



Review Fluorescence-Guided Surgery for Gliomas: Past, Present, and Future

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Simple Summary: Glioblastoma is an aggressive brain tumor with a poor prognosis, partly due to the challenge of completely removing all tumor cells during surgery. Fluorescence-guided surgery (FGS) is a technique that helps surgeons see tumor cells more clearly by using special dyes that light up under certain types of light. This approach has improved how much of the tumor can be removed safely, which can lead to better patient outcomes. In this paper, we review the history, current practices, and future possibilities for using FGS in brain tumor surgery, with a focus on glioblastoma. We discuss the different fluorescent agents being used and the technologies that enhance their effectiveness. Our goal is to provide a comprehensive overview of FGS, highlight recent advancements, and explore how further innovations could expand its use in neurosurgery, potentially improving patient survival and quality of life.

Abstract: Background/Objectives: Glioblastoma (GBM) is the most common primary malignant central nervous system tumor, accounting for 50.9% of malignant CNS diagnoses and carrying a median survival of 15 months despite maximal standard therapy. High recurrence rates are driven by residual infiltrative tumor cells at the resection margin. Fluorescence-guided surgery (FGS) has emerged as a key innovation to improve intraoperative tumor visualization and maximize the extent of resection (EOR). This review examines the historical development, current clinical applications, and future directions of FGS in GBM surgery. Methods: A comprehensive literature review was conducted, covering the evolution of fluorophores (fluorescein, indocyanine green [ICG], and 5-aminolevulinic acid [5-ALA]), visualization technologies (wide- and narrow-field modalities), therapeutic adjuncts (photodynamic and sonodynamic therapies), and clinical adoption patterns and outcomes. Results: Early intraoperative fluorescence using fluorescein dates to 1947. ICG angiography has broad surgical utility, while 5-ALA received FDA approval in 2017, with phase III trials demonstrating gross total resection rates of 65% versus 36% with white-light surgery. Adjunct technologies-3D exoscopes, FGS-compatible loupes, and quantitative spectroscopy probes-enhance detection of residual tumor. Preliminary studies of intraoperative photodynamic and sonodynamic therapies show feasibility and potential survival benefits. Global adoption of 5-ALA FGS exceeds 75% among surveyed neurosurgeons.



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Copyright: © 2025 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons.org/ licenses/by/4.0/). **Conclusions**: FGS significantly improves EOR in GBM surgery, translating into better patient outcomes. Ongoing clinical trials and technological refinements—novel fluorophores, quantitative imaging, and therapeutic applications—promise to further optimize tumor visualization and treatment.

Keywords: fluorescence-guided surgery; glioblastoma; 5-ALA; intraoperative imaging; high-grade glioma

1. Introduction

Glioblastoma, the most common primary malignant brain tumor, encompasses 50.9% of all malignant central nervous system (CNS) diagnoses and has a median survival of 15 months with the current standard of care [1,2]. This prognosis has remained dismal over decades due to the high tumor-recurrence rate, which is driven by residual and/or resistant tumor cells in the infiltrative margin that are left behind after resection [3,4]. For this reason, maximizing the extent of resection (EOR) and consequent cytoreduction is an integral component of standard of care, with a demonstrated survival benefit [5,6]. There is therefore great interest in efforts to improve intraoperative visualization of glioblastoma, particularly at the infiltrative edge, in order to maximize EOR [7,8]. One of the most significant innovations in this realm is the advent of fluorescence-guided surgery (FGS), which has revolutionized intraoperative visualization and improved the extent of resection rates in patients with glioblastoma, the most common high-grade glioma (HGG) [9–12].

FGS uses photoactive agents called fluorophores to illuminate structures of interest during surgery. These fluorophores absorb a range of wavelengths and emit a specific wavelength, with these excitation and emission wavelengths typically in the visible to near-infrared (NIR) spectrum (500–900 nm) [13–15]. Fluorescence in this spectrum is utilized to provide contrast between a target of interest and surrounding tissue. Fluorescence guidance is used throughout the medical field, from identifying native hepatobiliary anatomy to photodynamic diagnosis of bladder cancer to assessing coronary artery bypass graft patency and perfusion. In neurosurgery, FGS is predominantly used within vascular neurosurgery (visualizing blood vessels, such as in aneurysm and bypass surgery) and neurosurgical oncology, in which it is now approved and standard of care for glioblastoma surgery.

As FGS continues to advance, distinguishing between clinically established and experimental applications is essential. Experimental applications generally include investigational usage in existing clinical trials, off-label applications, or novel technologies absent of wide-scale validation. Clinically established uses are interpreted as those with regulatory approval and demonstrated safety and efficacy in phase III clinical trials or diffuse clinical application. In this review, we discuss historical perspectives, current practices, and future directions for FGS in neurosurgical oncology, with a focus on glioblastoma.

2. Historical Perspective

The use of fluorescence in the medical sciences began in the late 19th century, with the word "fluorescence" first entering the lexicon in 1852 [16,17]. The first fluorescent stain, fluorescein, was invented in 1871 [16,18]. Decades later, in 1947, it was discovered that fluorescein could be used to differentiate neoplastic and normal tissue during brain tumor surgery [19]. Intracranial lesions demonstrated this quality most consistently, with tumor tissue emitting robust yellow fluorescence upon exposure to ultraviolet (UV) light [19]. Unsurprisingly, neurosurgery became the first surgical field to explore the utility of intraoperative fluorescence when, the following year, neurosurgeons at the University of

Minnesota Medical School administered intravenous fluorescein to patients undergoing craniotomy for suspected brain tumors [20]. They reported that the fluorescing tissue was indeed found to be neoplastic upon neuropathological evaluation in 44 of 46 patients [21]. In the decades since, various fluorophores have been developed and studied for application in FGS, the most popular today being 5-aminolevulinic acid hydrochloride (5-ALA), indocyanine-green (ICG), and fluorescein. While 5-ALA is the only Food and Drug Administration (FDA)-approved agent specifically for glioma surgery in the United States [12,22], ICG and fluorescein are FDA approved for use in ophthalmologic and vascular procedures but remain off-label for neurosurgical tumor resection. In Europe, fluorescein has received CE marking and is more widely adopted for glioma surgery. The study that galvanized the eventual approval of 5-ALA was a landmark multicenter, phase III clinical trial from Dusseldorf, Germany, using 5-ALA-guided resection of malignant gliomas in 270 adult patients [12,23]. The control group was standard microsurgery using the optical microscope with white light, and the primary endpoint was the number of patients with residual enhancing tumor after resection. The study found that the tumor was resected completely (no contrast-enhancing residual tumor at 72 h post-resection) in 65% of the 5-ALA group compared to 36% of the control group. A timeline of key historical events in the advancement of FGS for neurosurgery is provided in Figure 1.

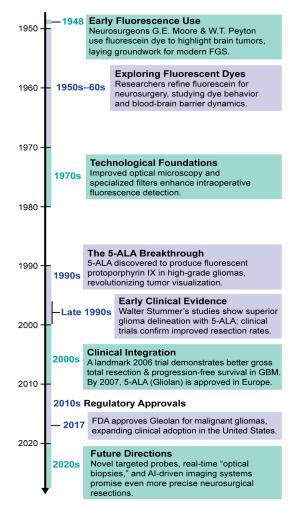


Figure 1. Key historical milestones in the development of fluorescence-guided surgery (FGS) in neurosurgery. The timeline highlights pivotal advancements, including the introduction of 5-ALA and fluorescein sodium, as well as major regulatory approvals and technological innovations that have shaped the clinical adoption of FGS techniques.

3. Fluorophores

3.1. Indocyanine Green (ICG)

ICG is the most commonly used fluorophore in surgery [20,24]. It is a water-soluble compound that is delivered intravenously in a reconstituted aqueous solution that binds serum albumin at the hydrophobic alpha- and beta-lipoprotein molecules [25–28]. These properties make ICG a great candidate for angiography as it distributes evenly and rapidly in the blood [29,30]. ICG is excited by wavelengths in the range of 750–800 nm, and it emits a light that peaks at 832 nm [29,31].

ICG was originally employed as a quantitative measurement of hepatic and cardiac function [32,33]. The fluorophore was used to quantify the amount of albumin in the serum of patients by measuring the decay of fluorescence. Albumin-bound ICG takes longer to decay; therefore, faster decay of fluorescence represents lower serum albumin levels [32]. ICG was later adopted into ophthalmology for infrared fundus angiograms and also as an intraoperative contrast agent to assist in performing vitrectomies to treat macular holes [34–37].

In today's medical practice, ICG is used in angiograms throughout the body. In general surgery, it is used to evaluate the integrity of surgical anastomosis in esophageal, gastric, small bowel, and colorectal surgery [38–43]. In neurosurgery, ICG is most commonly used as a contrast agent for ICG angiography (ICGA) during aneurysm surgery and to illuminate blood vessels during tumor surgery [44,45]. An example of cutting-edge ICG technology is the SPY Elite System (Stryker, Kalamazoo, MI, USA), a handheld, laser-assisted ICGA that has been used to monitor reperfusion of skin flaps in plastic surgery and track sentinel lymph nodes for breast cancer [46–49]. The University of Pennsylvania has also been productive in the ICG space, especially in the development of the second window technique, which will be described in depth below [50].

3.2. Fluorescein

Fluorescein sodium dye is an organic compound that is readily soluble in water and most often used in ophthalmology [51–53]. The aqueous solution is excited by light in the cobalt visible spectrum (465–490 nm) and fluoresces as bright green in the 520–530 nm range [54]. In ophthalmology, fluorescein can be applied directly to the eye using a stained paper strip; however, it is typically administered orally or intravenously [55].

Fluorescein dye was first used in humans to perform ophthalmological angiograms in 1959 [56]. Since then, the use of fluorescein in ophthalmology has expanded to assist in the "tear breakup time test" to quantify dry eyes, but it is still most applicable for retinal vasculature angiography [52,53]. In other disciplines of medicine, Fluorescein has been adopted to provide an assist in intraoperative imaging of peripheral nerves and colorectal epithelium [57,58]. Diagnostically, it is used to quantify occlusion of myocardial microcirculation for evaluation of CAD [59].

As far as its use in neurosurgery, intrathecal fluorescein is used to identify and localize CSF leaks during skull base surgeries or in the case of traumatic CSF leaks [60]. Additionally, it can be used for the identification of high-grade gliomas, although 5-aminolevulinic acid is more sensitive and specific. Lastly, fluorescein has been used to assess cerebral blood flow during aneurysm clipping, AVM resection, and other neurovascular procedures [61].

3.3. 5-Aminolevulinic Acid (5-ALA)

5-ALA is the newest fluorophore in clinical use and became FDA-approved for neurosurgical applications in 2017 [62,63]. 5-ALA is an amino acid that occurs naturally and serves as a precursor to heme and protoporphyrin IX (PpIX) [64]. The accumulation of PpIX in neoplastic tissue is directly responsible for the fluorescent capabilities of 5-ALA.

