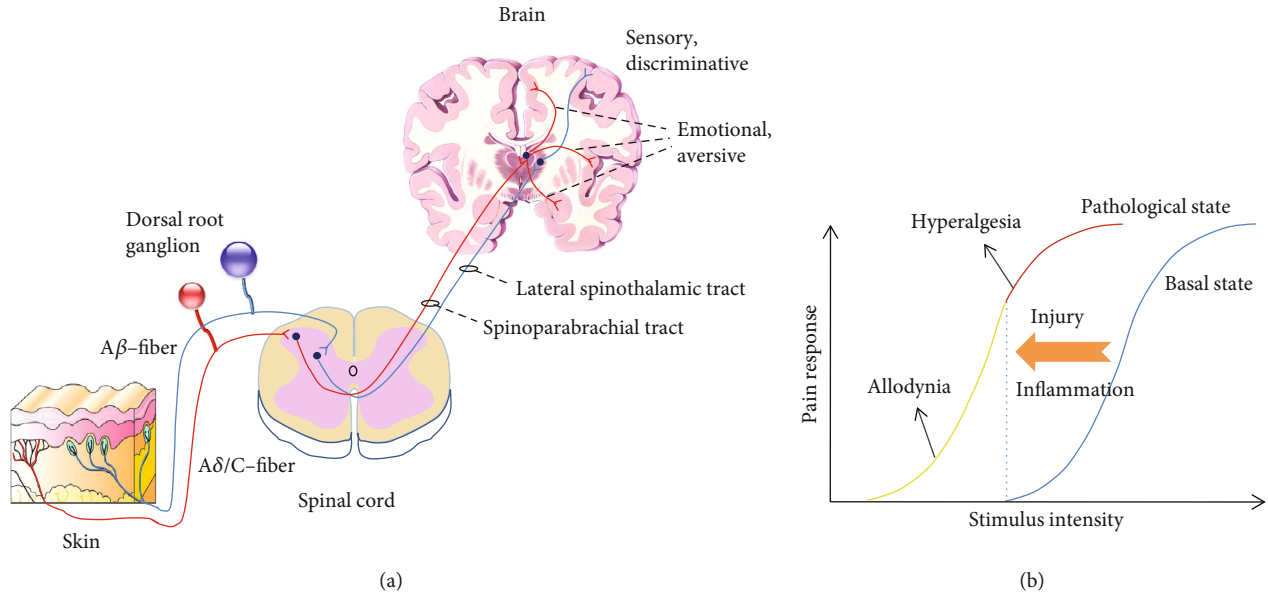


FIGURE 1: Nociceptive sensory detection and transmission in the ascending pain pathway. (a) Nociceptors detect external painful stimuli through a variety of receptors and transmit this noxious information from the periphery to the spinal cord and further central nervous system along axons, leading to sensory discrimination and affective motivation of pain. (b) Upon inflammation and injury, physiological pain converts to chronic, pathological pain, manifesting as spontaneous pain, hyperalgesia, and allodynia.

thermal and chemical stimuli that activate sensory neurons to produce acute or persistent pain [10–12]. Although TRPC subfamily was the first to be cloned among TRP genes, lacking of specific pharmacological tools targeting at TRPC subunits led to a much lagging in the exploration of a functional role of TRPC subfamily and its underlying mechanisms. With the establishment of specific TRPC subunit transgenic mouse models and discovery of selective pharmacological tools at TRPC subunits during the past few years, emerging evidence has accumulated that TRPC subfamily exert an important role in a variety of neuronal functions, including memory, motor coordination, fear, anxiety, Huntington's disease, neurite growth, and pain [13–26]. In this review, we will focus on the role of TRPC subfamily in nociception and the modulatory mechanisms of TRPC subfamily by inflammation or injury. Recent advances in the development of therapeutic strategies targeting against TRPC subfamily will also be reviewed.

## 2. Sensory Detection and Transmission in the Pain Pathway

The body detects various modalities of noxious stimuli through a specialized set of sensory nociceptive fibers innervating peripheral tissues: unmyelinated C fibers and thinly myelinated Aδ fibers, which are distinct from myelinated tactile sensors (Aβ fibers) and proprioceptors (Figure 1(a)). A variety of ion channels and receptors expressed on nociceptors, such as transient receptor potential ion channels (TRP channels), acid-sensing ion channels (ASIC channels), purinoreceptor, and serotonin receptors, transduce the physicochemical properties of noxious stimuli (e.g., heat, cold, pressure, and chemicals) into electrical activity—a membrane depolarization, which is further encoded as a train of

propagating action potentials by sodium channels. Nociceptive afferents carrying these peripheral signals terminate predominantly in the superficial laminae (I and II) of the spinal cord dorsal horn and form glutamatergic synapses onto second-order superficial spinal neurons, whereas nonnociceptive inputs terminate in deeper laminae (III–IV) (Figure 1(a)). The integrated nociceptive information in the superficial dorsal horn is further transmitted to projection neurons mostly located in lamina I and lamina V of the spinal dorsal horn, whose axons cross the midline and ascend to a variety of supraspinal targets. The spinal dorsal horn is therefore the site of the first synapse and processing center in the ascending pathway that conveys incoming pain information from the periphery to the central nervous system (CNS). Several pathways are demonstrated to carry the net output from spinal networks to distinct projection regions in the brain with one or several relay stations in their way so that pain is ultimately perceived in its multiple dimensions. For instance, the lateral spinothalamic tract projects multimodal sensory inputs from spinal wide-dynamic range (WDR) neurons to the lateral thalamus, the neurons from which in turn send sensory signals to the parietal somatosensory cortex. Important synapses are formed serially in the thalamus and somatosensory cortex along this ascending pathway, which has been implicated in processing sensory and discriminative aspects of pain. By contrast, the medial spinothalamic tract and the spinoparabrachial tract as well as the spinomesencephalic tract project nociceptive signals to the medial thalamus and limbic structures via monosynaptic or polysynaptic relay. From the medial thalamus, sensory inputs are further projected to widespread forebrain regions such as the anterior cingulate cortex (ACC), amygdala, and insular cortex, which are believed to mediate the emotional and aversive components of pain. The perceiving of pain in

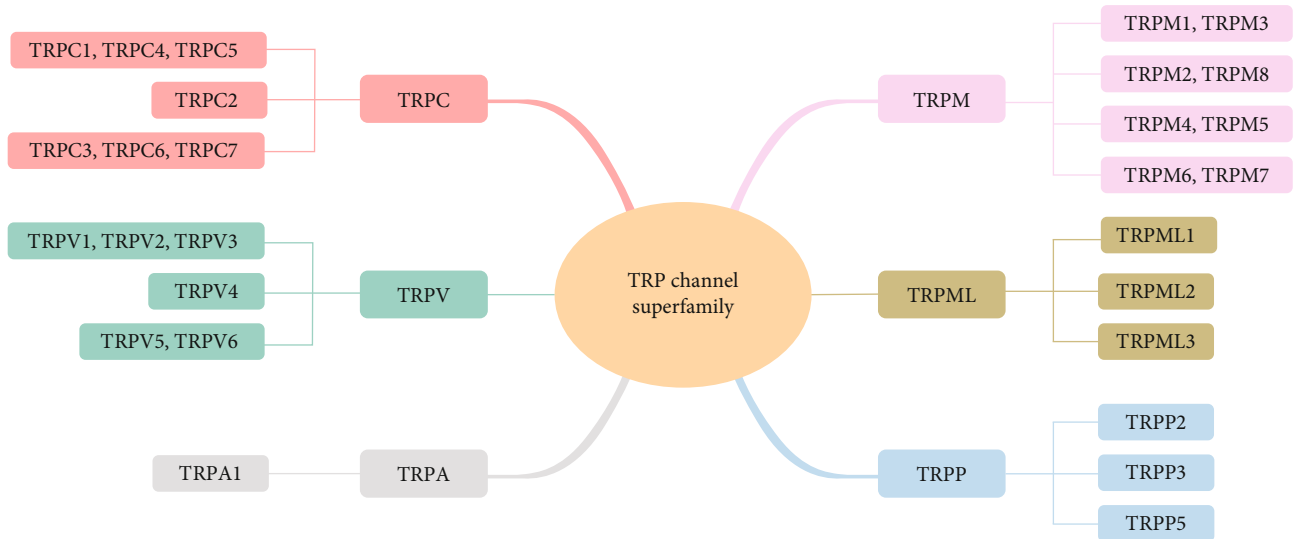


FIGURE 2: Composition of TRP channel superfamily and different TRP channel subfamily. TRP channel superfamily consists of six subfamilies, TRP canonical or classical (TRPC, 7 subunits), TRP vanilloid (TRPV, 6 subunits), TRP melastatin (TRPM, 8 subunits), TRP ankyrin (TRPA, 1 subunit), TRP polycystin (TRPP, 3 subunits), and TRP mucolipin (TRPML, 3 subunits). Based on amino acid sequence homology and functional similarities, TRPC subfamily members can be grouped into three major groups: TRPC1/4/5, TRPC2, and TRPC3/6/7.

the cortex accordingly initiates information to the spinal cord and enables withdrawal reflex from the noxious stimulus (Figure 1(a)).

