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Novel IKZF3 transcriptomic signature correlates with positive outcomes of skin cutaneous melanoma: A pan-cancer analysis

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To investigate the potential relationship between Ikaros family genes and skin cutaneous melanoma (SKCM), we undertook a pan-cancer analysis of the transcriptional signature and clinical data of melanoma through multiple databases. First, 10,327 transcriptomic samples from different cancers were included to determine the overall characteristics and clinical prognoses associated with Ikaros gene expression across cancer types. Second, differentially expressed genes analysis, prognostic evaluation, and gene set enrichment analysis were employed to investigate the role of Ikaros (*IKZF*) genes in SKCM. Third, we evaluated the relationship between Ikaros family genes and SKCM immune infiltrates and verified the findings using the GEO single-cell sequencing dataset. The results show that Ikaros genes were widely expressed among different cancer types with independently similar patterns as follows: 1. *IKZF1* and *IKZF3*, and 2. *IKZF2* and *IKZF4–5*. *IKZF2* and *IKZF5* were downregulated in the primary tumor, and *IKZF1–3* expression decreased significantly as the T-stage or metastasis increased in SKCM. Moreover, high *IKZF1–3* expression was associated with better overall survival, disease-specific survival, and progression-free interval. *IKZF3* is an independent prognostic factor of SKCM. Among Ikaros genes, the expression of *IKZF1* and *IKZF3* positively correlated with the infiltration level of CD4⁺ T cells and CD8⁺ T cells, B cells, and Tregs in SKCM and negatively correlated with the infiltration level of M0 and M1 macrophages. Moreover, single-cell sequencing data analysis revealed that *IKZF1* and *IKZF3* were mainly expressed by immune cells. Correlation analysis shows the immune factors and drug responses associated with *IKZF3* expression. In conclusion, the present study is the first, to our knowledge, to identify a pan-cancer genomic signature of the Ikaros gene family among different cancers. Expression of these family members, particularly high levels of *IKZF3*, indicate positive immunological status and beneficial clinical outcomes of SKCM. *IKZF3* may therefore serve as potential targets for immunotherapy of melanoma.

KEYWORDS

ikaros family genes, SKCM, immune infiltration, immunotherapy biomarker, pan-cancer analysis

Introduction

As one of the most serious cancers, SKCM easily metastasizes to other parts of body through the lymphatic and peripheral circulatory systems. In recent years, SKCM morbidity tends to affect more of the younger population and has become one of the tumors with the fastest increase in incidence across cancer types (Ferlay et al., 2015; Siegel et al., 2016). Early-stage patients with SKCM do not exhibit apparent symptoms or classical characteristics in pathological examinations, which hinders physicians from diagnosing and treating patients in a timely manner (Megahed et al., 2002).

For example, conventional surgical resection of local lesions and radiation therapy hardly achieve significantly positive outcomes of SKCM, particularly for multiorgan metastasis (Balch et al., 2001). In contrast, the antiapoptotic activity of SKCM renders it resistant to most chemotherapies and increases the risk of its recurrence (Soengas and Lowe, 2003; Nazarian et al., 2010). Although SKCM is currently treated with a combination of surgery, radiation, and other systematic modalities, the survival rate of patients with advanced melanoma metastasis remains extremely low.

However, with constant improvements in immunomodulatory technology, cancer immunotherapy appears to serve as an effective method to treat malignant tumors. Mounting studies highlight the role of tumor-infiltrating lymphocytes in eliminating tumors from the tumor microenvironment (TME), thus making it reasonable to treat SKCM with an immunotherapy strategy.

In tumor tissues, fibroblasts, infiltrating lymphocytes, and components such as the surrounding stromal cells and capillaries constitute the TME (Locy et al., 2018). As the “soil” for tumor cell growth, the TME mainly provides nutrients for tumor cell proliferation and invasion. However, once the dysfunctional infiltrated immune cells recover, the tumor microenvironment may become a battlefield to kill tumors. In recent years, mounting studies reveal that quiescent tumor-infiltrating lymphocytes (TILs) are the main forces that kill tumor cells after they respond to effective immunological stimulation *via* the TME (Lee et al., 2016; Fu et al., 2019). In SKCM, effector T cells, a member of the TIL population, continuously eradicate tumor cells once activated by antigens presented by other cells, thus improving the prognostic outcome of patients (Zhou et al., 2005).

