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# The effect of long non-coding RNAs in joint destruction of rheumatoid arthritis

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Rheumatoid arthritis (RA) is a systemic autoimmune disease accompanied with joint destruction. Serious joint destruction will eventually lead to disability and the decline of life quality in RA patients. At present, the therapeutic effect of drugs to alleviate joint destruction in RA is limited. Recently, accumulating evidences have shown that long non-coding RNAs (lncRNAs) play an important role in the pathogenesis of joint diseases. Therefore, this paper reviews the expression change and the action mechanism of lncRNAs in joint destruction of RA in recent years. A more comprehensive understanding of the role of lncRNAs in joint destruction will help the treatment of RA.

#### KEYWORDS

rheumatoid arthritis, IncRNAs, joint destruction, cells, ceRNA

# Introduction

Rheumatoid arthritis (RA) is a disease characterized by progressive symmetrical inflammation and joint destruction. In inflamed joints, chronic destructive synovitis forms, causing irreversible damage to cartilage and bone, which can eventually lead to joint deformities and affect the normal life of RA patients (Weyan and Goronzy. 2021). Up to now, there are limited drugs targeting joint destruction of RA. Therefore, it is necessary to further investigate the molecular mechanism responsible for joint destruction of RA so as to develop novel therapeutic strategies.

In eukaryotic cells, about 90% of RNAs cannot encode proteins, and these RNAs are called non-coding RNAs. In non-coding RNAs, those with a length greater than 200 nucleotides are called long non-coding RNAs (lncRNAs), and those with a length of about 22 nucleotides are called microRNAs (miRNAs) (Deviatkin et al., 2020). LncRNAs can interact with DNA, RNA and protein to regulate transcription, modify epigenetics, maintain the stability, translation and post-translational modification (Bridges et al., 2021).Current studies show that lncRNAs have the following action mode (Bhat et al., 2016; Dykes and Emanueli. 2017): lncRNAs can interact with chromatin-modifying enzymes and RNA-binding factors to activate or inhibit the

transcription of target genes. LncRNAs can be an enhancer or a decoy for the transcription factor to promote or repress the transcription. Besides, lncRNAs, as negative regulators, regulate miRNAs expression and function by specifically sponging their corresponding miRNAs, which is called as the competing endogenous RNAs (ceRNAs). This competitive relationship realized through is the combination of miRNA reaction elements (MREs) contained in lncRNAs or mRNAs and miRNAs seed sequences. LncRNAs are now known to participate in many biological processes such as cell proliferation, development and apoptosis, as well as disease progression including cancer, diabetes and cardiovascular disease (He et al., 2017; Simion et al., 2019; Nandwani et al., 2021). Recently, accumulating evidences have indicated that lncRNAs also play an important role in the pathogenesis of RA (Lodde et al., 2020; Huang et al., 2021). They can regulate the development and function of a variety of cells related to RA, including fibroblast-like synoviocytes (FLSs), osteoclasts, osteoblasts, and chondrocytes, etc. (Zhu et al., 2019; Guo et al., 2020; Li, et al., 2020; Aurilia et al., 2021), hence influencing the progression of joint destruction.

Therefore, this paper aimed to summarize the action mechanism of lncRNAs in various cells of RA joints and their important effect on joint destruction in recent years. The potential of lncRNAs as promising drug targets in joint destruction of RA was also discussed. We searched the original English articles from 2017 to 2022 through PubMed with the following keywords: rheumatoid arthritis and lncRNAs and joint destruction, rheumatoid arthritis and lncRNAs and T lymphocytes, rheumatoid arthritis and lncRNAs and B lymphocytes, rheumatoid arthritis and lncRNAs and macrophages, rheumatoid arthritis and lncRNAs and fibroblast-like synoviocytes, rheumatoid arthritis and lncRNAs and chondrocytes, rheumatoid arthritis and lncRNAs and osteoblasts, rheumatoid arthritis and lncRNAs and osteoclasts, rheumatoid arthritis and lncRNAs and treatment. Comments and articles in other languages were excluded. We hope this review will provide clues for further study of lncRNAs as biomarkers and effective therapeutic targets in treating joint destruction of RA.

# Joint destruction in RA

Joints are composed of osteoclasts, osteoblasts, chondrocytes, synovial cells and other types of immune cells such as T lymphocytes and B lymphocytes. These cells interact with each other and jointly maintain joint homeostasis.

In the maintenance of joint homeostasis, the balance of bone homeostasis is very important. In healthy joints, bone formation promoted by osteoblasts and bone resorption

mediated by osteoclasts jointly maintain the dynamic balance of bone tissue. However, in the joints of RA, this balanced relationship is broken, resulting in bone destruction (Zhao et al., 2020). Mature osteoclasts are formed from expressing CD14<sup>+</sup> and CD16<sup>-</sup> circulating monocytes/ macrophages and tissue-specific macrophages after fusion and multinucleation (Rana et al., 2018; Yao et al., 2021). The combination of receptor activator of nuclear factor  $\kappa B$ ligand (RANKL) and macrophage colony-stimulating factor (M-CSF) with their respective ligands activates downstream related signaling pathways affecting the formation and development of osteoclasts (Chen et al., 2018; Győri and Mocsal. 2020). The inflammatory environment of RA joint promotes osteoblasts and other cells to secrete RANKL, thereby promoting osteoclast-mediated bone resorption (Auréal et al., 2020). Osteoblasts are differentiated from mesenchymal stem cells by activating various cell growth factors, transcription factors, and multiple signaling pathways such as wingless/integrated (WNT), bone morphogenetic protein (BMP) signaling pathways (Neve et al., 2011). In the past, osteoclasts obtained intensive investigations in the field of bone destruction research in RA. In contrast, osteoblasts have received less attention. Recently, studies have reported that the maturation and mineralization of osteoblasts was compromised in RA, and regulating the signaling pathways of osteoblasts could effectively attenuate bone destruction and simultaneously promote bone formation in RA (Berardi et al., 2021).

Chondrocytes are the only cell in cartilage, which can produce and maintain cartilage matrix. In RA, the function of chondrocytes is dysregulated. On the one hand, cytokines such as tumor necrosis factor-alpha (TNF- $\alpha$ ) and interferon- $\gamma$  (IFN- $\gamma$ ) can interfere with chondrogenesis and promote chondrocyte apoptosis (Tseng et al., 2020). On the other hand, matrix metalloproteinases (MMPs) secreted by chondrocytes themselves can also accelerate the destruction of cartilage (Otero and Goldring. 2007).

Besides the osteoclasts, osteoblasts and chondrocytes, there are also other types of cells directly or indirectly involved in the pathological process of joint destruction in RA. FLSs, as the main non immune cells in synovial tissue, are generally believed to play central roles in RA progression. They can promote joint destruction in a variety of ways. FLSs participate in osteoclastogenesis by secreting RANKL and M-CSF, and they also have the ability to directly migrate and even invade articular cartilage (Harre and Schett. 2017; Yoshitomi 2019; Nygaard and Firestein. 2020). In addition, FLSs can accelerate cartilage destruction by secreting MMPs, such as MMP-3 and MMP-9. Clinical studies have shown that the level of MMP-3 in serum can effectively reflect the activity of RA disease and joint and bone injury (Lerner et al., 2018). MMP-9 is also proved to be closely related to bone resorption of RA (Cabral-Pacheco et al., 2020).