### 3. Transition from Acute Pain to Chronic Pain

Acute, physiological pain serves as an alarming system in our body and is characterized by a high threshold and is typically transient. However, tissue/nerve injury or inflammation often causes plastic changes of primary sensory neurons and synaptic transmission in the central nervous system, shifting physiological pain to chronic, pathological pain, which is characterized by a lowered threshold and persistent pain hypersensitivity. This includes spontaneous pain (pain experience in the absence of any obvious stimulus), hyperalgesia (an increased sensitivity to noxious stimuli), and allodynia (pain in response to normally innocuous stimuli) (Figure 1(b)) [2]. In addition, chronic pain is often accompanied by aversive emotions, such as anxiety and depression [1, 27]. The incidence of chronic pain is estimated to be 20-25% worldwide. Few patients with chronic pain obtain complete relief from the drugs that are currently available, and more than half of them report inadequate relief. Therefore, to understand the underlying mechanism that contribute to the transition from acute to chronic pain is crucial to develop better drugs to manage chronic pain. Accumulating experimental and clinical evidence indicates that both peripheral and central sensitization contribute to the development of chronic pain [1, 28, 29]. Following tissue/nerve injury, release of inflammatory mediators, increased activity of transient receptor potential channels, and dysregulation of voltage-gated ion channels are some of the changes contributing to neuronal hyperexcitability in the peripheral primary sensory neurons [28, 29]. Subsequently, this peripheral hyperexcitability sends ongoing nociceptive signals to the spinal dorsal

horn and further to supraspinal brain regions, where alterations of ion channel expression, microglia and astrocyte activation, long-term potentiation of synaptic transmission, dendritic spine remodeling, and impaired descending pain modulation are among the typical changes in the central nervous system [1]. Substrates that contribute to peripheral and central sensitization are therefore potential targets for treatment of chronic pain.

### 4. TRPC Subfamily Expression in the Pain Pathway

**4.1. Structure and Composition of TRPC Subfamily.** TRP channel superfamily consists of six transmembrane domains, termed S1-S6, with cytoplasmic N- and C-terminal regions and the pore region formed by S5 and S6 segments [5, 30]. On the basis of amino acid homology, TRP superfamily is divided into six subfamilies, i.e., TRPC (TRPC1-7), TRPV (TRPV1-6), TRPM (TRPM1-8), TRPA (TRPA1), TRPP (TRPP2, TRPP3, and TRPP5), and TRPML (TRPML1-3) [4-9] (Figure 2). The first subfamily of TRP gene cloned in mammals was TRPC subfamily [31]. So far, seven members of the TRPC subfamily have been identified, i.e., TRPC1-TRPC7. Seven members of the TRPC subfamily are structurally related to *Drosophila* TRP but also to each other (>30% within the first 750-900 amino acids) and differ mainly within the carboxyterminal region. Based on amino acid sequence homology and functional similarities, TRPC subfamily members can be grouped into three major groups: TRPC1/4/5, TRPC2, and TRPC3/6/7 [4, 32-37] (Figure 2). TRPC2 is a pseudogene in humans, which is related but is clearly distinct from the other two major groups [31]. TRPC1, TRPC4, and TRPC5 is believed to cluster together (TRPC1/4/5) to form homo- or heteromeric channels. TRPC4 and TRPC5 are each able to form homomeric

channels with similar biophysical characteristics to each other [33, 34]. In contrast, TRPC1 does not or poorly form homomeric channels, yet it readily forms heteromers with TRPC4 and TRPC5, generating ion channels with distinct biophysical characteristics [30, 38]. TRPC3, TRPC6, and TRPC7 cluster on the phylogenetic tree and can form either homotetramers or heterotetramers with variable calcium ion permeability [33, 34, 39]. In aggregate, these previous studies suggest that heteromeric interactions mainly occur within members of two groups of TRPCs: TRPC1/TRPC4/TRPC5 and TRPC3/TRPC6/TRPC7. However, exceptions to these findings have also been reported. For example, exogenously and endogenously expressed TRPC1 and TRPC3 have been found to coassemble to form a heteromeric complex [40, 41].

#### 4.2. Dorsal Root Ganglion: The Forefront of Pain Pathway.

Dorsal root ganglion (DRG) is an aggregate of the somata of primary sensory neurons, which play an essential role in initiating somatosensation by detecting sensory stimuli in the periphery via their peripheral terminals and sending signals to the spinal cord via their central terminals [42–44]. DRG neurons are highly diverse in terms of cell sizes, gene expression, myelination levels, etc. Small-diameter neurons are assumed to be pain-sensing neurons, while medium- to large-diameter neurons preferentially detect low-threshold, nonpainful mechanical stimulation [45, 46]. Tissue inflammation or nerve injury can sensitize DRG neurons, causing exaggerated pain sensitivity, often leading to the development of chronic pain. TRP channels have been shown to be expressed in DRG neurons and to be particularly important for sensory physiology. These include roles of TRP channels for thermosensation, chemosensation, mechanosensation, tactile sensation, and hearing.

In the somatic sensory DRG neurons, TRPV1 is mainly expressed in small- to medium-diameter neurons and is sensitive to noxious heat, protons, and irritant vanilloids [10, 47, 48]. TRPV1 marks a population of unmyelinated, slowly conducting neurons (C-fibers) and account for 30–50% of all DRG neurons within rodent sensory ganglia [49, 50]. TRPM8 is activated by cold stimuli as well as a variety of natural and synthetic cooling agents, such as menthol [51, 52]. TRPM8 is mostly expressed in small-diameter, unmyelinated C-fibers as well as a minor cohort of lightly myelinated A $\delta$ -fibers, which constitutes 15% of all somatosensory neurons. TRPA1 serves as a detector for chemical irritants, such as isothiocyanates and thiosulfonates that constitute pungent agents from mustard and allium plants [53–56]. TRPA1 is expressed exclusively by peptidergic C-fibers [49, 53] and plays a key role in chemonociception.

**4.3. TRPC Protein Expression in the DRG.** The expression of TRPC channels was mapped in the sensory neurons of rodents throughout an embryonic and postnatal development period. Various subtypes of TRPC channels are expressed in rat and mouse primary sensory neurons [24, 57–63]. In adult mice and rat, mRNAs for TRPC1–TRPC7 were present in the DRG, with TRPC1, TRPC3, and TRPC6 being the most predominant, TRPC2, TRPC4, and TRPC5

at low levels, and TRPC7 at extremely low levels. In the embryonic stage, TRPC2 mRNAs were expressed at high levels at embryonic (E) day 12 and E14 but greatly reduced to a quite low level in adult. In contrast, TRPC1, TRPC3, and TRPC6 expression levels increased progressively from E12 to adult. TRPC4, C5, and C7 expressions increased from E12 to peak levels at E18. TRPC1 and C2 were expressed in the neurofilament-200- (NF-200-) positive large-diameter neurons, while TRPC3 mRNA expression was almost exclusively present in nonpeptidergic isolectin B4- (IB4-) positive small size neurons that were largely TRPV1-negative, which stained up to 35% of DRG neurons. A strong expression of TRPC3 in DRG was also reported by Kunert-Keil et al. [64]. In addition, TRPC5 has been found to be localized to small and medium diameter sensory neurons [65]. A single cell RNA sequencing study also determined a subset of DRG neurons express TRPC subunits [66, 67]. Interestingly, the majority of the TRPC3-positive neurons did not show TRPV1 immunoreactivity [57]. The expression and regulation of TRPC channel mRNAs in the NG were unexpectedly similar to the DRG [57]. In comparison with the well-reported expression in DRG, the profile of TRPC proteins in the spinal cord and supraspinal regions which are involved in the pain pathway has been rarely studied. In addition, in human tissue, a recent study reported differential regulation of TRPC channel gene and protein expression by intervertebral disc (IVD) degeneration and back pain in human [68]. TRPC1, C3, C4, and C6 was observed to be expressed at mRNA and protein levels in the intervertebral disc. In the degenerated IVD, TRPC6 displayed enhanced gene and protein expressions as compared to nondegenerated IVDs, while TRPC1, C3, and C4 did not display difference between degenerated and nondegenerated IVD. This study has for the first time linked TRPC6 to the degeneration of IVD and development of low back pain in human. The thorough and detailed investigation of TRPC expression in the human pain pathway remains to be further conducted.

#### 4.4. Sensitivity of TRPC Proteins to Mechanical Stimuli.

Emerging evidence shows that TRPC proteins confer sensitivity to mechanical stimulation in the vertebrate cells. TRPC3 and TRPC6 have been shown to be essential for normal mechanotransduction in subsets of sensory neurons and cochlear hair cells [61]. TRPC3/TRPC6 double knockout mice caused deficits in light touch and silenced half of small-diameter sensory neurons expressing mechanically activated rapidly adapting mechanosensitive currents although TRPC3 or TRPC6 single knockout mice produced no behavioral phenotype. This indicates that TRPC3 and TRPC6 may show some compensation in function, consistent with their functional redundancy observed also in other system [69]. Similarly, double TRPC3/TRPC6 knockout mice also showed hearing impairment, vestibular deficits, and defective auditory brain stem responses to high frequency sounds. In addition, another study reported that TRPC1 mutant animals display a decrease in sensitivity to innocuous mechanical stimuli and show a reduction in down-hair A $\delta$  and slowly adapting A $\delta$  fiber firing in response to innocuous mechanical stimulation [70]. TRPC1 and TRPC5 are shown



to be sensitive to osmotically induced membrane stretch in cultured DRG neurons and HEK293 cells, respectively [71, 72]. TRPC6 is also activated by membrane stretch while both TRPC5 and TRPC6 activities are blocked by a tarantula toxin known to inhibit mechanosensitive channels [73]. However, it should be noted that although several TRPC channel members, including TRPC1 and TRPC6, have been proposed to contribute to mechanical sensitivity in the vertebrate cells, the functional expression of these subunits in heterologous systems remains problematic, manifesting as no alteration of mechanosensitive current by overexpression of these subunits [74]. Given this background, it is hard to see TRPC channels act as a direct mechanosensor. It is possible that TRPC proteins are necessary for regulating intracellular calcium levels that may modulate the activity of other direct mechanosensors, for example, Piezo, TMC, TACAN, or other proteins [75–78]. This possibility remains to be further explored.

