Research Article

Lei Zhao*, Congzhe Tian, Erbin Xiao, Jinduo Du, Jingwei Liang, Xianghong Chen, Weiwei Chi Clinical significance and potential mechanisms of *miR-223-3p* and *miR-204-5p* in squamous cell carcinoma of head and neck: a study based on TCGA and GEO

https://doi.org/10.1515/med-2020-0146 received April 13, 2020; accepted June 21, 2020

Abstract

Objective - To explore the clinical significance and mechanisms of altered miRNAs in squamous cell carcinoma of head and neck (SCCHN) and provide references for SCCHN diagnosis and prognosis.

Method - Differential expressed miRNAs (DEMs) in SCCHN were screened through gene expression omnibus (GEO) DataSets and verified by the cancer genome atlas (TCGA) database. Next, the overall survival analysis, receiver operating characteristics, and clinical correlation analysis were adopted to filter the miRNAs with diagnostic and prognostic values. Finally, functional enrichment analyses were conducted for inquiring into the mechanisms of miRNAs.

Results – Total 103 DEMs (p < 0.05, fold change ≥ 2) in SCCHN were screened out from GSE124566. Partly, the expression levels of the selected (12/17) miRNAs were verified by TCGA. Followed, of the 12 miRNAs, two miRNA expression levels were associated with the overall survival, and five miRNAs showed diagnostic values (AUC \geq 0.85). Besides, *miR-223-3p* and *miR-204-*5p expression levels were correlated to certain clinical features. Epithelial-mesenchymal transition (EMT) related biological process and energy metabolism controlling related AMPK signaling pathway might mediate the roles of miR-223-3p and miR-204-5p, respectively.

Conclusion - With diagnostic and prognostic values, miR-223-3p and miR-204-5p may be involved in the progression of SCCHN through EMT-related biological process and energy balance related AMPK signaling pathway, respectively.

Key words: head and neck tumor, miRNA, miR-223-3p, miR-204-5p, SCCHN

1 Introduction

As a heterogeneous malignant tumor, squamous cell carcinoma of head and neck (SCCHN) originates from lip, oral cavity, larynx, and pharynx, and is responsible for the high morbidity and mortality of malignant tumors worldwide [1]. According to GLOBOCAN 2018, approximately 8,34,860 new SCCHN cases registered and 4,31,131 cases died of it worldwide in 2018 [2]. Apart from the complicated anatomical features, an insufficient understanding of the pathogenesis and the absence of effective target therapy are also the pivotal reasons for the poor diagnosis and prognosis of SCCHN [1,3,4]. Hence, exploring the critical regulatory mechanisms and identifying the key effect molecules in the pathogenesis and progression of SCCHN are extensively concerned by researchers, and numerous achievements have been made, covering the DNA damage repair [5], effects of exosomes on tumor angiogenesis [6], roles of long non-coding in epithelial-mesenchymal transition (EMT) [7], positive or negative regulation of microRNAs (miRNAs) [8-10], and other aspects.

Here, the present study focused on exploring the regulatory functions of miRNAs in SCCHN for its pivotal biological activity. Previous studies have documented that miRNAs exerted significant biological effects on the occurrence and development of SCCHN. Certain miRNAs suppress SCCHN cellular growth, migration, and invasion ability, and reverse the progression of EMT through diverse mechanisms. For instance, the overexpression of the downregulated miR-4282 retards the progression of oral squamous cell carcinoma by regulating the LIN28B/ZBTB2 pathway in gain-of-function assays [11]. The downregulated miR-204-5p inhibits the SCCHN progression by forming a regulatory complex with SNAI2/SUZ12/HDAC1/STAT3 [12].

^{*} Corresponding author: Lei Zhao, Department of

Otorhinolaryngology, The Affiliated Hospital of Hebei University, Baoding, No. 212 Yuhua Road, Hebei Province, 071000, China, e-mail: erbiyanhouxue@126.com

Congzhe Tian, Erbin Xiao, Jinduo Du, Jingwei Liang, Xianghong Chen: Department of Otorhinolaryngology, The Affiliated Hospital of Hebei University, Baoding, No. 212 Yuhua Road, Hebei Province, 071000, China

Weiwei Chi: Department of Otorhinolaryngology, The First Hospital of Hebei Medical University, Shijiazhuang, Hebei Province, 050031, China

Moreover, the abnormally expressed miRNAs influence the cellular resistance against the radiotherapy or chemotherapy through PI3K-Akt signaling pathway, miR-936/ GPR78 or miR-372-ZBTB7A-TRAIL-R2 pathway in SCCHN [13-15]. Even altered miRNAs could attenuate the angiogenesis and metastasis [16], genomic stability [17], and immune evasion [9]. The same miRNA could regulate multiple mRNAs, such as the inhibitory effect of miR-129 on CDK4, CDK6, and MDM2 [18], and multiple miRNAs could also regulate the same mRNA simultaneously, such as the regulation of miR-21 and miR-499 on PDCD4 [19], which is another embodiment of complexity and diversity of miRNAs' regulatory mechanisms. Apart from the complicated and varied effect mechanisms, pan-cancer character is also responsible for its rising concern. For the same miRNA, it may be involved in various biological behaviors of multiple cancers, which accelerated the widespread development of the related studies, such as the regulatory effects of miR-129-5p on liver cancer, rectal adenocarcinoma, nasopharyngeal carcinoma, and prostate cancer [20-23].

