

REVIEW ARTICLE

Neuroprotective Role of Hypothermia in Hypoxic-ischemic Brain Injury: Combined Therapies using Estrogen

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Abstract: Hypoxic-ischemic brain injury is a complex network of factors, which is mainly characterized by a decrease in levels of oxygen concentration and blood flow, which lead to an inefficient supply of nutrients to the brain. Hypoxic-ischemic brain injury can be found in perinatal asphyxia and ischemic-stroke, which represent one of the main causes of mortality and morbidity in children and adults worldwide. Therefore, knowledge of underlying mechanisms triggering these insults may help establish neuroprotective treatments. Selective Estrogen Receptor Modulators and Selective Tissue Estrogenic Activity Regulators exert several neuroprotective effects, including a decrease of reactive oxygen species, maintenance of cell viability, mitochondrial survival, among others. However, these strategies represent a traditional approach of targeting a single factor of pathology without satisfactory results. Hence, combined therapies, such as the administration of therapeutic hypothermia with a complementary neuroprotective agent, constitute a promising alternative. In this sense, the present review summarizes the underlying mechanisms of hypoxic-ischemic brain injury and compiles several neuroprotective strategies, including Selective Estrogen Receptor Modulators and Selective Tissue Estrogenic Activity Regulators, which represent putative agents for combined therapies with therapeutic hypothermia.

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1. INTRODUCTION

The Central Nervous System (CNS), due to its own functional characteristics, is susceptible to changes in energy levels, especially in oxygen (O₂) and glucose concentration [1]. Oxygen and glucose consumption by the brain is 20 and 25 %, respectively [2, 3]. This high energy demand is attributed to functions such as the recycling of neurotransmitters, ion transport and the synaptic activity carried out by brain cells [2]. Therefore, an efficient metabolism of the brain allows an optimal neuronal functioning. Hypoxic-ischemic (HI) brain injury is mainly characterized by decreasing O₂ concentration and blood flow, which lead to an inefficient

supply of nutrients to the brain. This pathophysiological conditions lead to cell death by energy depletion and increased free radical generation [4], as it is observed in perinatal asphyxia (PA) and ischemic-stroke (IS) [5-7]. PA and IS represent one of the main causes of mortality and morbidity in children and adults worldwide [8, 9]. Therefore, knowledge of underlying mechanisms triggering these insults becomes essential for the establishment of efficient treatments.

Several neuroprotective strategies have been tested, including Selective Estrogen Receptor Modulators (SERMs) and Selective Tissue Estrogenic Activity Regulators (STEARS), which have shown the same benefits as estrogen, including the decrease of reactive oxygen species (ROS), maintenance of cell viability, mitochondrial survival, among others; without its negative side effects [10-12]. However, these therapies remain inconclusive, representing a traditional approach of targeting a single factor of pathology.

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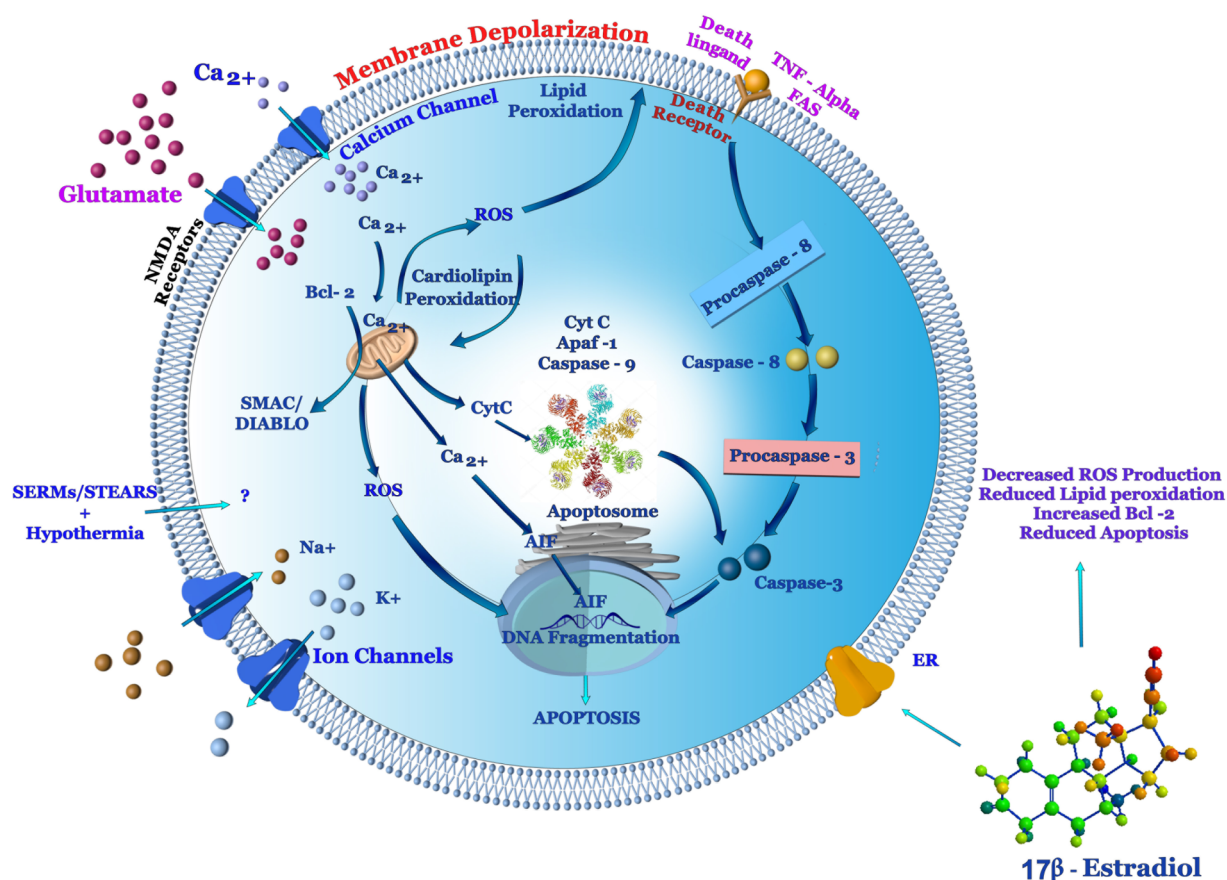


Fig. (1). Mechanisms of brain damage in hypoxia-ischemia and role of Neuroactive steroids and hypothermia. Calcium, Ca^{2+} ; Cytochrome-C, Cyt C; Estrogen receptor, ER; Reactive Oxygen species, ROS; Selective estrogen receptor modulators, SERMs; Selective tissue estrogen activity regulators, STEARs, Sodium, Na^+ ; Potassium, K^+ . (The color version of the figure is available in the electronic copy of the article).

Currently, the implementation of combined therapies has gained greater relevance [13]. Some combined therapies include therapeutic hypothermia and a complementary neuroprotective agent, since the former, which is the only treatment used in the clinical practice, does not exert complete neuroprotection [14]. In this sense, the present review summarizes the underlying mechanisms of HI brain injury and compiles several neuroprotective strategies, including SERMs and STEARs, which represent promising agents for combined therapies with therapeutic hypothermia.

2. MECHANISMS OF BRAIN DAMAGE IN HYPOXIA-ISCHEMIA

HI triggers an initial response immediately after the insult, followed by a secondary response some hours later (Fig. 1), and a third phase of persistent effects which lasts several years [15]. Global hypoxia affects the cerebral cortex, the thalamus and the basal ganglia, causing damage to deep gray matter [16]. While the complete pathogenic pathways of HI are not fully described, some mechanisms like increased glutamate, calcium overload, mitochondrial dysfunction and oxidative stress have been proposed to contribute to generate neuronal damage [17].

Primary response depends on the energetic failure causing primary brain injury. This is characterized by a reduction

of energy supply, generating an accumulation of ROS *via* lactate production augment, and making the cell susceptible to oxidative stress and mitochondrial dysfunction [16]. Low levels of adenosine triphosphate (ATP) derived from this energetical failure affect the integrity of cell membrane. Ion flux of sodium (Na^+)/potassium (K^+) is altered by the Na^+/K^+ pump dysfunction. In addition, calcium (Ca^{2+}) enters easily to the cell causing membrane depolarization and inhibiting glutamate storage inside the cell, being therefore accumulated in the extracellular space [18]. The second phase of the injury is caused by the recovery of blood flow and the reestablishment of brain metabolism, which are characterized by an inflammatory response, excitotoxicity and oxidative stress. These processes are responsible for cell death after HI and continue exerting deleterious effects for years [16].

2.1. Cell Death During the Second Phase of Injury

Apoptosis and necrosis are cell death pathways displayed after HI (Fig. 1). However, apoptosis is the most common one in the young brain unchained by mitochondrial failure [19]. Apoptosis can follow two pathways: the extrinsic pathway, triggered by external signals like Tumor Necrosis Factor- α (TNF- α) and Fatty Acid Synthase (FAS); and the intrinsic pathway, mediated by internal factors such as DNA damage or cell stress [20].

The extrinsic pathway is involved in the action of Caspase 8 and 10, which activate caspase effectors directly, interacting with the intrinsic pathway and triggering a permeabilization of mitochondrial membrane [20, 21]. The intrinsic pathway is mediated by the release of apoptotic factors such as cytochrome-c (cyt-c), second mitochondria-derived activator of caspase/Direct inhibitor of apoptosis-binding protein with low pI (Smac/Diablo), Apoptosis Inducing Factor (AIF), endonuclease G (endoG), Serine protease HTRA2, mitochondrial (Omi/HtrA2) after permeabilization of cell membrane [20,21]. These apoptotic factors may trigger cell death processes, which could be mediated by caspase-dependent pathways. Cyt-c interacts with Apoptosis protease-activating factor-1 (Apaf-1), creating the apoptosome. Smac/Diablo interacts with apoptosis inhibitors. AIF and endoG also act through a caspase- dependent pathway. These apoptotic factors are translocated to the nucleus, causing nuclear fragmentation [20, 21]. Hence, permeabilization of mitochondrial membrane has been proposed as a marker of a point of no return in HI.

2.2. Excitotoxicity and ROS Production

Another agent responsible for brain damage caused by HI is the increase in extracellular glutamate concentration as mentioned above. Glutamate activates N-Methyl-D-Aspartate (NMDA) receptors, causing the accumulation of Ca^{2+} and nitric oxide (NO), and the production of ROS in turn (Fig. 1) [22].

One of the main organelles affected by energy failure is mitochondria, which plays a key role in cell death due to the control of energy metabolism, production of ROS, and the release of apoptotic factors into the cytoplasm. The most prominent pro-apoptotic factor is cyt-c [23]. Apoptosis-inducing mitochondrial proteins such as endoG, Smac/Diablo and Omi/HtrA2 have also been described playing an important role in apoptosis regulation [24]. The release of these pro-apoptotic factors is not necessarily mediated by Mitochondrial Permeability Transition (MPT) pores, indicating that changes in mitochondrial membrane potential ($\Delta\psi_m$) are directly related to necrosis and apoptosis [25]. At this stage, cell apoptosis is beginning, specifically mitochondrial apoptosis, which is characterized by an excessive release of Ca^{2+} , hyperpolarization of $\Delta\psi_m$, and an excessive production of ROS [26].

Another consequence of cell death caused by oxidative stress is cardiolipin peroxidation, one of the most critical targets [27]. Cardiolipin is a unique phospholipid, which is found mostly in the inner mitochondrial membrane, where it has a very close association with the components of oxidative phosphorylation [27]. Cardiolipin plays a crucial role in the function of cytochrome-C, cytochrome- C oxidase and other phosphorylation complexes. This is required, therefore, for an optimal functioning of complex I (NADH: ubiquinone reductase), complex III (NADH: ubiquinone cytochrome-C oxidoreductase), complex IV (cytochrome-C oxidase) and complex V (ATP synthase) [28]. An alteration in the structure of cardiolipin is responsible for mitochondrial dysfunction. Therefore, the release of cytochrome-c depends on the integrity of cardiolipin, which is highly sensitive to peroxidation by ROS, due to its high content of fatty acids [28].

2.3. Glial Cells and Neuroinflammation

Glial cells play differential roles in HI brain injury, mainly in inflammation. If glial response becomes chronic, it may potentiate the mechanisms of damage mentioned above, due to the secretion of a large amount of proinflammatory cytokines and ROS.

2.3.1. Microglia

Microglia, the immune cells of the CNS, are the first to be activated after HI. Microglia migrate to the place of injury, where they change their morphology to amoeboid-like functional cells, acting in conjunction with monocytes and macrophages [29-31]. Microglia present a dual behavior on inflammation caused by HI. Microglial M1 phenotype releases proinflammatory agents to the environment such as ROS, cytokines Interleukin-1beta, Interleukin-6 (IL-1 β , IL-6, TNF- α), glutamate, and nitric oxide, creating a cytotoxic environment which triggers cell death [29-31]. In contrast, microglial M2 phenotype is responsible for mediating an anti-inflammatory response, exerting neuroprotective properties by the release of anti-inflammatory agents and growth factors (insulin-like growth factor (IGF), transforming growth factor (TGF β), Glial cell-derived neurotrophic factor (GDNF), and Interleukin-10 (IL-10)) [29].

2.3.2. Endothelial Cells

The extent of injury noted in HI is not only determined by the biochemical cascades that trigger the apoptosis-necrosis continuum of cell death in the brain parenchyma, but also by the pro-inflammatory factors of the Blood Brain Barrier (BBB), such as the endothelial cells [32]. Endothelial cells can sense variation in the Partial Oxygen Pressure (PO_2) through different mechanosensors. Then, they can adapt their metabolism to maintain ATP production, switching into a hypoxic metabolism. In this way, endothelial cells augment the production of ROS by making the respiratory chain slower, reduce the cytochrome-c capacity in order to trap O_2 , and alter the cellular redox potential [33].

2.3.3. Astrocytes

In the last 20 years, astrocytes have been granted multiple functions, such as providing support to neurons, helping in the maintenance of the neural microenvironment, and regulating cerebral blood flow, which is necessary for the adequate function of neurons [2, 34]. Astrocytes also contribute to the accurate metabolic function of the brain [35]. These glial cells take glucose from blood vessels and provide energy metabolites to the neuron [36]. In addition, astrocytes provide lactate to neurons as a substrate for the citric acid cycle, supplying in this way their energy requirements [37].

However, the role of astrocytes in injuries such as HI is not fully elucidated [38]. Like microglia, astrocytes respond differently depending on the severity of the injury. Immediately after HI, astrocytes exhibit an activated state, which eventually leads to the formation of a glial scar [39, 40]. Astrocytes may exacerbate cytotoxicity death after the secretion of inflammatory cytokines such as IL-1, IL-6, interferon- γ , and TNF- α ; and help the migration of immune cells to the CNS by the secretion of chemokines [30]. On the other hand, astrocytes play a neuroprotective effect by promoting toler-

ance to cerebral ischemic injury [41] and attenuating neuroinflammation [41, 42].