## 5. Functional Roles of TRPC Subfamily in the Nociception

*5.1. Plastic Changes of TRPC Proteins under Pathological States.* Following nerve injury or tissue inflammation, TRPC proteins displayed different change profile in the DRG. Staaf et al. reported that TRPC1-TRPC7 displayed different change profile in the DRG [79]. TRPC1 and TRPC6 were not differentially regulated throughout the test period after spared nerve injury (SNI). TRPC3 was downregulated at 4 days and back to normal at 15 days and 3 months after SNI. TRPC4 had reduced expression levels at all time points after SNI while TRPC5 was downregulated at 15 days and 3 months but less reduced at 4 days. TRPC2 and C7 were not detected. In contrast, in another study with nerve injury, TRPC4 was shown to be increased at both transcriptional and protein levels in the DRG [59]. In the synovia from animal and human inflammatory arthritis, TRPC5 expression demonstrated a reduction at mRNA level, while the other TRPC proteins were not altered [26]. Repeated cyclophosphamide injections induced marked cystitis and bladder hyperactivity, which is accompanied by a specific increase in the expression of TRPC1 and TRPC4 in bladder innervating sensory neurons and the sprouting of sensory fibers in the bladder mucosa [80]. In addition, TRPC1, TRPC3, and TRPC6 have also been upregulated in the spinal cord in mice developed with morphine analgesic tolerance and hyperalgesia after chronic morphine exposure [81].

*5.2. Functional Roles of TRPC Proteins in Pathological Pain.* Nonselective blockade of TRPC channels with SKF-96365 was shown to inhibit spontaneous nociception and inflammatory pain hypersensitivity [82]. A previous study showed that TRPC1 and TRPC6 are coexpressed with TRPV4 in DRG neurons. Using antisense to TRPC1 and TRPC6, it has been proposed that they may act in concert to mediate mechanical hypersensitivity induced by carrageenan and paclitaxel chemotherapy [24]. Chronic pain usually accompanies immune-related disorders with an elevated level of serum IgG immune complex (IgG-IC), but the underlying

mechanisms are obscure. A previous study by interfering TRPC3 with siRNA reported that TRPC3 is required for IgG immune complex-induced excitation of rat DRG neurons [60]. There is also evidence showing that TRPC4 is required for the detection and/or transmission of colonic MO visceral pain sensation [25]. Rats with a transposon-mediated TRPC4-knockout mutation displayed tolerance to visceral pain induced by colonic mustard oil (MO) exposure, but not somatic or neuropathic pain stimuli. Moreover, wild-type rats treated with a selective TRPC4 antagonist (ML-204) prior to MO exposure mimicked the behavioral responses observed in TRPC4-knockout rats.

Although the TRPC protein expression profile in the supraspinal regions responsible for pain perception is rarely investigated, a recent study by Wei et al. showed a facilitating regulation of neuropathic pain by amygdaloid TRPC4/C5 channels [23]. Chronic cannula for microinjection of ML-204, a TRPC4/C5 antagonist, produced a dose-dependent inhibition of pain hypersensitivity as well as anxiety-like behavior associated with nerve injury. This result indicates that amygdaloid TRPC4/C5 may contribute to the maintenance of pain hypersensitivity and pain-related aversion in neuropathy. In addition, spinal TRPC6 channels are shown to contribute to morphine-induced antinociceptive tolerance and hyperalgesia in rats [81]. Knockdown of TRPC6 in the spinal cord prevents the development of morphine-induced tolerance and hyperalgesia. In comparison with the pronociceptive action of other TRPC proteins, TRPC5 is recently reported to be able to protect against pain and vascular inflammation in arthritis and joint inflammation [26]. In this study, following complete Freund's adjuvant- (CFA-) induced unilateral arthritis, genetic ablation (TRPC5 KO mice) or pharmacological blockade of TRPC5 displays augmented weight-bearing asymmetry, enhanced secondary hyperalgesia, and increased cytokine concentrations in the synovial lavage fluid. This result suggests that the activation of TRPC5 may be associated with an endogenous anti-inflammatory/analgesic pathway in inflammatory joint conditions [26].

## 6. The Underlying Mechanisms for Functional Roles of TRPC Proteins in Pathological Pain

*6.1. Regulation of Calcium Homeostasis by TRPC Proteins in Primary Sensory Neurons.* It has recently been shown that an abnormal persistent increase in intracellular calcium levels mediates the transition from acute to chronic pain in inflammatory states [1, 83, 84]. Thus, the regulation of intracellular calcium level could be a key mechanism in preventing pain chronicity. TRPC channels (including subtypes 1-7), a family of  $\text{Ca}^{2+}$ -permeable nonselective cation channels, play a critical role in the regulation of intracellular  $\text{Ca}^{2+}$  homeostasis and membrane excitability in excitable cells [85]. It has been thought that TRPC channels contribute to receptor-operated calcium entry (ROCE) or store-operated calcium entry (SOCE) through PIP2 hydrolysis by PLC activation. Accumulated evidence has suggested that TRPC3, TRPC6, and TRPC7 form agonist-regulated channels that can be directly activated by DAG

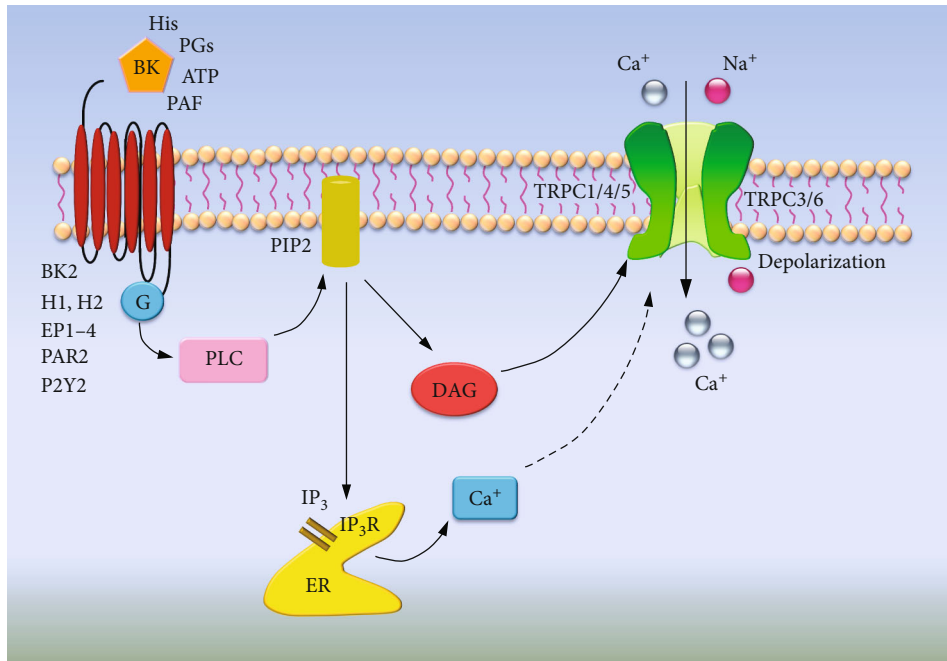


FIGURE 3: The proposed mechanisms underlying functional roles of TRPC channels in nociceptive hypersensitivity. Following inflammation or injury, a variety of proinflammatory mediators, such as bradykinin, prostaglandins, histamine, platelet-activating factor, and ATP, are released from the injured site and activates their corresponding Gq-protein-coupled receptors (GPCRs) in the peripheral nociceptors. These GPCRs couple with PLC and cleave PIP2 into DAG and IP3, involving the activation of TRPC3/6 directly by DAG, or the activation of TRPC3/6 or TRPC1/4/5 by IP3-induced depletion of endoplasmic reticulum calcium store. The resulting increase of calcium level via TRPC channels leads to depolarization of nociceptors, which in turn causes peripheral sensitization and nociceptive hypersensitivity. BK: bradykinin; His: histamine; PAF: platelet-activating factor; BK2: bradykinin receptor 2; PLC: phospholipase C; PIP2: phosphatidylinositol 4,5-bisphosphate; DAG: diacylglycerol; IP3: inositol trisphosphate.

in different biological system [86–88]. This inference is further supported by recent Cryo-EM structure analysis of TRPC3 and TRPC6 [39, 89, 90]. In contrast to good consensus regarding a direct binding of DAG to TRPC3/6/7 subfamily, there is a controversial debate on the role of TRPC channels as candidates of SOCE. Although TRPC1, TRPC4, and TRPC5 channels have been linked to SOCE in several different biological systems [91–93], none of the TRPC channels generate  $\text{Ca}^{2+}$  currents that resemble the  $I_{\text{SOC}}$  current [94]. Nevertheless, this does not exclude the possibility that TRPC channels also participate in SOCE under certain scenarios, such as the assembly with the STIM1-Orai1 complex, which have been identified as the key components of SOCE. There is evidence showing TRPC1 are assembled to form a dynamic STIM1-Orai1-TRPC1 ternary complex to drive the  $I_{\text{SOC}}$  current in human embryonic kidney (HEK) cells and human salivary gland cells [94–97]. However, both the cellular physiology of TRPC channels in primary sensory neurons and their underlying mechanisms in the regulation of nociception have yet to be clarified.