Based on the above-mentioned points, the present study concentrated on exploring the altered miRNAs and corresponding roles in SCCHN through mining public databases and re-analyses, hoping to provide valuable reference for further research studies on SCCHN. After a series of bioinformatics analyses, the present study screened out 103 ectopic expressed miRNAs in SCCHN and partly verified by the cancer genome atlas (TCGA) database. Further detailed analyses indicated that *miR-223-3p* and *miR-204-5p*, with potential diagnostic and prognostic values, might involve in the occurrence and development of SCCHN through the mediation of EMT-related biological process and energy metabolism controlling related *AMPK* signaling pathway.

2 Materials and methods

2.1 Ethics, consent, and permission

The present study was authorized by the Ethics Committee of the Affiliated Hospital of Hebei University (No. HDFY-LL-2020-008). Written consent and permission from the patients were not needed.

2.2 Expression profiles' mining and screening

Gene expression omnibus (GEO) DataSets (http://www. ncbi.nlm.nih.gov/geo/), a public and open accessible database for the expression profile storage [24], was applied to mine SCCHN-related miRNA expression profiles based on retrieval terms: (miRNA OR microRNA) AND (HNSCC OR SCCHN OR "head and neck squamous cell carcinoma" OR "squamous cell carcinoma of head and neck"). Screening criteria include: (1) study type: non-coding RNA profiling by array; (2) tissue: paired cancerous and adjacent non-cancerous samples; (3) retrieval duration: from database creation to retrieval of data on February 25, 2020.

2.3 Differential expressed miRNA screening

GEO2R (https://www.ncbi.nlm.nih.gov/geo/info/geo2r. html), an implanted online tool for comparing differential expressed genes in two or more groups of samples [25], was applied to screen the differential expressed miRNAs (DEMs) in the included GEO Series with the default parameters: Benjamini & Hochberg false discovery rate and p < 0.05. And then, the DEMs were further filtered manually according to adjustment of p < 0.05 and fold change ≥ 2 .

2.4 Acquisition of the miRNA expression profile for verification

The TCGA database (https://www.cancer.gov/tcga), a database for free and open access to genome, transcriptome, and other data [26], was adopted for the acquisition of a miRNA expression profile. And Genomic data commons data portal (https://portal.gdc.cancer.gov/), a data-driven platform for searching and downloading cancerous data [27], was used to implement the acquisition process according to the user's guide (https://docs.gdc.cancer.gov/).

2.5 miRNA targets prediction and filtering

TargetScanHuman 7.2 (http://www.targetscan.org/vert_ 72/), a free and open access online database for miRNA targets' prediction [28], miRDB (http://mirdb.org/), an online database to predict miRNA targets based on thousands of high-throughput sequencing experiments [29], and miRWalk (http://mirwalk.umm.uni-heidelberg. de/), an online database for predicting miRNA targets with a machine learning algorithm including the experimental verification of miRNA-target interactions [30], were applied for predicting potential miRNA targets. Further, Venn diagrams (http://bioinformatics.psb. ugent.be/beg/tools/venn-diagrams), an online tool for calculating the intersections of multiple datasets, was used for obtaining the intersections of the above three databases for selected miRNAs.

2.6 Functional annotation for miRNA targets

The database for annotation, visualization and integrated discovery (DAVID) v6.8 (https://david.ncifcrf. gov/), an online resource for providing comprehensively functional annotations [31], was used for exploring the potential mechanisms exerted by interested miRNAs. The analysis was completed following the typical analysis flow in the user's manuals (https://david.ncifcrf.gov/ content.jsp?file=functional_annotation.html). The detailed parameters were as follows: target mRNAs as the gene list for loading, official gene symbol as the retrieval identifier, and homo sapiens as the background.

2.7 Statistical analysis

Results were analyzed with SPSS 22.0 software (IBM SPSS, USA). Independent sample *t*-test or Mann–Whitney *U* test was used for measurement data and chi-square test was used for enumeration data. Related diagrams were completed by GraphPad Prism 7 software (GraphPad Software Inc., USA). A value of p < 0.05 means a statistic difference.

3 Results

3.1 SCCHN-related miRNA expression profiles in GEO DataSets

By searching the miRNA expression profiles of SCCHN in GEO DataSets based on retrieval terms, 15 profiles matching the retrieval terms were screened out. After a detailed review of the 15 profiles, some of them were excluded, according to the screening criteria. Of them, GSE58911 dataset was first eliminated as a de facto mRNA expression profile. Another five profiles (GSE10301, GSE25524, GSE32176, GSE33196, and

GSE79369) concerning SCCHN cell lines and three profiles (GSE11163, GSE34496, and GSE92595) concerning unpaired SCCHN tissues were also excluded, due to dissatisfaction with the criteria. In addition, GSE33232 and GSE79372, integrated datasets composed of four subseries separately, were further eliminated due to cell lines and unpaired tissue sources. Moreover, GSE31277, also a dataset, concerned cell lines and unpaired tissues and was composed of three platforms, which could be extracted using a miRNA expression profile of 14 paired tissues based on platform GPL9770, namely, GSE31277-GPL9770. Subsequently, GSE31277-GPL9770 and three other datasets based on paired tissues (GSE32115, GSE62819, and GSE124566) were enrolled in subsequent analyses (Figure 1).