Methods for transforming the TME from a quiescent to active immunological state range from intravenous injection of recombinant interleukin (Atkins et al., 2000) and adoptive T-cell therapy (Besser et al., 2010) to monoclonal antibody therapy (Weber, 2010). By blocking the PD-1/PD-L1-induced signaling pathway or activating the costimulatory receptor 4-1BB/4-1BBL,

monoclonal antibody therapy of tumors may improve clinical outcomes. However, most patients with SKCM do not positively respond to single-target therapy, particularly those with metastasized melanoma (Tsai et al., 2014). Therefore, it is vitally importance to investigate effective immunological therapies that target SKCM.

The Ikaros family of zinc-finger proteins plays a critical role in lymphatic development, differentiation, and homeostasis. There are five homologous members of the Ikaros gene family including Ikaros (*IKZF1*), Helios (*IKZF2*), Aiolos (*IKZF3*), Eos (*IKZF4*), and Pegasus (*IKZF5*), which are translated as distinct isoforms through alternative splicing (John and Ward, 2011). The Ikaros zinc finger-like N-terminal domain specifically binds DNA, and its zinc finger-like C-terminal domain mediates homologous binding as well as to other family members (Molnár et al., 1994; Sun et al., 1996). These features confer upon Ikaros family members the ability to regulate cell proliferation, differentiation, and apoptosis through the combination of Ikaros family members or other transcription factors (Yoshida and Georgopoulos, 2014).

IKZF4 and *IKZF5* are mainly expressed in skeletal muscle and other solid organs, and *IKZF1–3* expressed by lymphocytes participate in the regulation of the development and differentiation of these cells (Perdomo et al., 2000; Yoshida and Georgopoulos, 2014). It is therefore reasonable to conclude that Ikaros genes influence the immunological status in the TME to affect patients’ clinical outcomes. However, the potential relationship between TILs that express Ikaros genes in the TME and SKCM has not been comprehensively investigated.

In the present study, we first applied bioinformatics to analyze RNA-seq data for different cancers to determine the role of Ikaros genes in their pathogenesis. The potential function of Ikaros genes in SKCM was confirmed using gene set enrichment analysis (GSEA), survival evaluation, and other analytical tools. To determine the effects of Ikaros gene expression on the TIL population of SKCM, we performed immunological analyses using CIBERSORT and single-cell RNA-seq. Finally, a novel *IKZF1–3* transcriptomic signature was shown to correlate with positive outcomes of SKCM.

Methods

Data analysis

RNA sequence data and corresponding clinical data of patients with different carcinomas (33 types) were acquired

from the UCSC Xena database (<https://xenabrowser.net/datapages/>). The Ensemble IDs of the expression profile data were converted to symbol IDs through the human GTF file. Cancer types were included: ACC, BLCA, BRCA, COAD, DLBC, ESCA, GBM, HNSC, KICH, KIRC, KIRP, LAML, LGG, LIHC, LUAD, LUSC, OV, PAAD, PRAD, READ, SKCM, STAD, TGCT, THCA, THYM, UCEC, and UCS. At the same time, clinical data were included survival time, progression free survival time, survival status, clinical staging and other clinical follow-up data.

Ikaros transcriptional signature across cancer types

We determined the expression levels of *Ikaros* family members in all cancers and identified the individual expression characteristics of *Ikaros* family members through Pearson coefficient correlation analysis. In view of the lack of normal samples of SKCM in TCGA data and to evaluate the association between *Ikaros* and the progression of SKCM, differential expression levels of normal and tumor samples were retrieved and compared using the GEPIA database (<http://gepia.cancer-pku.cn>). Furthermore, we selected cancer types with more than three normal data samples and analyzed the differences between the expression of *Ikaros* family genes in cancer and normal tissues. $|\log_{2}FC| > 2$ and $p < 0.05$ were considered to indicate significant differences in expression.