TRPC channels integrate several types of intracellular stimuli, including PLC and PKC activity, DAG level, intracellular calcium levels, and PIP2 levels into changes in membrane potential and calcium entry [32]. Of note, TRPC proteins have been shown to be expressed in the primary sensory neurons along the somatosensory pathway which might contribute to inflammatory pain [24, 57–63]. Following

inflammation or injury, the inflamed area releases various proinflammatory mediators, which forms a soup to trigger oedema, itchiness, redness, and peripheral sensitization [28, 98]. This “inflammatory soup” comprises a variety of signaling molecules, such as bradykinin, prostaglandins, histamine, platelet-activating factor, and ATP, which are able to activate their corresponding receptors located in nociceptors and cause hyperexcitability of nociceptors [99]. Many of these receptors include well-characterized Gq-protein-coupled receptors (GPCR), such as bradykinin B2, histamine receptor H1 and H2, P2Y2 purinoceptor, and protease-activated PAR2 receptor [99]. The activation of these GPCRs by agonist binding and their coupling with phospholipase C (PLC) cleaves the phosphatidylinositol 4,5-bisphosphate (PIP2) into two bioactive products, diacylglycerol (DAG), and inositol trisphosphate (IP3). Both of these two compounds act to increase intracellular  $\text{Ca}^{2+}$  levels through two distinct mechanisms: one is referred to as receptor-operated  $\text{Ca}^{2+}$  entry (ROCE), involving the activation of calcium permeable channels directly by DAG [87, 88, 99]. The other is store-operated  $\text{Ca}^{2+}$  entry (SOCE), involving the activation of calcium channels by IP3-induced depletion of endoplasmic reticulum calcium stores. Hence, these two processes via PLC activation collectively result in both intracellular  $\text{Ca}^{2+}$  increase and enhanced extracellular  $\text{Ca}^{2+}$  influx across the plasma membrane following depletion of intracellular  $\text{Ca}^{2+}$  store (Figure 3). In a recent study by Alkhani et al., the authors identified TRPC3, a highly expressed TRPC

protein in DRG neurons, is coupled to several classes of pro-inflammatory metabotropic receptors and plays a significant role in  $\text{Ca}^{2+}$  homeostasis and sensitization in primary nociceptors, through its involvement in both SOCE and ROCE [62]. Indeed, TRPC3 has been shown to be required for the cellular response to IgG immune complex (IgG-IC), a pain-inducing inflammatory compound that binds to the GPCR Fc-gamma receptor I (Fc $\gamma$ RI), which in turn is coupled to TRPC3 through the Syk-PLC-IP3 pathway [60]. Similarly, with the use of nonselective TRPC inhibitor SKF-96365, TRPC channels were revealed to be involved in the inflammatory component melittin-induced  $\text{Ca}^{2+}$  increase and activation of primary nociceptors [100]. These results indicate that regulation of  $\text{Ca}^{2+}$  homeostasis in the primary sensory neurons by TRPC proteins provides a cellular basis for its role in nociception.

## 7. Novel Pharmacological Tools for TRPC Proteins

TRPC channels are widely expressed in the nervous system and permit the influx of  $\text{Na}^+$  and  $\text{Ca}^{2+}$  ions into the cells. The establishment of transgenic mouse models of different subtypes of TRPC channels and the development of subtype-specific pharmacological modulators for TRPC channels are crucial for the distinction of respective TRPC channels that play particular physiological and pathological roles in the native system. With these important tools, accumulating evidence has emerged that TRPC channels play pivotal roles in many physiological and pathological processes, including memory, cardiovascular diseases, neurological disorders, cancer, chronic kidney disease, pain, and other pathological conditions. However, it is important to point out that many of previous research on TRPC channels were based on genetic evidence through which phenotype was examined in animals or cells with naturally occurring mutations or gene knockout or knockdown. Given the apparent structural and functional redundancy observed in TRPC subunits, the genetic-based approaches may suffer from compensatory changes between subunits, which may mask the function of specific TRPC subunit under some circumstances. In recent years, there has been a great progress in the exploration of selective and specific compounds acting on different subtypes of TRPC channels. Here, we summarize several characterized important pharmacological tools to specifically activate or block TRPC channel subtype, with which for further exploration of the functional significance and translational potential of TRPC channels.

### 7.1. Broad Range TRPC Channel Blockers

**7.1.1. SKF-96365.** Organic synthetic blockers have been recognized to interfere with receptor-dependent and store-operated calcium entry mechanisms [101, 102]. SKF-96365, 1- $\{\beta$ -[3-(4-methoxyphenyl) propoxy]-4-methoxyphenethyl}-1H-imidazole hydrochloride, is an inhibitor of receptor-mediated as well as store-operated calcium entry mechanisms [101, 102]. The molecular identification, cloning, and functional characterization of a great variety of ion channels

enabled the identification of SKF-96365 as a broad range TRPC channel blocker. In rats with inflammatory pain induced by melittin injection, localized peripheral administration of SKF-96365 prevented the development and maintenance of spontaneous nociception and hyperalgesia [82]. Further mechanistic analysis revealed that SKF-96365-induced antinociception was mediated by inhibition of melittin-evoked calcium increase and hyperexcitability in primary sensory neurons [100].

### 7.2. Selective Modulators of TRPC3/6/7 Channels

**7.2.1. Hyperforin.** Hyperforin, a bicyclic polyprenylated acylphloroglucinol derivative, is the main active principle of St. John's wort extract responsible for its antidepressive profile. In contrast to classic, synthetic antidepressants, hyperforin reduces monoamine uptake by elevating the intracellular sodium concentration and subsequent elimination of the sodium gradient as required for neurotransmitter transporter function [103, 104]. The attempts to identify the molecular target of hyperforin by Harteneck and Gollasch led to the identification of TRPC6 [105]. Further studies revealed that hyperforin-induced cation entry was highly specific and related to TRPC6 and was suppressed in cells expressing a dominant negative mutant of TRPC6, whereas phylogenetically related channels, i.e., TRPC3, remained unaffected. The discovery of hyperforin as a selective TRPC6 channel activator is exciting because modulation of TRP channels by secondary plant compounds is not restricted to TRPA, TRPV, or TRPM subfamilies [105]. These data provided an important pharmacological tool to investigate TRPC6 function.

**7.2.2. GSK1702934A and Structural Analogs.** Additionally, a small 1,3-dihydro-2H-benzo [d]imidazol-2-one-based potent agonist, GSK1702934A, has been identified as a tool to directly activate TRPC3/6 channels. In HEK293 cells expressing human TRPC3 and TRPC6, GSK1702934A was shown to induce a current with EC<sub>50</sub> at 80 and 440 nM [106, 107]. Based on the main scaffold of GSK1702934A, an azobenzene photoswitch moiety was added to produce a light-sensitive TRPC agonist, OptoBI-1, which possesses the ability to preferentially activate TRPC3/6/7, but not TRPC4/5 [108]. It has been reported that light stimulation can suppress the action potential firing induced by current injection in hippocampal neurons exposed to OptoBI-1. In addition, a recent study synthesized a positive allosteric TRPC6 modulator, namely, C20 [109]. This compound shows higher selectivity at TRPC6 over closely related TRPC3 and TRPC7.

**7.2.3. Pyrazole Derivatives.** Kiyonaka et al. identifies a pyrazole compound (Pyr3) as a selective TRPC3 inhibitor [110]. The direct action of Pyr3 on the TRPC3 protein was demonstrated by electrophysiological and photoaffinity-labeling experiments. Moreover, Pyr3 was shown to be capable of eliminating B cell receptor-induced  $\text{Ca}^{2+}$  oscillation regulated by TRPC3-mediated  $\text{Ca}^{2+}$  influx and attenuating activation of nuclear factor of activated T cells as well as hypertrophic growth in cardiomyocytes. These findings are certainly exciting because Pyr3 enables a pharmacological



dissection of closely related TRPC subtypes and provides a powerful tool to study *in vivo* function of TRPC3. Previous studies reported the requirement of TRPC3 for IgG immune complex-induced excitation of rat DRG neurons under chronic pain states. A pyrazole compound, Pyr3, was shown to suppress the neuronal excitation induced by IgG immune complex in DRG [60]. However, later investigations questioned on the selectivity of pyrazole inhibitors and demonstrated Pyr3 inhibit STIM/Orai  $\text{Ca}^{2+}$  entry complexes [111]. Schleifer et al. identified other pyrazole derivatives, especially Pyr10, as a selective TRPC3 blocker, which is able to distinguish between TRPC3 and Orai-mediated SOCE. Pyr10 showed much more potency on TRPC3 than Orai-mediated SOCE [111]. A previous study demonstrated that TRPC3 channels contribute to SOCE and inflammatory transductions in primary nociceptors. Delivery of TRPC3 antagonist, Pyr10, was found to result in a substantial decrease of SOCE [62].

**7.2.4. GSK Compounds.** GSK compounds, i.e., GSK2332255B and GSK2833503A, have been shown to selectively block the activities of TRPC3 and TRPC6 with  $\text{IC}_{50}$  at 3-21 nM [112]. These two compounds display more than 100-fold selectivity over other calcium-permeable channels and 100-fold greater potency at TRPC3 compared with Pyr3. Functional analysis revealed that GSK2332255B and GSK2833503A possess the capability to inhibit pathological cardiac hypertrophy [112].

**7.2.5. BI 749327.** A recent study by Lin et al. developed an orally bioavailable TRPC6 antagonist, BI 749327 [113], and tested its effect in *in vivo* disease models. BI 749327 was reported to have an 85- and 42-fold selectivity on TRPC6 ( $\text{IC}_{50}$  13 nM against mouse TRPC6,  $t_{1/2}$  8.5-13.5 hours) over the most closely related channels, TRPC3 and TRPC7. *In vitro* and *in vivo* data collectively supported BI 749327 as a selective pharmacological TRPC6 inhibitor with oral bioavailability and suitable pharmacokinetics to ameliorate TRPC6-mediated disorders.

**7.2.6. Other TRPC6 Inhibitors.** Recently, SAR7334 synthesized from a series of aminoindanol derivatives was identified as a potent TRPC6 inhibitor with an  $\text{IC}_{50}$  at 7.9 nM [114]. This compound showed higher selectivity for TRPC6 over TRPC3/7 channels. In addition, another compound DS88790512, a bicyclo[4.3.0]nonane derivative, has also been demonstrated to inhibit TRPC6 with an  $\text{IC}_{50}$  at 11 nM [115]. Beyond this, some compounds derived from a natural product, i.e., SH045, larixyl acetate, have also the capability to inhibit TRPC6 channels with higher selectivity over TRPC3 and TRPC7 [109, 116].

### 7.3. Selective Modulators of TRPC1/4/5 Channels

**7.3.1. (-)-Englerin A.** (-)-Englerin A (EA) was found to be a highly efficient, fast-acting, potent, selective, and direct stimulator of TRPC4 and TRPC5 channels [117]. EA produced the rapid and selective killing of renal cancer cells through the activation of TRPC4/5 channels. The  $\text{EC}_{50}$  of EA for the activation of TRPC4 channels is 11.2 nM and 7.6 nM for TRPC5 channels [117]. A recent study identified an

(-)-englerin A analog, which antagonizes (-)-englerin A at TRPC1/4/5 channels [118].