3.2 Distribution of DEMs in each included dataset

For each included dataset, the DEMs were calculated with GEO2R and further filtered according to adjustment of the value of p < 0.05 and fold change ≥ 2 . Subsequently, total 76, 103, and 29 DEMs were acquired from GSE32115, GSE124566, and GSE31277-GPL9770, respectively. Notably, after screening by adjusting the value of p < 0.05, no eligible DEMs were screened out in GSE62819. Hence, GSE62819 was further eliminated from the present study. Further analysis indicated that there was no intersection among the remaining three datasets, but only a few DEMs that co-existed in two datasets. Considering that the amount of DEMs and integrity of annotation databases were affected by the time of publication, GSE32115 (published in 2011) and GSE31277-GPL9770 (published in 2014) were further removed from subsequent analyses. Ultimately, the recently published dataset GSE124566 (published in 2020) was selected for subsequent analyses with 103 DEMs (Figure 1).

3.3 Details of DEMs in GSE124566

Through the preliminary screening criteria (adjustment of p < 0.05 and fold change ≥ 2), total 103 DEMs (63 overexpressed and 40 downexpressed miRNAs) were screened out from GSE124566. For acquiring the more significant DEMs in GSE124566, the screening criteria were further narrowed down to adjust p < 0.01 and fold change ≥ 4 . Total 18 more significant DEMs (11



Figure 1: Flowchart for analyses. GEO, gene expression omnibus , DEMs, differential expressed miRNAs, TCGA, the cancer genome atlas, ROC, receiver operating characteristic.

overexpressed and 7 downexpressed miRNAs) were screened out and used for subsequent analyses (Figure 2).

3.4 SCCHN-related miRNA expression profile in TCGA

For verification, the miRNA expression levels of GSE124566 in SCCHN, a different miRNA expression profile, were acquired from TCGA, which consists of 524 cancerous tissues and 45 normal tissues of SCCHN and includes 2,242 miRNAs (Query strategy: Project Id IS TCGA-HNSC AND Access IS open AND Data Category IN (clinical transcriptome profiling) AND Data Format IS txt AND Data Type IS miRNA Expression Quantification). Based on the TCGA miRNA expression profile, the 18 DEMs of GSE124566 were well validated in expression tendency. Of the 18 miRNAs, miR-4324 is not included in the TCGA miRNA expression profile. For the remaining 17 miRNAs, total 12 miRNA expression trends in TCGA expression profile were consistent with GSE124566 (Figure 3). Nevertheless, miR-142-5p, miR-23b-5p, and miR-4647 showed no significant difference in the expression trend between cancerous and normal tissues.



Figure 2: Differential expressed miRNAs of GSE124566. Red triangles represent the 11 overexpressed miRNAs and green triangles represent the seven downexpressed miRNAs in cancerous tissues of SCCHN, respectively. Gray dots represent miRNAs with no significant difference (adjustment of p < 0.01 and fold change \geq 4). SCCHN, squamous cell carcinoma of head and neck.

Even *miR-513c-5p* and *miR-3654* showed an opposite expression tendency to GSE124566.

3.5 Relationships between the expression levels of 12 validated miRNAs and overall survival times

Based on the clinical data and miRNA expression levels of TCGA, survival analyses showed that most of the 12 validated miRNAs correlated with overall survival times with high hazard ratios (HR). However, merely miR-223-3p and miR-142-3p showed significant differences with lower p-values (p < 0.05) (Figure 4). Inconsistently, for an overexpressed miR-142-3p in cancerous tissues, it seems to present that patients with high expression levels of

miR-142-3p may have a long survival period. Consequently, survival analyses indicated that patients with high-level *miR-223-3p* have shorter survival times than patients with low expression levels (p = 0.006, HR = 1.440) (Logrank test).

3.6 Diagnostic values of the 12 validated DEMs

Through a receiver operating characteristic (ROC) curve analyzing the diagnostic values of 12 validated DEMs, the results presented that five out of 12 miRNAs may have potential diagnostic values with high area under ROC curves (AUC) (AUC \geq 0.85, p < 0.05), including *miR-503-5p*, *miR-424-5p*, *miR-204-5p*, and *miR-375-3p* (Figure 5).



Figure 3: Expression levels of miRNAs in SCCHN based on TCGA database. *X*-axis represents the types of tissues, C, cancerous tissues, N, normal tissues. The numbers in parentheses represent the number of samples. Due to the absence of some miRNA expression values in expression profile data, the sample sizes are unequal for different miRNAs. *Y*-axis represents the relative expression levels of miRNAs in SCCHN tissues. TPM, transcripts per million. A value of p < 0.05 means significant difference.



Figure 4: Overall survival analyses for differential expressed miRNAs. *X*-axis represents the overall survival time and *Y*-axis represents the survival rates at different times. High means the miRNA expression levels were higher than the median of total miRNA expression levels, while low the opposite. HR: hazard ratio. HR (high) means the hazard ratio of high expression group to low expression group, while HR (low) the opposite. A value of p < 0.05 and HR > 0.5 mean significant difference.