In RA, T lymphocytes infiltrate into the synovial membrane where they can initiate and inhibit the activities of other cells. T lymphocytes of subtypes such as helper T (Th)1 and Th17 cells can up-regulate the expression of receptor activator of nuclear factor-  $\kappa B$  (RANK) in osteoclast precursors. However, some subtypes of T cells can inhibit the development and maturation of osteoclast precursors by secreting IL-4 (Tanaka 2019; Deng et al., 2021). For osteoblasts, T lymphocytes can also regulate the differentiation of osteoblasts in both directions (Deng et al., 2021).

The role of B lymphocytes in joint destruction of RA has been emphasized in recent years. B lymphocytes can inhibit the differentiation of osteoblasts by producing TNF- $\alpha$  and C-C motif chemokine 3 (CCL3) (Sun et al., 2018).They can also produce RANKL to promote the differentiation of osteoclast precursors (Meednu et al., 2016). In addition, autoantibodies such as rheumatoid factor (RF) produced by B lymphocytes also participate in the pathogenesis of RA joint destruction. Harre U and their colleagues found that the immune complex formed by RF and autoantigens can induce osteoclasts differentiation through Fc $\gamma$  receptors (Fc $\gamma$ R) in RA (Harre et al., 2015).

# LncRNAs in joint destruction of RA

## LncRNAs and FLSs

FLSs are crucial participants in the joint destruction of RA. In FLSs of RA, the expression of a few of lncRNAs were abnormal. LncRNA PICSAR, lncRNA LINK-A and lncRNA ZFAS1 expression were up-regulated (Bi et al., 2019; Wang, et al., 2021; Wang, et al., 2021).While lncRNA THRIL expression was down-regulated (Zou et al., 2021). Inhibition of lncRNA PICSAR, lncRNA LINK-A and lncRNA ZFAS1 expression could weakened the invasion ability of FLSs, whereas lncRNA THRIL knockout showed the opposite results.

Besides influencing the invasion of FLSs, lncRNAs also modulate the production of MMPs in FLSs of RA. Research results explained that lncRNA NEAT1 promoted the secretion of MMP-9 in FLSs of RA *via* interacting with miR-410-3p and miR-204-5p (Wang et al., 2020; Xiao et al., 2021). Another study showed that silence of lncRNA THRIL inhibited MMP-3 production in FLSs of RA by regulating the phosphatidylinositol 3-kinases (PI3K)/protein kinase B (AKT) signaling pathway (Liang et al., 2020). One study indicated that overexpression of lncRNA OSER1-AS1 could decreased the production of MMP-3 in FLSs of RA. The mechanism was that lncRNA OSER1-AS1 served as a ceRNA through the sponge of miR-1298-5p and increased the expression of early two factor transcription factor 1 (E2F1) (Fu et al., 2022). In addition, another study found IncRNAS56464. One could bind to miR-152-3p, and then affect the expression of WNT1 in FLSs of RA (Jiang et al., 2021). As a secretory glycoprotein, WNT1 is a core member of WNT signaling pathway, which has been proved to be closely related to the progression and pathogenesis of RA by regulating articular chondrogenesis and bone destruction (Malysheva et al., 2016).

#### LncRNAs and osteoclasts

Osteoclasts are the major players directly responsible for the pathogenesis of joint destruction in RA. Numerous research have proved that the proliferation and differentiation of osteoclasts were promoted, leading to the enhancement of bone resorption in RA (Tateiwa et al., 2019; Kitaura et al., 2020).

One study reported that the expression of lncRNA X inactive specific transcript (Xist) was significantly increased in female patients with RA. LncRNA Xist could promote the proliferation of osteoclasts, and this promotion effect could be enhanced after overexpression of lncRNA Xist (Bost et al., 2022). Further, several researchers investigated the mechanism of lncRNA Xist in regulating the proliferation and development of osteoclasts. Shao et al. demonstrated that lncRNA Xist contributed to osteoclast differentiation through serving as a ceRNA of miR-590-3p to promote Tgif2 level (Shao et al., 2021). Other findings suggested that lncRNA Xist promoted osteoclast differentiation through sphingosine kinase 1 (SPHK1)/sphingosine 1-phosphate (S1P)/ extracellular signal-regulated kinase (ERK) signaling pathway. It could interact with fused in sarcoma (FUS) and increase the stability of SPHK1 (Zhang et al., 2022a). Although these articles reported the mechanism of action of lncRNA Xist in osteoclasts from different aspects, the role of lncRNA Xist in RA joint destruction still needs to be further verified by in vivo experimental data.

Besides lncRNA Xist, some other lncRNAs have been also proved to have the effect on regulating osteoclastogenesis. A study confirmed that lncRNA SNHG15 was highly expressed in THP-1 cells stimulated by M-CSF and RANKL. Overexpression of lncRNA SNHG15 could promote the proliferation, differentiation and metastasis of osteoclasts through sponging with miR-381-3p to upregulate the expression of never-in-mitosis-A-related kinase 2 (NEK2) (Wang, et al., 2022). Another research indicated that lncRNA NEAT1 also played an important role in osteoclast differentiation. Knockdown of lncRNA NEAT1 impaired osteoclastogenesis, whereas overexpression of lncRNA NEAT1 promoted osteoclastogenesis. Mechanistically, lncRNA NEAT1 sponged with miR-7 and blocked miR-7mediated regulation of protein tyrosine kinase 2 (PTK2). Moreover, this study showed that rs12789028 could act as a strong allele-specific functional enhancer for lncRNA NEAT1 (Zhang et al., 2020b).

Recently, with the advances in RNA sequencing (RNA-seq)) technology, lncRNA-mRNA expression profiles have become the research hotspot. A couple of lncRNA-mRNA co-expression network was established and analyzed. Through RNA-seq analysis, a previous study noted that a lot of lncRNAs and mRNAs differentially expressed in human osteoclast differentiation. Then, researchers constructed a lncRNA-mRNA network co-expression and found that lncRNA ENSG00000257764.2 obtained the highest number of interactions and interacted with tissue inhibitor of metalloproteinases 2 (TIMP2), which was an inhibitor of MMPs (Li, et al., 2020). Hypoxia-inducible factor  $1\alpha$  (HIF- $1\alpha$ ) has been implicated in the pathogenesis of RA and it participates in osteoclast differentiation by regulating nutrient and energy sensors (Fearon et al., 2016; Tang et al., 2019). Therefore, another study explored the lncRNA-mRNA expression profiles associated with HIF-1aknockout mouse osteoclast differentiation by RNA-seq (Tian 2022). The et al., results showed that lncRNA MSTRG.31769.2 and MSTRG.7566.12 were the two lncRNAs with the two highest numbers of interactions. The expression of these two lncRNAs had a strong negative correlation with MMP-9 and cathepsin K (CTSK) expression. However, most of these results are from in vitro experiments and lack of further validation in RA animal models and patients.

## LncRNAs and osteoblasts

Albeit osteoblasts are among the dominant cell types in the joint, their role in joint destruction of RA has only been received attention in recent years. Interference of the growth and differentiation of osteoblasts is one of the important factors causing joint destruction of RA.

The expression of lncRNA Colorectal Neoplasia Differentially Expressed (CRNDE) in various cancer tissues and plasma is sensitive and specific (Lu et al., 2020). Recent studies have demonstrated that lncRNA CRNDE could promote the proliferation and differentiation of osteoblasts through WNT signaling pathway (Maeda et al., 2019). Mice with lncRNA CRNDE knockout showed reduced bone mass. Moreover, the proliferation and differentiation of osteoblasts isolated from lncRNA CRNDE knockout mice were inhibited (Mulati et al., 2020). LncRNA differentiation antagonizing non-protein coding RNA (DANCR) play an important role in many diseases. A study demonstrated that lncRNA DANCR suppressed the differentiation of mesenchymal stem cells by promoting the degradation of Skp2-induced forkhead box O1 (FOXO1) ubiquitination. Silencing lncRNA DANCR could promote the maturation and mineralization of osteoblasts (Tang et al., 2018). Although the expression level of lncRNA DANCR was upregulated in the bone tissue of patients undergoing total hip

replacement, this expression change has not been confirmed in patients or animal models with RA.

