

TRANSFERRIN ENSURES SURVIVAL OF OVARIAN CARCINOMA CELLS SUBJECTED TO DEFEROXAMINE, TNFA, FASL, TRAIL OR MYC-ACTIVATION

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INTRODUCTION. Human ovarian adenocarcinoma N.1 cells apoptose upon *Myc*-activation when serum factors are limited (1). The downstream mechanism that is triggered by *Myc* is unknown, however. We discovered that *Myc*-activation and treatment with TNF α , FasL and TRAIL induces H-ferritin expression. H-ferritin is an intracellular iron chelator. Since also the pharmacological iron chelator deferoxamine induces apoptosis in N.1 cells, we investigated the regulation and the role of H-ferritin in response to *Myc*-activation and TNF α treatment under serum free, pro-apoptotic conditions, and identified transferrin as a survival factor in N.1 human ovarian carcinoma cells.

METHODS. Western blotting, northern blotting and DNA fragmentation and apoptosis assays were performed according to standard procedures and transfection and retroviral transduction according to standard protocols.

RESULTS. When N.1 cells are treated with TRAIL, FasL or TNF α under serum deprived cell culture conditions, which is a prerequisite to induce apoptosis, H-ferritin becomes upregulated (Fig. 1a). Under non-apoptotic conditions, i.e. when cells are maintained under 10% fetal calf serum, the treatment with these inducers prevents cell death (2) and also H-ferritin levels are not induced (Fig. 1b). Activation of an ectopic *Myc:ER* construct also induces H-ferritin expression when serum is withdrawn, not however in presence of serum. Therefore we assumed that intracellular iron sequestration might be a trigger mechanism of *Myc*-induced apoptosis. Hence, transferrin was substituted for serum and this, in fact, rescues from apoptosis and does not allow for H-ferritin induction (Fig. 1c). Substitution of transferrin for serum also aborts cell death elicited by TNF α , FasL and TRAIL (Fig. 2), and moreover, prevents downregulation of Akt during TNF α -treatment.

Fig 1

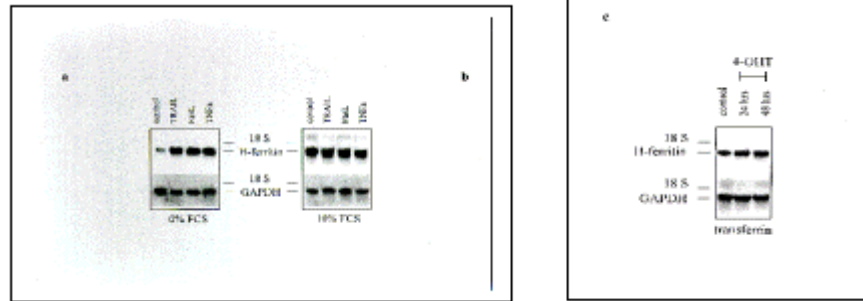
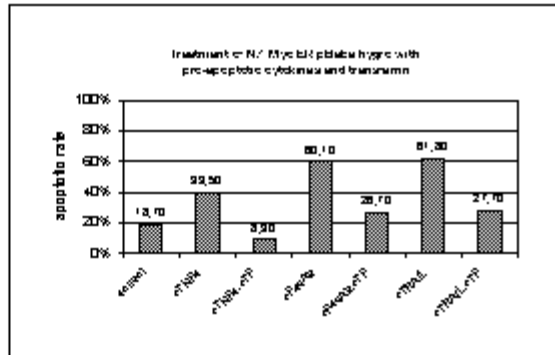


Fig 2

Cells were grown to 20% confluence, then serum was withdrawn. 24 hours after serum deprivation transferrin (TF; 10µg/ml) was added and after 5 hours TNFα (50ng/ml), inducing Fas antibody (FasAb; 80ng/ml) or TRAIL (75ng/ml; TRAIL-enhancer: 1µg/ml) were added for 72 hours. Cells floating in the supernatant were pooled with cells, which were trypsinized from the culture dish. Apoptosis rates were determined by HO/PI staining.



CONCLUSION. These data suggest that *Myc*-activation, FasL, TNFα and TRAIL disturb cellular iron homeostasis and that this is an apoptotic trigger. TNFα suppresses constitutive levels of Akt and phosphorylated Akt, which are restored in presence of transferrin. Downregulation of Akt might generate an apoptosis-prone state in N1 cells by inhibiting a major survival pathway.

It was shown recently that H-ferritin is a target of c-Myc which downregulates its expression under high serum conditions and thereby induces proliferation (3). It can be speculated whether the distinct regulation of H-ferritin by *Myc* is a switch for the dual pathway that either promotes *Myc*-triggered growth or apoptosis, depending on survival factor availability.

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