3. EXPERIMENTAL MODELS

For the study of HI, *in vivo* and *in vitro* models are used (Table 1). The murine unilateral common carotid artery occlusion (UCCAO) represents a widely used model of HI. The UCCAO is followed by an exposure to an O₂ atmosphere of 8% for 1-3 hours [43], reproducing the anatomical injury caused by HI in neonates, which damages gray matter in hippocampus, thalamus and basal ganglia, as well as in white matter [43,44]. Similarly, it reproduces metabolic damage in parameters such as cerebral acidosis, decreased cerebral blood flow, and decreased glucose uptake [29], and it is useful for studying the neuroprotective effect of hypothermia [45, 46]. Bilateral common carotid artery occlusion (BCCAO) is also used to accentuate damage in white matter [47, 48]. UCCAO and BCCAO have been also used in larger animals such as primates, sheep, pigs and rabbits in order to better replicate the conditions of a human fetus with HI, with the disadvantage of not permitting the performance of behavioral tests [29, 44, 49, 50].

Another murine model of HI excludes ligation of the common carotid, causing damage only by O₂ deprivation. This model is used to reproduce milder and severe lesions, in order to investigate the biochemical alterations of the brain [29]. Under this concept there are several methods, in which the times of exposure to hypoxia and the percentage of O₂ are variable [29]. Concentrations vary from 2.6% O₂ for 30 min [51, 52], and 5% O₂ / 95% nitrogen (N₂) for 15 min [53]. Other methods provide O₂ atmospheres that decrease from 7% to 4% [54], or 9% O₂ for 1 hour [55]. There are other models of PA in rats that differ from the previous ones by the way in which the whole procedure is performed. Rat pups inside the uterine horns are subjected to a severe asphyxia by submerging them in a water bath at 37°C for 19 min [56], reproducing in this way a intrauterine global asphyxia, which is highly prevalent in humans. In addition, this model has been widely used for studying synaptic alterations caused by PA [57]. The lack of methodological homogeneity constitutes a disadvantage since it makes difficult to contrast the results in an optimal way [29, 44, 49, 50].

3.1. *In Vitro* Approaches

The different methodological limitations of *in vivo* models make *in vitro* models relevant. To replicate the conditions that occur in the presence of deprivation or decrease in glucose and O₂ levels such as those present in HI, several studies have proposed a model of O₂ and glucose deprivation (OGD) (Table 1) This model has the ability to adjust to specific research needs and the versatility of including different cell lines, making possible the study of molecular and biochemical mechanisms of HI injury. However, methodological differences have been found in the implementation of this model, especially in the exposure time of hypoxia and reoxygenation, making this model dependent on the specific conditions of the tissue or cells used. Different studies suggest hypoxia times of 1, 1.5, 2.3, 4, 6, 8, 16, 48 h and reoxygenation of 1, 2, 8, 12, 24, 48h [78-81, 84, 85, 87, 106].

Another methodological approach used to study the effects of hypoxia *in vitro* include the use of chemical agents known as hypoxia-mimetic agents (HMAs). These are based on producing at molecular level the effects caused by the low concentration of O₂, mainly the effects involved in the expression of Hypoxia-inducible factor-1 (HIF-1) [92, 93]. The activation of this factor depends on O₂ concentration, and HIF-1 is involved in several cellular processes that trigger hypoxia [107-112]. Hypoxia can be induced in normoxia by different types of compounds, among them: iron chelators, such as desferrioxamine (DFO) [113, 114] and divalent metal ions such as cobalt and nickel [114]. These agents can inactivate different enzymes associated to hypoxic processes substituting the ferrous group in regulatory dioxygenases. In addition, cobalt and nickel block the delivery of ferrous ions into cells since they bind much more tightly to the membrane transporter DMT-1 (Divalent Metal Transporter 1) in comparison to iron ions [114].

Within the HMAs, multiple compounds have been evaluated among these: 2-aminoanthracene, benzo(b)fluoranthene, benzo(k)fluoranthene, cobalt sulfate, dibenz(a,h)anthracene, 7-diethylamino-4-methylcoumarin, 7,12-dimethylbenz(a)anthracene, iodochlorohydroxyquinoline, o-phenanthroline, prednisone, salicylazosulfapyridine and triamterene, among others [114]. However, DFO and cobalt chloride (CoCl₂) are the most widely used compounds in *in vitro* models. DFO and CoCl₂ are also used in different *in vivo* experimental models [94, 95, 97-99-101, 104, 115-117]. DFO has also shown protective properties against hypoxia [98], including *in vitro* models of HI (Hamrick *et al.*, 2005). It is administered in preconditioning models in order to attenuate the damage [99, 101, 102]. Similarly, in murine models of PA it has been used as a combined treatment with erythropoietin, reducing the number of cleaved caspase 3 (+) cells [103]. In addition, it has been used as a neuroprotective agent in dopaminergic neuronal death [96].

Summarizing, these compounds have a wide versatility to be used in HI models. However, the use of HMAs to reproduce an HI event presents limitations, one of which is related to the regulatory mechanisms at transcriptional and post-transcriptional levels, since they use distinct regulatory mechanisms through the interplay between DNA and mRNA elements [115]. These limitations should be taken into account at the moment of implementation.

4. NEUROPROTECTIVE AGENTS TESTED IN HI

Neuroprotective agents tested in HI are classified according to their mechanism of action: antioxidation, anti-inflammation, anti-excitotoxicity and anti-apoptosis [19]. Allopurinol and aminoguanidine are antioxidant; ammonium pyrrolidinedithiocarbamate (PDCT), anti-inflammatory; memantine and topiramate, antiexcitotoxic; and indomethacin, antiapoptotic [19].

Within most studied therapies, Xenon appears as a noble gas used as an anesthetic agent, showing neuroprotective effects in *in vitro* and *in vivo* experimental models of HI brain injury. It could augment B-cell lymphoma 2 (Bcl-2) and brain-derived neurotrophic factor (BDNF) in asphyctic rats [19, 118]. In addition, a phase 1 study reported that

Table 1. Experimental models for HI.

Refs.	Species	Animal Model	Outcomes
Large Animal Models			
[50]	Macaca nemestrina, near term	UCO	Poor weight gain and cerebellar growth, abnormal brain DTI, behavioral impairment, 43% develop CP
[58, 59]	Fetal sheep, near term	Bilateral CCAO	Shorter HI (<30 min): selective neuronal loss. Longer HI: cortical necrosis. Post-HI EEG suppression related to insult severity and pathology; prevented by hypothermia
[47]	Fetal sheep, midgestation	Bilateral CCAO	Necrosis of subcortical white matter, neuronal loss in thalamus and striatum similar to near term fetus. Little loss of final EEG amplitude
[60]	Fetal sheep, midgestation and near Term	UCO	Hippocampal neuronal loss only in near term group. Degree of injury associated with the severity of hypotension during UCO
[61]	Pigs, <24h old	CCAO + hypoxia	Secondary energy failure. Energy metabolism ameliorated by hypothermia (35°C for 12h) at 24h-48h
[62]	Pigs, P9	Hypotension + hypoxia	~60% fall in CBF, reduced cerebral O ₂ uptake, phosphorylated metabolites and pH and increased inorganic phosphate
[44]	Rabbits, 21–22 d gestation	Uterine ischemia	P1 pups: overt posture and tone after ischemia > 37 min, correlates with microgliosis in basal ganglia and thalamus. MRI: WMI in IC
Rodent Models with Global Hypoxic or Excitotoxic Component			
[63]	Mice at E8, P0 or P5	Ibotenate, i.c.v.	laminar neuronal depopulation of layer V–VIa. P5: neuronal loss in all cortical layers, formation of porencephalic cysts
[64]	Pregnant Sprague-Dawley rats, embryonic	Hypoxia E5-E20	White matter cysts in offspring P0–P7, increased lipid peroxidation, WMI and macrophages
Rodent Models with Hypoxia-Ischemia			
[65, 66]	Sprague Dawley rats, P1–P3	CCAL + hypoxia	Selective vulnerability of late OL progenitors, independent of age Death of sub-plate neurons, motor deficits, altered thalamocortical connections to somatosensory and visual cortex normal
[43]	Sprague-Dawley rats, P7	CCAL + hypoxia	Unilateral ischemic injury in the cortex, hippocampus, basal ganglia in > 90% of survivors
[67]	Wistar rat, P7	LPS, 4h prior to CCAL + hypoxia	Blocking lymphocyte trafficking reduced brain inflammation, BBB damage, and improved LPS-induced HI brain injury. No effect with pure HI
[68]	C57BL/6 WT, Tg SOD1, GPx1 over-expressing P7 mice	CCAL + hypoxia	Reduced injury in GPx1-Tg mice but not in SOD1-Tg or GPx1/SOD1. NOS inhibition did not improve outcome in SOD-Tg
[69, 70]	C57BL/6 WT and Gal-3 KO, P9	CCAL + hypoxia	Increased BBB permeability 2–24h, reduced BBB protein expression. Infarct volume reduction in Gal-3 KO mice
[71]	C57BL/6J and TRIF KO mice, P8–9	Poly I:C, 14h prior to CCAL + hypoxia	Increased infarct volume and WMI, prevented in TRIF KO. Injury linked to inflammatory response & decrease in M2-like microglia

(Table 1) contd....

Refs.	Species	Animal Model	Outcomes
Focal Ischemia Rodent Models			
[72]	Wistar rat, P7	Permanent MCAO + 1h CCAO	Infarcts in frontoparietal cortex at 3-month recovery. DNA fragmentation from 6–96h
[73-75]	Sprague Dawley rats, P7	Transient MCAO, 3h	Severe unilateral perfusion deficits, restoration of CBF upon suture removal. Decreased ADC associated with brain injury at 24h reperfusion. Demonstrated endogenous neuroprotective role of microglial cells after acute injury
[76]	Sprague Dawley rats, P10	Transient MCAO, 1.5h	Time resolved cell-type specific increase in HIF-1 α and VEGF expression, gliosis
[77]	C57/Bl6 mice, CD36 KO and WT, P9	Transient MCAO, 1.5h and 3h	Focal ischemia-reperfusion, increased injury and caspase-3 cleavage associated with apoptotic neuronal debris in CD36 KO. Effects independent of NF κ B activation
In Vitro Models			
Refs.	Cell Line	Experimental Model	Outcomes
[78]	PC12 cells	48 h OGD/ 2h reperfusion	Significant morphological cell changes
[79]	Primary cortical astrocyte	6 h OGD/ 0, 12, 24, 48 h reperfusion	Significantly increased 2- NBDG uptake by about 1.2 to 2.5 times in cells compared to control
[80]	Primary cerebral cortex neurons	3 h OGD/ 48 h Reperfusion	Damage to neuronal viability, dendrite branch number in neurons decreased significantly
[81]	Primary astrocyte	3, 5, 7 h OGD/ 24 h Reoxygenation	Increases in HMGB1 and TNF- α , induced phosphorylation of PI3K, promoted nuclear translocation of NF- κ B
[82]	primary cortical neurons	2 h OGD	Suppressed significantly cortical neurons proliferation
[83]	SH-SY-5Y cells	6 h OGD/ 1h reoxygenation	Caused significant mitochondrial fragmentation, excessive mitochondrial fission
[84]	Primary Cortical Neuron	OGD	Decrease in neurite outgrowth
[85]	Neural progenitor cell	6 h OGD	Increased apoptosis
[86]	mouse hippocampal neurons HT22	4 h OGD/ 24 h Reoxygenation	miR-144-3p expression was significantly downregulated in neurons following OGD/R treatment
[87]	Neuro 2a cells	4 h OGD/ 12 h Reoxygenation	Inhibited cell viability and cell proliferation, reduced phosphorylation levels of p38 MAPK and ERK1/2
[88]	SH-SY5Y cells and primary murine cortical neurons	4 h OGD	OGDR-induced mitochondrial depolarization, reactive oxygen species production, lipid peroxidation and DNA damages
[89]	Primary astrocytes and microglial cells	2 h OGD/ 48 h Reoxygenation	Induced abnormally opened hemichannels with increased ATP release and EtBr uptake but reduced GJIC permeability Astrocytic Cx43, hemichannels, and GJIC play critical roles in OGD/R injury-induced neuroinflammatory responses
[90]	Primary astrocytes	4 h OGD/ 3 h, 6 h, 12 h, 24h reoxygenation	Expression of Ski was proved to be up-regulated
[91]	Primary hippocampal neurons	2 h OGD/ 24 h reperfusion	Caspase-3 activity and expression increased in the first 24 h
HMA's Models			
Refs.	Cell Line/species	Experimental Model	Outcomes
[92]	multiple myeloma cell line U266	CoCl ₂	CoCl ₂ -mediated hypoxia affects the expression profiles of genes that are functionally related to apoptosis and angiogenesis
[93]	myeloid leukemic cell lines NB4 and U937	CoCl ₂ and DFO	Apoptosis with a loss of mitochondrial transmembrane potentials, activation of caspase-3/8 and cleavage of anti-apoptotic protein Mcl-1
[94]	U251 human glioblastoma cell line	CoCl ₂	Increases HIF-1 α gene expression

(Table 1) contd....

Refs.	Cell Line/species	Experimental Model	Outcomes
[95]	glioblastoma cell lines U373MG and DBTRG05MG	DFO	Activation of factors associated with ECM degradation and invasion of glioma cells
[96]	C57BL/6 mice	DFO	DFO up-regulated the expression of vascular endothelial growth factor (VEGF), HIF-1 α protein and growth associated protein 43 (GAP43) and down-regulated the expression of divalent metal transporter with iron-responsive element (DMT1+IRE), α -synuclein, and transferrin receptor (TFR)
[97]	Hippocampal neurons	DFO pretreatment/3h OGD	45% reduction in cell death
[98]	Sprague-Dawley rats	subarachnoid hemorrhage/DFO treatment	DFO-induced increase in HIF-1 protein level and activity exerts significant attenuation of BA vasospasm
[99]	Hippocampal cultures	Ppreconditioning CoCl ₂ , DFO or dimethylloxylalylglycine (DMOG), 3h OGD	Cobalt induced the transcription of the cytokine erythropoietin. cobalt and DFO, enhanced survival of neurons. DMOG exacerbates OGD-induced neuronal death
[100]	Sprague-Dawley rats	CCA/DFO treatment	Neural-protective and angiogenesis effects through regulating the levels of HIF-1 α
[101]	adipose-derived stem cells	DFO preconditioning	Restored neovascularization potential of ADSCs
[102]	Sprague – Dawley rats	MCA/DFO treatment	Preserved brain volumes, upregulation of HIF1 α
[103]	Wistar rats	MCAO/DFO+Erythropoietin treatment	Reduced the number of cleaved caspase 3-positive cells in the ipsilateral cerebral cortex.
[104]	Human mesenchymal stem cells	CoCl ₂ and DFO	Proliferation of hMSCs was inhibited by DFO and CoCl

Modified from [105].

breathing 50% of xenon for 18h didn't show adverse effects for newborns even after 18 months follow-up. Nevertheless, the high-cost and difficulty in the administration of xenon are the main barriers for its clinical use [19].

Melatonin was found to be protective in MCAO at post-natal day 12 (P12). This effect was probably mediated by inhibition of autophagy and neuronal cell death as well as enhanced Growth Associated Protein 43 (GAP43) expression [119]. Erythropoietin (EPO) is a naturally occurring hormone produced with angiogenic action. It presents neuroprotective, neurogenic, anti-inflammatory and anti-apoptotic properties [20]. In a rodent model of adult stroke, rhEPO increased vascular endothelial growth factor (VEGF) and brain-derived neurotrophic factor (BDNF), reducing the size of stroke infarct and improving functional outcomes [20].