Multiple studies have confirmed the high diagnostic accuracy of 5-ALA-induced fluorescence in HGGs. The accumulation of PpIX in HGGs results in an unprecedented high sensitivity and positive predictive value of fluorescence visualization correlating with tumor tissue [65–67]. One multicenter study completed in the US confirmed a positive predictive value of over 96% [65]. Initial use of 5-ALA as a fluorophore in glioma surgery was pioneered by Stummer et al. in 1998, demonstrating intraoperative tumor fluorescence in malignant gliomas [68]. This foundational work led to a pivotal multicenter, randomized Phase III trial published in 2006, in which 5-ALA-guided surgery achieved complete resection of contrast-enhancing tumor in 65% of patients. The 5-ALA group also experienced significantly improved 6-month progression-free survival (41% vs. 21%) [12]. Since then, FGS with 5-ALA has gained considerable popularity among neurosurgeons and is now the standard of care for glioblastoma surgery at many centers around the world [63].

5-ALA is administered orally and absorbed through the gastrointestinal tract into the bloodstream; it is rapidly metabolized in brain tumors, where it is converted to PpIX in the heme biosynthesis pathway [69]. PpIX accumulates within brain tumor cells and is excited by blue light (400 nm range), leading to red-violet fluorescence at 635 nm and 704 nm emission peaks [69,70]. Intraoperatively, normal brain parenchyma appears blue, while the PpIX accumulated in the solid tumor bulk tissue appears red and the tumor margin pink [71]. In addition to the original Stummer study, multiple other studies have confirmed the greater ability to more completely resect tumors. In a meta-analysis performed by Ejamel et al., a gross total resection rate (removal of at least 98% of contrast-enhancing tumor) of 75.4% (418/565 patients) was achieved with 5-ALA FGS [72]. More recently, a randomized multicenter trial in France confirmed in a comparison between 5-ALA FGS and conventional microsurgery that greater overall tumor resection could be performed with 5-ALA FGS in GBM patients [73]. These data highlight the utility of 5-ALA FGS in the resection of HGG and overall better experiences in the OR for neurosurgeons.

5-ALA is also being implemented in fields other than neurosurgery, including in other oncologic surgeries for dermatology, head and neck, gynecology oncology, and orthopedics. Filip et al. described the use of 5-ALA for resection of head and neck squamous cell carcinoma [74]. Bickels et al. demonstrated in a study of 24 patients that 5-ALA can help decrease the likelihood of tumor recurrence in the surgical removal of desmoid tumors, solitary fibrous tumors, and dermatofibrosarcoma protuberans [75]. There are also promising preclinical in vivo models studying the effectiveness of 5-ALA in bone cancers such as chondroblastoma [76]. An overview of all described fluorophores can be found within Table 1.

Table 1. Key Fluorophores Used in Glioma Surgery.

Fluorophore	Tumor Specificity	Depth of Penetration	GTR Rate	PFS (Months)	OS (Months)	Cost	Key Limitations
5-ALA	High	~1 to 2 mm	65–75%	6.8	15–20	\$\$\$	Phototoxicity, limited depth
Fluorescein	Moderate	~1 mm	~60 to 70%	~9.2	~15	\$	Non-specific, absorption interference
ICG (SWIG)	Low- Moderate	Deep (NIR, ~5 to 10 mm)	Not well defined	N/A	N/A	\$\$	Non-specific uptake, experimental

ICG: Indocyanine Green; SWIG: Second-Window Indocyanine Green; GTR: Gross Total Resection; PFS: Progression-Free Survival; OS: Overall Survival. Summary of key fluorophores used in glioma surgery, comparing regulatory status, optical characteristics, resection outcomes, and clinical limitations. Data reflect reported ranges from major trials and meta-analyses where available. The relative costliness of each compound is described as low (\$), moderate (\$\$), or high (\$\$\$).

4. Practical Clinical Application

Since its FDA approval in 2017, 5-ALA has been regularly implemented in the resection of HGG in the US. 5-ALA is administered orally anywhere from 2 to 4 h prior to surgery at a dose of 20 mg/kg body weight [12]. This provides ample time for the preoperative setup, anesthesia, and craniotomy so that peak intraoperative fluorescence will occur during tumor resection. The dosing is based upon experiments in rodents in which the peak fluorescence was observed 6 h after administration [77]. While Stummer et al. demonstrated effective administration and strong fluorescence with their protocol, Kaneko et al. found in a prospective study of 68 patients that maximal fluorescence was observed 7 to 8 h after administration of 5-ALA, with weaker fluorescence lasting as long as 8 to 9 h [12,78]. Others have also confirmed that 5-ALA tumor fluorescence may be better more, than 4 h after administration, with tumor fluorescence reported up to 24 h after administration [79]. The 2 to 4 h time window is the FDA-approved administration criterion, but it is important to further investigate if peak fluorescence extends further so that intraoperative fluorescence can be utilized despite inevitable logistical issues and delays in surgery [80]. Regardless, 5-ALA provides a suitable window for surgeons to clearly visualize tumor tissue in patients with HGG.

The intensity of red fluorescence correlates with the density of tumor cells [71]. The neurosurgeon then decides how much tissue can be safely resected. 5-ALA has excellent sensitivity for detecting tumors, but it has slightly lower specificity; in some cases, fluorescent tissue may therefore contain both tumor tissue and functional brain tissue. Moreover, fluorescence penetration can be limited to approximately 1–2 mm, and signal intensity can vary due to tumor heterogeneity. False positives may occur, particularly in areas of necrosis or inflammation. As a result, surgeons must therefore use clinical judgment in using FGS to guide resections, along with other intraoperative adjuncts such as neurophysiological monitoring [63].

While fluorescence-guided surgery with 5-ALA and other agents is generally well tolerated, it is important to be aware of associated adverse effects. 5-ALA can cause photosensitivity reactions, requiring patients to avoid direct sunlight or strong indoor lighting for 24 h postoperatively. Fluorescein has been linked to mild allergic responses and can cause temporary skin or urine discoloration [21]. Indocyanine green, although rare, has been associated with anaphylactic reactions, particularly in iodine-sensitive individuals [30]. While the incidence of serious adverse events is low, preoperative screening and patient counseling remain essential components of safe clinical practice.

4.1. Complementary Technologies

Since the FDA approval of 5-ALA for brain tumor resection, several technologies have been developed to complement and improve the capabilities of FGS. Fluorophore visualization utilizes a short-pass filter (~400 nm) to generate light to excite the fluorophore and a long-pass filter (~650 nm) to detect emitted light from the fluorophore [73,81]. The technologies that assist in the visualization of fluorescence can be widely separated into two field of view (FOV) categories: wide field of view (WFOV) and narrow field of view (NFOV) [82]. WFOV refers to technologies that illuminate the entire surgical cavity and broadly detect the emission of the fluorophore. This category includes the widely used visualization technologies like optical microscopes, loupes, and exoscopes, but with added filters. NFOV illumination focuses on a smaller field, which enhances resolution and allows for quantitative measurement of fluorescence at a cellular scale [83–88].

Optical microscopes were the first technology outfitted for compatibility with fluorescence, as described by Stummer and colleagues when they used an optical microscope with an excitation filter at 375–440 nm and an emission filter at >455 nm to visualize malignant gliomas with 5-ALA. More recently, the three-dimensional (3D) exoscope has gained popularity in the neurosurgical operating room (OR). Exoscopes are 3D high-definition camera systems that project onto a monitor in a neurosurgical OR to provide the entire staff with a visual of the surgical field [89–94]. The exoscope provides superior magnification and tumor tissue clarity with the use of LED lighting and a longer working distance and depth of field in comparison to the conventional operating microscope. The use of the exoscope for resection of GBM tumors has been reported with a high extent of tumor resections achieved [95]. These systems also have FGS compatibility to excite 5-ALA, fluorescein, and ICG [96,97]. The exoscope may have both visualization and ergonomic benefits; Dell Pepa et al. recorded ten procedures, five with an exoscope and five with an optical microscope [98]. They found that exoscopes provided superior visualization of vessels, parenchyma, surgical instruments, and fluorescence under the 5-ALA blue filters compared to the optical microscope. The exoscope was also better integrated into surgeon workflows, with less frequent blue-to-white light switching as compared to the optical microscope.

The last of the WFOV devices are FGS-compatible loupes. Multiple commercial systems exist. The REVEAL (Designs for Vision) loupes are equipped with a TriBeam light source that combines a light source with 420 nm and 475 nm filters for excitation and emission of 5-ALA [99,100]. Giatini-Larsen et al. presented a comparison of visualizing fluorescence in three patients using three different modalities: a blue flashlight with an optical microscope, a low-cost headlamp with an optical microscope, and the FGS-compatible loupes alone. The team found fluorescence with all three modalities, but the loupes outperformed the microscope in terms of visualization capabilities [100]. Zhang et al. recorded their experience using the FGS-compatible loops on a cohort of 11 consecutive patients and rated their experience with the loupes as excellent, without any difficulties incorporating the loupes into their workflow [99].

In contrast to WFOV visualization, LFOV optics can detect light emitted in a much smaller pixel, even down to a cellular level [82,101]. Intuitively, this higher resolution protects against the distorting effects of photobleaching from surrounding light and scattering effects off of tissue [86]. More importantly, it allows for a quantitative measurement of fluorescence as opposed to the subjective visual interpretation of the surgeon [66,84–88,102]. The use of fluorescence spectrometry to quantify the emission of fluorescence was pioneered by Diamond et al., who, building off of previous work defining a photon migration model to quantify fluorescence, developed a single-fiber optic probe for both light source and collection [103–105]. Similarly to WFOV, NFOV visualization relies on a blue-light excitation wavelength that is coupled with a broadband light sensor to detect fluorescent emission. However, with NFOV, both of these are localized on a much smaller probe [84].

Since their invention, these probes have been adopted by neurosurgeons for glioma surgery. Stummer and team used the probe to perform "fluorescence biopsies" in human subjects with HGG, quantifying the fluorescence of tissue as either "strong" or "weak" fluorescence prior to resection; the specimens were then histo-pathologically evaluated [66]. The group found that "strong" fluorescence had a 100% positive predictive value for tumor and a strong correlation to higher tumor cell density. "Weak" fluorescence had a 95% positive predictive value for tumor and a strong correlation to medium-to-low tumor cell density. Haj-Hosseini et al. developed a spectroscopy system with pulsed modulation to quantify fluorescence in the resection cavity and at the tumor margin before finishing HGG resection procedures [85]. During tumor resections, they used the spectroscopy system to survey residual malignant cells and were able to quantitatively detect 5-ALA fluorescence in the surgical cavity.