In addition to lncRNA CRNDE and lncRNA DANCR, increasing *in vitro* cell experiments showed the effects of lncRNAs in regulating the proliferation, differentiation and mineralization of osteoblasts. By sponging miR-33a, lncRNA MCF2L-AS1 positively regulated the expression of Runx2 and promoted the differentiation and calcium nodule formation in human bone marrow mesenchymal stem cells (BMSCs) (Chen, Wang, et al., 2020). Linc-ROR could promote osteogenesis of mesenchymal stem cells by functioning as a ceRNA for miR-138 and miR-145. The ALP activity and calcium nodules formation were increased in linc-ROR-overexpressing cells but decreased in linc-ROR-knockdown cells (Feng et al., 2018). Nevertheless, despite some reports of lncRNAs and osteoblasts, the function and mechanism of these lncRNAs in joint destruction of RA are still largely unknown, which deserve further studies.

### LncRNAs and chondrocytes

As an important part of the joint, chondrocytes also contribute to the progressive destructive process of RA. Chondrocytes are the target cells of multiple inflammatory factors. Simultaneously, chondrocytes also act as effector cells, directly or indirectly facilitate joint damage of RA (Tseng et al., 2020).

As a member of a family of isozymes, pyruvate dehydrogenase kinase 4 (PDK4) is one of the most important factors which direct carbon flux into glycolysis from oxidative phosphorylation (Li, et al., 2020). It can promote proliferation and migration of chondrocytes through the RANKL/RANK/osteoprotegerin (OPG) pathway. Recent studies indicated that PDK4 was decreased in chondrocytes of RA (Liu, et al., 2020). The results of one study pointed out that lncRNA GAS5 was overexpressed in LPS-induced chondrocytes. It decreased the proliferation of chondrocytes by sponging miR-361-5p to modulate PDK4 expression (Zhang et al., 2021). Another research reported that knockout of lncRNA GAS5 could prevented cartilage destruction and reduce the expression of MMP-13 and FGF21 in cartilage of antigeninduced arthritis (AIA) mice (Chen, et al., 2020). Besides, Gao et al. showed that lncRNA GAS5 could induce apoptosis of chondrocytes from osteoarthritis patients through downregulating miR-137 (Gao et al., 2020). Although lncRNA GAS was confirmed to be upregulated in cartilage and serum of patients with osteoarthritis, there is still a lack of validation in serum and cartilage of patients with RA.

Simultaneously, the function of several other lncRNAs in chondrocytes have also been investigated by *in vitro* experiments. LncRNA MEG3 and lncRNA ZNF667-AS1 were down-regulated in LPS-stimulated chondrocytes. These two lncRNAs could protect chondrocytes from LPS-induced damage and inhibit the secretion of inflammatory factors by chondrocytes (Li et al., 2019; Zhuo et al., 2021).

### LncRNAs and T lymphocytes

In normal environment, a variety of cytokines secreted by T cells maintain a dynamic balance in the proliferation, development, differentiation and apoptosis of osteoblasts, osteoclasts and chondrocytes. Many lncRNAs were found to be abnormal in T cells of patients with RA. LncRNA GAS5, lncRNA RMRP, lncRNA THRIL and lncRNA NEAT1 were found to be increased in T cells of RA patients (Moharamoghli et al., 2019; Shui et al., 2019; Liu et al., 2021).

As major subsets of CD4<sup>+</sup> Th cells, Th17 cells not only promoted joint inflammation, but also induced osteoclasts differentiation and bone destruction of RA. IL-17 secreted by Th17 cells can promote the production of RANKL by osteoblasts and synoviocytes, thus indirectly inducing osteoclasts differentiation (Hashimoto 2017; Yang et al., 2019; Tang et al., 2020). One study pointed out that the expression of lncRNA NEAT1 was up-regulated in Th17 cells differentiated by CD4+T cells of RA *in vitro* (Shui et al., 2019). Another study suggested that lncRNA NEAT1 in serum derived exosomes could promote the proliferation of CD4+T cells and differentiate into Th17 cells (Liu et al., 2021). Through exosomes, circulating lncRNAs can be delivered to recipient cells and participate in the pathogenesis of RA.

#### LncRNAs and B lymphocytes

In RA, a variety of cytokines and autoantibodies secreted by B cells can inhibit the differentiation of osteoblasts and chondrocytes, as well as promote the formation of osteoclasts. Although the role of B cells in the progression of RA disease has been extensive investigated, there are few studies on lncRNAs and B cells in RA. It is well known that glucocorticoids inhibit the survival of osteoblasts and promote the life span of osteoclasts through glucocorticoid receptors, leading to the risk of bone loss (Güler-Yüksel et al., 2018; Lee et al., 2021). One study found that the expression of lncRNA GAS5 in B cells of RA patients was obviously decreased. The pathogenesis of lncRNA GAS5 involved in RA may be related to the inhibition of glucocorticoid receptors via its decoy RNA "glucocorticoid response element" (Mayama et al., 2016).

## LncRNAs' multiple effects

Interestingly, not only different lncRNAs can regulate the same cell, a lncRNA can also act on different cells and perform different functions as well. For instance, one study showed that lncRNA GAS5 accelerated cartilage destruction by regulating PI3K/AKT/fibroblast growth factor 21 (FGF21) axis through sponging miR-103 (Chen et al., 2020). Other studies pointed that lncRNA GAS5 promoted apoptosis of FLSs and suppress

their proliferation by promoting histone deacetylase 4 (HDAC4) and sirtuin 1 (Sirt1) via inhibiting miR-128–3p and miR-222–3p (Peng et al., 2021; Yang et al., 2021). In addition, one study showed that through binding miR-181c-5p, lncRNA SNHG1 induced BMSCs differentiation into osteoclasts, whereas inhibited the osteogenic differentiation of BMSCs (Yu et al., 2021). Other studies reported that lncRNA MEG3 could promote the proliferation and differentiation of osteoblasts by activating WNT signaling pathway, and affect the differentiation of macrophages into osteoclasts by inhibiting interferon regulatory factor 8 (IRF8) (Li et al., 2019; Gao et al., 2022). The above results indicate that the functions of lncRNAs are variety and complex.