A novel drug, orientin, is under study in a rat cortical neurons primary culture model of OGD. This molecule is a water-soluble flavonoid C-glycoside, commonly extractable from different plants like the passion fruit with antioxidant properties. The pretreatment dose-dependently inhibited OGD/Reperfusion (RP) induced phosphorylation of c-Jun N-terminal kinases (JNK) and Extracellular signal-regulated kinases (ERK)1/2. In addition, JNK inhibitor SP600125 and ERK1/2 inhibitor PD98059 ameliorated OGD/RP-induced cell death and ROS generation. Further, orientin failed to protect cortical neurons with the JNK activator anisomycin or with the ERK1/2 activator FGF-2 [6], showing that the neuroprotective effects of orientin against OGD/RP-induced damage depend on down-regulation of JNK and ERK1/2

signaling pathways [6]. In a model of OGD/R injury in a primary co-culture of rat cortical capillary endothelial cells, astrocytes and neurons to allow simultaneous observations in the same system, it was found that pretreatment with Ginkgo biloba extract or Ginkgolide B increased neuronal viability, ameliorated cell injury, and inhibited cellular apoptotic rate through Bcl-2 and Bcl-2-associated X protein (Bax) expression [7]. On the other hand, allopurinol, a xanthine oxidase inhibitor and free radical scavenger, reduced cerebral edema and brain damage induced by perinatal HI in P7 rat pups. Nonetheless, a meta-analysis of three clinical trials showed no significant benefits for neonates with HI encephalopathy treated with allopurinol [19].

Stem cells have been used in clinical trials to treat stroke on adults. Keeping in mind that stroke in adults and HI in neonates share many symptoms, neuroprotective treatments for stroke might also be effective in neonates [120]. Umbilical Cord Blood Cells (UCBs) have shown anti-inflammatory and anti-apoptotic properties in animal models of HI encephalopathy. UCBs are composed of different cells such as lymphocytes, endothelial progenitors, hematopoietic stem cells, monocytes and mesenchymal stem cells (MSCs) [20]. Following an injury, MSCs promote neuronal differentiation, augment cellular regeneration and reduce immune cell response [20].

Norm baric hyperoxia (NBO) has been shown to exhibit neuro- and vaso-protective effects by improving tissue oxygenation when it is given during ischemia. However, the effect of NBO might diminish when the duration of ischemia and reperfusion is extended. For that reason, it should be

administered together with drugs that could enhance the protective effects [121].

4. 1. Therapeutic Hypothermia

Currently, the only available treatment for PA is therapeutic hypothermia (TH) [20]. TH is the unique treatment available after resuscitation from moderate- to-severe perinatal HI. It should be started as soon as possible, at least within 6 h after birth, and continued for 72 h [18]. Using TH before the onset of the secondary energy failure prevents the development of cytotoxic edema, cell death and brain damage [18, 122].

It has been shown that mild hypothermia could reduce or inhibit secondary impaired oxidative phosphorylation, augmenting ATP and correcting Protein C reactive (PCr) levels after HI [18]. TH protects the brain by reducing brain metabolism in almost 5% for every degree of temperature lowering [122]. Reducing cerebral metabolism provokes the attenuation of oxidative stress, excitotoxicity, inflammation, cell death pathways and the accumulation of essential amino acids due to the delay in the depolarization [18, 122]. The efficacy of TH can be clinically and experimentally tested with the use of prognostic 1H magnetic resonance spectroscopy (MRS) biomarkers [18].

Immediate use of TH after HI reduces caspase-3 expression in oligodendrocytes and cortical infarct at P7, an age when murine brain development presents similarities with the late preterm human infant [122]. By the same way, TH has shown reduction of apoptosis in neurons after 24 hours, decreasing the activation of caspases-3, -8, and -9, and the migration of cyt-c [122]. TH seems to have beneficial effects on astrocytes as well. In a neonatal-rat brain slice OGD model, an augment of glutamine was observed in the TH group. This change was registered after the injection of a mixture of glucose and acetate, which converts glucose into glutamine *via* the pyruvate carboxylase, and is related only to astrocytes metabolism [18, 123]. Similarly, intra ischemic hypothermia maintains mitochondrial activity in neonatal rats after 4 days of recovery. Nevertheless, it is unclear if TH directly protects the mitochondria or if this effect depends on the suppression of apoptosis and inflammation [122, 124].

Finally, a meta-analysis of randomized clinical trials showed that TH is beneficial for term infants with moderate-to-severe HI encephalopathy, producing a relative risk reduction of mortality or neurodevelopmental disabilities [18, 125]. However, mild hypothermia treatment after intravenous thrombolysis in patients with acute stroke did not show protective effects [126, 127]. Therefore, it becomes necessary to find drugs that enhance the efficacy of mild hypothermia [18, 127].

4.2. The Neuroprotective Role of 17 β -estradiol in HI

Neuroactive steroids (NASs) are naturally or artificially synthesized steroids which exert actions on the brain. Most of the naturally synthesized steroids are made as *de novo* from cholesterol [128, 129]. NASs are classified as pregnane, androstane, and sulfated neurosteroids (NSs) [129]. They regulate different processes interacting with: gamma-aminobutyric acid (GABA), microtubule-associated protein

2 (MAP-2), NMDA, serotonin, α -adrenoreceptors, transient receptor potential channels, voltage-gated calcium channels, voltage-dependent anion channels, neurotrophins, nerve growth factor, and σ 1 receptors [129]. Other molecules, such as synthetic steroid analogues, are used for exerting estrogenic effects with major stability. Several studies have shown 3 α -hydroxyl configuration is the key for binding and activating these molecules, but modifications in the steroid nucleus may emphasize different pharmacophores [130].

A derivate of estrogen, 17 β -estradiol, is a female sex hormone and NAS related to the development of secondary sexual characteristics, fat storage and regulation of menstrual cycle [131]. 17 β -estradiol, which was originally administered as a hormone replacement therapy in order to ameliorate climacteric symptoms, showed beneficial effects in verbal and visual memory performance [132]. The activity of 17 β -estradiol depends on its union with estrogen receptors (ERs) [35, 132, 133]. These receptors are classified in two subtypes: the estrogen receptor-alpha (ER- α) and the estrogen receptor-beta (ER- β). Locus from ER- α and ER- β are chromosome 6 and chromosome 14, respectively [132]. These ERs are transcription factors which present the peculiarity of being activated by a ligand. ER- α and ER- β have a similar structure, with a DNA-binding domain and a ligand-binding domain [134]. 17 β -estradiol binds to ERs and induces the activation and the homodimerization or heterodimerization of these receptors. Then, the ERs bind to estrogen-responsive elements (EREs) in the promoter region of specific genes through the DNA-binding domain, recruiting transcriptional co-activators and co-repressors [134, 135]. Classical ERs may also regulate gene transcription by acting as transcriptional partners at non-ERE sites, such as activating protein 1 (AP1) sites [136]. 17 β -estradiol can bind to membrane-associated non-classical ERs, such as G protein-coupled ERs (GPERs). GPER30, a member of the G protein-coupled receptor superfamily, regulates the activity of extracellular signal-regulated kinases (ERKs) and the phosphoinositide 3-kinase (PI3K) signaling pathway. This union allows the interaction with the signaling of other neuroprotective molecules [134, 137]. Another membrane-associated non-classical ER is G α q protein-coupled membrane ER (Gq-mER), which was originally identified in hypothalamic neurons, modulating μ -opioid and GABA neurotransmission [134, 138].

Estrogen and its derivatives may exert their activity through genomic and non-genomic responses [134]. However, these responses might be overlapped [139]. Genomic responses require the translocation to the nucleus, binding to EREs and transcription factors in order to achieve a long-lasting response [140, 141]. Estrogenic non-genomic response involves signaling mechanisms that allow estrogen to accomplish rapid and dramatic effects [142]. These responses act through a second messenger, generating several signal transduction cascades, such as ion fluxes (specially calcium), cyclic AMP modulation and protein kinase pathways, giving an acute response between seconds or minutes, mainly through PI3K or MAPK pathways [140, 143].

In a murine model of memory consolidation, 17 β -estradiol enhanced hippocampal memory *via* rapid activation of Erk MAPK and PI3K/Akt [144, 145], suggesting that 17 β -

estradiol could increase spinogenesis in hippocampus and learning experience through kinase cascades [146]. Tozzi et al. (2015) evaluated the interactions between ERs and dopamine (DA) receptors in dorsal striatum of adult male rats, showing that inhibition of 17 β -estradiol synthesis by aromatase P-450 inhibition prevented the induction of long-term potentiation (LTP) in both medium spiny neurons (MSNs) and cholinergic interneurons (ChIs). Downstream estrogen signaling pathway activated D1-like DA receptor/cAMP/Protein kinase A (PKA)-dependent pathway, restoring LTP, and exogenous 17 β -estradiol administration reversed the effect of the aromatase P-450 inhibition., confirming 17 β -estradiol is involved in LTP of MSNs [147].

Non-genomic actions of estrogens in different cell contexts usually are mediated by GPR30, functionally cross-reacting with diverse cell signaling systems such as the epidermal growth factor receptor (EGFR) pathway, Notch and mitogen-activated protein kinases (MAPK) signaling pathways [148]. 17 β -estradiol exerts neuroprotective effects against ischemia by activation of GPR30, which is linked to transactivation of Insuline-like growth factor 1 (IGF-1) receptor [149]. 17 β -estradiol increases the expression of basic fibroblast growth factor (bFGF) in astrocytes [150], inducing neuroprotection against ischemia and glutamate-induced excitotoxic neuronal cell death [151-153]. These findings have led to research on the neuroprotective properties of estrogen and its derivatives on brain injury.

In HI brain injury 17 β - estradiol has shown several neuroprotective effects, such as: reducing reactive gliosis, decreasing oxidative stress, ameliorating the release of pro inflammatory molecules, preventing cell death and mitochondrial dysfunction, releasing neurotrophic factors [154]. It has also been reported that 17 β - estradiol may exert protective actions against OGD-induced cell death in primary oligodendrocytes. In this way, 17 β - estradiol presents neuroprotective properties in hypoxic-ischemic and oxidative injury [155].

A repeated dosing paradigm (three doses of 17 β -estradiol) in neonates subjected to HI provided approximately 70% protection in the hippocampus, basal ganglia, and amygdala [156]. In addition, treatment with estradiol after PA augmented the expression of IGF-1 and its receptor (IGF-IR). The PI3K/Akt/GSK3 signaling pathway was activated as an increase in Akt and GSK3 phosphorylation [157]. However, it has been found that male sex is a well-established epidemiological risk factor for poor neurodevelopmental outcome after PA. While the mechanisms responsible for this gender difference are unknown, growing evidence has identified neuro-inflammation, oxidative stress and cell death pathways as key players in these differences [158].

Using a mice model of MCAO with a mutant form of ER- α , neuroprotection was absent, showing that protective properties depend on ER- α [159]. Similarly, after emulating hypoxia in the neuroblastoma cell line SH-SY5Y by using CoCl₂ (250 μ g/mL), a hypoxic mimetic agent, treatment with 17 β -estradiol (250 nM) exerted neuroprotection. Afterwards, using ER- α and ER- β agonist (PPT and DPN, respectively) without 17 β -estradiol treatment, results showed neu-

roprotection was mimicked by PPT and suggested that ER- α regulates this protective effect [157].

Likewise, in a model of astrocytic cells it was found that treatment with estradiol improved HI parameters such as cell viability and mitochondrial membrane potential, besides reducing ROS production and preventing the loss of mitochondrial mass [160]. Nevertheless, estrogen might have detrimental effects, such as the augmentation in breast and uterus cancer incidence [10-12]. In order to maintain the benefits of estrogen and avoid side effects, synthetic steroid analogues have been developed. These group of drugs includes SERMs, such as raloxifene and tamoxifene, and STEARs, like tibolone [10-12].

The mechanism of regulation of SERMs that determines if they act either as agonist or antagonist in a specific cell type depends on the predominant subtype of estrogen receptor α or β . The regions of union to the ERs are: activation function 1 (AF-1), capable of binding with the DNA, and activation function 2 (AF-2), a region of union with the ligand in order to achieve an estrogenic response. The ligand must bind to the AF-2 region of the ER and migrate to the nucleus to bind the response site. Co-activators, co-factors and helper proteins of each cell will determine the kind of response of the tissue exposed to SERMs [161, 162].

In rats subjected to MCAO and treated with estrogen and Raloxifene, neurogenesis in the ipsilateral subventricular zone (SVZ) after ischemia was significantly higher in estrogen and raloxifene-treated animals compared to rats treated with placebo. Tamoxifen did not show this enhancing effect on neurogenesis. However, both tamoxifen and raloxifene as well as estrogen, significantly reversed spine density loss observed in the ischemic cortex at day-5 post ischemia [163].

Tibolone action is given by the metabolization of tibolone to three different metabolites (δ -4 tibolone, α -hydroxy tibolone and 3- β -hydroxy tibolone). Each of them produces different responses. δ -4 tibolone is an agonist of androgen and progesterone receptor, while α -hidroxy and β -hidroxy tibolone are antagonists of those receptors but agonists of ERs [164]. Keeping this in mind, it was found tibolone could ameliorate the effects of GD in an *in vitro* model of astrocytes, making these molecules interesting for further research in an OGD model [11].

5. COMBINED THERAPIES

5.1. Combined Therapies Using TH

The lack of effective treatments against HI and the little success of clinical trials for different treatments, lead to a search for new and better therapies. Since TH is the most widespread therapy against the deleterious effects of HI injury, it has become the basis for the development of combined therapies or co-treatments [20]. Combined treatments aim to use the protective effects of each of its components (Table 2). TH can prevent the development of brain edema, as well as reducing the damage produced by inflammation, oxidative stress and excitotoxicity [18]. Similarly, TH has also shown the ability to improve mitochondrial autophagy [106] and prevent apoptosis by preventing the translocation of the apoptosis-inducing factor (AIF) to the nucleus [165].

Table 2. Summary of combined therapies.