Prognostic analysis

The prognostic outcomes of *Ikaros* expression among multiple cancers were determined using Cox univariate analysis with the indexes of hazard ratios (HRs) and 95% confidence intervals (CIs). The log-rank test and the Kaplan–Meier algorithm were used to determine the correlation between expression of individual *Ikaros* genes and a patient's survival. Several indicators of outcomes, such as overall survival (OS), progression-free interval (PFI), and disease-free survival (DSS) were applied to comprehensively evaluate the prognostic significance of *Ikaros* gene expression.

GSEA and protein-protein interaction network analysis

A computational method of GSEA (GSEA v.3.0 in the Java environment) was used to explore the potential mechanism of the regulation of *Ikaros* genes in the occurrence and development of SKCM. We ranked the levels of *Ikaros* genes expressed by SKCM samples from high to low for GSEA, which was used to identify signaling pathways correlated with *Ikaros*. Gene-set permutations were

performed 1,000 times for each analysis. The expression levels of *IKZF1–IKZF5* were used as phenotypic labels for all SKCM samples. A false discovery rate (FDR) < 0.25 and $p < 0.05$ were used to identify KEGG pathways significantly enriched in each phenotype. *Ikaros*-interacting proteins were determined using the GeneMANIA protein-protein interaction network (<http://genemania.org/>).

Analysis of immune infiltration

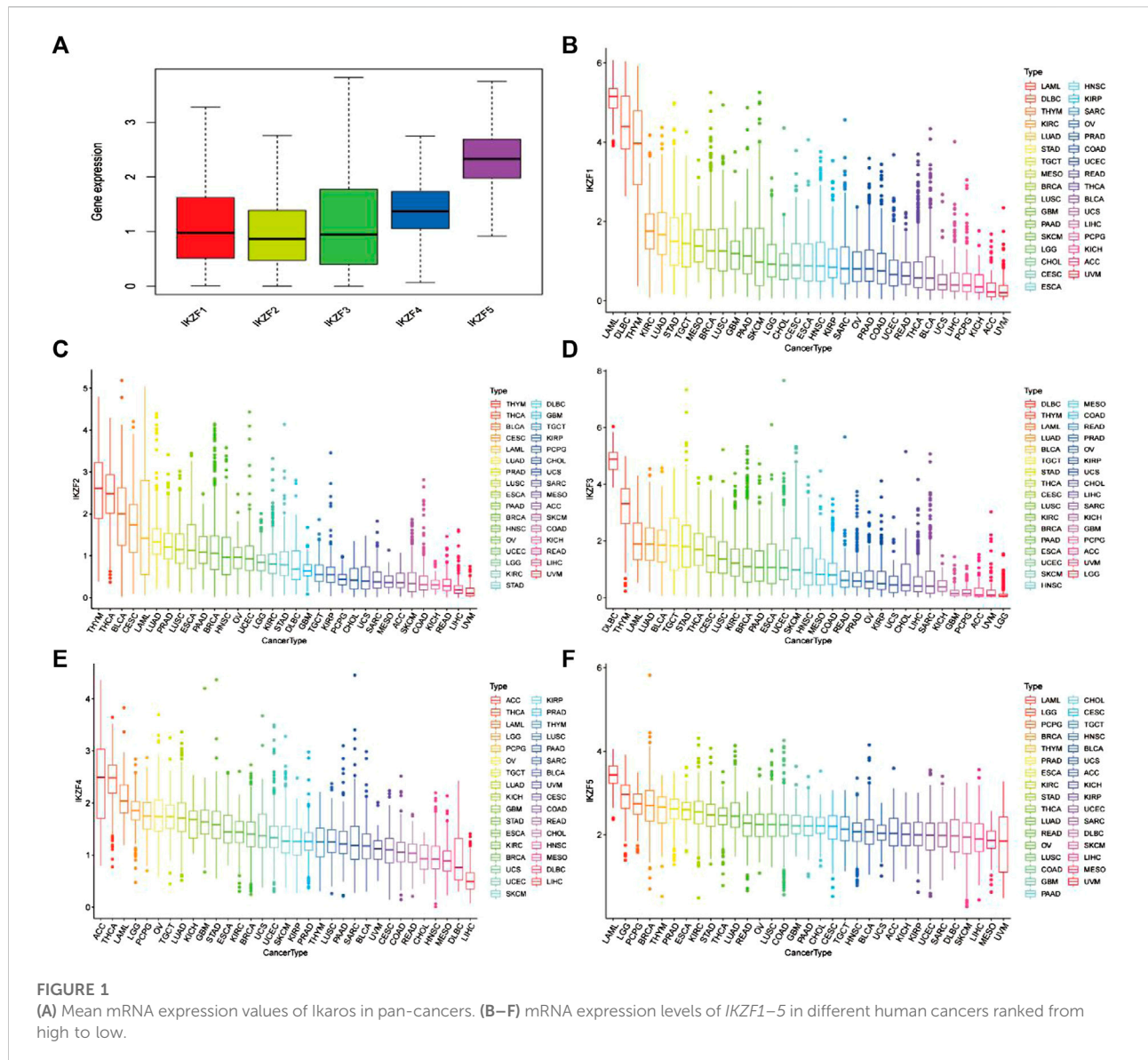
An algorithm of ESTIMATE (Yoshihara et al., 2013) was adopted to evaluate immune infiltration condition across cancer types using the indexes of stromal score, immune score, and tumor purity. The relationship between *Ikaros* gene expression and immunological reactivity was determined by comparing the scores and purity indexes among different carcinomas. CIBERSORT was used for characterizing the infiltrating immune cell composition of SKCM tissue from their gene expression profiles. We performed deconvolution calculations with the annotating file LM22. CIBERSORT annotates the abundance of 22 infiltrating immune cells through a 547-gene expression eigenmatrix (Newman et al., 2015). Through the results of CIBERSORT, the correlations between immune-infiltrating cells in the TME of SKCM and the expression of *Ikaros* genes were demonstrated.