**7.3.2. AM237.** Recent research has yielded specific and potent xanthine-based TRPC1/4/5 inhibitors, such as Pico145; a newly published study by Minard et al. reported the possibility of xanthine-based activation of TRPC5 channels [119]. AM237, an analog of the TRPC1/4/5 inhibitor Pico145, was synthesized and showed the capability to activate homomeric TRPC5 channels, but not other subtypes of TRP channels. This study supports the general principle that xanthine-based small molecules can activate, potentiate, or inhibit these channels depending on subunit composition.

**7.3.3. Benzothiadiazine Derivative and Methylprednisolone.** Recently, Beckmann et al. screened compound libraries and identified the glucocorticoid methylprednisolone and N-[3-(adamantan-2-yloxy)propyl]-3-(6-methyl-1,1-dioxo-2H-1 $\lambda$ 6,2,4-benzothiadiazin-3-yl)propanamide (BTD) as novel TRPC5 activators [120]. These compounds exhibited selectivity for TRPC5 channels or heteromeric channel complexes containing TRPC5 but failed to activate the other TRP channel subtypes, i.e., TRPC3, TRPC6, TRPC7, TRPA1, TRPV1, TRPV2, TRPV4, TRPM2, and TRPM3.

**7.3.4. Tonantzitolone.** TZL is a novel potent agonist for TRPC1/4/5 channels, which shows potency at a nanomolar level [118]. TZL activated overexpressed channels with  $\text{EC}_{50}$  values of 123 nM (TRPC4), 83 nM (TRPC5), 140 nM (TRPC4-TRPC1), and 61 nM (TRPC5-TRPC1). These effects of TZL were potently antagonized by the TRPC1/4/5 inhibitor Pico145. TZL failed to activate endogenous store-operated  $\text{Ca}^{2+}$  entry or overexpressed TRPC3, TRPV4, or TRPM2 channels in HEK293 cells.

**7.3.5. Pico145.** For exploration of the physiological significance and translational potential of TRPC1/4/5 channels, it is of great importance to develop pharmacological tools with which to specifically block the channels. A recent study by Rubaiy et al. characterized a potent, specific antagonist, Pico145 at TRPC1/4/5 channels [38]. The potency of Pico145 ranged from 9 to 1300 pM, depending on the TRPC1/4/5 subtype and activation mechanism. The heteromeric channels TRPC4/TRPC1 or TRPC5/TRPC1 concatemers showed more sensitivity to Pico145 with  $\text{IC}_{50}$  at 33 pM and 199 pM, respectively. Moreover, the other TRP channel subtypes were not affected by Pico145, such as TRPC3, TRPC6, TRPV1, TRPV4, TRPA1, TRPM2, TRPM8, and store-operated  $\text{Ca}^{2+}$  entry mediated by Orai1 [38, 121].

**7.3.6. ML204.** A study by Miller et al. reported the identification and characterization of ML204 as a novel, potent inhibitor for TRPC4 and TRPC5 channels [122]. ML204 was shown to inhibit TRPC4 $\beta$ -mediated intracellular  $\text{Ca}^{2+}$  elevation with  $\text{IC}_{50}$  at 0.96  $\mu\text{M}$  and exhibit 19-fold selectivity against muscarinic receptor-coupled TRPC6 channel activation. In DRG neurons, ML204 at 10-20  $\mu\text{M}$  showed no appreciable blockade of the other TRP channel subtype, including TRPV1, TRPV3, TRPA1, TRPM8, and KCNQ2 as well as native voltage-gated  $\text{Na}^+$ ,  $\text{K}^+$ , and  $\text{Ca}^{2+}$  channels. Of note,



native TRPC1/4/5 channels exist as heteromers, but ML204 was less sensitive to heteromeric channels, such as TRPC4/TRPC1. Whole-cell patch clamp recording demonstrated that ML204 display potent inhibition on homomeric TRPC4 channels. In a rat visceral pain model induced by colonic injection of mustard oil, intraperitoneal administration of ML204 produced a prominent antinociceptive effect [25]. Moreover, microinjection of ML204 in the amygdala led to a dose-dependent pain relief without obvious side effects. Additionally, amygdaloid administration of ML204 reduced aversive emotion associated with neuropathic pain [23].

**7.3.7. Benzimidazoles.** A recent study by Sharma *et al.* reported the synthesis and biological characterization of a series of N-heterocyclic-1-benzyl-1Hbenzo[d]imidazole-2-amines as selective TRPC5 inhibitors [123]. This team evaluated the benzimidazole scaffold and substituents resulting in the discovery of AC1903, a novel TRPC5 inhibitor that is active in multiple animal models of chronic kidney disease (CKD). This compound was shown to have an  $IC_{50}$  at  $4.06 \mu M$ , as determined using the SyncroPatch. An *in vivo* pharmacokinetic study demonstrated that this compound has PK values similar to ML204 and is selective for TRPC5 over TRPC4 and TRPC6. In addition, another 2-aminobenzimidazole derivative, M084, was identified to be an TRPC4/5 inhibitor. Although not as potent as ML204, M084 has better pharmacokinetic properties and similar potency at TRPC4 and TRPC5 channels with  $IC_{50}$  at about  $10 \mu M$ . Intraperitoneal injection of ML084 produced a profound antidepressant and anxiolytic effects in mice [124].

## 8. Concluding Remarks and Future Directions

Despite great progress that has been made in our understanding of the chronic pain in the last decades, chronic pain still represents a major public health problem. This mainly manifests as two unevadable facts: one, the underlying mechanism remains elusive, and two, treatment of chronic pain in clinic still faces serious challenges. TRPC channels form non-selective cation channels with higher permeability for  $Ca^{2+}$  and  $Na^+$  into the cells and expressed in the pain pathway, especially in the DRG—the first site for the detection of nociceptive signals [24, 28, 57–63]. Emerging evidence has indicated a potential role of TRPC channels in the detection of external stimuli and nociceptive hypersensitivity under pathological states. Nevertheless, our understanding of these ion channels in nociception and nociceptive hypersensitivity is far from being revealed. In recent years, the resolution revolution in cryoelectron microscopy brought several high-resolution structures of TRP channels. What excites us is that the electron cryomicroscopy structure of the TRPC3, TRPC6, and TRPC4 proteins has been resolved just recently [39, 89, 125, 126]. This can help to study TRPC channel pharmacology and activation mechanisms, which in turn offer the opportunity to develop more potent, selective pharmacological tools to explore the physiological and pathological functions of TRPC channels, including pain and sensitization.

## Conflicts of Interest

The authors declare no completing financial interests.

## Authors' Contributions

C.L. composed the idea and wrote the manuscript. Z.C.S. drafted the manuscript and prepared the figure. S.B.M. and W.G.C. discussed the draft and prepared the figure. D.J. provided constructive comments. Zhi-Chuan Sun, Sui-Bin Ma, and Wen-Guang Chu contributed equally to this work.

## Acknowledgments

This work was supported by the Natural Science Foundation of China (NSFC) grants to C.L. (Nos. 31671088 and 31471059) and grants from the Natural Science Foundation of Shaanxi Province (No. 2017ZDJC-01) to C.L.

## References

- [1] R. Kuner, "Central mechanisms of pathological pain," *Nature Medicine*, vol. 16, no. 11, pp. 1258–1266, 2010.
- [2] C. J. Woolf and M. W. Salter, "Neuronal plasticity: increasing the gain in pain," *Science*, vol. 288, no. 5472, pp. 1765–1768, 2000.
- [3] B. Nilius and A. Szallasi, "Transient receptor potential channels as drug targets: from the science of basic research to the art of medicine," *Pharmacological Reviews*, vol. 66, no. 3, pp. 676–814, 2014.
- [4] B. Nilius, G. Owsianik, T. Voets, and J. A. Peters, "Transient receptor potential cation channels in disease," *Physiological Reviews*, vol. 87, no. 1, pp. 165–217, 2007.
- [5] D. E. Clapham, "TRP channels as cellular sensors," *Nature*, vol. 426, no. 6966, pp. 517–524, 2003.
- [6] D. P. Corey, "New TRP channels in hearing and mechanosensation," *Neuron*, vol. 39, no. 4, pp. 585–588, 2003.
- [7] P. Delmas, "Polycystins: from mechanosensation to gene regulation," *Cell*, vol. 118, no. 2, pp. 145–148, 2004.
- [8] C. Montell, L. Birnbaumer, and V. Flockerzi, "The TRP channels, a remarkably functional family," *Cell*, vol. 108, no. 5, pp. 595–598, 2002.
- [9] M. M. Moran, H. Xu, and D. E. Clapham, "TRP ion channels in the nervous system," *Current Opinion in Neurobiology*, vol. 14, no. 3, pp. 362–369, 2004.
- [10] D. Julius, "TRP channels and pain," *Annual Review of Cell and Developmental Biology*, vol. 29, no. 1, pp. 355–384, 2013.
- [11] I. Jardín, J. J. López, R. Diez *et al.*, "TRPs in pain sensation," *Frontiers in Physiology*, vol. 8, p. 392, 2017.
- [12] C. Y. Hung and C. H. Tan, "TRP channels in nociception and pathological pain," *Advances in Experimental Medicine and Biology*, vol. 1099, pp. 13–27, 2018.
- [13] S. Selvaraj, Y. Sun, and B. B. Singh, "TRPC channels and their implication in neurological diseases," *CNS and Neurological Disorders - Drug Targets*, vol. 9, no. 1, pp. 94–104, 2010.
- [14] E. B. E. Becker, "The Moonwalker mouse: new insights into TRPC3 function, cerebellar development, and ataxia," *Cerebellum*, vol. 13, no. 5, pp. 628–636, 2014.
- [15] M. Trebak, "The puzzling role of TRPC3 channels in motor coordination," *Pflugers Arch*, vol. 459, no. 3, pp. 369–375, 2010.