Refs.	Therapy	Model	Outcomes
[178]	Erythropoietin+ Hypothermia	Randomized trials in infants	preterm infants assigned to receive rhEPO showed that the rate of moderate/severe neurological disability in the rhEPO group was (7.1%)significantly lower compared to the placebo group (18.8%; $p < 0.001$)
[179]	Melatonin + Hypothermia	Piglet model of HI	Improved white matter tract development and reduced apoptosis
[166]	Xenon + Hypothermia	Asphyxiated Piglets	Xenon with hypothermia, offers histopathological and functional neuroprotection
[167]	Mesenchymal stromal cells + Hypothermia	Primary Neurons from rats	shown neuroprotection in rat primary neurons preserving cell viability
[127]	Mild hypothermia + glibenclamide; dizocilpine; neuroglobin	Primary cortical Neurons	co-treatment of those three drugs and mild hypothermia decreased ROS and intracellular calcium accumulation and stabilized mitochondrial membrane potential (MMP)
[177]	17 β -estradiol + Hypothermia	Rats with HI	low dose of 17 β -estradiol after the ischemic event exerted neuroprotective effects for the transient global ischemia, and its effect is potentiated by therapeutic hypothermia
[180]	CIMT + EA	Rats with HI	-CIMT combined with EA significantly reduced motor asymmetry after Hi. augmented NeuN and reduced GFAP expression in the cortex
[181]	Minocycline + NBO	Rats on MCAO	Neuro- and vaso-protective effects by inhibiting matrix metallo-proteinase (MMP)-2/9-mediated occludin degradation and attenuation of caspase-dependent and independent apoptotic pathways
[182]	Ederavone + NBO	Mice HI reperfusion	Combined treatment reduced the infarct zone volume, getting better neurological functions, in cortex and subcortex after 22 hours of reperfusion
[183]	Cilostazol + NBO	Mice with HI	the combined therapy ameliorated the damage done by focal cerebral ischemia/reperfusion injury, by enhancing the blood flow to the damaged area after the reperfusion, and this effect was related to eNOS activity

Some combined treatments approved for clinical trials involve EPO combined with hypothermia. Neonates with PA treated with a combination of rhEPO and hypothermia showed improved neurological outcomes, as well as fewer white matter tract abnormalities. Melatonin combined with clinical hypothermia improved white matter tract development and reduced apoptosis in a piglet model of HI [20].

Xenon, a noble gas that exerts neuroprotective effect by up-regulating Bcl-2 pro-survival proteins [18], offers histopathological and functional neuroprotection in combination with hypothermia [166]. Another combined therapy that has shown promising results in rat primary neurons is the treatment with mesenchymal stromal Cells and hypothermia, which preserves cell viability [167]. Finally, a recent study evaluated the neuroprotective activity of 26 different medicines with neuroprotective potential in combination with TH in cortical neurons subjected to OGD, finding a significant neuroprotective effect using BDNF, glibenclamide (GBC), kallidinogenase (HUK), (1) -MK-801 ((1) -Dizocilpine hydrogen maleate (MK-801) and neuroglobin (Ngb).

Taking into account the few options of combined treatments to date, it is imperative to deepen the search for new therapies that help combat the negative effects caused by HI. SERMs and STEARs represent a promising option in conjunction with hypothermia. As for SERMs, in murine models of MCAO raloxifene increases neurogenesis in the ipsilateral

subventricular zone, and tamoxifen together with raloxifene and estrogen reverse the loss of spine density [163]. Similarly, raloxifene has shown neuroprotective G protein-coupled receptor 30 (GPR30) -dependent activity in an OGD model of rat neuronal cells [168]. Likewise, raloxifene exerted neuroprotective effects on neocortical cells in a caspase-3-independent manner [169, 170]. In addition, raloxifene and tamoxifen have the ability to reduce microglial activation in rats after HI brain injury [171].

With regards to STEARs, tibolone may exert neuroprotective effects in cerebellum and hippocampus, reducing tau hyper phosphorylation and modulating Glycogen synthase kinase-3b (GSK3b) phosphorylation [172]. Additionally, tibolone has shown a protective effect against oxidative stress and lipid peroxidation [173, 174]. It also has the ability to modulate reactive gliosis and reduce the number of microglial cells with a reactive phenotype, exerting beneficial effects on the homeostasis of cerebral cortex [175]. This STEAR is also capable of modulating the expression of Ngb in astrocytic cells in a model of GD [176].

Keeping in mind the neuroprotective effects exerted by the SERMs and STEARs, it is possible to suggest that the multifactorial neuroprotective action of TH may act synergistically in a possible combination treatment with these compounds, since they also exert their beneficial action from various points of action. Although research on the neuropro-

tective effect of TH combined with SERMs or STEARs is missing, combination treatment with 17 β -estradiol and TH has shown greater efficacy than monotherapies [177], suggesting NASSs might potentiate the protective effects of TH. The distinctive but complementary protective mechanisms of NASSs and TH might be responsible for the synergistic effect of a combined therapy [177].

5.2. Other Combined Therapies

Other combined therapies (Table 2) do not include TH but have not been approved for clinical trials yet. A combined therapy of constraint-induced movement therapy (CIMT) and electroacupuncture (EA) was tested in neonatal rats with HI-induced hemiplegia. CIMT consists in the inhibition of the normal limb, forcing the animals to use the damaged one. EA is based on using needles in acupuncture points and applying electrical stimulation. This combined therapy has shown effective results in stroke rehabilitation and hemiplegia [180]. CIMT combined with EA reduced motor asymmetry after HI, but HI lesion size was not improved. This treatment augmented NeuN and reduced GFAP expression in the cortex, suggesting it targets both neurons and astrocytes. In addition, CIMT combined with EA reduced the expression of cleaved caspase-3, an apoptotic mediator [180].

Combined therapy using NBO and minocycline provided greater neuro- and vaso-protective effects than monotherapy, by inhibiting matrix metallo-proteinase (MMP)-2/9-mediated occludin degradation and attenuating caspase-dependent and independent apoptotic pathways [121,181]. NBO combined with edaravone, a scavenger of hydroxyl radicals, exerted greater effects than monotherapies by reducing the infarct zone volume and improving neurological functions in cortex and subcortex after 22 hours reperfusion [121,182]. Cilostazol combined with NBO has been reported to protect the brain after cerebral ischemia in rodents, ameliorating the damage caused by focal cerebral ischemia/reperfusion injury. This effect was associated with an enhancement of blood flow to the damaged area after reperfusion, which is related to eNOS activity [121,183].

Tissue plasminogen activator (tPA) can solve blood clots to restore blood flow in the ischemic brain region. Until now, the administration of tPA within 3 or 4.5 hours after symptom onset is the only FDA approved treatment for acute ischemic stroke. However, its combination with NBO has the potential to expand the therapeutic time window for tPA administration and to reduce subsequent reperfusion induced injury [121]. Similarly, combination therapy of Umbilical cord blood cells (UCBs) and EPO resulted in significant improvements in motor and cognition, with associated improvements in structural and metabolic changes in the brain [20].

CONCLUSION

HI events exhibit a high impact on society. Over the years, research has intended to find effective treatments that counteract the damage caused by HI. Neuroprotective targets include specific points of damage caused by HI, such as oxidative stress, dysregulation of the cell cycle and energy ho-

meostasis [20]. Both in the initial damage phase and in the final one, the different neuroprotective agents may exert anti-inflammatory, antioxidant, anti-excitotoxicity or anti-apoptotic effects [19]. However, due to the complex network of factors that influence these pathologies, including cellular interactions inherent to the CNS as well as gender-dependent response [158], the effectiveness of neuroprotective treatments has not been optimal.

Several studies have focused on TH, which with its multifactorial action represents the only clinical treatment available for human use. However, TH has shown a relative success. Bearing this in mind, research has currently focused on the search for new neuroprotective agents that carry out their activity from different fronts, whereupon SERMs and STEARS arise as an alternative [10-12]. *In vitro* and *in vivo* studies should be developed to give a better understanding of the processes involved for a better implementation of the treatments.

Following this logic, the search for treatments that not only focus their attention on improving a specific parameter seem to be a better alternative. Combined treatments or co-treatments respond to this need. These assume TH as a therapeutic basis by combining it with different drugs [13]. Investigations of the mechanisms by which this combination therapy exerts its neuroprotection are currently taking place. Likewise, SERMs and STEARS emerge as promising candidates in the implementation of a combination therapy due to their benefits in different parameters. Further research is necessary for proving this hypothesis.

CONSENT FOR PUBLICATION

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CONFLICT OF INTEREST

The authors declare no conflict of interest, financial or otherwise.

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REFERENCES

- [1] Bélanger, M.; Allaman, I.; Magistretti, P.J. Brain energy metabolism: focus on astrocyte-neuron metabolic cooperation. *Cell Metab.*, **2011**, *14*(6), 724-738. [<http://dx.doi.org/10.1016/j.cmet.2011.08.016>] [PMID: 22152301]
- [2] Allaman, I.; Bélanger, M.; Magistretti, P.J. Astrocyte-neuron metabolic relationships: for better and for worse. *Trends Neurosci.*, **2011**, *34*(2), 76-87. [<http://dx.doi.org/10.1016/j.tins.2010.12.001>] [PMID: 21236501]
- [3] Dwyer, D.S.; Vannucci, S.J.; Simpson, I.A. Expression, regulation, and functional role of glucose transporters (GLUTs) in brain. *Int. Rev. Neurobiol.*, **2002**, *51*, 159-188. [[http://dx.doi.org/10.1016/S0074-7742\(02\)51005-9](http://dx.doi.org/10.1016/S0074-7742(02)51005-9)] [PMID: 12420359]
- [4] Vavilis, T.; Delivanoglou, N.; Aggelidou, E.; Stamoula, E.; Melidlis, K.; Kaidoglou, A.; Cheva, A.; Pourzitaki, C.; Chatzimeletiou,

- K.; Lazou, A.; Albani, M.; Kritis, A. Oxygen-glucose deprivation (OGD) modulates the unfolded protein response (UPR) and inflicts autophagy in a PC12 Hypoxia cell line model. *Cell. Mol. Neurobiol.*, **2016**, *36*(5), 701-712.
- [5] Salvador, E.; Burek, M.; Förster, C.Y. Stretch and/or oxygen glucose deprivation (OGD) in an *in vitro* traumatic brain injury (TBI) model induces calcium alteration and inflammatory cascade. *Front. Cell. Neurosci.*, **2015**, *9*, 323. [http://dx.doi.org/10.3389/fncel.2015.00323] [PMID: 26347611]
- [6] Tian, T.; Zeng, J.; Zhao, G.; Zhao, W.; Gao, S.; Liu, L. Neuroprotective effects of orientin on oxygen-glucose deprivation/reperfusion-induced cell injury in primary culture of rat cortical neurons. *Exp. Biol. Med. (Maywood)*, **2017**, *243* (1), 78-86. 10.1177/1535370217737983. [PMID: 29073777]
- [7] Yang, X.; Zheng, T.; Hong, H.; Cai, N.; Zhou, X.; Sun, C.; Wu, L.; Liu, S.; Zhao, Y.; Zhu, L.; Fan, M.; Zhou, X.; Jin, F. Neuroprotective effects of Ginkgo Biloba extract and ginkgolide B against oxygen-glucose deprivation/reoxygenation and Gglucose injury in a new *in vitro* multicellular network model. *Front. Med.* **2018**, *12*(3), 307-318.
- [8] Mozaffarian, D.; Benjamin, E. J.; Go, A. S.; Arnett, D. K.; Blaha, M. J.; Cushman, M.; de Ferranti, S.; Després, J.-P.; Fullerton, H. J.; Howard, V. J. Heart disease and stroke statistics—2015 update. *Circulation.*, **2015**, *131*(4), e29-322.
- [9] Northington, F.J.; Chavez-Valdez, R.; Martin, L.J. Neuronal cell death in neonatal hypoxia-ischemia. *Ann. Neurol.*, **2011**, *69*(5), 743-758. [http://dx.doi.org/10.1002/ana.22419] [PMID: 21520238]
- [10] Arevalo, M. A.; Santos-Galindo, M.; Lagunas, N.; Azcoitia, I.; Garcia-Segura, L. M. Selective estrogen receptor modulators as brain therapeutic agents. *J. Mol. Endocrinol.* **2011**, *46*(1), R1-R9.
- [11] Ávila, R.M.; Garcia-Segura, L. M.; Cabezas, R.; Torrente, D.; Capani, F.; Gonzalez, J.; Barreto, G. E. Tibolone protects T98G cells from glucose deprivation. *J. Steroid Biochem. Mol. Biol.*, **2014**, *144*(PART B), 294-303.
- [12] Garzón, D.; Cabezas, R.; Vega, N.; Ávila-Rodríguez, M.; Gonzalez, J.; Gómez, R.M.; Echeverría, V.; Aliev, G.; Barreto, G.E. Novel approaches in astrocyte protection: From experimental methods to computational approaches. *J. Mol. Neurosci.*, **2016**, *58*(4), 483-492. [http://dx.doi.org/10.1007/s12031-016-0719-6] [PMID: 26803310]
- [13] Cilio, M.R.; Ferriero, D.M. Synergistic neuroprotective therapies with hypothermia. *Semin. Fetal Neonatal Med.*, **2010**, *15*(5), 293-298. [http://dx.doi.org/10.1016/j.siny.2010.02.002] [PMID: 20207600]
- [14] Berger, H.R.; Brekke, E.; Widerøe, M.; Morken, T.S. Neuroprotective Treatments after perinatal hypoxic-ischemic brain injury evaluated with magnetic resonance spectroscopy. *Dev. Neurosci.*, **2017**, *39*(1-4), 36-48. [http://dx.doi.org/10.1159/000472709] [PMID: 28448965]
- [15] Fleiss, B.; Gressens, P. Tertiary mechanisms of brain damage: a new hope for treatment of cerebral palsy? *Lancet Neurol.*, **2012**, *11*(6), 556-566. [http://dx.doi.org/10.1016/S1474-4422(12)70058-3] [PMID: 22608669]
- [16] Li, B.; Concepcion, K.; Meng, X.; Zhang, L. Brain-immune interactions in perinatal hypoxic-ischemic brain injury. *Prog. Neurobiol.*, **2017**, *159*, 50-68. [http://dx.doi.org/10.1016/j.pneurobio.2017.10.006] [PMID: 29111451]
- [17] Chen, X.; Guo, C.; Kong, J. Oxidative stress in neurodegenerative diseases. *Neural Regen. Res.*, **2012**, *7*(5), 376-385. [PMID: 25774178]
- [18] Berger, H.R.; Brekke, E.; Widerøe, M.; Morken, T.S. Neuroprotective treatments after perinatal hypoxic-ischemic brain injury evaluated with magnetic resonance spectroscopy. *Dev. Neurosci.*, **2017**, *39*(1-4), 36-48. [http://dx.doi.org/10.1159/000472709] [PMID: 28448965]
- [19] Wu, Q.; Chen, W.; Sinha, B.; Tu, Y.; Manning, S.; Thomas, N.; Zhou, S.; Jiang, H.; Ma, H.; Kroessler, D. A.; Yao, J.; Liz, Z.; Inder, T. E.; Wang, X. Neuroprotective agents for neonatal hypoxic-ischemic brain injury. *Drug Discov. Today*, **2015**, *20*(11), 1372-1381.
- [20] Leaw, B.; Nair, S.; Lim, R.; Thornton, C.; Mallard, C.; Hagberg, H. Mitochondria, bioenergetics and excitotoxicity: New therapeutic targets in perinatal brain injury. *Front. Cell. Neurosci.*, **2017**, *11*, 199. [http://dx.doi.org/10.3389/fncel.2017.00199] [PMID: 28747873]
- [21] Descloux, C.; Ginot, V.; Clarke, P.G.H.; Puyal, J.; Truttmann, A.C. Neuronal death after perinatal cerebral hypoxia-ischemia: Focus on autophagy-mediated cell death. *Int. J. Dev. Neurosci.*, **2015**, *45*, 75-85. [http://dx.doi.org/10.1016/j.ijdevneu.2015.06.008] [PMID: 26225751]
- [22] Weber, J.T. Altered calcium signaling following traumatic brain injury. *Front. Pharmacol.*, **2012**, *3*, 60. [http://dx.doi.org/10.3389/fphar.2012.00060] [PMID: 22518104]
- [23] Ouyang, Y.-B.; Giffard, R.G. Cellular neuroprotective mechanisms in cerebral ischemia: Bcl-2 family proteins and protection of mitochondrial function. *Cell Calcium*, **2004**, *36*(3-4), 303-311. [http://dx.doi.org/10.1016/j.ceca.2004.02.015] [PMID: 15261486]
- [24] Wang, C.; Youle, R.J. The role of mitochondria in apoptosis*. *Annu. Rev. Genet.*, **2009**, *43*, 95-118. [http://dx.doi.org/10.1146/annurev-genet-102108-134850] [PMID: 19659442]
- [25] Jacobson, J.; Duchon, M.R. Mitochondrial oxidative stress and cell death in astrocytes—requirement for stored Ca²⁺ and sustained opening of the permeability transition pore. *J. Cell Sci.*, **2002**, *115*(Pt 6), 1175-1188. [PMID: 11884517]
- [26] Kadenbach, B.; Hüttemann, M.; Arnold, S.; Lee, I.; Bender, E. Mitochondrial energy metabolism is regulated via nuclear-coded subunits of cytochrome c oxidase. *Free Radic. Biol. Med.*, **2000**, *29*(3-4), 211-221. [http://dx.doi.org/10.1016/S0891-5849(00)00305-1] [PMID: 11035249]
- [27] Kagan, V.E.; Chu, C.T.; Tyurina, Y.Y.; Cheikhi, A.; Bayir, H. Cardiolipin asymmetry, oxidation and signaling. *Chem. Phys. Lipids*, **2014**, *179*, 64-69. [http://dx.doi.org/10.1016/j.chemphyslip.2013.11.010] [PMID: 24300280]
- [28] Paradies, G.; Petrosillo, G.; Paradies, V.; Ruggiero, F.M. Role of cardiolipin peroxidation and Ca²⁺ in mitochondrial dysfunction and disease. *Cell Calcium*, **2009**, *45*(6), 643-650. [http://dx.doi.org/10.1016/j.ceca.2009.03.012] [PMID: 19368971]
- [29] Millar, L.J.; Shi, L.; Hoerder-Suabedissen, A.; Molnár, Z. Neonatal hypoxia ischaemia: Mechanisms, models, and therapeutic challenges. *Front. Cell. Neurosci.*, **2017**, *11*, 78. [http://dx.doi.org/10.3389/fncel.2017.00078] [PMID: 28533743]
- [30] Rocha-Ferreira, E.; Hristova, M. Antimicrobial peptides and complement in neonatal hypoxia-ischemia induced brain damage. *Front. Immunol.*, **2015**, *6*, 56. [http://dx.doi.org/10.3389/fimmu.2015.00056] [PMID: 25729383]
- [31] Ziemka-Nalecz, M.; Jaworska, J.; Zaleska, T. Insights into the neuroinflammatory responses after neonatal hypoxia-ischemia. *J. Neuroinflammation. Exp. Neurol.*, **2017**, *76*(8), 644-654. [http://dx.doi.org/10.1093/jnen/nlx046] [PMID: 28789477]
- [32] Lee, W.L.A.; Michael-Titus, A.T.; Shah, D.K. Hypoxic-ischaemic encephalopathy and the blood-brain barrier in neonates. *Dev. Neurosci.*, **2017**, *39*(1-4), 49-58. [http://dx.doi.org/10.1159/000467392] [PMID: 28434009]
- [33] Paternotte, E.; Gaucher, C.; Labrude, P.; Stoltz, J.F.; Menu, P. Review: Behaviour of endothelial cells faced with hypoxia. *Bio-med. Mater. Eng.*, **2008**, *18*(4-5), 295-299. [PMID: 19065037]
- [34] Bélanger, M.; Magistretti, P.J. The role of astroglia in neuroprotection. *Dialogues Clin. Neurosci.*, **2009**, *11*(3), 281-295. [PMID: 19877496]
- [35] Karki, P.; Webb, A.; Zerguine, A.; Choi, J.; Son, D.S.; Lee, E. Mechanism of raloxifene-induced upregulation of glutamate transporters in rat primary astrocytes. *Glia*, **2014**, *62*(8), 1270-1283. [http://dx.doi.org/10.1002/glia.22679] [PMID: 24782323]
- [36] Guíllamón-Vivancos, T.; Gómez-Pinedo, U.; Matías-Guio, J. Astrocytes En Las enfermedades neurodegenerativas (I): Función y caracterización molecular. *Neurología*, **2015**, *30*(2), 119-129. [http://dx.doi.org/10.1016/j.nrl.2012.12.007] [PMID: 23465689]
- [37] Fuller, S.; Steele, M.; Münch, G. Activated astroglia during chronic inflammation in Alzheimer's disease—do they neglect their neurosupportive roles? *Mutat. Res.*, **2010**, *690*(1-2), 40-49. [http://dx.doi.org/10.1016/j.mrfmmm.2009.08.016] [PMID: 19748514]
- [38] Romero, J.; Muñoz, J.; Logica, T.T.; Holubiec, M.; González, J.; Barreto, G.E.; Guelman, L.; Lillig, C.H.; Blanco, E.; Capani, F. Dual role of astrocytes in perinatal asphyxia injury and neuropro-