3.7 Comparison of the expression levels of miRNAs with diagnostic or prognostic values in different clinical feature groups

For exploring the associations between the miRNA expression levels and clinical features, the expression levels of miRNAs (*miR-223-3p*, *miR-503-5p*, *miR-424-5p*, *miR-21-5p*, *miR-204-5p*, and *miR-375-3p*) in different groups with distinct clinical features were compared and analyzed. Analyses showed that *miR-223-3p*, *miR-503-5p*, and *miR-424-5p* expression levels varied with different genders (p < 0.05). Besides, *miR-223-3p* expression levels were changed in different clinical stages (p = 0.026) and *miR-204-5p* expression levels were also distinctive in

different differentiation degrees (p = 0.036). Added, specimens with smoking or without smoking showed different *miR-503-5p* expression levels (p = 0.000) (Table 1).

3.8 Prediction of target mRNAs and functional enrichment analyses for *miR*-223-3p and *miR*-204-5p

Comprehensively considering survival analyses, ROC curve and miRNA differential expression levels in distinct clinical feature groups, *miR-223-3p* and *miR-204-5p* were selected for further analyses to filter the significant miRNAs in



Figure 5: Receiver operating characteristic curves of differential expressed miRNAs. ROC, Receiver operating characteristic, AUC, area under ROC curve. CI, confidence interval. A value of p < 0.05 and AUC ≥ 0.85 mean effective diagnostic values.

SCCHN. Followed, the target mRNAs of miR-223-3p and miR-204-5p were predicted by TargetScanHuman 7.2, miRDB, and miRWalk separately. The intersection mRNAs of the three databases (103 mRNAs for miR-223-3p and 292 mRNAs for miR-204-5p) were used for subsequent analyses. Afterwards, functional enrichment analyses of intersection mRNAs showed that miR-223-3p or miR-204-5p may be involved in multiple biological processes, molecular functions, and signaling pathways, respectively. Analyses illustrated that *miR-223-3p* may participate in multiple biological processes (GO: 0001837 ~ epithelial-to-mesenchymal transition and GO: 0030177 ~ positive regulation of Wnt signaling pathway) and molecular functions (GO: 0046322 ~ SMAD binding and GO: 0008134 ~ transcription factor binding), which indicated that miR-223-3p may be involved in the regulation of EMT. Analyses also illustrated that miR-223-3p and miR-204-5p may be involved in the process of energy anabolism and catabolism mediated by the same signaling pathway (hsa04152: *AMPK* signaling pathway).

4 Discussion

The expression profile is a composite of multifarious valuable information, for instance, the difference in overall gene expression trend between the experimental group and the control group, the distribution of differential expressed genes in the experimental group or the control group, and the difference of a specific gene among different specimens. Since the above-mentioned characteristics, re-analyses of gene expression profiles acquired more valuable and significant references for further research studies [32–35].

| | | Gender | | Age (years) | | Drinking | | Smoking | | Stage | | Differentiation | | Lymph node metastasis | |
|----------------|----------------|--------|--------|-------------|-------|----------|-------|---------|--------|-------|--------|-----------------|-------|--------------------------|-------|
| | | Male | Female | <61 | ≥61 | No | Yes | No | Yes | I–II | III–IV | High- medium | Low | No | Yes |
| miR- | High (n) | 178 | 84 | 119 | 142 | 83 | 174 | 62 | 195 | 66 | 196 | 195 | 64 | 131 | 118 |
| 223-3p | Low (n) | 204 | 57 | 138 | 123 | 81 | 176 | 59 | 194 | 45 | 216 | 175 | 68 | 114 | 141 |
| | χ2 | | 6.938 | | 2.767 | | 0.036 | | 0.046 | | 4.942 | | 0.693 | | 3.151 |
| | р | | 0.008 | | 0.096 | | 0.850 | | 0.831 | | 0.026 | | 0.405 | | 0.076 |
| miR- | High (n) | 180 | 81 | 138 | 122 | 83 | 174 | 82 | 175 | 55 | 206 | 192 | 57 | 116 | 133 |
| 503-5p | Low (n) | 200 | 60 | 118 | 142 | 81 | 174 | 39 | 212 | 56 | 204 | 176 | 75 | 127 | 126 |
| | χ2 | | 4.178 | | 3.078 | | 0.017 | | 18.750 | | 0.017 | | 3.142 | | 0.655 |
| | р | | 0.041 | | 0.079 | | 0.898 | | 0.000 | | 0.897 | | 0.076 | | 0.418 |
| miR- | High (n) | 180 | 82 | 131 | 130 | 74 | 181 | 70 | 188 | 60 | 202 | 180 | 71 | 120 | 134 |
| 424-5p | Low (n) | 202 | 59 | 126 | 135 | 90 | 169 | 51 | 201 | 51 | 210 | 190 | 61 | 125 | 125 |
| | χ ² | | 5.017 | | 0.192 | | 1.941 | | 3.348 | | 0.883 | | 1.028 | | 0.383 |
| | р | | 0.025 | | 0.662 | | 0.164 | | 0.067 | | 0.347 | | 0.311 | | 0.536 |
| miR- | High (n) | 194 | 68 | 125 | 137 | 78 | 179 | 63 | 193 | 54 | 208 | 173 | 74 | 121 | 132 |
| 21-5p | Low (n) | 188 | 73 | 132 | 129 | 86 | 171 | 58 | 196 | 57 | 204 | 197 | 58 | 124 | 127 |
| | χ ² | | 0.270 | | 0.429 | | 0.573 | | 0.222 | | 0.118 | | 3.370 | | 0.125 |
| | р | | 0.604 | | 0.512 | | 0.449 | | 0.638 | | 0.731 | | 0.066 | | 0.723 |
| miR- 204-5p | High (n) | 155 | 69 | 103 | 120 | 75 | 145 | 55 | 163 | 57 | 167 | 147 | 67 | 114 | 103 |
| | Low (n) | 172 | 52 | 117 | 107 | 69 | 151 | 52 | 167 | 43 | 181 | 167 | 48 | 100 | 115 |
| | χ ² | | 3.272 | | 1.633 | | 0.372 | | 0.130 | | 2.523 | | 4.411 | | 1.567 |
| | р | | 0.071 | | 0.201 | | 0.542 | | 0.718 | | 0.112 | | 0.036 | | 0.211 |
| miR- | High (n) | 193 | 69 | 131 | 131 | 74 | 173 | 55 | 198 | 58 | 204 | 187 | 67 | 130 | 119 |
| 375-3p | Low (n) | 189 | 72 | 126 | 134 | 90 | 167 | 66 | 191 | 53 | 208 | 183 | 65 | 115 | 140 |
| | χ ² | | 0.104 | | 0.124 | | 1.469 | | 1.095 | | 0.262 | | 0.002 | | 2.550 |
| | p | | 0.747 | | 0.725 | | 0.226 | | 0.295 | | 0.609 | | 0.966 | | 0.110 |