The recent advancements in complementary visualization technologies, whether it is WFOV or NFOV, have served to increase surgeon adoption of FGS and improve patient outcomes. An overview of key intraoperative visualization technology is provided in Figure 2.

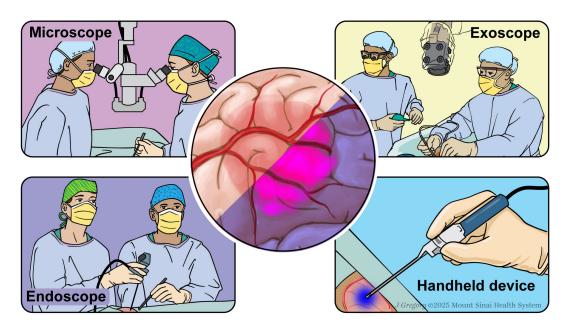


Figure 2. Contemporary applications of fluorescence-guided surgery (FGS) in neurosurgery. The illustration highlights the primary visualization platforms used intraoperatively, including surgical microscopes, exoscopes, endoscopes, and handheld devices. These modalities enable real-time identification of fluorescent tumor tissue, enhancing surgical precision and resection outcomes.

4.2. Therapeutic Applications

In addition to visualization aiding technologies, fluorophores have been incorporated into intraoperative therapies, such as photodynamic therapy (PDT) and sonodynamic therapy (SDT). These interventions capitalize on the reactive nature of PpIX to generate reactive oxygen species and kill cancer cells in the brain [106,107]. Therapeutic-oriented trials are examining possible targeted therapeutic effects of fluorophores when combined with light or low-intensity-focused ultrasound therapy (in PDT and SDT). In the case of intraoperative PDT, a different wavelength of light (635 nm) is administered to activate the 5-ALA metabolite, PpIX, to create reactive oxygen species (ROS), which are toxic to the tumor cells [108,109]. A balloon is placed in the resection cavity at the time of surgery with a laser fiber inserted to illuminate the surgical site and target residual tumor cells. A recent study was completed in France utilizing 5-ALA for intraoperative PDT during glioblastoma surgery. The authors reported the feasibility, efficacy, and safety of this approach in combination with standard of care fractionated external beam radiotherapy with concomitant and adjuvant chemotherapy [110]. In their initial 10-patient study, they found no serious adverse events related to the PDT treatment and, more importantly, 40% of patients ended up surviving over 5 years [111].

With sonodynamic therapy (SDT), focused ultrasound is used to target the intracellular PpIX in tumor cells to generate ROS [112]. SDT does not require any open surgery but uses low-intensity-focused ultrasound waves that are applied through the skull. Current clinical studies are underway in the US evaluating the efficacy of 5-ALA SDT in the treatment of recurrent HGG [113–115]. SDT is also being investigated for diffuse intrinsic pontine glioma (DIPG) in young adults and children [116]. Two clinical trials exploring PDT for glioblastoma are available in the US [117,118]. One intraoperative PDT trial is open for enrollment at the University of Pittsburgh Medical Center for newly diagnosed GBM patients utilizing the new PDT agent known as Pentalafen. Taken together, there are multiple active and recruiting clinical trials examining the use of fluorophores for the actual therapy of brain tumors underway in the US.

4.3. Adoption and Outcomes

Following the European, Asian, Canadian, and US FDA approval in 2017, 5-ALA FGS has become a standard of care during the resection of HGG worldwide. Multiple studies have confirmed 5-ALA FGS to be safe and effective, with minimal side effects. While there is a noted learning curve in the use of 5-ALA FGS for neurosurgeons, there is no question that more complete resections can be performed during resection of HGG tumors [70].

A comprehensive review by Mansouri et al. highlighted the growing body of evidence supporting the use of 5-ALA in glioma surgery, particularly emphasizing the increasing integration of FGS into neurosurgical training programs, facilitating wider adoption [119]. Ferraro et al. conducted a survey of neurosurgeons across Europe and found that 5-ALA was used by 78% of respondents for HGG surgeries, indicating its widespread acceptance in clinical practice [120].

The increasing adoption of FGS has also spurred technological innovations to enhance its utility. Kaneko and Kumagai described the development of quantitative fluorescence imaging systems that can provide more objective assessments of tumor fluorescence, potentially improving surgical decision-making [121]. Additionally, Widhalm et al. reported on the successful integration of FGS with other advanced imaging modalities such as PET, further refining the surgical approach to gliomas [122].

This growing body of evidence and widespread clinical adoption highlight the utility of 5-ALA FGS in the resection of high-grade gliomas, leading to improved extent of resection and potentially better outcomes for patients. As research continues to expand and technology evolves, FGS is poised to remain a cornerstone in the neurosurgical management of gliomas.

5. Future Directions

5.1. Improved Performance of 5-ALA

As 5-ALA has gained popularity among neurosurgeons in the resection of HGG, it has more recently been applied to other tumors, including meningiomas and pediatric brain tumors [123]. In 2014, Valdes et al. demonstrated excellent utility of 5-ALA in 12 out of 15 patients with meningiomas, with accuracy as high as 90% in differentiating tumor tissue from normal brain [124]. More recently, Wadiura et al. examined the use of 5-ALA in 191 samples from 85 surgeries and found that 5-ALA had a positive predictive value for tumor of 100% [125]. A multicenter study in the US utilizing 5-ALA for meningioma resection has been completed in the US. In children, Milos et al. demonstrated 5-ALA-induced fluorescence in five patients with low-grade gliomas (LGGs) [126]. LGGs are known not to fluoresce as avidly as HGGs. Another ongoing phase II trial in Germany is investigating the application of 5-ALA in pediatric brain tumors [127]. Given the success of 5-ALA in the resection of glioblastoma in adults, we anticipate seeing more clinical studies on pediatric brain tumor resections using 5-ALA in the coming years.

5.2. Exploring Other Fluorophores

Despite its great successes, 5-ALA does have some limitations. Its visible-light emission (635 nm) is significantly absorbed by other endogenous fluorophores, like heme, which can cause interference when identifying tumor margins [22,128]. This flaw is being addressed by incorporating fluorophores with unique emission spectra that do not experience as much endogenous noise. The most promising of these fluorophores is ICG. ICG has an emission peak at 832 nm, which is within the near infrared (NIR) spectrum [29,31,129]. This allows for maximum penetration depth and minimum interference from serosanguinous fluid [129]. Second-Window-ICG (SWIG) is a technique that takes advantage of the permeability of endothelial tissue within peritumoral tissue. A large bolus of ICG is administered, and over time, the ICG leaks through the permeable tumor vasculature and accumulates in the tumor and peritumoral space [22]. The next day, surgery is performed and ICG is visualized under NIR light to identify tumor margins. Lee et al. utilized SWIG in 15 patients by administering 5 mg/kg of ICG 22 h before surgery and found that 12 out of 15 tumors could be visualized using an NIR camera [130]. Using pathology as the gold standard, SWIG ICG had a sensitivity of 98% and a specificity of 45% in identifying tumor tissue. In 2022, Karsalia et al. visualized the glioblastoma resection cavity using an exoscope and SWIG under NIR light to detect residual tumor [131]. The group posited that fluorescence visualized in the cavity would represent residual tumor cells, so all fluorescing tissue was resected. They found that compared to gadolinium-enhanced postoperative MRI scans, SWIG and NIR had a higher sensitivity (96%), a higher negative predictive value (89%), and higher accuracy (91%) in detecting residual tumor.

Fluorescein is another popular fluorophore that has been proposed as an alternative or complement to 5-ALA in glioma resection. Unlike ICG, fluorescein does not address the endogenous fluorophore absorption problem of 5-ALA. Fluorescein fluoresces in the yellow-green spectrum at 520–530 nm, which still faces interference from endogenous fluorophores. However, the benefits of fluorescein lie in its cost-effectiveness and lack of phototoxicity. 5-ALA can be cost-prohibitive and can exhibit phototoxicity and photobleaching in situ [132]. For these reasons, surgeons may choose to use fluorescein off-label in glioma resections [133,134]. This has led to studies into the comparison of effectiveness between 5-ALA and fluorescein. Hansen et al. compared 209 patients who underwent resection for HGGs with 5-ALA (n = 58 patients) or fluorescein (n = 51 patients) and found that there was no statistically significant difference between the two groups in extent of resection, the percent of patients with residual tumor volume less than 0.175 cm³, or the median overall survival [132]. The only significant outcome difference was progression-free survival, in which fluorescein outperformed 5-ALA (9.2 months vs. 8.7 months).

Fluorescein has also been tested in conjunction with 5-ALA to improve tumor margin visualization, with the rationale of using 5-ALA to target the tumor and fluorescein to target the peritumoral space [22]. This dual fluorescence has been explored in multiple studies. Molina et al. combined fluorescein and 5-ALA to better visualize gliomas in six patients and concluded that the yellow-green background of fluorescein improved visualization of 5-ALA, which led to improved delineation of the tumor margin [135]. Similarly, Schwake et al. (2015) reported that the dual-labeling approach enhanced the visibility of tumor tissue in areas where 5-ALA fluorescence was weak, particularly at the tumor margins [136]. These studies suggest that combining fluorescein and 5-ALA may provide complementary information, potentially leading to more precise tumor delineation and improved extent of resection.

Emerging molecular strategies are also exploring the use of antibody–fluorophore or antibody–drug conjugates to improve tumor specificity. These conjugates bind to tumor-specific antigens and may allow for precise delivery of either fluorescent markers or cytotoxic agents to glioma cells, enabling both visualization and targeted therapy. Recent preclinical studies have shown promise in targeting glioma-specific markers, suggesting future integration of these biologics with FGS platforms [137].

5.3. Current Clinical US Trials and Therapeutic Adjuncts

The FDA approval of 5-ALA led to many additional clinical trials for FGS, many of which are active and/or still recruiting (Table 2). The aim of these trials can be split into two categories: visualization-oriented trials and therapeutic-oriented trials. Several of these trials evaluate investigational uses of fluorophores in therapeutic settings, including early-phase studies of 5-ALA combined with sonodynamic therapy and novel agents such as Photobac and Pentalafen.