- tection. *Neurosci. Lett.*, **2014**, *565*, 42-46. [http://dx.doi.org/10.1016/j.neulet.2013.10.046] [PMID: 24172702]
- [39] Lee, K.M.; MacLean, A.G. New advances on glial activation in health and disease. *World J. Virol.*, **2015**, *4*(2), 42-55. [http://dx.doi.org/10.5501/wjv.v4.i2.42] [PMID: 25964871]
- [40] Sullivan, S.M.; Björkman, S.T.; Miller, S.M.; Colditz, P.B.; Pow, D.V. Morphological changes in white matter astrocytes in response to hypoxia/ischemia in the neonatal pig. *Brain Res.*, **2010**, *1319*, 164-174. [http://dx.doi.org/10.1016/j.brainres.2010.01.010] [PMID: 20079338]
- [41] Hirayama, Y.; Koizumi, S. Astrocytes and ischemic tolerance. *Neurosci. Res.* **2018**, *126*, 53-59.
- [42] Sofroniew, M. V. Astrocyte barriers to neurotoxic inflammation. *Nat Rev. Neurosci.*, **2015**, *16*(5), 249-263. [http://dx.doi.org/10.1038/nrn3898]
- [43] Rice, J.E., III; Vannucci, R.C.; Brierley, J.B. The influence of immaturity on hypoxic-ischemic brain damage in the rat. *Ann. Neurol.*, **1981**, *9*(2), 131-141. [http://dx.doi.org/10.1002/ana.410090206] [PMID: 7235629]
- [44] Derrick, M.; Drobyshevsky, A.; Ji, X.; Tan, S. A model of cerebral palsy from fetal hypoxia-ischemia. *Stroke*, **2007**, *38*(2)(Suppl.), 731-735. [http://dx.doi.org/10.1161/01.STR.0000251445.94697.64] [PMID: 17261727]
- [45] Kida, H.; Nomura, S.; Shinoyama, M.; Ideguchi, M.; Owada, Y.; Suzuki, M. The effect of hypothermia therapy on cortical laminar disruption following ischemic injury in neonatal mice. *PLoS One*, **2013**, *8*(7), e68877. [http://dx.doi.org/10.1371/journal.pone.0068877] [PMID: 23894362]
- [46] Lin, E.P.; Miles, L.; Hughes, E.A.; McCann, J.C.; Vorhees, C.V.; McAuliffe, J.J.; Loepke, A.W. A combination of mild hypothermia and sevoflurane affords long-term protection in a modified neonatal mouse model of cerebral hypoxia-ischemia. *Anesth. Analg.*, **2014**, *119*(5), 1158-1173. [http://dx.doi.org/10.1213/ANE.0000000000000262] [PMID: 24878681]
- [47] Reddy, K.; Mallard, C.; Guan, J.; Marks, K.; Bennet, L.; Gunning, M.; Gunn, A.; Gluckman, P.; Williams, C. Maturation change in the cortical response to hypoperfusion injury in the fetal sheep. *Pediatr. Res.*, **1998**, *43*(5), 674-682. [http://dx.doi.org/10.1203/00006450-199805000-00017] [PMID: 9585015]
- [48] Domnguez, R.; Zitting, M.; Liu, Q.; Patel, A.; Babadjouni, R.; Hodis, D. M.; Chow, R. H.; Mack, W. J. Estradiol protects white matter of male C57BL/6J mice against experimental chronic cerebral hypoperfusion. *J. Stroke Cerebrovasc. Dis.* **2018**, *27*(7), 1743-1751.
- [49] Derrick, M.; Drobyshevsky, A.; Ji, X.; Chen, L.; Yang, Y.; Ji, H.; Silverman, R.B.; Tan, S. Hypoxia-ischemia causes persistent movement deficits in a perinatal rabbit model of cerebral palsy: assessed by a new swim test. *Int. J. Dev. Neurosci.*, **2009**, *27*(6), 549-557. [http://dx.doi.org/10.1016/j.ijdevneu.2009.06.008] [PMID: 19573586]
- [50] Traudt, C.M.; McPherson, R.J.; Bauer, L.A.; Richards, T.L.; Burbacher, T.M.; McAdams, R.M.; Juul, S.E. Concurrent erythropoietin and hypothermia treatment improve outcomes in a term nonhuman primate model of perinatal asphyxia. *Dev. Neurosci.*, **2013**, *35*(6), 491-503. [http://dx.doi.org/10.1159/000355460] [PMID: 24192275]
- [51] Anju, T.R.; Paulose, C.S. Amelioration of hypoxia-induced striatal 5-HT(2A) receptor, 5-HT transporter and HIF1 alterations by glucose, oxygen and epinephrine in neonatal rats. *Neurosci. Lett.*, **2011**, *502*(3), 129-132. [http://dx.doi.org/10.1016/j.neulet.2011.05.236] [PMID: 21683764]
- [52] Anju, T.R.; Paulose, C.S. Striatal cholinergic functional alterations in hypoxic neonatal rats: role of glucose, oxygen, and epinephrine resuscitation. *Biochem. Cell Biol.*, **2013**, *91*(5), 350-356. [http://dx.doi.org/10.1139/bcb-2012-0102] [PMID: 24032686]
- [53] Rodriguez-Alvarez, N.; Jimenez-Mateos, E.M.; Dunleavy, M.; Waddington, J.L.; Boylan, G.B.; Henshall, D.C. Effects of hypoxia-induced neonatal seizures on acute hippocampal injury and later-life seizure susceptibility and anxiety-related behavior in mice. *Neurobiol. Dis.*, **2015**, *83*, 100-114. [http://dx.doi.org/10.1016/j.nbd.2015.08.023] [PMID: 26341542]
- [54] Sampath, D.; Shmueli, D.; White, A.M.; Raol, Y.H. Flupirtine effectively prevents development of acute neonatal seizures in an animal model of global hypoxia. *Neurosci. Lett.*, **2015**, *607*, 46-51. [http://dx.doi.org/10.1016/j.neulet.2015.09.005] [PMID: 26365409]
- [55] Helmy, M.M.; Tolner, E.A.; Vanhatalo, S.; Voipio, J.; Kaila, K. Brain alkalosis causes birth asphyxia seizures, suggesting therapeutic strategy. *Ann. Neurol.*, **2011**, *69*(3), 493-500. [http://dx.doi.org/10.1002/ana.22223] [PMID: 21337602]
- [56] Herrera, M.I.; Udovin, L.D.; Toro-Urrego, N.; Kusnier, C.F.; Luaces, J.P.; Capani, F. Palmitoylethanolamide ameliorates hippocampal damage and behavioral dysfunction after perinatal asphyxia in the immature rat brain. *Front. Neurosci.*, **2018**, *12*, 145. [http://dx.doi.org/10.3389/fnins.2018.00145] [PMID: 29662433]
- [57] Herrera, M.I.; Otero-Losada, M.; Udovin, L.D.; Kusnier, C.; Kölliker-Frers, R.; de Souza, W.; Capani, F. Could perinatal asphyxia induce a synaptopathy? new highlights from an experimental model. *Neural Plast.*, **2017**, *2017*3436943 [http://dx.doi.org/10.1155/2017/3436943] [PMID: 28326198]
- [58] Tan, W.K.M.; Williams, C.E.; Gunn, A.J.; Mallard, C.E.; Gluckman, P.D. Suppression of postischemic epileptiform activity with MK-801 improves neural outcome in fetal sheep. *Ann. Neurol.*, **1992**, *32*(5), 677-682. [http://dx.doi.org/10.1002/ana.410320511] [PMID: 1449248]
- [59] Gunn, A.J.; Gunn, T.R.; de Haan, H.H.; Williams, C.E.; Gluckman, P.D. Dramatic neuronal rescue with prolonged selective head cooling after ischemia in fetal lambs. *J. Clin. Invest.*, **1997**, *99*(2), 248-256. [http://dx.doi.org/10.1172/JCI119153] [PMID: 9005993]
- [60] Mallard, E.C.; Williams, C.E.; Johnston, B.M.; Gluckman, P.D. Increased vulnerability to neuronal damage after umbilical cord occlusion in fetal sheep with advancing gestation. *Am. J. Obstet. Gynecol.*, **1994**, *170*(1 Pt 1), 206-214. [http://dx.doi.org/10.1016/S0002-9378(94)70409-0] [PMID: 8296824]
- [61] Thoresen, M.; Penrice, J.; Lorek, A.; Cady, E.B.; Wylezinska, M.; Kirkbride, V.; Cooper, C.E.; Brown, G.C.; Edwards, A.D.; Wyatt, J.S. Mild hypothermia after severe transient hypoxia-ischemia ameliorates delayed cerebral energy failure in the newborn piglet. *Pediatr. Res.*, **1995**, *37*(5), 667-670. [http://dx.doi.org/10.1203/00006450-199505000-00019] [PMID: 7603788]
- [62] Laptook, A.R.; Hassan, A.; Peterson, J.; Corbett, R.J.; Nunnally, R.L. Effects of repeated ischemia on cerebral blood flow and brain energy metabolism. *NMR Biomed.*, **1988**, *1*(2), 74-79. [http://dx.doi.org/10.1002/nbm.1940010204] [PMID: 3275028]
- [63] Gressens, P.; Marret, S.; Evrard, P. Developmental spectrum of the excitotoxic cascade induced by ibotenate: A model of hypoxic insults in fetuses and neonates. *Neuropathol. Appl. Neurobiol.*, **2018**, *22*(6), 498-502.
- [64] Baud, O.; Daire, J.-L.; Dalmaz, Y.; Fontaine, R.H.; Krueger, R.C.; Sebag, G.; Evrard, P.; Gressens, P.; Verney, C. Gestational hypoxia induces white matter damage in neonatal rats: a new model of periventricular leukomalacia. *Brain Pathol.*, **2004**, *14*(1), 1-10. [http://dx.doi.org/10.1111/j.1750-3639.2004.tb00492.x] [PMID: 14997932]
- [65] Sheldon, R.A.; Chuai, J.; Ferriero, D.M. A rat model for hypoxic-ischemic brain damage in very premature infants. *Biol. Neonate*, **1996**, *69*(5), 327-341. [http://dx.doi.org/10.1159/000244327] [PMID: 8790911]
- [66] Back, S.A.; Han, B.H.; Luo, N.L.; Chricton, C.A.; Xanthoudakis, S.; Tam, J.; Arvin, K.L.; Holtzman, D.M. Selective vulnerability of late oligodendrocyte progenitors to hypoxia-ischemia. *J. Neurosci.*, **2002**, *22*(2), 455-463. [http://dx.doi.org/10.1523/JNEUROSCI.22-02-00455.2002] [PMID: 11784790]
- [67] Yang, D.; Sun, Y.-Y.; Bhaumik, S. K.; Li, Y.; Baumann, J. M.; Lin, X.; Zhang, Y.; Lin, S.-H.; Dunn, R. S.; Liu, C.-Y.; Shie, F.S.; Lee, Y. H.; Wills-Karp, M.; Chougnat, C.A.; Kallapur, S.G.; Lewkowich, I.P.; Lindquist, D.M.; Murali-Krishna, K.; Kuan, C.Y. Blocking lymphocyte trafficking with FTY720 prevents inflammation-sensitized hypoxic-ischemic brain injury in newborns. *J. Neurosci.*, **2014**, *34*(49), 16467-16481.
- [68] Sheldon, R.A.; Jiang, X.; Francisco, C.; Christen, S.; Vexler, Z.S.; Täuber, M.G.; Ferriero, D.M. Manipulation of antioxidant pathways in neonatal murine brain. *Pediatr. Res.*, **2004**, *56*(4), 656-662. [http://dx.doi.org/10.1203/01.PDR.0000139413.27864.50] [PMID: 15295091]

