



Genome Sequence of *Flavobacterium akiainvivens* IK-1^T, Isolated from Decaying *Wikstroemia oahuensis*, an Endemic Hawaiian Shrub

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Flavobacterium spp. have been cultivated from diverse aquatic and terrestrial habitats. *F. akiainvivens* IK-1^T was cultivated from decaying wood of *Wikstroemia oahuensis*, an endemic Hawaiian shrub. The strain's genome sequence may provide insights into niche adaptation and evolution of the genus in a mid-ocean archipelago.

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A mong the more than 130 *Flavobacterium* spp., some are implicated in diseases in animals and humans (1-3), while some protect plants against pathogens (4). We report the draft genome sequence of *F. akiainvivens* IK-1^T, isolated from decaying wood of *Wikstroemia oahuensis*, an endemic Hawaiian flowering shrub (5).

Genomic DNA was isolated with the Wizard Genomic DNA purification kit (Promega, USA), with an additional CTAB extraction step to remove abundant exopolysaccharides. Shotgun and 8-kb paired-end libraries sequenced in a Roche 454 GS FLX+ platform generated 157.1 Mb of shotgun sequences and 64.4 Mb of 8-kb paired-end sequences, providing ~49× genome coverage; 99.4% of all bases were assigned Phred quality scores above Q20, and all scored above Q10 in the ea-utils program (6). Five scaffolds totaling 4,532,192 bp (scaffold $N_{50} = 4,520,429$ bp) were built in Newbler version 2.8. In silico gap closure proceeded through shredding 454 reads into 100-bp sequences with 50-bp overlap in EMBOSS (7). BGI GapCloser version 1.12 (http://soap.genomics .org.cn) closed gaps covering ~31 kb; full 454 reads were mapped to scaffolds using bowtie2 (8) to confirm the closed gaps were supported by 454 reads, but ~2.3 kb of gaps remain. The genome's G+C content is 43.8%.

The genome was annotated in the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) (9), and the Rapid Annotation using Subsystem Technology (RAST) server (10, 11). PGAP identified 3,841 protein-coding genes, 175 pseudogenes, 48 tRNA coding regions, and 9 rRNA coding regions. RAST identified 4,067 protein-coding open reading frames, and 338 subsystems. No flagella or chemotaxis systems were identified by RAST. The former is consistent with the strain's formal description (5). No phage or prophage regions were identified, although 39 proteins were predicted as similar to conjugative transposons in Bacteroidales by RAST. One CRISPR (clustered regularly interspaced short palindromic repeats) region containing 29 spacers was identified with CRISPRFinder (12, 13). Further manual annotation of the *F. akiainvivens* IK-1^T genome and a comparative analysis with those of other *Flavobacterium* spp. and phylum *Bacteroidetes* spp. will shed light on potential habitat-specific traits and genome evolution in the phylum (14).

Nucleotide sequence accession numbers. This whole-genome shotgun project has been deposited in DDBJ/EMBL/GenBank under the accession number LIYD00000000. The version described in this paper is the first version, LIYD01000000.

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REFERENCES

- Mudarris M, Austin B, Segers P, Vancanneyt M, Hoste B, Bernardet JF. 1994. Flavobacterium scophthalmum sp. nov., a pathogen of turbot (Scophthalmus maximus L.). Int J Syst Bacteriol 44:447–453. http://dx.doi.org/ 10.1099/00207713-44-3-447.
- 2. Bernardet J-F, Bowman JP. 2006. The genus *Flavobacterium*, p 481–531. In The prokaryotes, a handbook on the biology of bacteria, 3rd ed., vol. 7. Springer, New York, NY.
- Manfredi R, Nanetti A, Ferri M, Mastroianni A, Coronado OV, Chiodo F. 1999. *Flavobacterium* spp. organisms as opportunistic bacterial pathogens during advanced HIV disease. J Infect 39:146–152. http://dx.doi.org/ 10.1016/S0163-4453(99)90007-5.
- Sang MK, Kim KD. 2012. The volatile-producing *Flavobacterium john-soniae* strain GSE09 shows biocontrol activity against *Phytophthora capsici* in pepper. J Appl Microbiol 113:383–398. http://dx.doi.org/10.1111/j.1365-2672.2012.05330.x.
- Kuo I, Saw J, Kapan DD, Christensen S, Kaneshiro KY, Donachie SP. 2013. Flavobacterium akiainvivens sp. nov., from decaying wood of Wikstroemia oahuensis, Hawai'i, and emended description of the genus Flavobacterium. Int J Syst Evol Microbiol 63:3280–3286. http://dx.doi.org/ 10.1099/ijs.0.047217-0.
- 6. Aronesty E. 2011. Ea-utils: "command-line tools for processing biological sequencing data". http://code.google.com/p/ea-utils.
- Rice P, Longden I, Bleasby A. 2000. EMBOSS: The European Molecular Biology Open Software Suite. Trends Genet 16:276–277. http:// dx.doi.org/10.1016/S0168-9525(00)02024-2.
- Langmead B, Salzberg SL. 2012. Fast gapped-read alignment with bowtie 2. Nat Methods 9:357–359. http://dx.doi.org/10.1038/nmeth.1923.
- 9. Angiuoli SV, Gussman A, Klimke W, Cochrane G, Field D, Garrity GM, Kodira CD, Kyrpides N, Madupu R, Markowitz V, Tatusova T, Thom-

son N, White O. 2008. Toward an online repository of standard operating procedures (SOPs) for (meta) genomic annotation. OMICS **12:**137–141. http://dx.doi.org/10.1089/omi.2008.0017.

- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. BMC Genomics 9:75. http://dx.doi.org/10.1186/ 1471-2164-9-75.
- 11. Overbeek R, Olson R, Pusch GD, Olsen GJ, Davis JJ, Disz T, Edwards RA, Gerdes S, Parrello B, Shukla M, Vonstein V, Wattam AR, Xia F, Stevens R. 2014. The SEED and the rapid annotation of microbial ge-

nomes using subsystems technology (RAST). Nucleic Acids Res 42: D206–D214. http://dx.doi.org/10.1093/nar/gkt1226.

- 12. Grissa I, Vergnaud G, Pourcel C. 2007. The CRISPRdb database and tools to display CRISPRs and to generate dictionaries of spacers and repeats. BMC Bioinformatics 8:172. http://dx.doi.org/10.1186/1471-2105-8-172.
- 13. Grissa I, Vergnaud G, Pourcel C. 2007. CRISPRFinder: a web tool to identify clustered regularly interspaced short palindromic repeats. Nucleic Acids Res 35:W52–W57. http://dx.doi.org/10.1093/nar/gkm360.
- 14. Gupta RS, Lorenzini E. 2007. Phylogeny and molecular signatures (conserved proteins and indels) that are specific for the *Bacteroidetes* and *Chlorobi* species. BMC Evol Biol 7:71. http://dx.doi.org/10.1186/1471-2148-7-71.