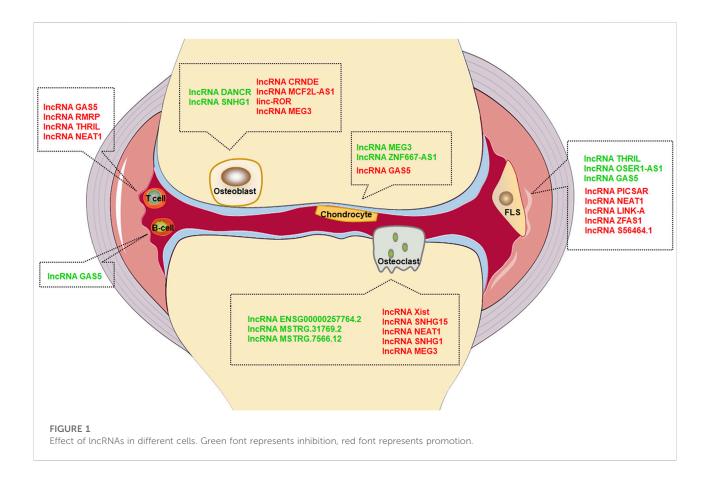
# Treatment

Given that lncRNAs participate in the development of joint destruction, focusing on lncRNAs could be a promising strategy for treatment of RA. Bioactive glass nanoparticles (BGN) are attractive for orthopedic applications (Kong et al., 2018). A study reported that BGN induced BMSCs to secrete extracellular vesicles containing lncRNA NRON, which could prevent osteoclasts differentiation by blocking the nuclear translocation of nuclear factor of activated T cell cytoplasmic 1 (NFATc1), a core member of the nuclear factor of activated T cell (NFAT) family of transcription factors that involved in osteoclasts differentiation (Gu et al., 2020; Yang et al., 2022). This study provides a hopeful strategy for the treatment of bone-related disease including RA. Besides that, some drugs for RA have also been found to regulate the expression of lncRNAs involved in joint destruction. Methotrexate is the guideline recommended first-line treatment for RA (Smolen et al., 2020). Iguratimod is a new drug that can promote bone formation and inhibit bone resorption (Xie et al., 2020).Combined treatment with methotrexate and iguratimod reduced the expression of lncRNA HOTAIR in RA patients (Tan et al., 2021). Recently, research has reported that lncRNA HOTAIR could promote osteoclasts differentiation, inhibit osteoclasts apoptosis and osteoblasts differentiation (Shen et al., 2019; Dong et al., 2022).

In addition to being a drug target, lncRNAs can also be used as a biomarker to judge the efficacy of drugs. Etanercept, the first TNF inhibitor used in the treatment of RA, can effectively improve bone loss and inhibit joint destruction in RA patients (Gulyás et al., 2020).Some researchers reported that after etanercept treatment in RA patients, there were significant differences in the expression of some lncRNAs between the responders and non-responders. These lncRNAs were enriched into osteoclasts differentiation signaling pathways, etc. (Wang et al., 2022). Tripterysium Glycosides Tablets

#### TABLE 1 Roles of IncRNAs in joint destruction of RA.

lncRNAs	Model	Targets	Signaling	Roles	References
lncRNA PICSAR	FLSs	miR-4701-5p	_	promote FLSs proliferation, migration and invasion	Bi et al. (2019)
lncRNA LINK-A	FLSs	miR-1262	_	promote FLSs invasion	Wang et al. (2021)
lncRNA ZFAS1	FLSs	miR-3926	_	promote FLSs proliferation, migration, invasion	Wang et al. (2021)
lncRNA NEAT1	FLSs	miR-410-3p; miR-204-5p	_	promote FLSs migration, invasion, and inflammatory cytokines secretion	Wang et al. (2020); Xiao et al (2021)
lncRNA \$56464.1	FLSs	miR-152-3p	WNT signaling pathway	promote FLSs proliferation	Jiang et al. (2021)
lncRNA THRIL	FLSs	MMP3	PI3K/AKT signaling pathway	inhibit FLSs proliferation, migration, invasion	Liang et al. (2020); Zou et al. (2021)
lncRNA OSER1-AS1	FLSs	miR-1298-5p	_	inhibit FLSs propagation, release of inflammatory factor	Fu et al. (2022)
lncRNA GAS5	FLSs	miR-128–3p; miR-222–3p	_	inhibit FLSs proliferation	Peng et al. (2021); Yang et al (2021)
lncRNA Xist	osteoclasts from RA women; RAW264.7 cells; BMMs	miR- 590–3p; FUS	SPHK1/S1P/ERK signaling pathway	promote the proliferation and differentiation of osteoclasts	Shao et al. (2021); Bost et al. (2022); Zhang et al. (2022)
lncRNA SNHG15	THP-1 cells	miR-381–3p	_	promote the proliferation, differentiation and metastasis of osteoclasts	Wang et al. (2022)
lncRNA NEAT1	BMMs and PBMCs	miR-7	_	promote osteoclast differentiation	Zhang et al., 2022
lncRNA ENSG00000257764.2	CD14 monocytes	TIMP2	_	-	Liu et al. (2020)
lncRNA MSTRG.31769.2	RAW264.7 cells	_	_	_	Tian et al. (2022)
lncRNA MSTRG.7566.12	RAW264.7 cells	_	_	_	Tian et al. (2022)
lncRNA MEG3	RAW264.7 cells	IRF8	_	affect osteoclasts differentiation	Gao et al. (2022)
lncRNA SNHG1	BMSCs	miR-181c-5p	WNT signaling pathway	inhibit osteoclast differentiation and induce osteoblast differentiation	Yu et al. (2021)
lncRNA CRNDE	MC3T3-E1 cells and CRNDE knockout mice	_	WNT signaling pathway	promote osteoblast proliferation and differentiation	Mulati et al. (2020)
lncRNA MCF2L-AS1	BMSCs	miR-33a	—	promote osteogenic differentiation	Chen et al. (2020)
linc-ROR	BMSCs	miR-138; miR-145	WNT signaling pathway	promote osteogenic differentiation	Feng et al. (2018)
lncRNA DANCR	BMSCs	FOXO1	_	inhibit osteoblast differentiation	Tang et al. (2018)
lncRNA MEG3	MC3T3-E1 cells	_	WNT signaling pathway	promote proliferation and differentiation of osteoblasts	Li, et al. (2019)
lncRNA GAS5	chondrocytes	miR-361–5p; miR-137	_	involve in chondrocytes proliferation and apoptosis	Gao et al. (2020); Zhang et al (2021)
lncRNA MEG3	chondrocytes	miR-141	AKT/mTOR signaling pathway	promote chondrocytes proliferation	Li et al. (2019)
lncRNA ZNF667-AS1	chondrocytes	miR-523-3p	JAK/STAT signaling pathway	promote chondrocytes proliferation	Zhuo et al. (2021)
lncRNA GAS5	T lymphocytes	_	_	_	Moharamoghli et al. (2019)
lncRNA THRIL	T lymphocytes	_	_	_	Moharamoghli et al. (2019)
lncRNA RMRP	T lymphocytes	_	_	_	Moharamoghli et al. (2019)
lncRNA NEAT1	T lymphocytes	STAT3; miR- 144–3p	WNT signaling pathway	promote CD4+T cells to differentiate into Th17 cells	Shui et al. (2019); Liu et al. (2021)
lncRNA GAS5	B lymphocytes	_	_	_	Mayama et al. (2016)



(TGT), derived from Tripterygium wilfordii Hook F, have been used to treat RA in clinic for many years. TGT can effectively improve the tissue architecture of joints and prevent the bone destruction in RA (Wang et al., 2017). However, clinical evidence shows that only 70% of RA patients respond to TGT. Recently, researchers found a TGT response-related lncRNA. Their study showed that the expression of lncRNA ENST000000494760 in the effective group of TGT was significantly down-regulated compared with that in the ineffective group. Further studies indicated that overexpressing **lncRNA** ENST00000494760 promoted C1qC expression by sponging with miR-654-5p (Zhang, Wang, et al., 2020). C1qC is one of polypeptide chains of C1q. A study showed that C1q could enhance osteoclasts development (Teo et al., 2012).

Collectively, these studies indicated that lncRNAs might provide targets and evaluation indicators for personalized drug therapy of joint destruction in RA.