Title	Clinical Trial Number	Sponsor	Indication	Fluorescence Type	Enrollment	Phase	Company
Second Window Indocyanine Green for All Nervous System Tumors	NCT05746104	Abramson Cancer Center at Penn Medicine	All CNS Tumors	ICG	105	1	TumorGlow (Pennsylvania, USA)
Study to Evaluate 5-ALA Combined With CV01 Delivery of Ultrasound in Recurrent High-Grade Glioma	NCT05362409	Alpheus Medical, Inc.	HGG	5-ALA	48	1	Alpheus Medical, Inc. (Minnesota, USA)
Diagnostic Performance of Fluorescein as an Intraoperative Brain Tumor Biomarker	NCT02691923	David W. Roberts, Dartmouth- Hitchcock Medical Center	HGG & LGG	Fluorescein	30	2	-
Sonodynamic Therapy in Patients With Recurrent GBM	NCT06039709	Shayan Moosa, MD, University of Virginia	rGBM	5-ALA	11	1	-
A Study of Sonodynamic Therapy Using SONALA-001 and Exablate 4000 Type 2.0 in Subjects With Recurrent GBM	NCT05370508	SonALAsense, Inc.	rGBM	5-ALA	44	1 and 2	SonALAsense, Inc. (California USA)
A Phase 2 Study of Sonodynamic Therapy Using SONALA-001 and Exablate 4000 Type 2.0 in Patients With DIPG	NCT05123534	SonALAsense, Inc.	DIPG	5-ALA	27	1 and 2	SonALAsense, Inc.
ALA-Induced PpIX Fluorescence During Brain Tumor Resection	NCT02191488	David W. Roberts, Dartmouth- Hitchcock Medical Center	HGG, LGG, rHGG, Mets, Meningioma	5-ALA	540	1	-
Study of Sonodynamic Therapy in Participants With Recurrent High-Grade Glioma	NCT04559685	Nader Sanai, St. Joseph's Hospital and Medical Center, Phoenix	rHGG	5-ALA	30	1	-
The Role of 5-Aminolevulinic Acid Fluorescence-Guided Surgery in Head and Neck Cancers: a Pilot Trial	NCT05101798	Alfred-Marc Iloreta, Icahn School of Medicine at Mount Sinai	Recurrent Head, Neck, or Skull Base	5-ALA	26	2	-
Loupe-Based Intraoperative Fluorescence Imaging	NCT04780009	Guoqiang Yu, University of Kentucky	GBM and AA HGG, GBM,	Fluorescein and 5-ALA	30	Observat	ional -
Evaluation of the CONVIVO System	NCT05139277	Linton T. Evans, Dartmouth- Hitchcock Medical Center	Mets, Meningioma, Acoustic Neuroma, Pituitary Adenoma	Fluorescein	30	Pre- Clinical	Zeiss (Oberkochen, Germany)
Intracavitary Photodynamic Therapy as an Adjuvant to Resection of Glioblastoma or Gliosarcoma Using IV Photobac	NCT05363826	Photolitec LLC	GBM Gliosarcoma	Photobac	30	1	Photolitec LLC (New York, USA)
A Dose-escalation Clinical Study of Intraoperative Photodynamic Therapy of Glioblastoma	NCT05736406	Hemerion Therapeutics	Newly diagnosed GBM	Pentalafen	12	1	Hemerion Therapeutics (Villeneuve- d'Ascq, France)

Table 2. Ongoing and Recent Clinical Trials Involving Intraoperative Fluorescence Imaging and Therapy in Brain and Head/Neck Tumors.

A summary of the key clinical trials evaluating various intraoperative fluorescence agents and technologies that include ICG, 5-ALA, and fluorescein in the surgical management of nervous system tumors. Trials are listed by indication, fluorescence type, sponsor, enrollment size, and phase.

Visualization-oriented trials are seeking to expand indications for FGS. One clinical trial at the Icahn School of Medicine at Mount Sinai is exploring the use of 5-ALA in treating head and neck cancers [138]. At Dartmouth-Hitchcock Medical Center, there is a study investigating the use of 5-ALA in multiple primary and metastatic brain tumors including HGG, recurrent HGG, LGG, meningioma, and brain metastases [139]. There are also many clinical trials considering other fluorophores. A group at the University of Pennsylvania is testing the efficacy of SWIG in all patients with CNS tumors [140]. At Dartmouth-Hitchcock Medical Center, fluorescein is being evaluated in HGG and LGG [141]. Additionally, there are multiple visualization technologies that are currently being examined in clinical trials. A recently completed clinical trial at the University of Pittsburgh utilized a modified NICO® Myriad handpiece (Stryker Corp., Kalamazoo, MI, USA) with a blue-light attachment for better visualization of fluorescent tumor tissue that is correlated with histopathology from tissue specimens harvested at the time of surgery. The NICO handpiece known as the SPECTRA was utilized in combination with the operative microscope or exoscope. The CONVIVO confocal endomicroscope is being studied in vivo for identifying HGG tumor tissue type using fluorescein [142]. Finally, bioptic loupes are being tested against optical microscopes for their ability to accurately identify fluorescing tumor tissue [143].

Artificial intelligence (AI) may soon augment fluorescence-guided surgery by automating intraoperative decision-making. Machine learning algorithms trained on intraoperative imaging and histopathologic datasets could assist in delineating tumor margins, identifying subtle fluorescence, or predicting tumor infiltration zones in real time. Early research supports the feasibility of integrating AI into fluorescence-based workflows [144].

6. Conclusions

FGS is a promising innovation within neurosurgical oncology for enhancing intraoperative visualization to improve the extent of resection and patient outcomes for glioma surgery. Adoption has increased dramatically in recent years and appears poised to continue increasing across centers doing brain tumor surgery. Ongoing trials hold promise for expanding indications for FGS as well as evaluating the possible therapeutic efficacy of fluorophores when used in combination with acoustic and light energy.

Despite growing clinical adoption in high-resource settings, global implementation of FGS remains limited by several barriers. These include the high cost of agents like 5-ALA, limited access to compatible imaging equipment in low- and middle-income countries (LMICs), and a lack of formalized training in fluorescence-based neurosurgical techniques. Overcoming these disparities will require coordinated global efforts, including the development of cost-effective portable imaging platforms and international training programs that leverage virtual surgical education and mentorship.

As research continues to expand and technology evolves, FGS is poised to remain a cornerstone in the neurosurgical management of gliomas. Its integration into standard practice reflects not only its clinical utility but also its potential to transform brain tumor surgery into a more precise, data-driven, and patient-tailored discipline. With continued innovation and investment, fluorescence-guided techniques hold the promise to elevate surgical outcomes for patients around the world.

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References

- Ostrom, Q.T.; Price, M.; Neff, C.; Cioffi, G.; Waite, K.A.; Kruchko, C.; Barnholtz-Sloan, J.S. CBTRUS Statistical Report: Primary Brain and Other Central Nervous System Tumors Diagnosed in the United States in 2016–2020. *Neuro Oncol.* 2023, 25, iv1–iv99. [CrossRef] [PubMed]
- Stupp, R.; Mason, W.P.; van den Bent, M.J.; Weller, M.; Fisher, B.; Taphoorn, M.J.B.; Belanger, K.; Brandes, A.A.; Marosi, C.; Bogdahn, U.; et al. Radiotherapy plus Concomitant and Adjuvant Temozolomide for Glioblastoma. *N. Engl. J. Med.* 2005, 352, 987–996. [CrossRef] [PubMed]
- Chen, J.; Li, Y.; Yu, T.-S.; McKay, R.M.; Burns, D.K.; Kernie, S.G.; Parada, L.F. A Restricted Cell Population Propagates Glioblastoma Growth after Chemotherapy. *Nature* 2012, 488, 522–526. [CrossRef] [PubMed]
- Glas, M.; Rath, B.H.; Simon, M.; Reinartz, R.; Schramme, A.; Trageser, D.; Eisenreich, R.; Leinhaas, A.; Keller, M.; Schildhaus, H.-U.; et al. Residual Tumor Cells Are Unique Cellular Targets in Glioblastoma. *Ann. Neurol.* 2010, 68, 264–269. [CrossRef]
- Oppenlander, M.E.; Wolf, A.B.; Snyder, L.A.; Bina, R.; Wilson, J.R.; Coons, S.W.; Ashby, L.S.; Brachman, D.; Nakaji, P.; Porter, R.W.; et al. An Extent of Resection Threshold for Recurrent Glioblastoma and Its Risk for Neurological Morbidity. *J. Neurosurg.* 2014, 120, 846–853. [CrossRef]
- Chaichana, K.L.; Zadnik, P.; Weingart, J.D.; Olivi, A.; Gallia, G.L.; Blakeley, J.; Lim, M.; Brem, H.; Quiñones-Hinojosa, A. Multiple Resections for Patients with Glioblastoma: Prolonging Survival. *J. Neurosurg.* 2013, 118, 812–820. [CrossRef]
- Schupper, A.J.; Hadjipanayis, C.G. Novel Approaches to Targeting Gliomas at the Leading/Cutting Edge. J. Neurosurg. 2023, 139, 760–768. [CrossRef]
- 8. Jiang, S.; Chai, H.; Tang, Q. Advances in the Intraoperative Delineation of Malignant Glioma Margin. *Front. Oncol.* **2023**, *13*, 1114450. [CrossRef]
- Molinaro, A.M.; Hervey-Jumper, S.; Morshed, R.A.; Young, J.; Han, S.J.; Chunduru, P.; Zhang, Y.; Phillips, J.J.; Shai, A.; Lafontaine, M.; et al. Association of Maximal Extent of Resection of Contrast-Enhanced and Non-Contrast-Enhanced Tumor With Survival Within Molecular Subgroups of Patients With Newly Diagnosed Glioblastoma. *JAMA Oncol.* 2020, *6*, 495–503. [CrossRef]
- Chohan, M.O.; Berger, M.S. 5-Aminolevulinic Acid Fluorescence Guided Surgery for Recurrent High-Grade Gliomas. J. Neurooncol. 2019, 141, 517–522. [CrossRef]
- 11. Acerbi, F.; Cavallo, C.; Broggi, M.; Cordella, R.; Anghileri, E.; Eoli, M.; Schiariti, M.; Broggi, G.; Ferroli, P. Fluorescein-Guided Surgery for Malignant Gliomas: A Review. *Neurosurg. Rev.* **2014**, *37*, 547–557. [CrossRef] [PubMed]
- Stummer, W.; Pichlmeier, U.; Meinel, T.; Wiestler, O.D.; Zanella, F.; Reulen, H.-J.; ALA-Glioma Study Group. Fluorescence-Guided Surgery with 5-Aminolevulinic Acid for Resection of Malignant Glioma: A Randomised Controlled Multicentre Phase III Trial. *Lancet Oncol.* 2006, 7, 392–401. [CrossRef] [PubMed]
- 13. Richards-Kortum, R.; Sevick-Muraca, E. Quantitative Optical Spectroscopy for Tissue Diagnosis. *Annu. Rev. Phys. Chem.* **1996**, 47, 555–606. [CrossRef] [PubMed]
- 14. Hilderbrand, S.A.; Weissleder, R. Near-Infrared Fluorescence: Application to in Vivo Molecular Imaging. *Curr. Opin. Chem. Biol.* **2010**, 14, 71–79. [CrossRef]
- 15. Chance, B. Near-Infrared Images Using Continuous, Phase-Modulated, and Pulsed Light with Quantitation of Blood and Blood Oxygenation. *Ann. N. Y. Acad. Sci.* **1998**, *838*, 29–45. [CrossRef]
- 16. Wollman, A.J.M.; Nudd, R.; Hedlund, E.G.; Leake, M.C. From Animaculum to Single Molecules: 300 Years of the Light Microscope. *Open Biol.* **2015**, *5*, 150019. [CrossRef]
- 17. Stokes, G.G. XXX. On the Change of Refrangibility of Light. Philos. Trans. R. Soc. Lond. 1997, 142, 463–562.
- 18. Baeyer, A. Ueber Eine Neue Klasse von Farbstoffen. Ber. Dtsch. Chem. Ges. 1871, 4, 555–558. [CrossRef]
- 19. Moore, G.E. Fluorescein as an Agent in the Differentiation of Normal and Malignant Tissues. Science 1947, 106, 130–131. [CrossRef]
- 20. Nagaya, T.; Nakamura, Y.A.; Choyke, P.L.; Kobayashi, H. Fluorescence-Guided Surgery. Front. Oncol. 2017, 7, 314. [CrossRef]
- 21. Moore, G.E.; Peyton, W.T. The Clinical Use of Fluorescein in Neurosurgery; the Localization of Brain Tumors. *J. Neurosurg.* **1948**, *5*, 392–398. [CrossRef] [PubMed]