Table 1: Comparison of miRNA expressions in different clinical features

Samples were divided into high expression levels and low expression levels according to the median of expression levels for each miRNA. Bold represents significant difference. Sixty-one years represent the median age.

Hitherto, numerous profiles, involving coding and noncoding RNA expression profiles, genome variation profiles, protein profiles, and methylation profiles, have been published and reserved in GEO [24], TCGA [26], or Oncomine (https://www.oncomine.org/) [36], which could be re-analyzed. A study elaborated the significant prognostic values of multiple genes for SCCHN patients through re-analyses based on TCGA and GEO profiles [37]. Our previous studies also completed the screening of SCCHNrelated hub genes through re-analyses of mRNA expression profiles [33], and mined a significantly overexpressed longnon-coding RNA LINCOO668 for further study of noncoding RNA profiles [34]. Here, the present study further explored the miRNAs with diagnostic and prognostic values in SCCHN with the method of re-analyses of miRNA expression profiles.

After rigorous screening, a recently published miRNA expression profile of SCCHN (GSE124566) was included in the present analyses. The original research study, which published the dataset GSE124566, proposed that total 43 deregulated miRNAs (fold change >2, adjusted p < 0.01) were screened out from included SCCHN tissue specimens [12]. In the present study, total 18 DEMs were screened out from GSE124566 (fold change \geq 4, adjusted *p* < 0.01), all of which were included in the above-mentioned 43 deregulated miRNAs. Of the 18 miRNAs, 12 miRNA expression trends were verified by TCGA, including the downregulated *miR-204-5p* in cancerous tissues, a proved tumor suppressor by the original research study [12]. The consistency between the re-analyses and the original study further confirmed the reliability of the present study and promoted the progression of the study. Followed survival analyses presented that the high levels of miR-223-3p were related to the poor prognosis of SCCHN patients, which indicated that miR-223-3p may be a prognostic marker for SCCHN. In addition, the survival analyses also suggested that the patients with high expression levels

of *miR-142-3p* presented significantly long survival times. This phenomenon seems to embody a protective effect of *miR-142-3p* on SCCHN patients, but this effect is contrary to the overexpression of miR-142-3p in cancerous tissues. This result needs a thorough study to explore the definite role of miR-142-3p in SCCHN. Certain miRNAs are helpful for cancer diagnoses [38]. In view of this, the present study distinguished five miRNAs with possible diagnostic values, which consisted of miR-503-5p, miR-424-5p, miR-21-5p, miR-204-5p, and miR-375-3p. All of the five miRNAs are worthy of further study on cancerous diagnosis for identifying an effective tumor marker for diagnosis. Of the above-mentioned six miRNAs with prognostic or diagnostic values, miR-223-3p and miR-204-5p presented significant distinct expression levels in different clinical stages or tissue differentiation, respectively. Notably, for miR-204-5p, the original study indicated that the patients with high expression levels of miR-204-5p showed higher clinical stages and positive neck lymph node metastasis [12]. However, apart from tissue differentiation degrees, the present study has not been able to ascertain the correlations between miR-204-5p expression levels and clinical stages or lymph node metastasis. This inconsistency may be related to the heterogeneity of tumor tissues, and it is likely to be eliminated following the implementation of follow-up studies. However, the prognostic value of miR-204-5p and its correlation with clinical features suggested that miR-204-5p may be an effective tumor marker or target for diagnostic and prognostic evaluation or therapy, which has been elaborated and confirmed in the original research study [12]. The present analyses also indicated that the miR-503-5p expression levels were related to smoking history, an agreed risk factor for the occurrence and development of SCCHN [39]. This finding suggested that *miR-503-5p* may play an unknown role in the effect of tobacco on SCCHN.