# Conclusion and prospect

At present, the treatment of bone destruction in RA is still a big challenge in clinic. The research progress of lncRNAs in bone destruction of RA provides us with a new therapeutic idea. The role of lncRNAs in RA bone destruction is complex and diverse. LncRNAs can regulate various type of cells in joints of RA through different ways, ultimately leading to joint destruction. However, there are still some limitations in the current research on lncRNAs' role in bone destruction of RA. On the one hand, majority of studies about lncRNAs in RA joint destruction are limited to cellular levels, lacking further verification from in vivo experiments and clinical trials. On the other hand, the role of lncRNAs in regulating some important cells such as macrophages in the joint of RA are still unclear. This might be one of the next research directions of lncRNAs in RA. Furthermore, current studies on lncRNAs in joint destruction mostly focus on evaluating the change of expression levels and exploring the mechanisms of action, with less experimental data on pathological implications as well as diagnosis and treatment potential, which deserve deeper research in future. Additionally, owing to the complex and diverse role of lncRNAs in different pathways and different cells, some new technologies such as multiomics and systemic biology are best introduced in in-depth studies.

To sum up, we reviewed the potential and action mechanism of lncRNAs participating in joint destruction and suggested that joint destruction of RA can be alleviated by regulating lncRNAs (Table 1 and Figure 1). LncRNAs can serve as potential biomarker and therapeutic target for RA joint destruction.

# Author contributions

HZ wrote the draft of the manuscript. LL and NZ contributed to the literature research for the manuscript. AL, CL, and XH revised the manuscript. All authors contributed to the article and approved the submitted version.

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# References

Auréal, M., Machuca-Gayet, I., and Coury, F. (2020). Rheumatoid arthritis in the view of osteoimmunology. *Biomolecules* 11, 48. doi:10.3390/biom11010048

Aurilia, C., Donati, S., Palmini, G., Miglietta, F., Iantomasi, T., and Brandi, M. L. (2021). The involvement of long non-coding RNAs in bone. *Int. J. Mol. Sci.* 22, 3909. doi:10.3390/ijms22083909

Berardi, S., Corrado, A., Maruotti, N., Cici, D., and Cantatore, F. P. (2021). Osteoblast role in the pathogenesis of rheumatoid arthritis. *Mol. Biol. Rep.* 48, 2843–2852. doi:10.1007/s11033-021-06288-y

Bhat, S. A., Ahmad, S. M., Mumtaz, P. T., Malik, A. A., Dar, M. A., Urwat, U., et al. (2016). Long non-coding RNAs: Mechanism of action and functional utility. *Noncoding. RNA Res.* 1, 43–50. doi:10.1016/j.ncrna.2016.11.002

Bi, X., Guo, X. H., Mo, B. Y., Wang, M. L., Luo, X. Q., Chen, Y. X., et al. (2019). LncRNA PICSAR promotes cell proliferation, migration and invasion of fibroblastlike synoviocytes by sponging miRNA-4701-5p in rheumatoid arthritis. *EBioMedicine* 50, 408–420. doi:10.1016/j.ebiom.2019.11.024

Bost, C., Arleevskaya, M. I., Brooks, W. H., Plaza, S., Guery, J. C., and Renaudineau, Y. (2022). Long non-coding RNA Xist contribution in systemic lupus erythematosus and rheumatoid arthritis. *Clin. Immunol.* 236, 108937. doi:10. 1016/j.clim.2022.108937

Bridges, M. C., Daulagala, A. C., and Kourtidis, A. (2021). LNCcation: lncRNA localization and function. J. Cell Biol. 220, e202009045. doi:10.1083/jcb.202009045

Cabral-Pacheco, G. A., Garza-Veloz, I., Castruita-De la Rosa, C., Ramirez-Acuña, J. M., Perez-Romero, B. A., Guerrero-Rodriguez, J. F., et al. (2020). The roles of matrix metalloproteinases and their inhibitors in human diseases. *Int. J. Mol. Sci.* 21, E9739. doi:10.3390/ijms21249739

Chen, H., He, C., Liu, Y., Li, X., Zhang, C., Qin, Q., et al. (2020). LncRNA-GAS5 inhibits expression of miR 103 and ameliorates the articular cartilage in adjuvant-induced arthritis in obese mice. *Dose. Response.* 18, 1559325820942718. doi:10. 1177/1559325820942718

Chen, Q., Wang, M., and Wu, S. (2020). The lncRNA MCF2L-AS1 controls osteogenic differentiation by regulating miR-33a. *Cell Cycle* 19, 1059–1065. doi:10. 1080/15384101.2020.1747776

Chen, X., Wang, Z., Duan, N., Zhu, G., Schwarz, E. M., and Xie, C. (2018). Osteoblast-osteoclast interactions. *Connect. Tissue Res.* 59, 99–107. doi:10.1080/ 03008207.2017.1290085

Deng, Z., Zhang, Q., Zhao, Z., Li, Y., Chen, X., Lin, Z., et al. (2021). Crosstalk between immune cells and bone cells or chondrocytes. *Int. Immunopharmacol.* 101, 108179. doi:10.1016/j.intimp.2021.108179

Deviatkin, A. A., Vakulenko, Y. A., Akhmadishina, L. V., Tarasov, V. V., Beloukhova, M. I., Zamyatnin, A. A., Jr., et al. (2020). Emerging concepts and challenges in rheumatoid arthritis gene therapy. *Biomedicines* 8, E9. doi:10.3390/biomedicines8010009

# **Conflict of interest**

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Dong, C., Liu, X., Li, J., Lan, D., and Zheng, S. (2022). Dysregulation of the HOTAIR-miR-152-camkiia Axis in craniosynostosis results in impaired osteoclast differentiation. *Front. Genet.* 13, 787734. doi:10.3389/fgene.2022.787734

Dykes, I. M., and Emanueli, C. (2017). Transcriptional and post-transcriptional gene regulation by long non-coding RNA. *Genomics Proteomics Bioinforma*. 15, 177–186. doi:10.1016/j.gpb.2016.12.005

Fearon, U., Canavan, M., Biniecka, M., and Veale, D. J. (2016). Hypoxia, mitochondrial dysfunction and synovial invasiveness in rheumatoid arthritis. *Nat. Rev. Rheumatol.* 12, 385–397. doi:10.1038/nrrheum.2016.69

Feng, L., Shi, L., Lu, Y. F., Wang, B., Tang, T., Fu, W. M., et al. (2018). Linc-ROR promotes osteogenic differentiation of mesenchymal stem cells by functioning as a competing endogenous RNA for miR-138 and miR-145. *Mol. Ther. Nucleic Acids* 11, 345–353. doi:10.1016/j.omtn.2018.03.004

Fu, Q., Song, M. J., and Fang, J. (2022). LncRNA OSER1-AS1 regulates the inflammation and apoptosis of rheumatoid arthritis fibroblast like synoviocytes via regulating miR-1298-5p/E2F1 axis. *Bioengineered* 13, 4951–4963. doi:10.1080/21655979.2022.2037854

Gao, S. T., Yu, Y. M., Wan, L. P., Liu, Z. M., and Lin, J. X. (2020). LncRNA GAS5 induces chondrocyte apoptosis by down-regulating miR-137. *Eur. Rev. Med. Pharmacol. Sci.* 24, 10984–10991. doi:10.26355/eurrev\_202011\_23582

Gao, X., Ge, J., Zhou, W., Xu, L., and Geng, D. (2022). IL-10 inhibits osteoclast differentiation and osteolysis through MEG3/IRF8 pathway. *Cell. Signal.* 95, 110353. doi:10.1016/j.cellsig.2022.110353