- 22. Cho, S.S.; Salinas, R.; Lee, J.Y.K. Indocyanine-Green for Fluorescence-Guided Surgery of Brain Tumors: Evidence, Techniques, and Practical Experience. *Front. Surg.* **2019**, *6*, 11. [CrossRef] [PubMed]
- 23. Medac GmbH. Fluorescence-Guided Resection of Malignant Gliomas with 5-Aminolevulinic Acid. Available online: https://clinicaltrials.gov/study/NCT00241670 (accessed on 21 August 2024).
- 24. Sutton, P.A.; van Dam, M.A.; Cahill, R.A.; Mieog, S.; Polom, K.; Vahrmeijer, A.L.; van der Vorst, J. Fluorescence-Guided Surgery: Comprehensive Review. *BJS Open* **2023**, *7*, zrad049. [CrossRef] [PubMed]
- DailyMed. INDOCYANINE GREEN Kit INDOCYANINE GREEN Injection, Powder, Lyophilized, for Solution. Available online: https://dailymed.nlm.nih.gov/dailymed/lookup.cfm?setid=c1031b17-b6e3-4153-869f-30d9c7e66bc3 (accessed on 21 August 2024).
- 26. Engel, E.; Schraml, R.; Maisch, T.; Kobuch, K.; König, B.; Szeimies, R.-M.; Hillenkamp, J.; Bäumler, W.; Vasold, R. Light-Induced Decomposition of Indocyanine Green. *Investig. Ophthalmol. Vis. Sci.* **2008**, *49*, 1777–1783. [CrossRef]
- 27. Desmettre, T.; Devoisselle, J.M.; Mordon, S. Fluorescence Properties and Metabolic Features of Indocyanine Green (ICG) as Related to Angiography. *Surv. Ophthalmol.* **2000**, *45*, 15–27. [CrossRef]
- 28. Landsman, M.L.; Kwant, G.; Mook, G.A.; Zijlstra, W.G. Light-Absorbing Properties, Stability, and Spectral Stabilization of Indocyanine Green. J. Appl. Physiol. 1976, 40, 575–583. [CrossRef]
- 29. Alander, J.T.; Kaartinen, I.; Laakso, A.; Pätilä, T.; Spillmann, T.; Tuchin, V.V.; Venermo, M.; Välisuo, P. A Review of Indocyanine Green Fluorescent Imaging in Surgery. *Int. J. Biomed. Imaging* **2012**, 2012, 940585. [CrossRef]
- 30. Reinhart, M.B.; Huntington, C.R.; Blair, L.J.; Heniford, B.T.; Augenstein, V.A. Indocyanine Green: Historical Context, Current Applications, and Future Considerations. *Surg. Innov.* **2016**, *23*, 166–175. [CrossRef]
- Mordon, S.; Devoisselle, J.M.; Soulie-Begu, S.; Desmettre, T. Indocyanine Green: Physicochemical Factors Affecting Its Fluorescence in Vivo. *Microvasc. Res.* 1998, 55, 146–152. [CrossRef]
- 32. Cherrick, G.R.; Stein, S.W.; Leevy, C.M.; Davidson, C.S. Indocyanine Green: Observations on Its Physical Properties, Plasma Decay, and Hepatic Extraction. *J. Clin. Investig.* **1960**, *39*, 592–600. [CrossRef]
- Burchell, H.B. Assessment of Clinical Value: Symposium on Diagnostic Applications of Indicator-Dilution Technics. Proc. Staff Meet. Mayo Clin. 1957, 32, 551–553. [PubMed]
- 34. Kogure, K.; Choromokos, E. Infrared Absorption Angiography. J. Appl. Physiol. 1969, 26, 154–157. [CrossRef] [PubMed]
- 35. Hochheimer, B.F. Angiography of the Retina with Indocyanine Green. Arch. Ophthalmol. 1971, 86, 564–565. [CrossRef] [PubMed]
- 36. Flower, R.W. Infrared Absorption Angiography of the Choroid and Some Observations on the Effects of High Intraocular Pressures. *Am. J. Ophthalmol.* **1972**, *74*, 600–614. [CrossRef]
- 37. Gandorfer, A.; Haritoglou, C.; Gass, C.A.; Ulbig, M.W.; Kampik, A. Indocyanine Green-Assisted Peeling of the Internal Limiting Membrane May Cause Retinal Damage. *Am. J. Ophthalmol.* **2001**, *132*, 431–433. [CrossRef]
- 38. Iinuma, Y.; Hirayama, Y.; Yokoyama, N.; Otani, T.; Nitta, K.; Hashidate, H.; Yoshida, M.; Iida, H.; Masui, D.; Manabe, S. Intraoperative Near-Infrared Indocyanine Green Fluorescence Angiography (NIR-ICG AG) Can Predict Delayed Small Bowel Stricture after Ischemic Intestinal Injury: Report of a Case. J. Pediatr. Surg. 2013, 48, 1123–1128. [CrossRef]
- 39. Carus, T.; Dammer, R. Laparoscop Fluorescence Angiography with Indocyanine Green to Control the Perfusion of Gastrointestinal Anastomoses Intraoperatively. *Surg. Technol. Int.* **2012**, *22*, 27–32.
- Jafari, M.D.; Lee, K.H.; Halabi, W.J.; Mills, S.D.; Carmichael, J.C.; Stamos, M.J.; Pigazzi, A. The Use of Indocyanine Green Fluorescence to Assess Anastomotic Perfusion during Robotic Assisted Laparoscopic Rectal Surgery. *Surg. Endosc.* 2013, 27, 3003–3008. [CrossRef]
- Ishiguro, T.; Kumagai, Y.; Ono, T.; Imaizumi, H.; Honjo, H.; Suzuki, O.; Ito, T.; Haga, N.; Kuwabara, K.; Sobajima, J.; et al. Usefulness of Indocyanine Green Angiography for Evaluation of Blood Supply in a Reconstructed Gastric Tube during Esophagectomy. *Int. Surg.* 2012, *97*, 340–344. [CrossRef]
- Kumagai, Y.; Ishiguro, T.; Haga, N.; Kuwabara, K.; Kawano, T.; Ishida, H. Hemodynamics of the Reconstructed Gastric Tube during Esophagectomy: Assessment of Outcomes with Indocyanine Green Fluorescence. *World J. Surg.* 2014, *38*, 138–143. [CrossRef]
- Rino, Y.; Yukawa, N.; Sato, T.; Yamamoto, N.; Tamagawa, H.; Hasegawa, S.; Oshima, T.; Yoshikawa, T.; Masuda, M.; Imada, T. Visualization of Blood Supply Route to the Reconstructed Stomach by Indocyanine Green Fluorescence Imaging during Esophagectomy. *BMC Med. Imaging* 2014, 14, 18. [CrossRef] [PubMed]
- Raabe, A.; Nakaji, P.; Beck, J.; Kim, L.J.; Hsu, F.P.K.; Kamerman, J.D.; Seifert, V.; Spetzler, R.F. Prospective Evaluation of Surgical Microscope-Integrated Intraoperative near-Infrared Indocyanine Green Videoangiography during Aneurysm Surgery. J. Neurosurg. 2005, 103, 982–989. [CrossRef] [PubMed]
- Norat, P.; Soldozy, S.; Elsarrag, M.; Sokolowski, J.; Yağmurlu, K.; Park, M.S.; Tvrdik, P.; Kalani, M.Y.S. Application of Indocyanine Green Videoangiography in Aneurysm Surgery: Evidence, Techniques, Practical Tips. *Front. Surg.* 2019, *6*, 34. [CrossRef] [PubMed]