- [16] J. Hartmann, E. Dragicevic, H. Adelsberger et al., "TRPC3 channels are required for synaptic transmission and motor coordination," *Neuron*, vol. 59, no. 3, pp. 392–398, 2008.
- [17] Y. Li, Y. C. Jia, K. Cui et al., "Essential role of TRPC channels in the guidance of nerve growth cones by brain-derived neurotrophic factor," *Nature*, vol. 434, no. 7035, pp. 894–898, 2005.
- [18] J. Bröker-Lai, A. Kollwe, B. Schindeldecker et al., "Heteromeric channels formed by TRPC1, TRPC4 and TRPC5 define hippocampal synaptic transmission and working memory," *The EMBO Journal*, vol. 36, no. 18, pp. 2770–2789, 2017.
- [19] A. Riccio, Y. Li, J. Moon et al., "Essential role for TRPC5 in amygdala function and fear-related behavior," *Cell*, vol. 137, no. 4, pp. 761–772, 2009.
- [20] C. Hong, H. Seo, M. Kwak et al., "Increased TRPC5 glutathionylation contributes to striatal neuron loss in Huntington's disease," *Brain*, vol. 138, no. 10, pp. 3030–3047, 2015.
- [21] M. Li, C. Chen, Z. Zhou, S. Xu, and Z. Yu, "A TRPC1-mediated increase in store-operated  $\text{Ca}^{2+}$  entry is required for the proliferation of adult hippocampal neural progenitor cells," *Cell Calcium*, vol. 51, no. 6, pp. 486–496, 2012.
- [22] A. Greka, B. Navarro, E. Oancea, A. Duggan, and D. E. Clapham, "TRPC5 is a regulator of hippocampal neurite length and growth cone morphology," *Nature Neuroscience*, vol. 6, no. 8, pp. 837–845, 2003.
- [23] H. Wei, B. Sagalajev, M. A. Yüzer, A. Koivisto, and A. Pertovaara, "Regulation of neuropathic pain behavior by amygdaloid TRPC4/C5 channels," *Neuroscience Letters*, vol. 608, pp. 12–17, 2015.
- [24] N. Alessandri-Haber, O. A. Dina, X. Chen, and J. D. Levine, "TRPC1 and TRPC6 channels cooperate with TRPV4 to mediate mechanical hyperalgesia and nociceptor sensitization," *Journal of Neuroscience*, vol. 29, no. 19, pp. 6217–6228, 2009.
- [25] K. N. Westlund, L. P. Zhang, F. Ma et al., "A rat knockout model implicates TRPC4 in visceral pain sensation," *Neuroscience*, vol. 262, pp. 165–175, 2014.
- [26] K. M. Alawi, F. A. Russell, A. A. Aubdool et al., "Transient receptor potential canonical 5 (TRPC5) protects against pain and vascular inflammation in arthritis and joint inflammation," *Annals of the Rheumatic Diseases*, vol. 76, no. 1, pp. 252–260, 2017.
- [27] J. N. Campbell and R. A. Meyer, "Mechanisms of neuropathic pain," *Neuron*, vol. 52, no. 1, pp. 77–92, 2006.
- [28] C. J. Woolf and Q. Ma, "Nociceptors—Noxious stimulus detectors," *Neuron*, vol. 55, no. 3, pp. 353–364, 2007.
- [29] M. S. Gold and G. F. Gebhart, "Nociceptor sensitization in pain pathogenesis," *Nature Medicine*, vol. 16, no. 11, pp. 1248–1257, 2010.
- [30] D. J. Beech, "Characteristics of transient receptor potential canonical calcium-permeable channels and their relevance to vascular physiology and disease," *Circulation Journal*, vol. 77, no. 3, pp. 570–579, 2013.
- [31] P. D. Wes, J. Chevesich, A. Jeromin, C. Rosenberg, G. Stetten, and C. Montell, "TRPC1, a human homolog of a *Drosophila* store-operated channel," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 92, no. 21, pp. 9652–9656, 1995.
- [32] I. S. Ramsey, M. Delling, and D. E. Clapham, "An introduction to TRP channels," *Annual Review of Physiology*, vol. 68, no. 1, pp. 619–647, 2006.
- [33] T. Hofmann, M. Schaefer, G. Schultz, and T. Gudermann, "Subunit composition of mammalian transient receptor potential channels in living cells," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 99, no. 11, pp. 7461–7466, 2002.
- [34] M. Goel, W. G. Sinkins, and W. P. Schilling, "Selective association of TRPC channel subunits in rat brain synaptosomes," *Journal of Biological Chemistry*, vol. 277, no. 50, pp. 48303–48310, 2002.
- [35] C. Strübing, G. Krapivinsky, L. Krapivinsky, and D. E. Clapham, "TRPC1 and TRPC5 form a novel cation channel in mammalian brain," *Neuron*, vol. 29, no. 3, pp. 645–655, 2001.
- [36] C. Strübing, G. Krapivinsky, L. Krapivinsky, and D. E. Clapham, "Formation of novel TRPC channels by complex subunit interactions in embryonic brain," *Journal of Biological Chemistry*, vol. 278, no. 40, pp. 39014–39019, 2003.
- [37] M. Freichel, R. Vennekens, J. Olausson et al., "Functional role of TRPC proteins in native systems: implications from knockout and knock-down studies," *Journal of Physiology*, vol. 567, no. 1, pp. 59–66, 2005.
- [38] H. N. Rubaiy, M. J. Ludlow, M. Henrot et al., "Picomolar, selective, and subtype-specific small-molecule inhibition of TRPC1/4/5 channels," *Journal of Biological Chemistry*, vol. 292, no. 20, pp. 8158–8173, 2017.
- [39] Q. Tang, W. Guo, L. Zheng et al., "Structure of the receptor-activated human TRPC6 and TRPC3 ion channels," *Cell Research*, vol. 28, no. 7, pp. 746–755, 2018.
- [40] X.-Z. S. Xu, H.-S. Li, W. B. Guggino, and C. Montell, "Coassembly of TRP and TRPL produces a distinct store-operated conductance," *Cell*, vol. 89, no. 7, pp. 1155–1164, 1997.
- [41] X. Liu, B. C. Bandyopadhyay, B. B. Singh, K. Groschner, and I. S. Ambudkar, "Molecular analysis of a store-operated and 2-acetyl-sn-glycerol-sensitive non-selective cation channel. Heteromeric assembly of TRPC1-TRPC3," *Journal of Biological Chemistry*, vol. 280, no. 22, pp. 21600–21606, 2005.
- [42] H. Schmalbruch, "The number of neurons in dorsal root ganglia L4-L6 of the rat," *The Anatomical Record*, vol. 219, no. 3, pp. 315–322, 1987.
- [43] B. Sørensen, T. Tandrup, M. Koltzenburg, and J. Jakobsen, "No further loss of dorsal root ganglion cells after axotomy in p75 neurotrophin receptor knockout mice," *Journal of Comparative Neurology*, vol. 459, no. 3, pp. 242–250, 2003.
- [44] A. I. Basbaum, D. M. Bautista, G. Scherrer, and D. Julius, "Cellular and molecular mechanisms of pain," *Cell*, vol. 139, no. 2, pp. 267–284, 2009.
- [45] A. I. Basbaum and C. J. Woolf, "Pain," *Current Biology*, vol. 9, no. 12, pp. R429–R431, 1999.
- [46] Y. Liu and Q. Ma, "Generation of somatic sensory neuron diversity and implications on sensory coding," *Current Opinion in Neurobiology*, vol. 21, no. 1, pp. 52–60, 2011.
- [47] M. J. Caterina, M. A. Schumacher, M. Tominaga, T. A. Rosen, J. D. Levine, and D. Julius, "The capsaicin receptor: a heat-activated ion channel in the pain pathway," *Nature*, vol. 389, no. 6653, pp. 816–824, 1997.
- [48] M. J. Caterina, A. Leffler, A. B. Malmberg et al., "Impaired nociception and pain sensation in mice lacking the capsaicin receptor," *Science*, vol. 288, no. 5464, pp. 306–313, 2000.
- [49] K. Kobayashi, T. Fukuoka, K. Obata et al., "Distinct expression of TRPM8, TRPA1, and TRPV1 mRNAs in rat primary afferent neurons with  $\text{A}\delta/\text{C}$ -fibers and colocalization with trk