Gu, W., Wang, Z., Sun, Z., Bao, Z., Zhang, L., Geng, Y., et al. (2020). Role of NFATc1 in the bone-vascular Axis calcification paradox. *J. Cardiovasc. Pharmacol.* 75, 200–207. doi:10.1097/fjc.000000000000788

Güler-Yüksel, M., Hoes, J. N., Bultink, I. E. M., and Lems, W. F. (2018). Glucocorticoids, inflammation and bone. *Calcif. Tissue Int.* 102, 592–606. doi:10.1007/s00223-017-0335-7

Gulyás, K., Horváth, Á., Végh, E., Pusztai, A., Szentpétery, Á., Pethö, Z., et al. (2020). Effects of 1-year anti-TNF- $\alpha$  therapies on bone mineral density and bone biomarkers in rheumatoid arthritis and ankylosing spondylitis. *Clin. Rheumatol.* 39, 167–175. doi:10.1007/s10067-019-04771-3

Guo, Q., Guo, Q., Xiao, Y., Li, C., Huang, Y., and Luo, X. (2020). Regulation of bone marrow mesenchymal stem cell fate by long non-coding RNA. *Bone* 141, 115617. doi:10.1016/j.bone.2020.115617

Győri, D. S., and Mócsai, A. (2020). Osteoclast signal transduction during bone metastasis formation. *Front. Cell Dev. Biol.* 8, 507. doi:10.3389/fcell.2020. 00507

Harre, U., Lang, S. C., Pfeifle, R., Rombouts, Y., Frühbeißer, S., Amara, K., et al. (2015). Glycosylation of immunoglobulin G determines osteoclast differentiation and bone loss. *Nat. Commun.* 6, 6651. doi:10.1038/ncomms7651

Harre, U., and Schett, G. (2017). Cellular and molecular pathways of structural damage in rheumatoid arthritis. *Semin. Immunopathol.* 39, 355–363. doi:10.1007/s00281-017-0634-0

Hashimoto, M. (2017). Th17 in animal models of rheumatoid arthritis. J. Clin. Med. 6, E73. doi:10.3390/jcm6070073

He, X., Ou, C., Xiao, Y., Han, Q., Li, H., and Zhou, S. (2017). LncRNAs: Key players and novel insights into diabetes mellitus. *Oncotarget* 8, 71325–71341. doi:10. 18632/oncotarget.19921

Huang, H., Xing, D., Zhang, Q., Li, H., Lin, J., He, Z., et al. (2021). LncRNAs as a new regulator of chronic musculoskeletal disorder. *Cell Prolif.* 54, e13113. doi:10. 1111/cpr.13113

Jiang, H., Liu, J., Fan, C., Wang, J., and Li, W. (2021). lncRNAS56464.1 as a ceRNA promotes the proliferation of fibroblast-like synoviocytes in experimental arthritis via the Wnt signaling pathway and sponges miR-152-3p. *Int. J. Mol. Med.* 47, 17. doi:10.3892/ijmm.2021.4850

Kitaura, H., Marahleh, A., Ohori, F., Noguchi, T., Shen, W. R., Qi, J., et al. (2020). Osteocyte-related cytokines regulate osteoclast formation and bone resorption. *Int. J. Mol. Sci.* 21, E5169. doi:10.3390/ijms21145169

Kong, C. H., Steffi, C., Shi, Z., and Wang, W. (2018). Development of mesoporous bioactive glass nanoparticles and its use in bone tissue engineering. *J. Biomed. Mat. Res. B Appl. Biomater.* 106, 2878–2887. doi:10. 1002/jbm.b.34143

Lee, S., Krüger, B. T., Ignatius, A., and Tuckermann, J. (2021). Distinct glucocorticoid receptor actions in bone homeostasis and bone diseases. *Front. Endocrinol.* 12, 815386. doi:10.3389/fendo.2021.815386

Lerner, A., Neidhöfer, S., Reuter, S., and Matthias, T. (2018). MMP3 is a reliable marker for disease activity, radiological monitoring, disease outcome predictability, and therapeutic response in rheumatoid arthritis. *Best. Pract. Res. Clin. Rheumatol.* 32, 550–562. doi:10.1016/j.berh.2019.01.006

Li, D., Yang, C., Yin, C., Zhao, F., Chen, Z., Tian, Y., et al. (2020). LncRNA, important player in bone development and disease. *Endocr. Metab. Immune Disord. Drug Targets* 20, 50–66. doi:10.2174/1871530319666190904161707

Li, G., Liu, Y., Meng, F., Xia, Z., Wu, X., Fang, Y., et al. (2019). LncRNA MEG3 inhibits rheumatoid arthritis through miR-141 and inactivation of AKT/ mTOR signalling pathway. *J. Cell. Mol. Med.* 23, 7116–7120. doi:10.1111/jcmm. 14591

Li, X. G., Liu, S. C., Qiao, X. F., Kong, Y., Liu, J. G., Peng, X. M., et al. (2019). LncRNA MEG3 promotes proliferation and differentiation of osteoblasts through Wnt/β-catenin signaling pathway. *Eur. Rev. Med. Pharmacol. Sci.* 23, 4521–4529. doi:10.26355/eurrev\_201906\_18027

Li, Z., Peng, Y., Li, J., Chen, Z., Chen, F., Tu, J., et al. (2020). N(6)methyladenosine regulates glycolysis of cancer cells through PDK4. *Nat. Commun.* 11, 2578. doi:10.1038/s41467-020-16306-5

Liang, Y., Li, H., Gong, X., and Ding, C. (2020). Long non-coding RNA THRIL mediates cell growth and inflammatory response of fibroblast-like synoviocytes by activating PI3K/AKT signals in rheumatoid arthritis. *Inflammation* 43, 1044–1053. doi:10.1007/s10753-020-01189-x

Liu, D., Fang, Y., Rao, Y., Tan, W., Zhou, W., Wu, X., et al. (2020). Synovial fibroblast-derived exosomal microRNA-106b suppresses chondrocyte proliferation and migration in rheumatoid arthritis via down-regulation of PDK4. *J. Mol. Med.* 98, 409–423. doi:10.1007/s00109-020-01882-2

Liu, R., Jiang, C., Li, J., Li, X., Zhao, L., Yun, H., et al. (2021). Serum-derived exosomes containing NEAT1 promote the occurrence of rheumatoid arthritis through regulation of miR-144-3p/ROCK2 axis. *Ther. Adv. Chronic Dis.* 12, 2040622321991705. doi:10.1177/2040622321991705

Liu, W., Li, Z., Cai, Z., Xie, Z., Li, J., Li, M., et al. (2020). LncRNA-mRNA expression profiles and functional networks in osteoclast differentiation. *J. Cell. Mol. Med.* 24, 9786–9797. doi:10.1111/jcmm.15560

Lodde, V., Murgia, G., Simula, E. R., Steri, M., Floris, M., and Idda, M. L. (2020). Long noncoding RNAs and circular RNAs in autoimmune diseases. *Biomolecules* 10, E1044. doi:10.3390/biom10071044

Lu, Y., Sha, H., Sun, X., Zhang, Y., Wu, Y., Zhang, J., et al. (2020). Crnde: An oncogenic long non-coding RNA in cancers. *Cancer Cell Int.* 20, 162. doi:10.1186/s12935-020-01246-3