- 46. Gurtner, G.C.; Jones, G.E.; Neligan, P.C.; Newman, M.I.; Phillips, B.T.; Sacks, J.M.; Zenn, M.R. Intraoperative Laser Angiography Using the SPY System: Review of the Literature and Recommendations for Use. *Ann. Surg. Innov. Res.* **2013**, *7*, 1. [CrossRef]
- 47. Holm, C.; Mayr, M.; Höfter, E.; Becker, A.; Pfeiffer, U.J.; Mühlbauer, W. Intraoperative Evaluation of Skin-Flap Viability Using Laser-Induced Fluorescence of Indocyanine Green. *Br. J. Plast. Surg.* **2002**, *55*, 635–644. [CrossRef]
- 48. Moyer, H.R.; Losken, A. Predicting Mastectomy Skin Flap Necrosis with Indocyanine Green Angiography: The Gray Area Defined. *Plast. Reconstr. Surg.* **2012**, *129*, 1043–1048. [CrossRef]
- 49. Kitai, T.; Inomoto, T.; Miwa, M.; Shikayama, T. Fluorescence Navigation with Indocyanine Green for Detecting Sentinel Lymph Nodes in Breast Cancer. *Breast Cancer* 2005, *12*, 211–215. [CrossRef]
- 50. Muhammad, N.; Ajmera, S.; Lee, J.Y.K. Intraoperative Visualization of Cranial Nerve Schwannomas Using Second-Window Indocyanine Green: A Case Series. *Clin. Neurol. Neurosurg.* **2024**, 240, 108241. [CrossRef]
- 51. Pothen, A.-G.; Parmar, M. Fluorescein; StatPearls Publishing: Treasure Island, FL, USA, 2023.
- Jin, E.; Yin, H.; Gui, Y.; Chen, J.; Zhang, J.; Liang, J.; Li, X.-X.; Zhao, M. Fluorescein Angiographic Findings of Peripheral Retinal Vasculature after Intravitreal Conbercept versus Ranibizumab for Retinopathy of Prematurity. J. Ophthalmol. 2019, 2019, 3935945.
 [CrossRef]
- 53. Paugh, J.R.; Tse, J.; Nguyen, T.; Sasai, A.; Chen, E.; De Jesus, M.T.; Kwan, J.; Nguyen, A.L.; Farid, M.; Garg, S.; et al. Efficacy of the Fluorescein Tear Breakup Time Test in Dry Eye. *Cornea* 2020, *39*, 92–98. [CrossRef]
- Olson, J.L.; Mandava, N. Chapter 1—Fluorescein Angiography. In *Retinal Imaging*; Huang, D., Kaiser, P.K., Lowder, C.Y., Traboulsi, E.I., Eds.; Mosby: Philadelphia, PA, USA, 2006; pp. 3–21, ISBN 9780323023467.
- 55. Food and Drug Administration. *FLUORESCITE®* (*Fluorescein Injection, USP*) 10%; Department of Health & Human Services: Rockville, MD, USA, 2006.
- 56. Novotny, H.R.; Alvis, D.L. A Method of Photographing Fluorescence in Circulating Blood in the Human Retina. *Circulation* **1961**, 24, 82–86. [CrossRef] [PubMed]
- 57. Orosco, R.K.; Tsien, R.Y.; Nguyen, Q.T. Fluorescence Imaging in Surgery. *IEEE Rev. Biomed. Eng.* 2013, *6*, 178–187. [CrossRef] [PubMed]
- Prieto, S.P.; Lai, K.K.; Laryea, J.A.; Mizell, J.S.; Mustain, W.C.; Muldoon, T.J. Fluorescein as a Topical Fluorescent Contrast Agent for Quantitative Microendoscopic Inspection of Colorectal Epithelium. *Biomed. Opt. Express* 2017, *8*, 2324–2338. [CrossRef] [PubMed]
- 59. Uchida, Y.; Uchida, Y. Dye-Staining Angioscopy for Coronary Artery Disease. Curr. Cardiovasc. Imaging Rep. 2015, 8, 10. [CrossRef]
- Save, A.V.; Gill, B.J.; D'amico, R.S.; Canoll, P.; Bruce, J.N. Fluorescein-Guided Resection of Gliomas. J. Neurosurg. Sci. 2019, 63, 648–655. [CrossRef]
- 61. Zhao, X.; Belykh, E.; Cavallo, C.; Valli, D.; Gandhi, S.; Preul, M.C.; Vajkoczy, P.; Lawton, M.T.; Nakaji, P. Application of Fluorescein Fluorescence in Vascular Neurosurgery. *Front. Surg.* **2019**, *6*, 52. [CrossRef]
- Lakomkin, N.; Hadjipanayis, C.G. Fluorescence-Guided Surgery for High-Grade Gliomas. J. Surg. Oncol. 2018, 118, 356–361. [CrossRef]
- Díez Valle, R.; Hadjipanayis, C.G.; Stummer, W. Established and Emerging Uses of 5-ALA in the Brain: An Overview. J. Neurooncol. 2019, 141, 487–494. [CrossRef]
- 64. Fujino, M.; Nishio, Y.; Ito, H.; Tanaka, T.; Li, X.-K. 5-Aminolevulinic Acid Regulates the Inflammatory Response and Alloimmune Reaction. *Int. Immunopharmacol.* **2016**, *37*, 71–78. [CrossRef]
- Schupper, A.J.; Baron, R.B.; Cheung, W.; Rodriguez, J.; Kalkanis, S.N.; Chohan, M.O.; Andersen, B.J.; Chamoun, R.; Nahed, B.V.; Zacharia, B.E.; et al. 5-Aminolevulinic Acid for Enhanced Surgical Visualization of High-Grade Gliomas: A Prospective, Multicenter Study. *J. Neurosurg.* 2022, *136*, 1525–1534. [CrossRef]
- 66. Stummer, W.; Tonn, J.-C.; Goetz, C.; Ullrich, W.; Stepp, H.; Bink, A.; Pietsch, T.; Pichlmeier, U. 5-Aminolevulinic Acid-Derived Tumor Fluorescence: The Diagnostic Accuracy of Visible Fluorescence Qualities as Corroborated by Spectrometry and Histology and Postoperative Imaging. *Neurosurgery* 2014, 74, 310–319; discussion 319–320. [CrossRef] [PubMed]
- Coburger, J.; Engelke, J.; Scheuerle, A.; Thal, D.R.; Hlavac, M.; Wirtz, C.R.; König, R. Tumor Detection with 5-Aminolevulinic Acid Fluorescence and Gd-DTPA-Enhanced Intraoperative MRI at the Border of Contrast-Enhancing Lesions: A Prospective Study Based on Histopathological Assessment. *Neurosurg. Focus* 2014, *36*, E3. [CrossRef] [PubMed]
- Stummer, W.; Stepp, H.; Möller, G.; Ehrhardt, A.; Leonhard, M.; Reulen, H.J. Technical Principles for Protoporphyrin-IX-Fluorescence Guided Microsurgical Resection of Malignant Glioma Tissue. *Acta Neurochir.* 1998, 140, 995–1000. [CrossRef] [PubMed]
- Stummer, W.; Novotny, A.; Stepp, H.; Goetz, C.; Bise, K.; Reulen, H.J. Fluorescence-Guided Resection of Glioblastoma Multiforme by Using 5-Aminolevulinic Acid-Induced Porphyrins: A Prospective Study in 52 Consecutive Patients. *J. Neurosurg.* 2000, 93, 1003–1013. [CrossRef]
- Hadjipanayis, C.G.; Widhalm, G.; Stummer, W. What Is the Surgical Benefit of Utilizing 5-Aminolevulinic Acid for Fluorescence-Guided Surgery of Malignant Gliomas? *Neurosurgery* 2015, 77, 663–673. [CrossRef]

- 71. Stepp, H.; Stummer, W. 5-ALA in the Management of Malignant Glioma. Lasers Surg. Med. 2018, 50, 399–419. [CrossRef]
- 72. Eljamel, S. 5-ALA Fluorescence Image Guided Resection of Glioblastoma Multiforme: A Meta-Analysis of the Literature. *Int. J. Mol. Sci.* 2015, *16*, 10443–10456. [CrossRef]
- Picart, T.; Pallud, J.; Berthiller, J.; Dumot, C.; Berhouma, M.; Ducray, F.; Armoiry, X.; Margier, J.; Guerre, P.; Varlet, P.; et al. Use of 5-ALA Fluorescence-Guided Surgery versus White-Light Conventional Microsurgery for the Resection of Newly Diagnosed Glioblastomas (RESECT Study): A French Multicenter Randomized Phase III Study. J. Neurosurg. 2024, 140, 987–1000. [CrossRef]
- Filip, P.; Lerner, D.K.; Kominsky, E.; Schupper, A.; Liu, K.; Khan, N.M.; Roof, S.; Hadjipanayis, C.; Genden, E.; Iloreta, A.M.C.
 5-Aminolevulinic Acid Fluorescence-Guided Surgery in Head and Neck Squamous Cell Carcinoma. *Laryngoscope* 2024, 134, 741–748. [CrossRef]
- 75. Bickels, J.; Gortzak, Y.; Sternheim, A. 5-ALA Photodynamic Ablation of Fibroblastic Soft-Tissue Tumors. *Photodiagn. Photodyn. Ther.* **2023**, *42*, 103624. [CrossRef]
- Guder, W.K.; Hartmann, W.; Buhles, C.; Burdack, M.; Busch, M.; Dünker, N.; Hardes, J.; Dirksen, U.; Bauer, S.; Streitbürger, A. 5-ALA-Mediated Fluorescence of Musculoskeletal Tumors in a Chick Chorio-Allantoic Membrane Model: Preclinical in Vivo Qualification Analysis as a Fluorescence-Guided Surgery Agent in Orthopedic Oncology. J. Orthop. Surg. Res. 2022, 17, 34. [CrossRef] [PubMed]
- Stummer, W.; Stocker, S.; Novotny, A.; Heimann, A.; Sauer, O.; Kempski, O.; Plesnila, N.; Wietzorrek, J.; Reulen, H.J. In Vitro and in Vivo Porphyrin Accumulation by C6 Glioma Cells after Exposure to 5-Aminolevulinic Acid. *J. Photochem. Photobiol. B* 1998, 45, 160–169. [CrossRef] [PubMed]
- Kaneko, S.; Suero Molina, E.; Ewelt, C.; Warneke, N.; Stummer, W. Fluorescence-Based Measurement of Real-Time Kinetics of Protoporphyrin IX After 5-Aminolevulinic Acid Administration in Human In Situ Malignant Gliomas. *Neurosurgery* 2019, 85, E739–E746. [CrossRef] [PubMed]
- 79. Maragkos, G.A.; Schüpper, A.J.; Lakomkin, N.; Sideras, P.; Price, G.; Baron, R.; Hamilton, T.; Haider, S.; Lee, I.Y.; Hadjipanayis, C.G.; et al. Fluorescence-Guided High-Grade Glioma Surgery More Than Four Hours After 5-Aminolevulinic Acid Administration. *Front. Neurol.* **2021**, *12*, 644804. [CrossRef]
- 80. Schupper, A.J.; Rao, M.; Mohammadi, N.; Baron, R.; Lee, J.Y.K.; Acerbi, F.; Hadjipanayis, C.G. Fluorescence-Guided Surgery: A Review on Timing and Use in Brain Tumor Surgery. *Front. Neurol.* **2021**, *12*, 682151. [CrossRef]
- Guyotat, J.; Pallud, J.; Armoiry, X.; Pavlov, V.; Metellus, P. 5-Aminolevulinic Acid–Protoporphyrin IX Fluorescence-Guided Surgery of High-Grade Gliomas: A Systematic Review. In *Advances and Technical Standards in Neurosurgery*; Schramm, J., Ed.; Springer International Publishing: Cham, Switzerland, 2016; Volume 43, pp. 61–90, ISBN 9783319213590.
- 82. Wei, L.; Roberts, D.W.; Sanai, N.; Liu, J.T.C. Visualization Technologies for 5-ALA-Based Fluorescence-Guided Surgeries. J. *Neurooncol.* **2019**, 141, 495–505. [CrossRef]
- 83. Utzinger, U.; Richards-Kortum, R.R. Fiber Optic Probes for Biomedical Optical Spectroscopy. J. Biomed. Opt. 2003, 8, 121–147. [CrossRef]
- Valdés, P.A.; Leblond, F.; Kim, A.; Harris, B.T.; Wilson, B.C.; Fan, X.; Tosteson, T.D.; Hartov, A.; Ji, S.; Erkmen, K.; et al. Quantitative Fluorescence in Intracranial Tumor: Implications for ALA-Induced PpIX as an Intraoperative Biomarker. J. Neurosurg. 2011, 115, 11–17. [CrossRef]
- 85. Haj-Hosseini, N.; Richter, J.; Andersson-Engels, S.; Wårdell, K. Optical Touch Pointer for Fluorescence Guided Glioblastoma Resection Using 5-Aminolevulinic Acid. *Lasers Surg. Med.* **2010**, *42*, 9–14. [CrossRef]
- 86. Kim, A.; Khurana, M.; Moriyama, Y.; Wilson, B.C. Quantification of in Vivo Fluorescence Decoupled from the Effects of Tissue Optical Properties Using Fiber-Optic Spectroscopy Measurements. *J. Biomed. Opt.* **2010**, *15*, 067006. [CrossRef]
- Ishihara, R.; Katayama, Y.; Watanabe, T.; Yoshino, A.; Fukushima, T.; Sakatani, K. Quantitative Spectroscopic Analysis of 5-Aminolevulinic Acid-Induced Protoporphyrin IX Fluorescence Intensity in Diffusely Infiltrating Astrocytomas. *Neurol. Med. Chir.* 2007, 47, 53–57; discussion 57. [CrossRef] [PubMed]
- Utsuki, S.; Oka, H.; Sato, S.; Suzuki, S.; Shimizu, S.; Tanaka, S.; Fujii, K. Possibility of Using Laser Spectroscopy for the Intraoperative Detection of Nonfluorescing Brain Tumors and the Boundaries of Brain Tumor Infiltrates. *J. Neurosurg.* 2006, 104, 618–620. [CrossRef] [PubMed]
- 89. Montemurro, N.; Scerrati, A.; Ricciardi, L.; Trevisi, G. The Exoscope in Neurosurgery: An Overview of the Current Literature of Intraoperative Use in Brain and Spine Surgery. *J. Clin. Med. Res.* **2021**, *11*, 223. [CrossRef] [PubMed]
- 90. Schupper, A.J.; Roa, J.A.; Hadjipanayis, C.G. Contemporary Intraoperative Visualization for GBM with Use of Exoscope, 5-ALA Fluorescence-Guided Surgery and Tractography. *Neurosurg. Focus Video* **2022**, *6*, V5. [CrossRef]
- Muscas, G.; Battista, F.; Boschi, A.; Morone, F.; Della Puppa, A. A Single-Center Experience with the Olympus ORBEYE 4K-3D Exoscope for Microsurgery of Complex Cranial Cases: Technical Nuances and Learning Curve. J. Neurol. Surg. A Cent. Eur. Neurosurg. 2021, 82, 484–489. [CrossRef]
- 92. Schupper, A.J.; Price, G.; Hadjipanayis, C.G. Robotic-Assisted Digital Exoscope for Resection of Cerebral Metastases: A Case Series. *Oper. Neurosurg.* **2021**, *21*, 436–444. [CrossRef]