- receptors," *Journal of Comparative Neurology*, vol. 493, no. 4, pp. 596–606, 2005.
- [50] M. Tominaga, M. J. Caterina, A. B. Malmberg et al., "The cloned capsaicin receptor integrates multiple pain-producing stimuli," *Neuron*, vol. 21, no. 3, pp. 531–543, 1998.
- [51] D. D. McKemy, W. M. Neuhauser, and D. Julius, "Identification of a cold receptor reveals a general role for TRP channels in thermosensation," *Nature*, vol. 416, no. 6876, pp. 52–58, 2002.
- [52] A. M. Peier, A. Moqrich, A. C. Hergarden et al., "A TRP channel that senses cold stimuli and menthol," *Cell*, vol. 108, no. 5, pp. 705–715, 2002.
- [53] D. M. Bautista, P. Movahed, A. Hinman et al., "Pungent products from garlic activate the sensory ion channel TRPA1," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 102, no. 34, pp. 12248–12252, 2005.
- [54] D. M. Bautista, M. Pellegrino, and M. Tsunozaki, "TRPA1: a gatekeeper for inflammation," *Annual Review of Physiology*, vol. 75, no. 1, pp. 181–200, 2013.
- [55] S. E. Jordt, D. M. Bautista, H. H. Chuang et al., "Mustard oils and cannabinoids excite sensory nerve fibres through the TRP channel ANKTM1," *Nature*, vol. 427, no. 6971, pp. 260–265, 2004.
- [56] L. J. Macpherson, B. H. Geierstanger, V. Viswanath et al., "The pungency of garlic: activation of TRPA1 and TRPV1 in response to allicin," *Current Biology*, vol. 15, no. 10, pp. 929–934, 2005.
- [57] S. Elg, F. Marmigere, J. P. Mattsson, and P. Ernfors, "Cellular subtype distribution and developmental regulation of TRPC channel members in the mouse dorsal root ganglion," *Journal of Comparative Neurology*, vol. 503, no. 1, pp. 35–46, 2007.
- [58] M. Kress, J. Karasek, A. V. Ferrer-Montiel, N. Scherbakov, and R. V. Haberberger, "TRPC channels and diacylglycerol dependent calcium signaling in rat sensory neurons," *Histochemistry and Cell Biology*, vol. 130, no. 4, pp. 655–667, 2008.
- [59] D. Wu, W. Huang, P. M. Richardson, J. V. Priestley, and M. Liu, "TRPC4 in rat dorsal root ganglion neurons is increased after nerve injury and is necessary for neurite outgrowth," *Journal of Biological Chemistry*, vol. 283, no. 1, pp. 416–426, 2008.
- [60] L. Qu, Y. Li, X. Pan, P. Zhang, R. H. LaMotte, and C. Ma, "Transient receptor potential canonical 3 (TRPC3) is required for IgG immune complex-induced excitation of the rat dorsal root ganglion neurons," *Journal of Neuroscience*, vol. 32, no. 28, pp. 9554–9562, 2012.
- [61] K. Quick, J. Zhao, N. Eijkelkamp et al., "TRPC3 and TRPC6 are essential for normal mechanotransduction in subsets of sensory neurons and cochlear hair cells," *Open Biology*, vol. 2, no. 5, article 120068, 2012.
- [62] H. Alkhani, A. R. Ase, R. Grant, D. O'Donnell, K. Groschner, and P. Séguéla, "Contribution of TRPC3 to store-operated calcium entry and inflammatory transductions in primary nociceptors," *Molecular Pain*, vol. 10, pp. 1744–8069, 2014.
- [63] I. Vandewauw, G. Owsianik, and T. Voets, "Systematic and quantitative mRNA expression analysis of TRP channel genes at the single trigeminal and dorsal root ganglion level in mouse," *BMC Neuroscience*, vol. 14, no. 1, p. 21, 2013.
- [64] C. Kunert-Keil, F. Bisping, J. Krüger, and H. Brinkmeier, "Tissue-specific expression of TRP channel genes in the mouse and its variation in three different mouse strains," *BMC Genomics*, vol. 7, no. 1, p. 159, 2006.
- [65] K. Zimmermann, J. K. Lennerz, A. Hein et al., "Transient receptor potential cation channel, subfamily C, member 5 (TRPC5) is a cold-transducer in the peripheral nervous system," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 108, no. 44, pp. 18114–18119, 2011.
- [66] C. L. Li, K. C. Li, D. Wu et al., "Erratum: Somatosensory neuron types identified by high-coverage single-cell RNA-sequencing and functional heterogeneity," *Cell Research*, vol. 26, no. 8, p. 967, 2016.
- [67] D. Usoskin, A. Furlan, S. Islam et al., "Unbiased classification of sensory neuron types by large-scale single-cell RNA sequencing," *Nature Neuroscience*, vol. 18, no. 1, pp. 145–153, 2015.
- [68] A. Sadowska, W. Hitzl, A. Karol et al., "Differential regulation of TRP channel gene and protein expression by intervertebral disc degeneration and back pain," *Scientific Reports*, vol. 9, no. 1, article 18889, 2019.
- [69] A. Dietrich, M. Mederos y Schnitzler, M. Gollasch et al., "Increased vascular smooth muscle contractility in TRPC6-/- mice," *Molecular and Cellular Biology*, vol. 25, no. 16, pp. 6980–6989, 2005.
- [70] S. R. Garrison, A. Dietrich, and C. L. Stucky, "TRPC1 contributes to light-touch sensation and mechanical responses in low-threshold cutaneous sensory neurons," *Journal of Neurophysiology*, vol. 107, no. 3, pp. 913–922, 2012.
- [71] A. Gomis, S. Soriano, C. Belmonte, and F. Viana, "Hypoosmotic- and pressure-induced membrane stretch activate TRPC5 channels," *Journal of Physiology*, vol. 586, no. 23, pp. 5633–5649, 2008.
- [72] S. Staaf, I. Maxvall, U. Lind et al., "Down regulation of TRPC1 by shRNA reduces mechanosensitivity in mouse dorsal root ganglion neurons *in vitro*," *Neuroscience Letters*, vol. 457, no. 1, pp. 3–7, 2009.
- [73] M. A. Spassova, T. Hewavitharana, W. Xu, J. Soboloff, and D. L. Gill, "A common mechanism underlies stretch activation and receptor activation of TRPC6 channels," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 103, no. 44, pp. 16586–16591, 2006.
- [74] P. Gottlieb, J. Folgering, R. Maroto et al., "Revisiting TRPC1 and TRPC6 mechanosensitivity," *Pflügers Archiv*, vol. 455, no. 6, pp. 1097–1103, 2008.
- [75] L. Beaulieu-Laroche, M. Christin, A. Donoghue et al., "TACAN is an ion channel involved in sensing mechanical pain," *Cell*, vol. 180, no. 5, pp. 956–967.e17, 2020.
- [76] B. Coste, J. Mathur, M. Schmidt et al., "Piezo1 and Piezo2 are essential components of distinct mechanically activated cation channels," *Science*, vol. 330, no. 6000, pp. 55–60, 2010.
- [77] Y. Kawashima, G. S. G. Géléoc, K. Kurima et al., "Mechanotransduction in mouse inner ear hair cells requires transmembrane channel-like genes," *Journal of Clinical Investigation*, vol. 121, no. 12, pp. 4796–4809, 2011.
- [78] B. Pan, G. S. Géléoc, Y. Asai et al., "TMC1 and TMC2 are components of the mechanotransduction channel in hair cells of the mammalian inner ear," *Neuron*, vol. 79, no. 3, pp. 504–515, 2013.
- [79] S. Staaf, S. Oerther, G. Lucas, J. P. Mattsson, and P. Ernfors, "Differential regulation of TRP channels in a rat model of neuropathic pain," *Pain*, vol. 144, no. 1, pp. 187–199, 2009.