- [69] Doverhag, C.; Hedtj rn, M.; Poirier, F.; Mallard, C.; Hagberg, H.; Karlsson, A.; S vman, K. Galectin-3 contributes to neonatal hypoxic-ischemic brain injury. *Neurobiol. Dis.*, **2010**, *38*(1), 36-46. [http://dx.doi.org/10.1016/j.nbd.2009.12.024] [PMID: 20053377]
- [70] Ek, C.J.; D'Angelo, B.; Baburamani, A.A.; Lehner, C.; Leverin, A.-L.; Smith, P.L.; Nilsson, H.; Svedin, P.; Hagberg, H.; Mallard, C. Brain barrier properties and cerebral blood flow in neonatal mice exposed to cerebral hypoxia-ischemia. *J. Cereb. Blood Flow Metab.*, **2015**, *35*(5), 818-827. [http://dx.doi.org/10.1038/jcbfm.2014.255] [PMID: 25627141]
- [71] Hagberg, H.; Mallard, C.; Ferriero, D.M.; Vannucci, S.J.; Levison, S.W.; Vexler, Z.S.; Gressens, P. The role of inflammation in perinatal brain injury. *Nat. Rev. Neurol.*, **2015**, *11*(4), 192-208. [http://dx.doi.org/10.1038/nrneurol.2015.13] [PMID: 25686754]
- [72] Renolleau, S.; Aggoun-Zouaoui, D.; Ben-Ari, Y.; Charriaut-Marlangue, C. A model of transient unilateral focal ischemia with reperfusion in the P7 neonatal rat: morphological changes indicative of apoptosis. *Stroke*, **1998**, *29*(7), 1454-1460. [http://dx.doi.org/10.1161/01.STR.29.7.1454] [PMID: 9660403]
- [73] Derugin, N.; Ferriero, D.M.; Vexler, Z.S. Neonatal reversible focal cerebral ischemia: a new model. *Neurosci. Res.*, **1998**, *32*(4), 349-353. [http://dx.doi.org/10.1016/S0168-0102(98)00096-0] [PMID: 9950062]
- [74] Fern andez-L pez, D.; Faustino, J.; Daneman, R.; Zhou, L.; Lee, S.Y.; Derugin, N.; Wendland, M.F.; Vexler, Z.S. Blood-brain barrier permeability is increased after acute adult stroke but not neonatal stroke in the rat. *J. Neurosci.*, **2012**, *32*(28), 9588-9600. [http://dx.doi.org/10.1523/JNEUROSCI.5977-11.2012] [PMID: 22787045]
- [75] Faustino, J.V.; Wang, X.; Johnson, C.E.; Klibanov, A.; Derugin, N.; Wendland, M.F.; Vexler, Z.S. Microglial cells contribute to endogenous brain defenses after acute neonatal focal stroke. *J. Neurosci.*, **2011**, *31*(36), 12992-13001. [http://dx.doi.org/10.1523/JNEUROSCI.2102-11.2011] [PMID: 21900578]
- [76] Mu, D.; Jiang, X.; Sheldon, R.A.; Fox, C.K.; Hamrick, S.E.G.; Vexler, Z.S.; Ferriero, D.M. Regulation of hypoxia-inducible factor 1 α and induction of vascular endothelial growth factor in a rat neonatal stroke model. *Neurobiol. Dis.*, **2003**, *14*(3), 524-534. [http://dx.doi.org/10.1016/j.nbd.2003.08.020] [PMID: 14678768]
- [77] Woo, M.-S.; Wang, X.; Faustino, J.V.; Derugin, N.; Wendland, M.F.; Zhou, P.; Iadecola, C.; Vexler, Z.S. Genetic deletion of CD36 enhances injury after acute neonatal stroke. *Ann. Neurol.*, **2012**, *72*(6), 961-970. [http://dx.doi.org/10.1002/ana.23727] [PMID: 23280844]
- [78] Elkordy, A.; Mishima, E.; Niizuma, K.; Akiyama, Y.; Fujimura, M.; Tominaga, T.; Abe, T. Stress-induced tRNA cleavage and tRNA generation in rat neuronal PC12 cells. *J. Neurochem.*, **2018**, *146*(5), 560-569. [http://dx.doi.org/10.1111/jnc.14321] [PMID: 29431851]
- [79] Chen, Y.; Zhang, J.; Zhang, X.Y. 2-NBDG as a marker for detecting glucose uptake in reactive astrocytes exposed to oxygen-glucose deprivation *in vitro*. *J. Mol. Neurosci.*, **2015**, *55*(1), 126-130. [http://dx.doi.org/10.1007/s12031-014-0385-5] [PMID: 25091860]
- [80] Cui, X.; Fu, Z.; Wang, M.; Nan, X.; Zhang, B. Pitavastatin treatment induces neuroprotection through the BDNF-TrkB signalling pathway in cultured cerebral neurons after oxygen-glucose deprivation. *Neurol. Res.* **2018**, *40*(5), 391-397.
- [81] Dong, Y.-F.; Guo, R.-B.; Ji, J.; Cao, L.-L.; Zhang, L.; Chen, Z.-Z.; Huang, J.-Y.; Wu, J.; Lu, J.; Sun, X.-L. S1PR3 is essential for phosphorylated fingolimod to protect Astrocytes against oxygen-glucose deprivation-induced neuroinflammation via inhibiting TLR2/4-NF κ B signalling. *J. Cell. Mol. Med.*, **2018**, *22*(6), 3159-3166.
- [82] Feng, S.-J.; Zhang, X.-Q.; Li, J.-T.; Dai, X.-M.; Zhao, F. miRNA-223 regulates ischemic neuronal injury by targeting the type 1 insulin-like growth factor receptor (IGF1R). *Folia Neuropathol.*, **2018**, *56*(1), 49-57. [http://dx.doi.org/10.5114/fn.2018.74659] [PMID: 29663740]
- [83] Guo, M.; Wang, X.; Zhao, Y.; Yang, Q.; Ding, H.; Dong, Q.; Chen, X.; Cui, M. Ketogenic diet improves brain ischemic tolerance and inhibits NLRP3 inflammasome activation by preventing Drp1-mediated mitochondrial fission and endoplasmic reticulum Stress. *Front. Mol. Neurosci.*, **2018**, *11*, 86. [http://dx.doi.org/10.3389/fnmol.2018.00086] [PMID: 29662437]
- [84] He, W.; Liu, Y.; Tian, X. Rosuvastatin improves neurite outgrowth of cortical neurons against oxygen-glucose deprivation *via* Notch1-mediated mitochondrial biogenesis and functional improvement. *Front. Cell. Neurosci.*, **2018**, *12*, 6. [http://dx.doi.org/10.3389/fncel.2018.00006] [PMID: 29387001]
- [85] Kim, M.; Jung, K.; Kim, I.-S.; Lee, I.-S.; Ko, Y.; Shin, J.E.; Park, K.I. TNF- α induces human neural progenitor cell survival after oxygen-glucose deprivation by activating the NF- κ B pathway. *Exp. Mol. Med.*, **2018**, *50*(4), 14. [http://dx.doi.org/10.1038/s12276-018-0033-1] [PMID: 29622770]
- [86] Li, Y.; Zhao, Y.; Cheng, M.; Qiao, Y.; Wang, Y.; Xiong, W.; Yue, W. Suppression of microRNA-144-3p attenuates oxygen-glucose deprivation/reoxygenation-induced neuronal injury by promoting Brg1/Nrf2/ARE signaling. *J. Biochem. Mol. Toxicol.*, **2018**, *32*(4), e22044. [http://dx.doi.org/10.1002/jbt.22044] [PMID: 29457851]
- [87] Wang, K.; Zhu, Y. Dexmedetomidine protects against oxygen-glucose deprivation/reoxygenation injury-induced apoptosis *via* the p38 MAPK/ERK signalling pathway. *J. Int. Med. Res.*, **2018**, *46*(2), 675-686. [http://dx.doi.org/10.1177/0300060517734460] [PMID: 29210287]
- [88] Weng, Y.; Lin, J.; Liu, H.; Wu, H.; Yan, Z.; Zhao, J. AMPK activation by Tanshinone IIA protects neuronal cells from oxygen-glucose deprivation. *Oncotarget*, **2017**, *9*(4), 4511-4521. [PMID: 29435120]
- [89] Yin, X.; Feng, L.; Ma, D.; Yin, P.; Wang, X.; Hou, S.; Hao, Y.; Zhang, J.; Xin, M.; Feng, J. Roles of astrocytic connexin-43, hemichannels, and gap junctions in oxygen-glucose deprivation/reperfusion injury induced neuroinflammation and the possible regulatory mechanisms of salvianolic acid B and carbenoxolone. *J. Neuroinflamm.*, **2018**, *15*(1), 97. [http://dx.doi.org/10.1186/s12974-018-1127-3] [PMID: 29587860]
- [90] Zhao, X.; Zhou, K.-S.; Li, Z.-H.; Nan, W.; Wang, J.; Xia, Y.-Y.; Zhang, H.-H. Knockdown of Ski decreased the reactive astrocytes proliferation *in vitro* induced by oxygen-glucose deprivation/reoxygenation. *J. Cell. Biochem.*, **2018**, *119*(6), 4548-4558. [http://dx.doi.org/10.1002/jcb.26597] [PMID: 29236326]
- [91] Zhou, T.; Lin, H.; Jiang, L.; Yu, T.; Zeng, C.; Liu, J.; Yang, Z. Mild hypothermia protects hippocampal neurons from oxygen-glucose deprivation injury through inhibiting caspase-3 activation. *Cryobiology*, **2018**, *80*, 55-61.
- [92] Bae, S.; Jeong, H.J.; Cha, H.J.; Kim, K.; Choi, Y.M.; An, I.S.; Koh, H.J.; Lim, D.J.; Lee, S.J.; An, S. The hypoxia-mimetic agent cobalt chloride induces cell cycle arrest and alters gene expression in U266 multiple myeloma cells. *Int. J. Mol. Med.*, **2012**, *30*(5), 1180-1186. [http://dx.doi.org/10.3892/ijmm.2012.1115] [PMID: 22941251]
- [93] Guo, M.; Song, L.-P.; Jiang, Y.; Liu, W.; Yu, Y.; Chen, G.-Q. Hypoxia-mimetic agents desferrioxamine and cobalt chloride induce leukemic cell apoptosis through different hypoxia-inducible factor-1 α independent mechanisms. *Apoptosis*, **2006**, *11*(1), 67-77. [http://dx.doi.org/10.1007/s10495-005-3085-3] [PMID: 16374551]
- [94] Al Okail, M.S. Cobalt chloride, a chemical inducer of hypoxia-inducible factor-1 α in U251 human glioblastoma cell line. *J. Saudi Chem. Soc.*, **2010**, *14*(2), 197-201. [http://dx.doi.org/10.1016/j.jscs.2010.02.005]
- [95] Elstner, A.; Holtkamp, N.; von Deimling, A. Involvement of Hif-1 in desferrioxamine-induced invasion of glioblastoma cells. *Clin. Exp. Metastasis*, **2007**, *24*(1), 57-66. [http://dx.doi.org/10.1007/s10585-007-9057-y] [PMID: 17357815]
- [96] Guo, C.; Hao, L.-J.; Yang, Z.-H.; Chai, R.; Zhang, S.; Gu, Y.; Gao, H.-L.; Zhong, M.-L.; Wang, T.; Li, J.-Y.; Wang, Z. Y. Deferoxamine-mediated up-regulation of HIF-1 α prevents dopaminergic neuronal death *via* the activation of MAPK family proteins in MPTP-treated mice. *Exp. Neurol.*, **2016**, *280*, 13-23.
- [97] Hamrick, S.E.G.; McQuillen, P.S.; Jiang, X.; Mu, D.; Madan, A.; Ferriero, D.M. A role for hypoxia-inducible factor-1 α in desferoxamine neuroprotection. *Neurosci. Lett.*, **2005**, *379*(2), 96-100. [http://dx.doi.org/10.1016/j.neulet.2004.12.080] [PMID: 15823423]
- [98] Hishikawa, T.; Ono, S.; Ogawa, T.; Tokunaga, K.; Sugiu, K.; Date, I. Effects of deferoxamine-activated hypoxia-inducible factor-1 on the brainstem after subarachnoid hemorrhage in rats. *Neurosurgery*, **2008**, *62*(1), 232-240.