- 93. Pafitanis, G.; Hadjiandreou, M.; Alamri, A.; Uff, C.; Walsh, D.; Myers, S. The Exoscope versus Operating Microscope in Microvascular Surgery: A Simulation Non-Inferiority Trial. *Arch. Plast. Surg.* **2020**, *47*, 242–249. [CrossRef]
- Abunimer, A.M.; Abou-Al-Shaar, H.; White, T.G.; Park, J.; Schulder, M. The Utility of High-Definition 2-Dimensional Stereotactic Exoscope in Cranial and Spinal Procedures. *World Neurosurg*. 2022, 158, e231–e236. [CrossRef]
- Baron, R.B.; Lakomkin, N.; Schupper, A.J.; Nistal, D.; Nael, K.; Price, G.; Hadjipanayis, C.G. Postoperative Outcomes Following Glioblastoma Resection Using a Robot-Assisted Digital Surgical Exoscope: A Case Series. J. Neurooncol. 2020, 148, 519–527. [CrossRef]
- 96. Witten, A.J.; Ben-Shalom, N.; Ellis, J.A.; Boockvar, J.A.; D'Amico, R.S. Optimization of Novel Exoscopic Blue Light Filter during Fluorescence-Guided Resection of Glioblastoma. *J. Neurooncol.* **2023**, *161*, 617–623. [CrossRef]
- 97. Goehre, F.; Ludtka, C.; Schwan, S. Ergonomics of Surgical Microscopes for the Sitting Position as Determined by Ocular-Corpus Length. *Surg. Neurol. Int.* 2020, *11*, 244. [CrossRef] [PubMed]
- Della Pepa, G.M.; Mattogno, P.; Menna, G.; Agostini, L.; Olivi, A.; Doglietto, F. A Comparative Analysis with Exoscope and Optical Microscope for Intraoperative Visualization and Surgical Workflow in 5-Aminolevulinic Acid-Guided Resection of High-Grade Gliomas. *World Neurosurg.* 2023, 170, 133–137. [CrossRef] [PubMed]
- 99. Zhang, X.; Jaman, E.; Habib, A.; Ozpinar, A.; Andrews, E.; Amankulor, N.M.; Zinn, P.O. A Novel 5-Aminolevulinic Acid-Enabled Surgical Loupe System-A Consecutive Brain Tumor Series of 11 Cases. *Oper. Neurosurg.* **2022**, *22*, 298–304. [CrossRef] [PubMed]
- Giantini-Larsen, A.M.; Parker, W.E.; Cho, S.S.; Goldberg, J.L.; Carnevale, J.A.; Michael, A.P.; Teng, C.W.; De Ravin, E.; Brennan, C.W.; Lee, J.Y.K.; et al. The Evolution of 5-Aminolevulinic Acid Fluorescence Visualization: Time for a Headlamp/Loupe Combination. *World Neurosurg.* 2022, 159, 136–143. [CrossRef]
- Tonn, J.-C.; Stummer, W. Fluorescence-Guided Resection of Malignant Gliomas Using 5-Aminolevulinic Acid: Practical Use, Risks, and Pitfalls. *Clin. Neurosurg.* 2008, 55, 20–26.
- 102. Valdés, P.A.; Leblond, F.; Jacobs, V.L.; Wilson, B.C.; Paulsen, K.D.; Roberts, D.W. Quantitative, Spectrally-Resolved Intraoperative Fluorescence Imaging. *Sci. Rep.* **2012**, *2*, 798. [CrossRef]
- 103. Diamond, K.R.; Patterson, M.S.; Farrell, T.J. Quantification of Fluorophore Concentration in Tissue-Simulating Media by Fluorescence Measurements with a Single Optical Fiber. *Appl. Opt.* **2003**, *42*, 2436–2442. [CrossRef]
- 104. Müller, M.G.; Georgakoudi, I.; Zhang, Q.; Wu, J.; Feld, M.S. Intrinsic Fluorescence Spectroscopy in Turbid Media: Disentangling Effects of Scattering and Absorption. *Appl. Opt.* **2001**, *40*, 4633–4646. [CrossRef]
- 105. Wu, J.; Feld, M.S.; Rava, R.P. Analytical Model for Extracting Intrinsic Fluorescence in Turbid Media. *Appl. Opt.* **1993**, *32*, 3585–3595. [CrossRef]
- Mahmoudi, K.; Garvey, K.L.; Bouras, A.; Cramer, G.; Stepp, H.; Jesu Raj, J.G.; Bozec, D.; Busch, T.M.; Hadjipanayis, C.G.
 5-Aminolevulinic Acid Photodynamic Therapy for the Treatment of High-Grade Gliomas. *J. Neurooncol.* 2019, 141, 595–607. [CrossRef]
- 107. Wu, S.-K.; Santos, M.A.; Marcus, S.L.; Hynynen, K. MR-Guided Focused Ultrasound Facilitates Sonodynamic Therapy with 5-Aminolevulinic Acid in a Rat Glioma Model. *Sci. Rep.* **2019**, *9*, 10465. [CrossRef] [PubMed]
- 108. Vermandel, M.; Quidet, M.; Vignion-Dewalle, A.-S.; Leroy, H.-A.; Leroux, B.; Mordon, S.; Reyns, N. Comparison of Different Treatment Schemes in 5-ALA Interstitial Photodynamic Therapy for High-Grade Glioma in a Preclinical Model: An MRI Study. *Photodiagn. Photodyn. Ther.* 2019, 25, 166–176. [CrossRef] [PubMed]
- Leroy, H.-A.; Vermandel, M.; Leroux, B.; Duhamel, A.; Lejeune, J.-P.; Mordon, S.; Reyns, N. MRI Assessment of Treatment Delivery for Interstitial Photodynamic Therapy of High-Grade Glioma in a Preclinical Model. *Lasers Surg. Med.* 2018, 50, 460–468. [CrossRef] [PubMed]
- Vermandel, M.; Dupont, C.; Lecomte, F.; Leroy, H.-A.; Tuleasca, C.; Mordon, S.; Hadjipanayis, C.G.; Reyns, N. Standardized Intraoperative 5-ALA Photodynamic Therapy for Newly Diagnosed Glioblastoma Patients: A Preliminary Analysis of the INDYGO Clinical Trial. J. Neurooncol. 2021, 152, 501–514. [CrossRef]
- 111. Peciu-Florianu, I.; Vannod-Michel, Q.; Vauleon, E.; Bonneterre, M.-E.; Reyns, N. Long Term Follow-up of Patients with Newly Diagnosed Glioblastoma Treated by Intraoperative Photodynamic Therapy: An Update from the INDYGO Trial (NCT03048240). J. Neurooncol. 2024, 168, 495–505. [CrossRef]
- 112. Scanlon, S.E.; Shanahan, R.M.; Bin-Alamer, O.; Bouras, A.; Mattioli, M.; Huq, S.; Hadjipanayis, C.G. Sonodynamic Therapy for Adult-Type Diffuse Gliomas: Past, Present, and Future. *J. Neurooncol.* **2024**, *169*, 507–516. [CrossRef]
- 113. Shayan Moosa, MD, University of Virginia Sonodynamic Therapy in Patients with Recurrent GBM (GBM 001). Available online: https://clinicaltrials.gov/study/NCT06039709?intr=5-ALA&aggFilters=status:act%20rec%20not&term=brain&locStr=USA&country=United%20States&rank=3 (accessed on 21 August 2024).
- 114. SonALAsense, Inc. A Study of Sonodynamic Therapy Using SONALA-001 and Exablate 4000 Type 2.0 in Subjects with Recurrent GBM. Available online: https://clinicaltrials.gov/study/NCT05370508?intr=5-ALA&aggFilters=status:act%20rec%20not&term=brain&locStr=USA&country=United%20States&rank=4 (accessed on 21 August 2024).