Single-cell RNA-seq analysis

The Tumor Immune-single Cell Hub (TISCH) is an scRNA-Seq database focused on the TME, which provides detailed cellular annotations at the level of a single cell that enables exploration of the TMEs of different cancer types (Sun et al., 2021). GSE72056, a series of 4,645 single-cell scRNA-seq data from 19 melanoma samples included in the Gene Expression Omnibus by Tirosh et al. was further analyzed using TISCH to determine *Ikaros* gene expression levels in immunocytes.

Correlation of *IKZF3* expression and immune factors

The TISIDB database (<http://cis.hku.hk/TISIDB>) is a web portal for tumour and immune system interaction, which integrates multiple heterogeneous data types. Gene signatures of Tumor-infiltrating lymphocytes (TILs) were obtained from TISIDB database. Here, we further analyzed associations for *IKZF3* with immunomodulators, chemokines and receptors. Immunomodulators can be divided into three categories, including immunoinhibitors, immunostimulators and major histocompatibility complex (MHC) molecules.



Correlation of *IKZF3* expression and drug responses

The cellminer database contains (NCI)-60 gene expression and drug sensitivity data. We obtained the cellminer database including drug data approved by the US Food and Drug Administration and drug data evaluated in clinical trials. The Pearson correlation coefficient was used to evaluate the correlation between *IKZF3* expression and drug response.

Statistical analysis

The Wilcoxon rank-sum test and the Kruskal–Wallis test were used to compare two and multiple groups, respectively.

Pearson analysis was used to perform correlation analysis among members of the Ikaros gene family. $p < 0.05$ indicates a significant difference. All statistical analyses were performed using R (version 3.6.0).