Maeda, K., Kobayashi, Y., Koide, M., Uehara, S., Okamoto, M., Ishihara, A., et al. (2019). The regulation of bone metabolism and disorders by wnt signaling. *Int. J. Mol. Sci.* 20, E5525. doi:10.3390/ijms20225525

Malysheva, K., de Rooij, K., Lowik, C. W., Baeten, D. L., Rose-John, S., Stoika, R., et al. (2016). Interleukin 6/wnt interactions in rheumatoid arthritis: Interleukin 6 inhibits wnt signaling in synovial fibroblasts and osteoblasts. *Croat. Med. J.* 57, 89–98. doi:10.3325/cmj.2016.57.89

Mayama, T., Marr, A. K., and Kino, T. (2016). Differential expression of glucocorticoid receptor noncoding RNA repressor Gas5 in autoimmune and inflammatory diseases. *Horm. Metab. Res.* 48, 550–557. doi:10.1055/s-0042-106898

Meednu, N., Zhang, H., Owen, T., Sun, W., Wang, V., Cistrone, C., et al. (2016). Production of RANKL by memory B cells: A link between B cells and bone erosion in rheumatoid arthritis. *Arthritis Rheumatol.* 68, 805–816. doi:10.1002/art.39489

Moharamoghli, M., Hassan-Zadeh, V., Dolatshahi, E., Alizadeh, Z., and Farazmand, A. (2019). The expression of GAS5, THRIL, and RMRP lncRNAs is increased in T cells of patients with rheumatoid arthritis. *Clin. Rheumatol.* 38, 3073–3080. doi:10.1007/s10067-019-04694-z

Mulati, M., Kobayashi, Y., Takahashi, A., Numata, H., Saito, M., Hiraoka, Y., et al. (2020). The long noncoding RNA Crnde regulates osteoblast proliferation through the Wnt/ $\beta$ -catenin signaling pathway in mice. *Bone* 130, 115076. doi:10.1016/j. bone.2019.115076

Nandwani, A., Rathore, S., and Datta, M. (2021). LncRNAs in cancer: Regulatory and therapeutic implications. *Cancer Lett.* 501, 162–171. doi:10.1016/j.canlet.2020.11.048

Neve, A., Corrado, A., and Cantatore, F. P. (2011). Osteoblast physiology in normal and pathological conditions. *Cell Tissue Res.* 343, 289–302. doi:10.1007/s00441-010-1086-1

Nygaard, G., and Firestein, G. S. (2020). Restoring synovial homeostasis in rheumatoid arthritis by targeting fibroblast-like synoviocytes. *Nat. Rev. Rheumatol.* 16, 316–333. doi:10.1038/s41584-020-0413-5

Otero, M., and Goldring, M. B. (2007). Cells of the synovium in rheumatoid arthritis. Chondrocytes. Arthritis Res. Ther. 9, 220. doi:10.1186/ar2292

Peng, T., Ji, D., and Jiang, Y. (2021). Long non-coding RNA GAS5 suppresses rheumatoid arthritis progression via miR-128-3p/HDAC4 axis. *Mol. Cell. Biochem.* 476, 2491–2501. doi:10.1007/s11010-021-04098-1

Rana, A. K., Li, Y., Dang, Q., and Yang, F. (2018). Monocytes in rheumatoid arthritis: Circulating precursors of macrophages and osteoclasts and, their heterogeneity and plasticity role in RA pathogenesis. *Int. Immunopharmacol.* 65, 348–359. doi:10.1016/j.intimp.2018.10.016

Shao, Y., Hu, X., and Wu, X. (2021). LncRNA X inactive-specific transcript promotes osteoclast differentiation through Tgif2 by acting as a ceRNA of miR-590-3p in a murine model. *Regen. Med.* 16, 643–653. doi:10.2217/rme-2020-0174

Shen, J. J., Zhang, C. H., Chen, Z. W., Wang, Z. X., Yang, D. C., Zhang, F. L., et al. (2019). LncRNA HOTAIR inhibited osteogenic differentiation of BMSCs by regulating Wnt/β-catenin pathway. *Eur. Rev. Med. Pharmacol. Sci.* 23, 7232–7246. doi:10.26355/eurrev 201909 18826

Shui, X., Chen, S., Lin, J., Kong, J., Zhou, C., and Wu, J. (2019). Knockdown of lncRNA NEAT1 inhibits Th17/CD4(+) T cell differentiation through reducing the STAT3 protein level. *J. Cell. Physiol.* 234, 22477–22484. doi:10.1002/jcp.28811

Simion, V., Haemmig, S., and Feinberg, M. W. (2019). LncRNAs in vascular biology and disease. Vasc. Pharmacol. 114, 145–156. doi:10.1016/j.vph.2018.01.003

Smolen, J. S., Landewé, R. B. M., Bijlsma, J. W. J., Burmester, G. R., Dougados, M., Kerschbaumer, A., et al. (2020). EULAR recommendations for the management of rheumatoid arthritis with synthetic and biological disease-modifying antirheumatic drugs: 2019 update. *Ann. Rheum. Dis.* 79, 685–699. doi:10.1136/annrheumdis-2019-216655

Sun, W., Meednu, N., Rosenberg, A., Rangel-Moreno, J., Wang, V., Glanzman, J., et al. (2018). B cells inhibit bone formation in rheumatoid arthritis by suppressing osteoblast differentiation. *Nat. Commun.* 9, 5127. doi:10.1038/s41467-018-07626-8

Tan, J., Dan, J., and Liu, Y. (2021). Clinical efficacy of methotrexate combined with iguratimod on patients with rheumatoid arthritis and its influence on the expression levels of HOTAIR in serum. *Biomed. Res. Int.* 2021, 2486617. doi:10. 1155/2021/2486617

Tanaka, Y. (2019). Clinical immunity in bone and joints. J. Bone Min. Metab. 37, 2–8. doi:10.1007/s00774-018-0965-5

Tang, M., Lu, L., and Yu, X. (2020). Interleukin-17A interweaves the skeletal and immune systems. *Front. Immunol.* 11, 625034. doi:10.3389/fimmu.2020.625034

Tang, Y., Zhu, J., Huang, D., Hu, X., Cai, Y., Song, X., et al. (2019). Mandibular osteotomy-induced hypoxia enhances osteoclast activation and acid secretion by increasing glycolysis. *J. Cell. Physiol.* 234, 11165–11175. doi:10.1002/jcp.27765

Tang, Z., Gong, Z., and Sun, X. (2018). LncRNA DANCR involved osteolysis after total hip arthroplasty by regulating FOXO1 expression to inhibit osteoblast differentiation. *J. Biomed. Sci.* 25, 4. doi:10.1186/s12929-018-0406-8

Tateiwa, D., Yoshikawa, H., and Kaito, T.(2019).Cartilage and bone destruction in arthritis: Pathogenesis and treatment strategy: A literature review. Cells 8. doi:10. 3390/cells8080818

Teo, B. H., Bobryshev, Y. V., Teh, B. K., Wong, S. H., and Lu, J. (2012). Complement C1q production by osteoclasts and its regulation of osteoclast development. *Biochem. J.* 447, 229–237. doi:10.1042/bj20120888 Tian, Y., Shao, Q., Gu, J., Tang, Y., Bie, M., Zhou, Y., et al. (2022). LncRNAmRNA expression profiles of osteoclast after conditional knockout HIF-1a. *Front. Genet.* 13, 909095. doi:10.3389/fgene.2022.909095