- 115. Nader Sanai, St. Joseph's Hospital and Medical Center, Phoenix Study of Sonodynamic Therapy in Participants with Recurrent High-Grade Glioma. Available online: https://clinicaltrials.gov/study/NCT04559685?intr=5-ALA&aggFilters=status:act%20 rec%20not&term=brain&locStr=USA&country=United%20States&rank=7 (accessed on 21 August 2024).
- 116. SonALAsense, Inc. A Phase 2 Study of Sonodynamic Therapy Using SONALA-001 and Exablate 4000 Type 2.0 in Patients with DIPG. Available online: https://clinicaltrials.gov/study/NCT05123534?intr=5-ALA&aggFilters=status:act%20rec%20not&term=brain&locStr=USA&country=United%20States&rank=5 (accessed on 21 August 2024).
- 117. Photolitec, L.L.C. Intracavitary Photodynamic Therapy as an Adjuvant to Resection of Glioblastoma or Gliosarcoma Using IV Photobac[®]. Available online: https://clinicaltrials.gov/study/NCT05363826?intr=photodynamic%20therapy&aggFilters=status: act%20rec%20not&term=brain&locStr=USA&country=United%20States&rank=1 (accessed on 21 August 2024).
- 118. Therapeutics, H. A Dose-Escalation Clinical Study of Intraoperative Photodynamic Therapy of Glioblastoma. Available online: https://clinicaltrials.gov/study/NCT05736406 (accessed on 21 August 2024).
- 119. Mansouri, A.; Mansouri, S.; Hachem, L.D.; Klironomos, G.; Vogelbaum, M.A.; Bernstein, M.; Zadeh, G. The Role of 5-Aminolevulinic Acid in Enhancing Surgery for High-Grade Glioma, Its Current Boundaries, and Future Perspectives: A Systematic Review. *Cancer* 2016, 122, 2469–2478. [CrossRef]
- Ferraro, N.; Barbarite, E.; Albert, T.R.; Berchmans, E.; Shah, A.H.; Bregy, A.; Ivan, M.E.; Brown, T.; Komotar, R.J. The Role of 5-Aminolevulinic Acid in Brain Tumor Surgery: A Systematic Review. *Neurosurg. Rev.* 2016, 39, 545–555. [CrossRef]
- Zhang, C.; Boop, F.A.; Ruge, J. The Use of 5-Aminolevulinic Acid in Resection of Pediatric Brain Tumors: A Critical Review. J. Neurooncol. 2019, 141, 567–573. [CrossRef]
- 122. Widhalm, G.; Kiesel, B.; Woehrer, A.; Traub-Weidinger, T.; Preusser, M.; Marosi, C.; Prayer, D.; Hainfellner, J.A.; Knosp, E.; Wolfsberger, S. 5-Aminolevulinic Acid Induced Fluorescence Is a Powerful Intraoperative Marker for Precise Histopathological Grading of Gliomas with Non-Significant Contrast-Enhancement. *PLoS ONE* 2013, *8*, e76988. [CrossRef]
- 123. Boschi, A.; Della Puppa, A. 5-ALA Fluorescence on Tumors Different from Malignant Gliomas. Review of the Literature and Our Experience. *J. Neurosurg. Sci.* 2019, 63, 661–669. [CrossRef] [PubMed]
- Valdes, P.A.; Bekelis, K.; Harris, B.T.; Wilson, B.C.; Leblond, F.; Kim, A.; Simmons, N.E.; Erkmen, K.; Paulsen, K.D.; Roberts, D.W.
 5-Aminolevulinic Acid-Induced Protoporphyrin IX Fluorescence in Meningioma: Qualitative and Quantitative Measurements in Vivo. *Neurosurgery* 2014, *10* (Suppl. S1), 74–82; discussion 82–83. [CrossRef] [PubMed]
- 125. Wadiura, L.I.; Millesi, M.; Makolli, J.; Wais, J.; Kiesel, B.; Mischkulnig, M.; Mercea, P.A.; Roetzer, T.; Knosp, E.; Rössler, K.; et al. High Diagnostic Accuracy of Visible 5-ALA Fluorescence in Meningioma Surgery According to Histopathological Analysis of Tumor Bulk and Peritumoral Tissue. *Lasers Surg. Med.* 2021, *53*, 300–308. [CrossRef] [PubMed]
- 126. Milos, P.; Haj-Hosseini, N.; Hillman, J.; Wårdell, K. 5-ALA Fluorescence in Randomly Selected Pediatric Brain Tumors Assessed by Spectroscopy and Surgical Microscope. *Acta Neurochir.* **2023**, *165*, 71–81. [CrossRef]
- 127. Universität Münster. Clinical Safety Study on 5-Aminolevulinic Acid (5-ALA) in Children and Adolescents With Supratentorial Brain Tumors. Available online: https://clinicaltrials.gov/study/NCT04738162?cond=brain%20tumor&intr=5-ALA&rank=2 (accessed on 21 August 2024).
- 128. Alston, L.; Mahieu-Williame, L.; Hebert, M.; Kantapareddy, P.; Meyronet, D.; Rousseau, D.; Guyotat, J.; Montcel, B. Spectral Complexity of 5-ALA Induced PpIX Fluorescence in Guided Surgery: A Clinical Study towards the Discrimination of Healthy Tissue and Margin Boundaries in High and Low Grade Gliomas. *Biomed. Opt. Express* 2019, 10, 2478–2492. [CrossRef]
- 129. Muraleedharan, S.; Tripathy, K. Indocyanine Green (ICG) Angiography. In *StatPearls*; StatPearls Publishing: Treasure Island, FL, USA, 2023.
- Lee, J.Y.K.; Thawani, J.P.; Pierce, J.; Zeh, R.; Martinez-Lage, M.; Chanin, M.; Venegas, O.; Nims, S.; Learned, K.; Keating, J.; et al. Intraoperative Near-Infrared Optical Imaging Can Localize Gadolinium-Enhancing Gliomas During Surgery. *Neurosurgery* 2016, 79, 856–871. [CrossRef]
- 131. Karsalia, R.; Cheng, N.H.; Teng, C.W.; Cho, S.S.; Harmsen, S.; Lee, J.Y.K. Second Window ICG Predicts Postoperative MRI Gadolinium Enhancement in High Grade Gliomas and Brain Metastases. *Neurosurg. Focus Video* **2022**, *6*, V8. [CrossRef]
- 132. Hansen, R.W.; Pedersen, C.B.; Halle, B.; Korshoej, A.R.; Schulz, M.K.; Kristensen, B.W.; Poulsen, F.R. Comparison of 5-Aminolevulinic Acid and Sodium Fluorescein for Intraoperative Tumor Visualization in Patients with High-Grade Gliomas: A Single-Center Retrospective Study. J. Neurosurg. 2020, 133, 1324–1331. [CrossRef]
- 133. Ahrens, L.C.; Krabbenhøft, M.G.; Hansen, R.W.; Mikic, N.; Pedersen, C.B.; Poulsen, F.R.; Korshoej, A.R. Effect of 5-Aminolevulinic Acid and Sodium Fluorescein on the Extent of Resection in High-Grade Gliomas and Brain Metastasis. *Cancers* 2022, 14, 617. [CrossRef]
- 134. Xi, C.; Jinli, S.; Jianyao, M.; Yan, C.; Huijuan, L.; Zhongjie, S.; Zhangyu, L.; Liwei, Z.; Yukui, L.; Sifang, C.; et al. Fluorescein-Guided Surgery for High-Grade Glioma Resection: A Five-Year-Long Retrospective Study at Our Institute. *Front. Oncol.* 2023, 13, 1191470. [CrossRef]

- 135. Suero Molina, E.; Wölfer, J.; Ewelt, C.; Ehrhardt, A.; Brokinkel, B.; Stummer, W. Dual-Labeling with 5-Aminolevulinic Acid and Fluorescein for Fluorescence-Guided Resection of High-Grade Gliomas: Technical Note. J. Neurosurg. 2018, 128, 399–405. [CrossRef] [PubMed]
- Schwake, M.; Stummer, W.; Suero Molina, E.J.; Wölfer, J. Simultaneous Fluorescein Sodium and 5-ALA in Fluorescence-Guided Glioma Surgery. *Acta Neurochir.* 2015, 157, 877–879. [CrossRef] [PubMed]
- Parakh, S.; Nicolazzo, J.; Scott, A.M.; Gan, H.K. Antibody Drug Conjugates in Glioblastoma—Is there a Future for Them? *Front.* Oncol. 2021, 11, 718590. [CrossRef] [PubMed]
- 138. Alfred-Marc Iloreta, Icahn School of Medicine at Mount Sinai. The Role of 5-Aminolevulinic Acid Fluorescence-Guided Surgery in Head and Neck Cancers: A Pilot Trial. Available online: https://clinicaltrials.gov/study/NCT05101798?intr=5-ALA&aggFilters=status:act%20rec%20not&term=brain&locStr=USA&country=United%20States&rank=8 (accessed on 21 August 2024).
- 139. Roberts, D.W. Dartmouth-Hitchcock Medical Center ALA-Induced PpIX Fluorescence During Brain Tumor Resection. Available online: https://clinicaltrials.gov/study/NCT02191488?intr=5-ALA&aggFilters=status:act%20rec%20not&term=brain&locStr=USA&country=United%20States&rank=6 (accessed on 21 August 2024).
- 140. Abramson Cancer Center at Penn Medicine. Second Window Indocyanine Green for All Nervous System Tumors. Available online: https://clinicaltrials.gov/study/NCT05746104?intr=icg&aggFilters=status:act%20rec%20not&term=brain&rank=1 (accessed on 21 August 2024).
- 141. Roberts, D.W. Dartmouth-Hitchcock Medical Center Diagnostic Performance of Fluorescein as an Intraoperative Brain Tumor Biomarker. Available online: https://clinicaltrials.gov/study/NCT02691923?intr=5-ALA&aggFilters=status:act%20rec%20not& term=brain&locStr=USA&country=United%20States&rank=2 (accessed on 21 August 2024).
- 142. Linton, T. Evans, Dartmouth-Hitchcock Medical Center Evaluation of the CONVIVO System. Available online: https://clinicaltrials.gov/study/NCT05139277?intr=fluorescein&aggFilters=status:act%20rec%20not&term=brain&locStr= USA&country=United%20States&rank=3 (accessed on 21 August 2024).
- 143. Yu, G.; University of Kentucky. Loupe-Based Intraoperative Fluorescence Imaging. Available online: https://clinicaltrials. gov/study/NCT04780009?intr=5-ALA&aggFilters=status:act%20rec%20not&term=brain&locStr=USA&country=United%20 States&rank=9 (accessed on 21 August 2024).
- 144. Chen, H.; Xu, H.; Peng, B.; Huang, X.; Hu, Y.; Zheng, C.; Zhang, Z. Illuminating the future of precision cancer surgery with fluorescence imaging and artificial intelligence convergence. *Npj Precis. Oncol.* **2024**, *8*, 196. [CrossRef]

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