



# Complete Genome Sequence of Hospital-Acquired Methicillin-Resistant *Staphylococcus aureus* Strain WCUH29

Ting Lei,<sup>a</sup> Ying Zhang,<sup>b</sup> Junshu Yang,<sup>a</sup> Kevin Silverstein,<sup>b</sup> Yinduo Ji<sup>a</sup>

<sup>a</sup>Department of Veterinary Biomedical Science, College of Veterinary Medicine, University of Minnesota, St. Paul, Minnesota, USA

<sup>b</sup>Minnesota Supercomputing Institute, University of Minnesota, Minneapolis, Minnesota, USA

**ABSTRACT** The hospital-acquired methicillin-resistant *Staphylococcus aureus* (HA-MRSA) strain WCUH29 has been intensively and widely used as a model system for identification and evaluation of novel antibacterial targets and pathogenicity. In this announcement, we report the complete genome sequence of HA-MRSA WCUH29 (NCIMB 40771).

*Staphylococcus aureus* is an important pathogen that can cause a variety of diseases worldwide. The continuing emergence of methicillin-resistant *S. aureus* (MRSA) associated with both hospital-acquired and community-acquired infections has resulted in serious health care problems due to the limited effective treatment options (1, 2).

The *S. aureus* WCUH29 strain is a human clinical isolate from the Children's University Hospital in Warsaw, Poland (3). WCUH29 is a MRSA strain that is resistant to multiple antibiotics, including methicillin, amoxicillin, and kanamycin (4, 5). This organism has been successfully used to investigate the pathogenesis of *S. aureus* in animal models of infection, including a murine hematogenous pyelonephritis infection model (3, 6, 7), and an intraperitoneal infection that can cause mortality to infected mice (7, 8). This strain has been intensively utilized for validation of antibacterial agents (7, 9, 10) and potential target genes for the development of novel antibacterial agents (11). We used this strain as a model system for comprehensive identification of genes essential for bacterial survival (12). In this study, we determined the complete genome sequence of WCUH29, which enables us to further perform comparative genomics studies and determine the impact of genetic background on a given gene's function.

A single colony of WCUH29 was picked up from a sheep blood agar plate, inoculated into tryptic soy broth, and incubated at 37°C with shaking (225 rpm). Genomic DNA was purified from the bacterial cells of stationary-phase culture using a genomic DNA purification kit (Promega, Wisconsin WI). PicoGreen DNA quantitation, library construction with a TruSeq Nano DNA sample preparation kit (Illumina), and DNA sequencing were performed at the University of Minnesota Genomic Center (UMGC). One hundred paired-end cycles of DNA sequencing were conducted using a HiSeq 2500 platform (Illumina). The sequence data were transferred to the Minnesota Supercomputing Institute (MSI) for storage.

The genome was sequenced using a PacBio RS II system on two single-molecule real-time (SMRT) cells, which generated 184,878 filtered reads with a mean length of 6,255 bp. *De novo* assembly was performed using the Hierarchical Genome Assembly Process (HGAP) version 3 (13) with the settings filter out subreads shorter than 5,000 and set the minimum seed read to 16,000, the target coverage to 30, and the genome size to 2.9 Mb, which produced one assembled sequence (2,909,904 bp). Independent Illumina sequencing of the same strain produced 9,985,766 100-bp-long paired DNA reads. Pilon version 1.10 (14) was used to polish the PacBio assembly using this set of

**Citation** Lei T, Zhang Y, Yang J, Silverstein K, Ji Y. 2019. Complete genome sequence of hospital-acquired methicillin-resistant *Staphylococcus aureus* strain WCUH29. *Microbiol Resour Announc* 8:e00551-19. <https://doi.org/10.1128/MRA.00551-19>.

**Editor** Steven R. Gill, University of Rochester School of Medicine and Dentistry

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Address correspondence to Yinduo Ji, [jixxx002@umn.edu](mailto:jixxx002@umn.edu).

**Received** 13 May 2019

**Accepted** 13 May 2019

**Published** 6 June 2019

short DNA reads. It corrected 1 single base and inserted 29 1-bp indels genome-wide. By comparing the sequencing to 48 known *S. aureus* genomes, such as those of N315, MW2, Mu3, Mu50, COL, and MSSA476, we determined the origin of replication and reorganized the genome sequence to start from the origin of replication. Since we did not identify overlapping sequences, we used an “N” to fill the junction between the original start and end positions. The final assembly harbors a circular chromosome of 2,909,904 bp with a G+C content of 32.9%.

The Rapid Annotation Transfer Tool (RATT; time-stamp, 14 December 2011) (15) was used to annotate the assembled WCUH29 genome via comparisons with those of 4 known *S. aureus* strains (GenBank accession numbers AP009324, BA000017, BA000018, and CP001844). Rapid Annotations using Subsystems Technology (RAST) (myRAST version 36) (16) was also used to annotate the genome in order to predict WCUH29-specific genes. The final annotation of WCUH29 includes 3,004 coding genes, 35 rRNA genes, 124 tRNA genes, 2 transfer-messenger RNA (tmRNA) genes, and 2 pathogenic islands.

**Data availability.** The genome sequence of *Staphylococcus aureus* WCUH29 has been deposited in NCBI GenBank under the accession number CP039156 and Bio-Project number PRJNA531521.

## ACKNOWLEDGMENT

This study was partially supported by award MIN-63-075 from the General Agricultural Research fund for the EZID Signature Program project in the College of Veterinary Medicine at the University of Minnesota.