- [80] M. Boudes, P. Uvin, S. Pinto et al., "Crucial role of TRPC1 and TRPC4 in cystitis-induced neuronal sprouting and bladder overactivity," *PLoS One*, vol. 8, no. 7, article e69550, 2013.
- [81] H. Jin, Y. T. Sun, G. Q. Guo et al., "Spinal TRPC6 channels contributes to morphine-induced antinociceptive tolerance and hyperalgesia in rats," *Neuroscience Letters*, vol. 639, pp. 138–145, 2017.
- [82] J. Ding, J. R. Zhang, Y. Wang et al., "Effects of a non-selective TRPC channel blocker, SKF-96365, on melittin-induced spontaneous persistent nociception and inflammatory pain hypersensitivity," *Neuroscience Bulletin*, vol. 28, no. 2, pp. 173–181, 2012.
- [83] C. Luo, P. H. Seeburg, R. Sprengel, and R. Kuner, "Activity-dependent potentiation of calcium signals in spinal sensory networks in inflammatory pain states," *Pain*, vol. 140, no. 2, pp. 358–367, 2008.
- [84] S. G. Lu and M. S. Gold, "Inflammation-induced increase in evoked calcium transients in subpopulations of rat dorsal root ganglion neurons," *Neuroscience*, vol. 153, no. 1, pp. 279–288, 2008.
- [85] S. F. Pedersen, G. Owsianik, and B. Nilius, "TRP channels: an overview," *Cell Calcium*, vol. 38, no. 3-4, pp. 233–252, 2005.
- [86] M. Trebak, G. Vazquez, G. S. J. Bird, and J. W. Putney Jr., "The TRPC3/6/7 subfamily of cation channels," *Cell Calcium*, vol. 33, no. 5-6, pp. 451–461, 2003.
- [87] T. Hofmann, A. G. Obukhov, M. Schaefer, C. Harteneck, T. Gudermann, and G. Schultz, "Direct activation of human TRPC6 and TRPC3 channels by diacylglycerol," *Nature*, vol. 397, no. 6716, pp. 259–263, 1999.
- [88] M. Lichtenegger, O. Tiapko, B. Svobodova et al., "An optically controlled probe identifies lipid-gating fenestrations within the TRPC3 channel," *Nature Chemical Biology*, vol. 14, no. 4, pp. 396–404, 2018.
- [89] C. M. Azumaya, F. Sierra-Valdez, J. F. Cordero-Morales, and T. Nakagawa, "Cryo-EM structure of the cytoplasmic domain of murine transient receptor potential cation channel subfamily C member 6 (TRPC6)," *Journal of Biological Chemistry*, vol. 293, no. 26, pp. 10381–10391, 2018.
- [90] S. Fiedler, U. Storch, S. Erdogmus, T. Gudermann, M. Mederos y Schnitzler, and A. Dietrich, "Small fluorescein arsenical hairpin-based Förster resonance energy transfer analysis reveals changes in amino- to carboxyl-terminal interactions upon OAG activation of classical transient receptor potential 6," *Molecular Pharmacology*, vol. 96, no. 1, pp. 90–98, 2019.
- [91] X. Liu, W. Wang, B. B. Singh et al., "Trp1, a candidate protein for the store-operated  $Ca^{2+}$  influx mechanism in salivary gland cells," *Journal of Biological Chemistry*, vol. 275, no. 5, pp. 3403–3411, 2000.
- [92] M. Freichel, S. H. Suh, A. Pfeifer et al., "Lack of an endothelial store-operated  $Ca^{2+}$  current impairs agonist-dependent vasorelaxation in TRP4<sup>-/-</sup> mice," *Nature Cell Biology*, vol. 3, no. 2, pp. 121–127, 2001.
- [93] D. J. Beech, S. Z. Xu, D. McHugh, and R. Flemming, "TRPC1 store-operated cationic channel subunit," *Cell Calcium*, vol. 33, no. 5-6, pp. 433–440, 2003.
- [94] J. J. Lopez, I. Jardin, J. Sanchez-Collado, G. M. Salido, T. Smani, and J. A. Rosado, "TRPC channels in the SOCE scenario," *Cells*, vol. 9, no. 1, p. 126, 2020.
- [95] H. L. Ong, K. T. Cheng, X. Liu et al., "Dynamic assembly of TRPC1-STIM1-Orai1 ternary complex is involved in store-operated calcium influx. Evidence for similarities in store-operated and calcium release-activated calcium channel components," *Journal of Biological Chemistry*, vol. 282, no. 12, pp. 9105–9116, 2007.
- [96] K. T. Cheng, X. Liu, H. L. Ong, and I. S. Ambudkar, "Functional requirement for Orai1 in store-operated TRPC1-STIM1 channels," *Journal of Biological Chemistry*, vol. 283, no. 19, pp. 12935–12940, 2008.
- [97] M. S. Kim, W. Zeng, J. P. Yuan, D. M. Shin, P. F. Worley, and S. Muallem, "Native store-operated  $Ca^{2+}$  influx requires the channel function of orai1 and TRPC1," *Journal of Biological Chemistry*, vol. 284, no. 15, pp. 9733–9741, 2009.
- [98] A. Coutaux, F. Adam, J. C. Willer, and D. le Bars, "Hyperalgesia and allodynia: peripheral mechanisms," *Joint Bone Spine*, vol. 72, no. 5, pp. 359–371, 2005.
- [99] R. A. Meyer, M. Ringkamp, J. N. Campbell, S. N. Raja, and N. R. Srinivasa, "Peripheral mechanisms of cutaneous nociception," in *Textbook of Pain*, W. A. Melzack's, Ed., Churchill Livingstone, 5th edition, 2006.
- [100] J. Ding, Y. Xiao, D. Lu, Y. R. du, X. Y. Cui, and J. Chen, "Effects of SKF-96365, a TRPC inhibitor, on melittin-induced inward current and intracellular  $Ca^{2+}$  rise in primary sensory cells," *Neuroscience Bulletin*, vol. 27, no. 3, pp. 135–142, 2011.
- [101] E. Clementi and J. Meldolesi, "Pharmacological and functional properties of voltagedependent  $Ca^{2+}$  channels," *Cell Calcium*, vol. 19, no. 4, pp. 269–279, 1996.
- [102] Y. M. Leung and C. Y. Kwan, "Current perspectives in the pharmacological studies of store-operated  $Ca^{2+}$  entry blockers," *Japanese Journal of Pharmacology*, vol. 81, no. 3, pp. 253–258, 1999.
- [103] A. Singer, M. Wonnemann, and W. E. Muller, "Hyperforin, a major antidepressant constituent of St. John's Wort, inhibits serotonin uptake by elevating free intracellular  $Na^{+}$ ," *Journal of Pharmacology and Experimental Therapeutics*, vol. 290, no. 3, pp. 1363–1368, 1999.
- [104] M. Wonnemann, A. Singer, and W. E. Muller, "Inhibition of synaptosomal uptake of 3H-L-glutamate and 3H-GABA by hyperforin, a major constituent of St. John's Wort: the role of amiloride sensitive sodium conductive pathways," *Neuropharmacology*, vol. 23, no. 2, pp. 188–197, 2000.
- [105] C. Harteneck and M. Gollasch, "Pharmacological modulation of diacylglycerol-sensitive TRPC3/6/7 channels," *Current Pharmaceutical Biotechnology*, vol. 12, no. 1, pp. 35–41, 2011.
- [106] X. Xu, I. Lozinskaya, M. Costell et al., "Characterization of small molecule TRPC3 and TRPC6 agonist and antagonists," *Biophysical Journal*, vol. 104, no. 2, article 454a, 2013.
- [107] H. Wang, X. Cheng, J. Tian et al., "TRPC channels: structure, function, regulation and recent advances in small molecular probes," *Pharmacology and Therapeutics*, p. 107497, 2020.
- [108] O. Tiapko, N. Shrestha, S. Lindinger et al., "Lipid-independent control of endothelial and neuronal TRPC3 channels by light," *Chemical Science*, vol. 10, no. 9, pp. 2837–2842, 2019.
- [109] S. Häfner, N. Urban, and M. Schaefer, "Discovery and characterization of a positive allosteric modulator of transient receptor potential canonical 6 (TRPC6) channels," *Cell Calcium*, vol. 78, pp. 26–34, 2019.
- [110] S. Kiyonaka, K. Kato, M. Nishida et al., "Selective and direct inhibition of TRPC3 channels underlies biological activities of a pyrazole compound," *Proceedings of the National*



- Academy of Sciences of the United States of America*, vol. 106, no. 13, pp. 5400–5405, 2009.
- [111] H. Schleifer, B. Doleschal, M. Lichtenegger et al., “Novel pyrazole compounds for pharmacological discrimination between receptor-operated and store-operated  $\text{Ca}^{2+}$  entry pathways,” *British Journal of Pharmacology*, vol. 167, no. 8, pp. 1712–1722, 2012.
- [112] K. Seo, P. P. Rainer, V. Shalkey Hahn et al., “Combined TRPC3 and TRPC6 blockade by selective small-molecule or genetic deletion inhibits pathological cardiac hypertrophy,” *Proceedings of the National Academy of Sciences of the United States of America*, vol. 111, no. 4, pp. 1551–1556, 2014.
- [113] B. L. Lin, D. Matera, J. F. Doerner et al., “In vivo selective inhibition of TRPC6 by antagonist BI 749327 ameliorates fibrosis and dysfunction in cardiac and renal disease,” *Proceedings of the National Academy of Sciences of the United States of America*, vol. 116, no. 20, pp. 10156–10161, 2019.
- [114] T. Maier, M. Follmann, G. Hessler et al., “Discovery and pharmacological characterization of a novel potent inhibitor of diacylglycerol-sensitive TRPC cation channels,” *British Journal of Pharmacology*, vol. 172, no. 14, pp. 3650–3660, 2015.
- [115] K. Motoyama, T. Nagata, J. Kobayashi et al., “Discovery of a bicyclo[4.3.0]nonane derivative DS88790512 as a potent, selective, and orally bioavailable blocker of transient receptor potential canonical 6 (TRPC6),” *Bioorganic & Medicinal Chemistry Letters*, vol. 28, no. 12, pp. 2222–2227, 2018.
- [116] N. Urban, L. Wang, S. Kwiek, J. Rademann, W. M. Kuebler, and M. Schaefer, “Identification and validation of larixyl acetate as a potent TRPC6 inhibitor,” *Molecular Pharmacology*, vol. 89, no. 1, pp. 197–213, 2016.
- [117] Y. Akbulut, H. J. Gaunt, K. Muraki et al., “(-)-Englerin a is a potent and selective activator of TRPC4 and TRPC5 calcium channels,” *Angewandte Chemie International Edition*, vol. 54, no. 12, pp. 3787–3791, 2015.
- [118] H. N. Rubaiy, M. J. Ludlow, K. Siems et al., “Tonantzitolone is a nanomolar potency activator of transient receptor potential canonical 1/4/5 channels,” *British Journal of Pharmacology*, vol. 175, no. 16, pp. 3361–3368, 2018.
- [119] A. Minard, C. C. Bauer, E. Chuntharpursat-Bon et al., “Potent, selective and subunit-dependent activation of TRPC5 channels by a xanthine derivative,” *British Journal of Pharmacology*, vol. 176, no. 20, pp. 3924–3938, 2019.
- [120] H. Beckmann, J. Richter, K. Hill, N. Urban, H. Lemoine, and M. Schaefer, “A benzothiadiazine derivative and methylprednisolone are novel and selective activators of transient receptor potential canonical 5 (TRPC5) channels,” *Cell Calcium*, vol. 66, pp. 10–18, 2017.
- [121] H. N. Rubaiy, M. J. Ludlow, R. S. Bon, and D. J. Beech, “Pico145 - powerful new tool for TRPC1/4/5 channels,” *Channels*, vol. 11, no. 5, pp. 362–364, 2017.
- [122] M. Miller, J. Shi, Y. Zhu et al., “Identification of ML204, a novel potent antagonist that selectively modulates native TRPC4/C5 ion channels,” *Journal of Biological Chemistry*, vol. 286, no. 38, pp. 33436–33446, 2011.
- [123] S. H. Sharma, J. L. Pablo, M. S. Montesinos, A. Greka, and C. R. Hopkins, “Design, synthesis and characterization of novel *N*-heterocyclic-1-benzyl-1*H*-benzo[*d*]imidazole-2-amines as selective TRPC5 inhibitors leading to the identification of the selective compound, AC1903,” *Bioorganic & Medicinal Chemistry Letters*, vol. 29, no. 2, pp. 155–159, 2019.
- [124] L. P. Yang, F. J. Jiang, G. S. Wu et al., “Acute treatment with a novel TRPC4/C5 channel inhibitor produces antidepressant and anxiolytic-like effects in mice,” *PLoS One*, vol. 10, no. 8, article e136255, 2015.
- [125] D. Vinayagam, T. Mager, A. Apelbaum et al., “Electron cryo-microscopy structure of the canonical TRPC4 ion channel,” *ELife*, vol. 7, 2018.
- [126] J. Duan, J. Li, G. L. Chen et al., “Cryo-EM structure of TRPC5 at 2.8-Å resolution reveals unique and conserved structural elements essential for channel function,” *Science Advances*, vol. 5, no. 7, article eaaw7935, 2019.