- [http://dx.doi.org/10.1227/01.NEU.0000311082.88766.33] [PMID: 18300912]
- [99] Jones, S.M.; Novak, A.E.; Elliott, J.P. The role of HIF in cobalt-induced ischemic tolerance. *Neuroscience*, **2013**, *252*, 420-430. [http://dx.doi.org/10.1016/j.neuroscience.2013.07.060] [PMID: 23916558]
- [100] Li, L.; Yin, X.; Ma, N.; Lin, F.; Kong, X.; Chi, J.; Feng, Z. Desferrioxamine regulates HIF-1 alpha expression in neonatal rat brain after hypoxia-ischemia. *Am. J. Transl. Res.*, **2014**, *6*(4), 377-383. [PMID: 25075254]
- [101] Mehrabani, M.; Najafi, M.; Kamarul, T.; Mansouri, K.; Iranpour, M.; Nematollahi, M.H.; Ghazi-Khansari, M.; Sharifi, A.M. Deferoxamine preconditioning to restore impaired HIF-1 α -mediated angiogenic mechanisms in adipose-derived stem cells from STZ-induced type 1 diabetic rats. *Cell Prolif.*, **2015**, *48*(5), 532-549. [http://dx.doi.org/10.1111/cpr.12209] [PMID: 26332145]
- [102] Mu, D.; Chang, Y.S.; Vexler, Z.S.; Ferriero, D.M. Hypoxia-inducible factor 1 α and erythropoietin upregulation with deferoxamine salvage after neonatal stroke. *Exp. Neurol.*, **2005**, *195*(2), 407-415. [http://dx.doi.org/10.1016/j.expneurol.2005.06.001] [PMID: 16023639]
- [103] van der Kooij, M.A.; Groenendaal, F.; Kavelaars, A.; Heijnen, C.J.; van Bel, F. Combination of deferoxamine and erythropoietin: Therapy for hypoxia-ischemia-induced brain injury in the neonatal rat? *Neurosci. Lett.*, **2009**, *451*(2), 109-113. [http://dx.doi.org/10.1016/j.neulet.2008.12.013] [PMID: 19103262]
- [104] Zeng, H.-L.; Zhong, Q.; Qin, Y.-L.; Bu, Q.-Q.; Han, X.-A.; Jia, H.-T.; Liu, H.-W. Hypoxia-mimetic agents inhibit proliferation and alter the morphology of human umbilical cord-derived mesenchymal stem cells. *BMC Cell Biol.*, **2011**, *12*(1), 32. [http://dx.doi.org/10.1186/1471-2121-12-32] [PMID: 21827650]
- [105] Mallard, C.; Vexler, Z.S. Modeling ischemia in the immature brain: How translational are animal models? *Stroke*, **2015**, *46*(10), 3006-3011. [http://dx.doi.org/10.1161/STROKEAHA.115.007776] [PMID: 26272384]
- [106] Wang, Z.; Guo, L.M.; Wang, Y.; Zhou, H.K.; Wang, S.C.; Chen, D.; Huang, J.F.; Xiong, K. Inhibition of HSP90 α protects cultured neurons from oxygen-glucose deprivation induced necrosis by decreasing RIP3 expression. *J. Cell. Physiol.*, **2018**, *233*(6), 4864-4884. [http://dx.doi.org/10.1002/jcp.26294] [PMID: 29334122]
- [107] Bordt, E.A. The importance of controlling *in vitro* oxygen tension to accurately model *in vivo* neurophysiology. *Neurotoxicology*, **2018**, *66*, 213-220. [http://dx.doi.org/10.1016/j.neuro.2017.10.008] [PMID: 29102646]
- [108] Khan, M.; Khan, H.; Singh, I.; Singh, A.K. Hypoxia inducible factor-1 alpha stabilization for regenerative therapy in traumatic brain injury. *Neural Regen. Res.*, **2017**, *12*(5), 696-701. [http://dx.doi.org/10.4103/1673-5374.206632] [PMID: 28616019]
- [109] Semenza, G.L. Hypoxia-inducible factor 1: master regulator of O₂ homeostasis. *Curr. Opin. Genet. Dev.*, **1998**, *8*(5), 588-594. [http://dx.doi.org/10.1016/S0959-437X(98)80016-6] [PMID: 9794818]
- [110] Ke, Q.; Costa, M. Hypoxia-inducible factor-1 (HIF-1). *Mol. Pharmacol.*, **2006**, *70*(5), 1469-1480.
- [111] Huang, L.E.; Gu, J.; Schau, M.; Bunn, H.F. Regulation of hypoxia-inducible factor 1 α is mediated by an O₂-dependent degradation domain via the ubiquitin-proteasome pathway. *Proc. Natl. Acad. Sci. USA*, **1998**, *95*(14), 7987-7992. [http://dx.doi.org/10.1073/pnas.95.14.7987] [PMID: 9653127]
- [112] Wenger, R.H.; Gassmann, M. Oxygen(es) and the hypoxia-inducible factor-1. *Biol. Chem.*, **1997**, *378*(7), 609-616. [PMID: 9278140]
- [113] Wang, G.L.; Semenza, G.L. Desferrioxamine induces erythropoietin gene expression and hypoxia-inducible factor 1 DNA-binding activity: implications for models of hypoxia signal transduction. *Blood*, **1993**, *82*(12), 3610-3615. [PMID: 8260699]
- [114] Xia, M.; Huang, R.; Sun, Y.; Semenza, G.L.; Aldred, S.F.; Witt, K.L.; Inglese, J.; Tice, R.R.; Austin, C.P. Identification of chemical compounds that induce HIF-1 α activity. *Toxicol. Sci.*, **2009**, *112*(1), 153-163. [http://dx.doi.org/10.1093/toxsci/kfp123] [PMID: 19502547]
- [115] Huang, B.-W.; Miyazawa, M.; Tsuji, Y. Distinct regulatory mechanisms of the human ferritin gene by hypoxia and hypoxia mimetic cobalt chloride at the transcriptional and post-transcriptional levels. *Cell. Signal.*, **2014**, *26*(12), 2702-2709. [http://dx.doi.org/10.1016/j.cellsig.2014.08.018] [PMID: 25172425]
- [116] Müller, A.S.; Artner, M.; Janjić, K.; Edelmayer, M.; Kurzmann, C.; Moritz, A.; Agis, H. Synthetic clay-based hypoxia mimetic hydrogel for pulp regeneration: The impact on cell activity and release kinetics based on dental pulp-derived cells *In Vitro. J. Endod.*, **2018**, *44*(8), 1263-1269. [http://dx.doi.org/10.1016/j.joen.2018.04.010] [PMID: 29958677]
- [117] Yao, Q.; Liu, Y.; Tao, J.; Baumgarten, K.M.; Sun, H. Hypoxia-mimicking nanofibrous scaffolds promote endogenous bone regeneration. *ACS Appl. Mater. Interfaces*, **2016**, *8*(47), 32450-32459. [http://dx.doi.org/10.1021/acsami.6b10538] [PMID: 27809470]
- [118] Ma, D.; Hossain, M.; Pettet, G.K.; Luo, Y.; Lim, T.; Akimov, S.; Sanders, R.D.; Franks, N.P.; Maze, M. Xenon preconditioning reduces brain damage from neonatal asphyxia in rats. *J. Cereb. Blood Flow Metab.*, **2006**, *26*(2), 199-208. [http://dx.doi.org/10.1038/sj.jcbfm.9600184] [PMID: 16034370]
- [119] Hu, Y.; Wang, Z.; Liu, Y.; Pan, S.; Zhang, H.; Fang, M.; Jiang, H.; Yin, J.; Zou, S.; Li, Z.; Zhang, H.; Lin, Z.; Xiao, J. Melatonin Reduces hypoxic-ischaemic (HI) induced autophagy and apoptosis: An *in Vivo* and *in Vitro* investigation in experimental models of neonatal HI brain injury. *Neurosci. Lett.*, **2017**, *653*, 105-112. [PMID: 28616019]
- [120] Pabon, M.M.; Borlongan, C.V. Advances in the cell-based treatment of neonatal hypoxic-ischemic brain injury. *Future Neurol.*, **2013**, *8*(2), 193-203. [http://dx.doi.org/10.2217/fnl.12.85] [PMID: 23565051]
- [121] Liang, L.; Yang, J.; Jin, X. Cocktail Treatment, a Promising Strategy to Treat Acute Cerebral Ischemic Stroke? *Med. Gas. Res.*, **2016**, *6*(1), 33-38. <http://www.medgasres.com/article.asp?issn=2045-9912;year=2016;volume=6;issue=1;page=33;epage=38;aulast=Liang;=5>
- [122] Drury, P.P.; Gunn, E.R.; Bennet, L.; Gunn, A.J. Mechanisms of hypothermic neuroprotection. *Clin. Perinatol.*, **2014**, *41*(1), 161-175. [http://dx.doi.org/10.1016/j.clp.2013.10.005] [PMID: 24524453]
- [123] Liu, J.; Segal, M.R.; Kelly, M.J.S.; Pelton, J.G.; Kim, M.; James, T.L.; Litt, L. ¹³C NMR metabolomic evaluation of immediate and delayed mild hypothermia in cerebrocortical slices after oxygen-glucose deprivation. *Anesthesiology*, **2013**, *119*(5), 1120-1136. [http://dx.doi.org/10.1097/ALN.0b013e31829c2d90] [PMID: 23748856]
- [124] Nakai, A.; Shibasaki, Y.; Taniuchi, Y.; Oya, A.; Asakura, H.; Kuroda, S.; Koshino, T.; Araki, T. Influence of mild hypothermia on delayed mitochondrial dysfunction after transient intrauterine ischemia in the immature rat brain. *Brain Res. Dev. Brain Res.*, **2001**, *128*(1), 1-7. [http://dx.doi.org/10.1016/S0165-3806(01)00138-9] [PMID: 11356256]
- [125] Jacobs, S.E.; Berg, M.; Hunt, R.; Tarnow-Mordi, W.O.; Inder, T.E.; Davis, P.G. Cooling for newborns with hypoxic ischaemic encephalopathy. *Cochrane Database Syst. Rev.*, **2013**, (1)CD003311 [http://dx.doi.org/10.1002/14651858.CD003311.pub3] [PMID: 23440789]
- [126] Piironen, K.; Tiainen, M.; Mustanoja, S.; Kaukonen, K.-M.; Mere-toja, A.; Tatlisumak, T.; Kaste, M. Mild hypothermia after intravenous thrombolysis in patients with acute stroke: a randomized controlled trial. *Stroke*, **2014**, *45*(2), 486-491. [http://dx.doi.org/10.1161/STROKEAHA.113.003180] [PMID: 24436240]
- [127] Gao, X.Y.; Huang, J.O.; Hu, Y.F.; Gu, Y.; Zhu, S.Z.; Huang, K.B.; Chen, J.Y.; Pan, S.Y. Combination of mild hypothermia with neuroprotectants has greater neuroprotective effects during oxygen-glucose deprivation and reoxygenation-mediated neuronal injury. *Sci. Rep.*, **2014**, *4*, 7091. [http://dx.doi.org/10.1038/srep07091] [PMID: 25404538]
- [128] Dubrovsky, B.O. Steroids, neuroactive steroids and neurosteroids in psychopathology. *Prog. Neuropsychopharmacol. Biol. Psychiatry*, **2005**, *29*(2), 169-192. [http://dx.doi.org/10.1016/j.pnpbp.2004.11.001] [PMID: 15694225]
- [129] Tuem, K.B.; Atey, T.M. Neuroactive steroids: Receptor interactions and responses. *Front. Neurol.*, **2017**, *8*, 442. [http://dx.doi.org/10.3389/fneur.2017.00442] [PMID: 28894435]
- [130] Rey, M.; Coirini, H. Synthetic neurosteroids on brain protection. *Neural Regen. Res.*, **2015**, *10*(1), 17-21. [http://dx.doi.org/10.4103/1673-5374.150640] [PMID: 25788907]