Tseng, C. C., Chen, Y. J., Chang, W. A., Tsai, W. C., Ou, T. T., Wu, C. C., et al. (2020). Dual role of chondrocytes in rheumatoid arthritis: The chicken and the egg. *Int. J. Mol. Sci.* 21, E1071. doi:10.3390/ijms21031071

Wang, J., Shen, C., Li, R., Wang, C., Xiao, Y., Kuang, Y., et al. (2021). Increased long noncoding RNA LINK-A contributes to rheumatoid synovial inflammation and aggression. *JCI Insight* 6, e146757. doi:10.1172/jci.insight.146757

Wang, Q., Chu, P., Yu, X., Li, J., Zhang, W., and Gong, M. (2021). ZFAS1 knockdown inhibits ,broblast-like synoviocyte proliferation, migration, invasion and inflammation, and promotes apoptosis via miR-3926/FSTL1 in rheumatoid arthritis.,broblast-like synoviocyte proliferation, migration, invasion and inflammation, and promotes apoptosis via miR-3926/FSTL1 in rheumatoid arthritis. *Exp. Ther. Med.* 22, 914. doi:10.3892/etm.2021.10346

Wang, Q., Huang, X., Shao, Y., Liu, Q., Shen, J., Xia, J., et al. (2022). The implication of long non-coding RNA expression profile in rheumatoid arthritis: Correlation with treatment response to tumor necrosis factor inhibitor. *Mod. Rheumatol.*, roab128. doi:10.1093/mr/roab128

Wang, X., He, X., Zhang, C. F., Guo, C. R., Wang, C. Z., and Yuan, C. S. (2017). Anti-arthritic effect of berberine on adjuvant-induced rheumatoid arthritis in rats. *Biomed. Pharmacother.* 89, 887–893. doi:10.1016/j.biopha.2017.02.099

Wang, Y., Hou, L., Yuan, X., Xu, N., Zhao, S., Yang, L., et al. (2020). LncRNA NEAT1 targets fibroblast-like synoviocytes in rheumatoid arthritis via the miR-410-3p/YY1 Axis. *Front. Immunol.* 11, 1975. doi:10.3389/fimmu. 2020.01975

Wang, Y., Zhu, G., Pei, F., and Zhao, Z. (2022). lncRNA SNHG15 as a ceRNA modulates osteoclast differentiation, proliferation, and metastasis by sponging miR-381-3p/NEK2 Axis. J. Immunol. Res. 2022, 8634820. doi:10.1155/2022/8634820

Weyand, C. M., and Goronzy, J. J. (2021). The immunology of rheumatoid arthritis. *Nat. Immunol.* 22, 10–18. doi:10.1038/s41590-020-00816-x

Xiao, J., Wang, R., Zhou, W., Cai, X., and Ye, Z. (2021). LncRNA NEAT1 regulates the proliferation and production of the inflammatory cytokines in rheumatoid arthritis fibroblast-like synoviocytes by targeting miR-204-5p. *Hum. Cell* 34, 372–382. doi:10.1007/s13577-020-00461-4

Xie, S., Li, S., Tian, J., and Li, F. (2020). Iguratimod as a new drug for rheumatoid arthritis: Current landscape. *Front. Pharmacol.* 11, 73. doi:10.3389/fphar.2020. 00073

Yang, P., Qian, F. Y., Zhang, M. F., Xu, A. L., Wang, X., Jiang, B. P., et al. (2019). Th17 cell pathogenicity and plasticity in rheumatoid arthritis. *J. Leukoc. Biol.* 106, 1233–1240. doi:10.1002/jlb.4ru0619-197r

Yang, Z., Lin, S. D., Zhan, F., Liu, Y., and Zhan, Y. W. (2021). LncRNA GAS5 alleviates rheumatoid arthritis through regulating miR-222-3p/ Sirt1 signalling axis. *Autoimmunity* 54, 13–22. doi:10.1080/08916934.2020.1846183 Yang, Z., Liu, X., Zhao, F., Yao, M., Lin, Z., Yang, Z., et al. (2022). Bioactive glass nanoparticles inhibit osteoclast differentiation and osteoporotic bone loss by activating lncRNA NRON expression in the extracellular vesicles derived from bone marrow mesenchymal stem cells. *Biomaterials* 283, 121438. doi:10.1016/j. biomaterials.2022.121438

Yao, Y., Cai, X., Ren, F., Ye, Y., Wang, F., Zheng, C., et al. (2021). The macrophage-osteoclast Axis in osteoimmunity and osteo-related diseases. *Front. Immunol.* 12, 664871. doi:10.3389/fimmu.2021.664871

Yoshitomi, H. (2019). Regulation of immune responses and chronic inflammation by fibroblast-like synoviocytes. *Front. Immunol.* 10, 1395. doi:10. 3389/fimmu.2019.01395

Yu, X., Rong, P. Z., Song, M. S., Shi, Z. W., Feng, G., Chen, X. J., et al. (2021). IncRNA SNHG1 induced by SP1 regulates bone remodeling and angiogenesis via sponging miR-181c-5p and modulating SFRP1/Wnt signaling pathway. *Mol. Med.* 27, 141. doi:10.1186/s10020-021-00392-2

Zhang, D. W., Wang, H. G., Zhang, K. B., Guo, Y. Q., Yang, L. J., and Lv, H. (2022). LncRNA XIST facilitates S1P-mediated osteoclast differentiation via interacting with FUS. J. Bone Min. Metab. 40, 240–250. doi:10.1007/s00774-021-01294-3

Zhang, W., Li, B., Xia, N., Zhu, L., Zhang, Z., Ren, Z., et al. (2021). lncRNA GAS5 suppresses rheumatoid arthritis by inhibiting miR-361-5p and increasing PDK4. *Biochem. Biophys. Res. Commun.* 583, 7–13. doi:10.1016/j.bbrc.2021.10.040

Zhang, Y., Chen, X. F., Li, J., He, F., Li, X., and Guo, Y. (2020a). lncRNA Neat1 stimulates osteoclastogenesis via sponging miR-7. *J. Bone Min. Res.* 35, 1772–1781. doi:10.1002/jbmr.4039

Zhang, Y., Wang, X., Li, W., Wang, H., Yin, X., Jiang, F., et al. (2020b). Inferences of individual differences in response to tripterysium glycosides across patients with Rheumatoid arthritis using a novel ceRNA regulatory axis. *Clin. Transl. Med.* 10, e185. doi:10.1002/ctm2.185

Zhao, H., Lu, A., and He, X. (2020). Roles of MicroRNAs in bone destruction of rheumatoid arthritis. *Front. Cell Dev. Biol.* 8, 600867. doi:10.3389/fcell.2020.600867

Zhu, J., Yu, W., Wang, Y., Xia, K., Huang, Y., Xu, A., et al. (2019). lncRNAs: function and mechanism in cartilage development, degeneration, and regeneration. *Stem Cell Res. Ther.* 10, 344. doi:10.1186/s13287-019-1458-8

Zhuo, Q., Wei, L., Yin, X., Li, H., Qin, G., Li, S., et al. (2021). LncRNA ZNF667-AS1 alleviates rheumatoid arthritis by sponging miR-523-3p and inactivating the JAK/STAT signalling pathway. *Autoimmunity* 54, 406–414. doi:10.1080/08916934. 2021.1966770

Zou, Y., Shen, C., Shen, T., Wang, J., Zhang, X., Zhang, Q., et al. (2021). LncRNA THRIL is involved in the proliferation, migration, and invasion of rheumatoid fibroblast-like synoviocytes. *Ann. Transl. Med.* 9, 1368. doi:10. 21037/atm-21-1362