In general, miRNAs exert diverse regulatory roles through target differential mRNAs [11,14,16,22,40], same for *miR-223-3p* and *miR-204-5p*. Considering the potential clinical significance of *miR-223-3p* and *miR-204-5p*, it is necessary and worthy to define the possible targets for providing reference for the followed study. Therefore, TargetScanHuman 7.2, miRDB, and miRWalk jointly predicted the targets of *miR-223-3p* and *miR-204-5p*, respectively. The joint prediction of multiple databases greatly heightened the dependability of target mRNAs and the feasibility of subsequent study. Functional analyses based on target mRNAs speculated that *miR-223-3p* could be involved in the progression of SCCHN through EMT, a confirmed regulatory mechanism for the

development of SCCHN [41–43]. It may be a novel angle to elucidate the pathogenesis of SCCHN from the perspective of *miR-223-3p* regulating the EMT. Analyses also suggested that *miR-223-3p* and *miR-204-5p* may affect the energy balance of SCCHN through *AMPK* signaling pathway, a proved regulatory signaling pathway for the energy metabolism of tumors [44–47]. Energy metabolism plays important roles in cancer progression and the regulation of energy metabolism balance may become one of the effective anti-tumor treatment methods [48,49].

The present study screened out the altered 18 miRNAs in SCCHN through a series of analyses and identified the six miRNAs (*miR-223-3p*, *miR-503-5p*, *miR-424-5p*, *miR-21-5p*, *miR-204-5p*, and *miR-375-3p*) with significant prognostic or diagnostic values. Especially for *miR-223-3p* and *miR-204-5p*, the present analyses speculated that *miR-223-3p* may modulate the progression of SCCHN through the mediation of EMT. Meanwhile, *miR-223-3p* and *miR-204-5p* may also involve in the regulation of cancer energy metabolism through *AMPK* signaling pathway. It is worthy to further explore the potential molecular mechanisms for them, in consideration of the prognostic, diagnostic, and clinical significance.

5 Abbreviations

| AUC | area under ROC curve | | | | |
|--------|--|--|--|--|--|
| DAVID | The Database for Annotation, Visualization | | | | |
| | and Integrated Discovery | | | | |
| DEMs | differential expressed miRNAs | | | | |
| EMT | epithelial-mesenchymal transition | | | | |
| GDC | Genomic Data Commons | | | | |
| GEO | Gene Expression Omnibus | | | | |
| HR | hazard ratio | | | | |
| miRNAs | microRNAs | | | | |
| ROC | receiver operating characteristic | | | | |
| SCCHN | squamous cell carcinoma of head and neck | | | | |
| TCGA | The Cancer Genome Atlas | | | | |

Acknowledgments: The original data here are derived from the NCBI/GEO Datasets (http://www.ncbi.nlm.nih. gov/geo/) and the TCGA Research Network (https:// www.cancer.gov/tcga). This study was funded by the Key Program of Hebei Research and Development (No. 172777177) and Hebei Medical Science Research Program (No. 20200575).

Authors contributions: (I) Conception and design: Lei Zhao; (II) administrative support: Lei Zhao and Congzhe Tian; (III) collection and assembly of data: Erbin Xiao, Jinduo Du, and Jingwei Liang; (IV) data analysis and interpretation: Lei Zhao, Weiwei Chi, and Xianghong Chen; (V) manuscript writing and revision: All authors.

Conflict of interest: All authors have no competing interests.

References

- Leemans CR, Snijders PJF, Brakenhoff RH. The molecular landscape of head and neck cancer. Nat Rev Cancer. 2018;18(5):269–82.
- [2] Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin. 2018;68(6):394–424.
- [3] Solomon B, Young RJ, Rischin D. Head and neck squamous cell carcinoma: genomics and emerging biomarkers for immunomodulatory cancer treatments. Semin Cancer Biol. 2018;52(Pt 2):228–40.
- [4] Magnes T, Egle A, Greil R, Melchardt T. Update on squamous cell carcinoma of the head and neck: ASCO annual meeting 2017. Memo. 2017;10(4):220–3.
- [5] Schotz U, Balzer V, Brandt FW, Ziemann F, Subtil FSB, Rieckmann T, et al. Dual PI3K/mTOR inhibitor NVP-BEZ235 enhances radiosensitivity of head and neck squamous cell carcinoma (HNSCC) cell lines due to suppressed double-strand break (DSB) repair by nonhomologous end joining. Cancer. 2020;12(2):467.
- [6] Ludwig N, Yerneni SS, Razzo BM, Whiteside TL. Exosomes from HNSCC promote angiogenesis through reprogramming of endothelial cells. Mol Cancer Res. 2018;16(11):1798–808.
- Jiang Y, Cao W, Wu K, Qin X, Wang X, Li Y, et al. LncRNA LINC00460 promotes EMT in head and neck squamous cell carcinoma by facilitating peroxiredoxin-1 into the nucleus. J Exp Clin Cancer Res. 2019;38(1):365.
- [8] Kolenda T, Guglas K, Teresiak A, Blizniak R, Lamperska K. Low let-7d and high miR-205 expression levels positively influence HNSCC patient outcome. J Biomed Sci. 2019;26(1):17.
- [9] Yu D, Liu X, Han G, Liu Y, Zhao X, Wang D, et al. The let-7 family of microRNAs suppresses immune evasion in head and neck squamous cell carcinoma by promoting PD-L1 degradation. Cell Commun Signal. 2019;17(1):173.
- [10] Saleh AD, Cheng H, Martin SE, Si H, Ormanoglu P, Carlson S, et al. Integrated genomic and functional microRNA analysis identifies miR-30-5p as a tumor suppressor and potential therapeutic nanomedicine in head and neck cancer. Clin Cancer Res. 2019;25(9):2860–73.
- [11] Zhang Y, Zhang Z, Huang W, Zeng J. MiR-4282 inhibits tumor progression through down-regulation of ZBTB2 by targeting LIN28B in oral squamous cell carcinoma. J Cell Physiol. 2020. doi: 10.1002/jcp.29458.