- [131] Lizcano, F.; Guzmán, G. Estrogen deficiency and the origin of obesity during menopause. *Biomed. Res. Int.*, **2014**, *2014*, 757461.
- [132] Zhao, L.; O'Neill, K.; Diaz, B. R. Selective estrogen receptor modulators (SERMs) for the brain: Current status and remaining challenges for developing neuroSERMs. *Brain Res. Brain Res. Rev.*, **2005**, *49*(3), 472-493. [http://dx.doi.org/10.1016/j.brainresrev.2005.01.009] [PMID: 16269315]
- [133] Paterni, I.; Granchi, C.; Katzenellenbogen, J. A.; Minutolo, F. Estrogen receptors alpha (ER α) and beta (ER β): Subtype-selective ligands and clinical potential. *Steroids* **2014**, *90*, 13-29.
- [134] Arevalo, M.A.; Azcoitia, I.; Garcia-Segura, L.M. The neuroprotective actions of oestradiol and oestrogen receptors. *Nat. Rev. Neurosci.*, **2015**, *16*(1), 17-29. [http://dx.doi.org/10.1038/nrn3856] [PMID: 25423896]
- [135] Shang, Y.; Hu, X.; DiRenzo, J.; Lazar, M. A.; Brown, M. Cofactor dynamics and sufficiency in estrogen receptor-regulated transcription. *Cell* **2000**, *103* (6), 843-852.
- [136] Safe, S.; Kim, K. Non classical genomic ER/Sp and ER/AP-1 signaling pathways. *J. Mol. Endocrinol.*, **2008**, *41*(5), 263-275. [http://dx.doi.org/10.1677/JME-08-0103] [PMID: 18772268]
- [137] Ruiz-Palmero, I.; Hernandez, M.; Garcia-Segura, L.M.; Arevalo, M.A.G. G protein-coupled estrogen receptor is required for the neurotogenic mechanism of 17 β -estradiol in developing hippocampal neurons. *Mol. Cell. Endocrinol.*, **2013**, *372*(1-2), 105-115. [http://dx.doi.org/10.1016/j.mce.2013.03.018] [PMID: 23545157]
- [138] Qiu, J.; Bosch, M.A.; Tobias, S.C.; Grandy, D.K.; Scanlan, T.S.; Ronnekleiv, O.K.; Kelly, M.J. Rapid signaling of estrogen in hypothalamic neurons involves a novel G-protein-coupled estrogen receptor that activates protein kinase C. *J. Neurosci.*, **2003**, *23*(29), 9529-9540. [http://dx.doi.org/10.1523/JNEUROSCI.23-29-09529.2003] [PMID: 14573532]
- [139] Hammes, S.R.; Davis, P.J. Overlapping nongenomic and genomic actions of thyroid hormone and steroids. *Best Pract. Res. Clin. Endocrinol. Metab.*, **2015**, *29*(4), 581-593. [http://dx.doi.org/10.1016/j.beem.2015.04.001] [PMID: 26303085]
- [140] Harvey, B. J.; Condliffe, S.; Doolan, C. M. Sex and salt hormones: Rapid effects in epithelia. *News Physiol. Sci.*, **2001**, *16*, 174-177.
- [141] Simoncini, T.; Mannella, P.; Fornari, L.; Caruso, A.; Varone, G.; Genazzani, A.R. Genomic and non-genomic effects of estrogens on endothelial cells. *Steroids*, **2004**, *69*(8-9), 537-542. [http://dx.doi.org/10.1016/j.steroids.2004.05.009] [PMID: 15288766]
- [142] Meldrum, D.R. G-protein-coupled receptor 30 mediates estrogen's nongenomic effects after hemorrhagic shock and trauma. *Am. J. Pathol.*, **2007**, *170*(4), 1148-1151. [http://dx.doi.org/10.2353/ajpath.2007.070025] [PMID: 17392155]
- [143] Lösel, R.; Wehling, M. Nongenomic actions of steroid hormones. *Nat. Rev. Mol. Cell Biol.*, **2003**, *4*(1), 46-56. [http://dx.doi.org/10.1038/nrm1009] [PMID: 12511868]
- [144] Fernandez, S. M.; Lewis, M. C.; Pechenino, A. S.; Harburger, L. L.; Orr, P. T.; Gresack, J. E.; Schafe, G. E.; Frick, K. M. Estradiol-induced enhancement of object memory consolidation involves hippocampal extracellular signal-regulated kinase activation and membrane-bound estrogen receptors. *J. Neurosci.*, **2008**, *28*(35), 8660-8667.
- [145] Fan, L.; Zhao, Z.; Orr, P. T.; Chambers, C. H.; Lewis, M. C.; Frick, K. M. Nongenomic actions of steroid hormones. *Nat. Rev. Mol. Cell Biol.*, **2003**, *4*(1), 46-56.
- [146] Hojo, Y.; Kawato, S. Neurosteroids in adult hippocampus of male and female rodents: Biosynthesis and actions of sex steroids. *Front. Endocrinol. (Lausanne)*, **2018**, *9*(APR), 183. [http://dx.doi.org/10.3389/fendo.2018.00183] [PMID: 29740398]
- [147] Tozzi, A.; de Iure, A.; Tantucci, M.; Durante, V.; Quiroga-Varela, A.; Giampà, C.; Di Mauro, M.; Mazzocchetti, P.; Costa, C.; Di Filippo, M.; Grassi, S.; Pettorossi, V.E.; Calabresi, P. Endogenous 17 β -estradiol is required for activity-dependent long-term potentiation in the striatum: interaction with the dopaminergic system. *Front. Cell. Neurosci.*, **2015**, *9*, 192. [http://dx.doi.org/10.3389/fncel.2015.00192] [PMID: 26074768]
- [148] Pupo, M.; Maggolini, M.; Musti, A.M. GPER mediates nongenomic effects of estrogen BT - estrogen receptors: Methods and protocols; eyster, K. M.; York, S.N., Ed.; New York, NY, **2016**, pp. 471-488. [http://dx.doi.org/10.1007/978-1-4939-3127-9_37]
- [149] Lebesgue, D.; Chevaleyre, V.; Zukin, R.S.; Etgen, A.M. Estradiol rescues neurons from global ischemia-induced cell death: multiple cellular pathways of neuroprotection. *Steroids*, **2009**, *74*(7), 555-561. [http://dx.doi.org/10.1016/j.steroids.2009.01.003] [PMID: 19428444]
- [150] Galbiati, M.; Martini, L.; Melcangi, R.C. Oestrogens, via transforming growth factor α , modulate basic fibroblast growth factor synthesis in hypothalamic astrocytes: in vitro observations. *J. Neuroendocrinol.*, **2002**, *14*(10), 829-835. [http://dx.doi.org/10.1046/j.1365-2826.2002.00852.x] [PMID: 12372008]
- [151] Karki, P.; Smith, K.; Johnson, J., Jr.; Lee, E. Astrocyte-derived growth factors and estrogen neuroprotection: role of transforming growth factor- α in estrogen-induced upregulation of glutamate transporters in astrocytes. *Mol. Cell. Endocrinol.*, **2014**, *389*(1-2), 58-64. [http://dx.doi.org/10.1016/j.mce.2014.01.010] [PMID: 24447465]
- [152] Kirschner, P.B.; Henshaw, R.; Weise, J.; Trubetsky, V.; Finklestein, S.; Schulz, J.B.; Beal, M.F. Basic fibroblast growth factor protects against excitotoxicity and chemical hypoxia in both neonatal and adult rats. *J. Cereb. Blood Flow Metab.*, **1995**, *15*(4), 619-623. [http://dx.doi.org/10.1038/jcbfm.1995.76] [PMID: 7790410]
- [153] Nozaki, K.; Finklestein, S.P.; Beal, M.F. Basic fibroblast growth factor protects against hypoxia-ischemia and NMDA neurotoxicity in neonatal rats. *J. Cereb. Blood Flow Metab.*, **1993**, *13*(2), 221-228. [http://dx.doi.org/10.1038/jcbfm.1993.27] [PMID: 8436614]
- [154] Herrera, M. I.; Mucci, S.; Barreto, G. E.; Kolliker-Frers, R.; Capani, F. Neuroprotection in hypoxic-ischemic brain injury targeting glial cells. *Curr. Pharm. Des.* **2017**, *23*(26), 3899-3906.
- [155] Gerstner, B.; Lee, J.; DeSilva, T.M.; Jensen, F.E.; Volpe, J.J.; Rosenberg, P.A. 17 β -estradiol protects against hypoxic/ischemic white matter damage in the neonatal rat brain. *J. Neurosci. Res.*, **2009**, *87*(9), 2078-2086. [http://dx.doi.org/10.1002/jnr.22023] [PMID: 19224575]
- [156] Nuñez, J.; Yang, Z.; Jiang, Y.; Grandys, T.; Mark, I.; Levison, S.W. 17 β -estradiol protects the neonatal brain from hypoxia-ischemia. *Exp. Neurol.*, **2007**, *208*(2), 269-276. [http://dx.doi.org/10.1016/j.expneurol.2007.08.020] [PMID: 17950281]
- [157] Barreto, G.; Saraceno, E.; Gonzalez, J.; Kolliker, R.; Castilla, R.; Capani, F. Neuroprotection with estradiol in experimental perinatal asphyxia: A new approach A2 - duncan, Kelli, A. B.T - Estrogen effects on traumatic brain injury. In: *Academic Press*; San Diego, **2015**, pp. 113-124. [http://dx.doi.org/10.1016/B978-0-12-801479-0.00008-5]
- [158] Charriaut-Marlangue, C.; Besson, V.C.; Baud, O. Sexually dimorphic outcomes after neonatal stroke and hypoxia-ischemia. *Int. J. Mol. Sci.*, **2017**, *19*(1)E61 [http://dx.doi.org/10.3390/ijms19010061] [PMID: 29278365]
- [159] Elzer, J.G.; Muhammad, S.; Wintermantel, T.M.; Regnier-Vigouroux, A.; Ludwig, J.; Schütz, G.; Schwaninger, M. Neuronal estrogen receptor- α mediates neuroprotection by 17 β -estradiol. *J. Cereb. Blood Flow Metab.*, **2010**, *30*(5), 935-942. [http://dx.doi.org/10.1038/jcbfm.2009.258] [PMID: 20010956]
- [160] Toro-Urrego, N.; Garcia-Segura, L.M.; Echeverria, V.; Barreto, G.E. Testosterone protects mitochondrial function and regulates neuroglobin expression in astrocytic cells exposed to glucose deprivation. *Front. Aging Neurosci.*, **2016**, *8*, 152. [http://dx.doi.org/10.3389/fnagi.2016.00152] [PMID: 27445795]
- [161] Nelson, E.R.; Wardell, S.E.; McDonnell, D.P. The molecular mechanisms underlying the pharmacological actions of estrogens, SERMs and oxysterols: implications for the treatment and prevention of osteoporosis. *Bone*, **2013**, *53*(1), 42-50. [http://dx.doi.org/10.1016/j.bone.2012.11.011] [PMID: 23168292]
- [162] Marin, F.; Barbancho, M.C. Action of selective estrogen receptor modulators (SERMs) through the classical mechanism of estrogen action. In: *Selective Estrogen Receptor Modulators*, **2006**, pp. 71-77.
- [163] Khan, M.M.; Wakade, C.; de Sevilla, L.; Brann, D.W. Selective estrogen receptor modulators (SERMs) enhance neurogenesis and spine density following focal cerebral ischemia. *J. Steroid Biochem. Mol. Biol.*, **2015**, *146*, 38-47. [http://dx.doi.org/10.1016/j.jsbmb.2014.05.001] [PMID: 24815952]

- [164] Lopez-Rodriguez, A. B.; Ávila-Rodriguez, M.; Vega-vela, N. E.; Capani, F.; Gonzalez, J.; García-Segura, L. M.; Barreto, G. E. *Estrogen Effects on Traumatic Brain Injury*, **2015**.
- [165] Gao, Y.; Wang, Z.; He, W.; Ma, W.; Ni, X. Mild hypothermia protects neurons against oxygen glucose deprivation via Poly (ADP-Ribose) signaling. *J. Matern. Neonatal Med.*, **2017**, 1-7.
- [166] Chakkarapani, E.; Dingley, J.; Liu, X.; Hoque, N.; Aquilina, K.; Porter, H.; Thoresen, M. Xenon enhances hypothermic neuroprotection in asphyxiated newborn pigs. *Ann. Neurol.*, **2010**, *68*(3), 330-341. [<http://dx.doi.org/10.1002/ana.22016>] [PMID: 20658563]
- [167] Kaneko, Y.; Tajiri, N.; Su, T.-P.; Wang, Y.; Borlongan, C.V. Combination treatment of hypothermia and mesenchymal stromal cells amplifies neuroprotection in primary rat neurons exposed to hypoxic-ischemic-like injury *in vitro*: role of the opioid system. *PLoS One*, **2012**, *7*(10), e47583. [<http://dx.doi.org/10.1371/journal.pone.0047583>] [PMID: 23077646]
- [168] Abdelhamid, R.; Luo, J.; Vandevrede, L.; Kundu, I.; Michalsen, B.; Litosh, V. A.; Schiefer, I. T.; Gherezghiher, T.; Yao, P.; Qin, Z.; Thatcher, G. R. Benzothioephene selective estrogen receptor modulators provide neuroprotection by a Novel GPR30-dependent mechanism. *ACS Chem. Neurosci.* **2011**, *2*(5), 256-268.
- [169] Rzemieniec, J.; Litwa, E.; Wnuk, A.; Lason, W.; Golas, A.; Krzeptowski, W.; Kajta, M. Neuroprotective action of raloxifene against hypoxia-induced damage in mouse hippocampal cells depends on ER α but not ER β or GPR30 signalling. *J. Steroid Biochem. Mol. Biol.*, **2015**, *146*, 26-37. [<http://dx.doi.org/10.1016/j.jsbmb.2014.05.005>] [PMID: 24846829]
- [170] Rzemieniec, J.; Litwa, E.; Wnuk, A.; Lason, W.; Kajta, M. Bazedoxifene and raloxifene protect neocortical neurons undergoing hypoxia via targeting ER α and PPAR- γ . *Mol. Cell. Endocrinol.*, **2018**, *461*, 64-78. [<http://dx.doi.org/10.1016/j.mce.2017.08.014>] [PMID: 28859903]
- [171] Barreto, G.E.; Santos-Galindo, M.; Garcia-Segura, L.M. Selective estrogen receptor modulators regulate reactive microglia after penetrating brain injury. *Front. Aging Neurosci.*, **2014**, *6*, 132. [<http://dx.doi.org/10.3389/fnagi.2014.00132>] [PMID: 24999330]
- [172] Pinto-Almazán, R.; Calzada-Mendoza, C.C.; Campos-Lara, M.G.; Guerra-Araiza, C. Effect of chronic administration of estradiol, progesterone, and tibolone on the expression and phosphorylation of glycogen synthase kinase-3 β and the microtubule-associated protein tau in the hippocampus and cerebellum of female rat. *J. Neurosci. Res.*, **2012**, *90*(4), 878-886. [<http://dx.doi.org/10.1002/jnr.22808>] [PMID: 22183707]
- [173] de Aguiar, R.B.; Dickel, O.E.; Cunha, R.W.; Monserrat, J.M.; Barros, D.M.; Martinez, P.E. Estradiol valerate and tibolone: effects upon brain oxidative stress and blood biochemistry during aging in female rats. *Biogerontology*, **2008**, *9*(5), 285-298. [<http://dx.doi.org/10.1007/s10522-008-9137-7>] [PMID: 18386154]
- [174] Stark, J.; Varbiro, S.; Sipos, M.; Tulassay, Z.; Sara, L.; Adler, I.; Dinya, E.; Magyar, Z.; Szekacs, B.; Marczell, I.; Kloosterboer, H.J.; Racz, K.; Bekesi, G. Antioxidant effect of the active metabolites of tibolone. *Gynecol. Endocrinol.*, **2015**, *31*(1), 31-35.
- [175] Crespo-Castrillo, A.; Yanguas-Casás, N.; Arevalo, M. A.; Azcoitia, I.; Barreto, G. E.; Garcia-Segura, L. M. The synthetic steroid tibolone decreases reactive gliosis and neuronal death in the cerebral cortex of female mice after a stab wound injury. *Mol. Neurobiol.* **2018**, *55*(11), 8651-8667.
- [176] Avila-Rodriguez, M.; Garcia-Segura, L.M.; Hidalgo-Lanussa, O.; Baez, E.; Gonzalez, J.; Barreto, G.E. Tibolone protects astrocytic cells from glucose deprivation through a mechanism involving estrogen receptor beta and the upregulation of neuroglobin expression. *Mol. Cell. Endocrinol.*, **2016**, *433*, 35-46. [<http://dx.doi.org/10.1016/j.mce.2016.05.024>] [PMID: 27250720]
- [177] Oh, J.S.; Kim, S.W.; Cho, H.J.; Kyong, Y.Y.; Oh, Y.M.; Choi, S.M.; Choi, K.H.; Park, K.N. Combination treatment with 17 β -estradiol and therapeutic hypothermia for transient global cerebral ischemia in rats. *Am. J. Emerg. Med.*, **2013**, *31*(1), 154-160. [<http://dx.doi.org/10.1016/j.ajem.2012.06.033>] [PMID: 22980365]
- [178] Song, J.; Sun, H.; Xu, F.; Kang, W.; Gao, L.; Guo, J.; Zhang, Y.; Xia, L.; Wang, X.; Zhu, C. Recombinant human erythropoietin improves neurological outcomes in very preterm infants. *Ann. Neurol.*, **2016**, *80*(1), 24-34. [<http://dx.doi.org/10.1002/ana.24677>] [PMID: 27130143]
- [179] Robertson, N. J.; Faulkner, S.; Fleiss, B.; Bainbridge, A.; Aandorka, C.; Price, D.; Powell, E.; Lecky-Thompson, L.; Thei, L.; Chandrasekaran, M.; Hristova, M.; Cady, E.B.; Gressens, P.; Gollay, X.; Raivich, G. Melatonin augments hypothermic neuroprotection in a perinatal *Asphyxia Model*. *Brain*, **2013**, *136*(1), 90-105.
- [180] Kim, H.; Koo, Y. S.; Shin, M. J.; Kim, S.; Shin, Y. B.; Choi, B. T.; Yun, Y. J.; Lee, S.; Shin, H. K. Combination of constraint-induced movement therapy with electroacupuncture improves functional recovery following neonatal hypoxic-ischemic brain injury in rats. *Biomed. Res. Int.*, **2018**, 8638294. [doi: 10.1155/2018/8638294].
- [181] Jin, X.; Liu, J.; Liu, K.J.; Rosenberg, G.A.; Yang, Y.; Liu, W. Normobaric hyperoxia combined with minocycline provides greater neuroprotection than either alone in transient focal cerebral ischemia. *Exp. Neurol.*, **2013**, *240*, 9-16. [<http://dx.doi.org/10.1016/j.expneurol.2012.11.018>] [PMID: 23195595]
- [182] Nonaka, Y.; Shimazawa, M.; Yoshimura, S.; Iwama, T.; Hara, H. Combination effects of normobaric hyperoxia and edaravone on focal cerebral ischemia-induced neuronal damage in mice. *Neurosci. Lett.*, **2008**, *441*(2), 224-228. [<http://dx.doi.org/10.1016/j.neulet.2008.06.033>] [PMID: 18577423]
- [183] Nonaka, Y.; Koumura, A.; Hyakkoku, K.; Shimazawa, M.; Yoshimura, S.; Iwama, T.; Hara, H. Combination treatment with normobaric hyperoxia and cilostazol protects mice against focal cerebral ischemia-induced neuronal damage better than each treatment alone. *J. Pharmacol. Exp. Ther.*, **2009**, *330*(1), 13-22.

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