## REFERENCES

- Hassoun A, Linden PK, Friedman B. 2017. Incidence, prevalence, and management of MRSA bacteremia across patient populations—a review of recent developments in MRSA management and treatment. *Crit Care* 21:211. <https://doi.org/10.1186/s13054-017-1801-3>.
- Rodvold KA, McConeghy KW. 2014. Methicillin-resistant *Staphylococcus aureus* therapy: past, present, and future. *Clin Infect Dis* 58(Suppl 1): S20–7. <https://doi.org/10.1093/cid/cit614>.
- Chalker FA, Ingraham AK, Lunsford RD, Bryant PA, Bryant J, Wallis GN, Broskey PJ, Pearson CS, Holmes JD. 2000. The *bacA* gene, which determines bacitracin susceptibility in *Streptococcus pneumoniae* and *Staphylococcus aureus*, is also required for virulence. *Microbiology* 146: 1547–1553. <https://doi.org/10.1099/00221287-146-7-1547>.
- Payne DJ, Miller WH, Berry V, Brosky J, Burgess WJ, Chen E, DeWolf WE, Jr, Fosberry AP, Greenwood R, Head MS, Heerding DA, Janson CA, Jaworski DD, Keller PM, Manley PJ, Moore TD, Newlander KA, Pearson S, Polizzi BJ, Qiu X, Rittenhouse SF, Slater-Radosti C, Salyers KL, Seefeld MA, Smyth MG, Takata DT, Uzinskas IN, Vaidya K, Wallis NG, Winram SB, Yuan CC, Huffman WF. 2002. Discovery of a novel and potent class of FabI-directed antibacterial agents. *Antimicrob Agents Chemother* 46:3118–3124. <https://doi.org/10.1128/aac.46.10.3118-3124.2002>.
- Bax BD, Chan PF, Eggleston DS, Fosberry A, Gentry DR, Gorrec F, Giordano I, Hann MM, Hennessy A, Hibbs M, Huang J, Jones E, Jones J, Brown KK, Lewis CJ, May EW, Saunders MR, Singh O, Spitzfaden CE, Shen C, Shillings A, Theobald AJ, Wohlkonig A, Pearson ND, Gwynn MN. 2010. Type IIA topoisomerase inhibition by a new class of antibacterial agents. *Nature* 466:935–940. <https://doi.org/10.1038/nature09197>.
- Liang X, Yu C, Sun J, Liu H, Landwehr C, Holmes D, Ji Y. 2006. Inactivation of a two-component signal transduction system, SaeRS, eliminates adherence and attenuates virulence of *Staphylococcus aureus*. *Infect Immun* 74:4655–4665. <https://doi.org/10.1128/IAI.00322-06>.
- Lewandowski T, Huang J, Fan F, Rogers S, Gentry D, Holland R, DeMarsh P, Aubart K, Zalacain M. 2013. *Staphylococcus aureus* formyl-methionyl transferase mutants demonstrate reduced virulence factor production and pathogenicity. *Antimicrob Agents Chemother* 57:2929–2936. <https://doi.org/10.1128/AAC.00162-13>.
- Ji Y, Marra A, Rosenberg M, Woodnutt G. 1999. Regulated antisense RNA eliminates alpha-toxin virulence in *Staphylococcus aureus* infection. *J Bacteriol* 181:6585–6590.
- Herbert S, Barry P, Novick RP. 2001. Subinhibitory clindamycin differentially inhibits transcription of exoprotein genes in *Staphylococcus aureus*. *Infect Immun* 69:2996–3003. <https://doi.org/10.1128/IAI.69.5.2996-3003.2001>.
- Miles TJ, Axten JM, Barfoot C, Brooks G, Brown P, Chen D, Dabbs S, Davies DT, Downie DL, Eyrisch S, Gallagher T, Giordano I, Gwynn MN, Hennessy A, Hoover J, Huang J, Jones G, Markwell R, Miller WH, Minthorn EA, Rittenhouse S, Seefeld M, Pearson N. 2011. Novel amino-piperidines as potent antibacterials targeting bacterial type IIA topoisomerases. *Bioorg Med Chem Lett* 21:7489–7495. <https://doi.org/10.1016/j.bmcl.2011.09.117>.
- Lei T, Yang J, Ji Y. 2015. Determination of essentiality and regulatory function of staphylococcal YeaZ in branched-chain amino acid biosynthesis. *Virulence* 6:75–84. <https://doi.org/10.4161/21505594.2014.986415>.
- Ji Y, Zhang B, Van SF, Warren P, Woodnutt G, Burnham M, Rosenberg M. 2001. Identification of critical staphylococcal genes using conditional phenotypes generated by antisense RNA. *Science* 293:2266–2269. <https://doi.org/10.1126/science.1063566>.
- Chin CS, Alexander DH, Marks P, Klammer AA, Drake J, Heiner C, Clum A, Copeland A, Huddleston J, Eichler EE, Turner SW, Korlach J. 2013. Non-hybrid, finished microbial genome assemblies from long-read SMRT sequencing data. *Nat Methods* 10:563–569. <https://doi.org/10.1038/nmeth.2474>.
- Walker BJ, Abeel T, Shea T, Priest M, Abouelliel A, Sakthikumar S, Cuomo CA, Zeng Q, Wortman J, Young SK, Earl AM. 2014. Pilon: an integrated tool for comprehensive microbial variant detection and genome assembly improvement. *PLoS One* 9:e112963. <https://doi.org/10.1371/journal.pone.0112963>.
- Otto TD, Dillon GP, Degraeve WS, Berriman M. 2011. RATT: Rapid Annotation Transfer Tool. *Nucleic Acids Res* 39:e57. <https://doi.org/10.1093/nar/gkq1268>.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. *BMC Genomics* 9:75. <https://doi.org/10.1186/1471-2164-9-75>.