- [12] Zhuang Z, Yu P, Xie N, Wu Y, Liu H, Zhang M, et al. MicroRNA-204-5p is a tumor suppressor and potential therapeutic target in head and neck squamous cell carcinoma. Theranostics. 2020;10(3):1433–53.
- [13] Vahabi M, Pulito C, Sacconi A, Donzelli S, D'Andrea M, Manciocco V, et al. miR-96-5p targets PTEN expression affecting radio-chemosensitivity of HNSCC cells. J Exp Clin Cancer Res. 2019;38(1):141.
- [14] Lin XJ, Liu H, Li P, Wang HF, Yang AK, Di JM, et al. miR-936 suppresses cell proliferation, invasion, and drug resistance of laryngeal squamous cell carcinoma and targets GPR78. Front Oncol. 2020;10:60.
- [15] Yeh LY, Yang CC, Wu HL, Kao SY, Liu CJ, Chen YF, et al. The miR-372-ZBTB7A oncogenic axis suppresses TRAIL-R2 associated drug sensitivity in oral carcinoma. Front Oncol. 2020;10:47.
- [16] Zhang S, Li G, Liu C, Lu S, Jing Q, Chen X, et al. miR-30e-5p represses angiogenesis and metastasis by directly targeting AEG-1 in squamous cell carcinoma of the head and neck. Cancer Sci. 2020;111(2):356–68.
- [17] Valenti F, Sacconi A, Ganci F, Grasso G, Strano S, Blandino G, et al. The miR-205-5p/BRCA1/RAD17 axis promotes genomic instability in head and neck squamous cell carcinomas. Cancers. 2019;11(9):1347.
- [18] Moradimotlagh A, Arefian E, Rezazadeh Valojerdi R, Ghaemi S, Jamshidi Adegani F, Soleimani M. MicroRNA-129 inhibits glioma cell growth by targeting CDK4, CDK6, and MDM2. Mol Ther Nucleic Acids. 2020;19:759–64.
- [19] Ajuyah P, Hill M, Ahadi A, Lu J, Hutvagner G, Tran N. MicroRNA (miRNA)-to-miRNA regulation of programmed cell death 4 (PDCD4). Mol Cell Biol. 2019;39(18):e00086.
- [20] Li Z, Lu J, Zeng G, Pang J, Zheng X, Feng J, et al. MiR-129-5p inhibits liver cancer growth by targeting calcium calmodulindependent protein kinase IV (CAMK4). Cell Death Dis. 2019;10(11):789.
- [21] Wan P, Bai X, Yang C, He T, Luo L, Wang Y, et al. miR-129-5p inhibits proliferation, migration, and invasion in rectal adenocarcinoma cells through targeting E2F7. J Cell Physiol. 2020;235(7–8):5689–701.
- [22] Yu D, Han GH, Zhao X, Liu X, Xue K, Wang D, et al. MicroRNA-129-5p suppresses nasopharyngeal carcinoma lymphangiogenesis and lymph node metastasis by targeting ZIC2. Cell Oncol. 2020;43(2):249–61.
- [23] Wu C, Miao C, Tang Q, Zhou X, Xi P, Chang P, et al. MiR-129-5p promotes docetaxel resistance in prostate cancer by downregulating CAMK2N1 expression. J Cell Mol Med. 2020;24(3):2098–108.
- [24] Barrett T, Troup DB, Wilhite SE, Ledoux P, Rudnev D, Evangelista C, et al. NCBI GEO: mining tens of millions of expression profiles – database and tools update. Nucleic Acids Res. 2007;35(Database issue):D760–5.
- [25] Davis S, Meltzer PS. GEOquery: a bridge between the gene expression omnibus (GEO) and bioconductor. Bioinformatics. 2007;23(14):1846-7.
- [26] Gao GF, Parker JS, Reynolds SM, Silva TC, Wang LB, Zhou W, et al. Before and after: comparison of legacy and harmonized TCGA genomic data commons' data. Cell Syst. 2019;9(1):24–4.e10.
- [27] Jensen MA, Ferretti V, Grossman RL, Staudt LM. The NCI genomic data commons as an engine for precision medicine. Blood. 2017;130(4):453–9.

- [28] Agarwal V, Bell GW, Nam JW, Bartel DP. Predicting effective microRNA target sites in mammalian mRNAs. Elife. 2015;4:e05005.
- [29] Chen Y, Wang X. miRDB: an online database for prediction of functional microRNA targets. Nucleic Acids Res. 2020;48(D1):D127-31.
- [30] Sticht C, De La Torre C, Parveen A, Gretz N. miRWalk: an online resource for prediction of microRNA binding sites. PLoS One. 2018;13(10):e0206239.
- [31] Huang da W, Sherman BT, Lempicki RA. Systematic and integrative analysis of large gene lists using DAVID bioinformatics resources. Nat Protoc. 2009;4(1):44–57.
- [32] Yang M, Huang W, Sun Y, Liang H, Chen M, Wu X, et al. Prognosis and modulation mechanisms of COMMD6 in human tumours based on expression profiling and comprehensive bioinformatics analysis. Br J Cancer. 2019;121(8):699–709.
- [33] Zhao L, Chi W, Cao H, Cui W, Meng W, Guo W, et al. Screening and clinical significance of tumor markers in head and neck squamous cell carcinoma through bioinformatics analysis. Mol Med Rep. 2019;19(1):143–54.
- [34] Zhao L, Cao H, Chi W, Meng W, Cui W, Guo W, et al. Expression profile analysis identifies the long non-coding RNA landscape and the potential carcinogenic functions of LINC00668 in laryngeal squamous cell carcinoma. Gene. 2019;687:47–55.
- [35] Feng B, Shen Y, Pastor X, Bieg M, Plath M, Ishaque N, et al. Integrative analysis of multi-omics data identified EGFR and PTGS2 as key nodes in a gene regulatory network related to immune phenotypes in head and neck cancer. Clin Cancer Res. 2020;26(14):3616–28.
- [36] Rhodes DR, Yu J, Shanker K, Deshpande N, Varambally R, Ghosh D, et al. ONCOMINE: a cancer microarray database and integrated data-mining platform. Neoplasia. 2004;6(1):1–6.
- [37] Wang J, Chen X, Tian Y, Zhu G, Qin Y, Chen X, et al. Six-gene signature for predicting survival in patients with head and neck squamous cell carcinoma. Aging. 2020;12(1):767–83.
- [38] Liu C, Yu Z, Huang S, Zhao Q, Sun Z, Fletcher C, et al. Combined identification of three miRNAs in serum as effective diagnostic biomarkers for HNSCC. EBioMedicine. 2019;50:135–43.
- [39] Jethwa AR, Khariwala SS. Tobacco-related carcinogenesis in head and neck cancer. Cancer Metastasis Rev. 2017;36(3):411-23.

- [40] Chen J, Zhong Y, Li L. miR-124 and miR-203 synergistically inactivate EMT pathway via coregulation of ZEB2 in clear cell renal cell carcinoma (ccRCC). J Transl Med. 2020;18(1):69.
- [41] You X, Zhou Z, Chen W, Wei X, Zhou H, Luo W. MicroRNA-495 confers inhibitory effects on cancer stem cells in oral squamous cell carcinoma through the HOXC6-mediated TGF-beta signaling pathway. Stem Cell Res Ther. 2020;11(1):117.
- [42] Pai S, Bamodu OA, Lin YK, Lin CS, Chu PY, Chien MH, et al. CD47-SIRPalpha signaling induces epithelial-mesenchymal transition and cancer stemness and links to a poor prognosis in patients with oral squamous cell carcinoma. Cells. 2019;8(12):1658.
- [43] Lin C, Ren Z, Yang X, Yang R, Chen Y, Liu Z, et al. Nerve growth factor (NGF)-TrkA axis in head and neck squamous cell carcinoma triggers EMT and confers resistance to the EGFR inhibitor erlotinib. Cancer Lett. 2020;472:81–96.
- [44] Li Y, Liang R, Sun M, Li Z, Sheng H, Wang J, et al. AMPKdependent phosphorylation of HDAC8 triggers PGM1 expression to promote lung cancer cell survival under glucose starvation. Cancer Lett. 2020;478:82–92.
- [45] Zhou C, Lyu LH, Miao HK, Bahr T, Zhang QY, Liang T, et al. Redox regulation by SOD2 modulates colorectal cancer tumorigenesis through AMPK-mediated energy metabolism. Mol Carcinog. 2020;59(5):545–56.
- [46] Barisciano G, Colangelo T, Rosato V, Muccillo L, Letizia Taddei M, Ippolito L, et al. miR-27a is a master regulator of metabolic reprogramming and chemoresistance in colorectal cancer. Br J Cancer. 2020;122(9):1354–66.
- [47] Huang YJ, Jan YH, Chang YC, Tsai HF, Wu AT, Chen CL, et al. ATP synthase subunit epsilon overexpression promotes metastasis by modulating AMPK signaling to induce epithelial-to-mesenchymal transition and is a poor prognostic marker in colorectal cancer patients. J Clin Med. 2019;8(7):1070.
- [48] Yang B, Ding L, Chen Y, Shi J. Augmenting tumor-starvation therapy by cancer cell autophagy inhibition. Adv Sci. 2020;7(6):1902847.
- [49] Chen YY, Wang WH, Che L, Lan Y, Zhang LY, Zhan DL, et al. BNIP3L-dependent mitophagy promotes HBx-induced cancer stemness of hepatocellular carcinoma cells via glycolysis metabolism reprogramming. Cancers. 2020;12(3):655.