Results

Transcriptomic signatures of ikaros genes across cancer types

Differential transcriptomic levels of Ikaros members (*IKZF1–5*) were determined through the analysis of 11,057 samples of 33 tumors (10,327 tumor and 730 normal samples). To determine the expression

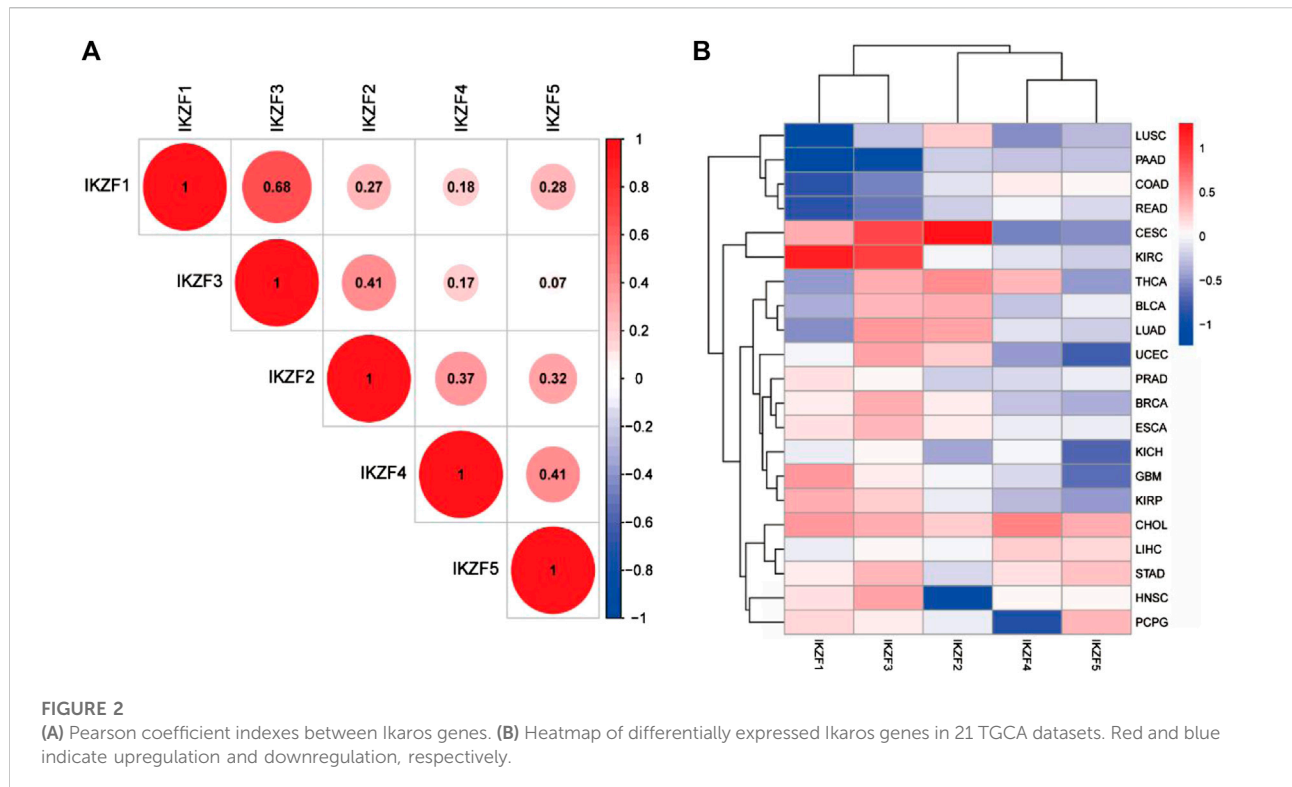


FIGURE 2 (A) Pearson coefficient indexes between Ikaros genes. (B) Heatmap of differentially expressed Ikaros genes in 21 TCGA datasets. Red and blue indicate upregulation and downregulation, respectively.

characteristics of Ikaros members, we first calculated the mean expression levels of *IKZF1–5* in pan-cancers. Among them, the mean expression levels of *IKZF1–3* were relatively consistent and that of *IKZF5* was the highest (Figure 1A). Furthermore, the transcriptional signatures of individual Ikaros genes in different cancer types revealed that *IKZF1–5* were mainly overexpressed in hematological tumors such as LAML and DLBC, but were expressed at low levels in digestive tumors, for example, LIHC. The detailed expression levels of Ikaros genes in different cancer types are shown in Figures 1B–F.

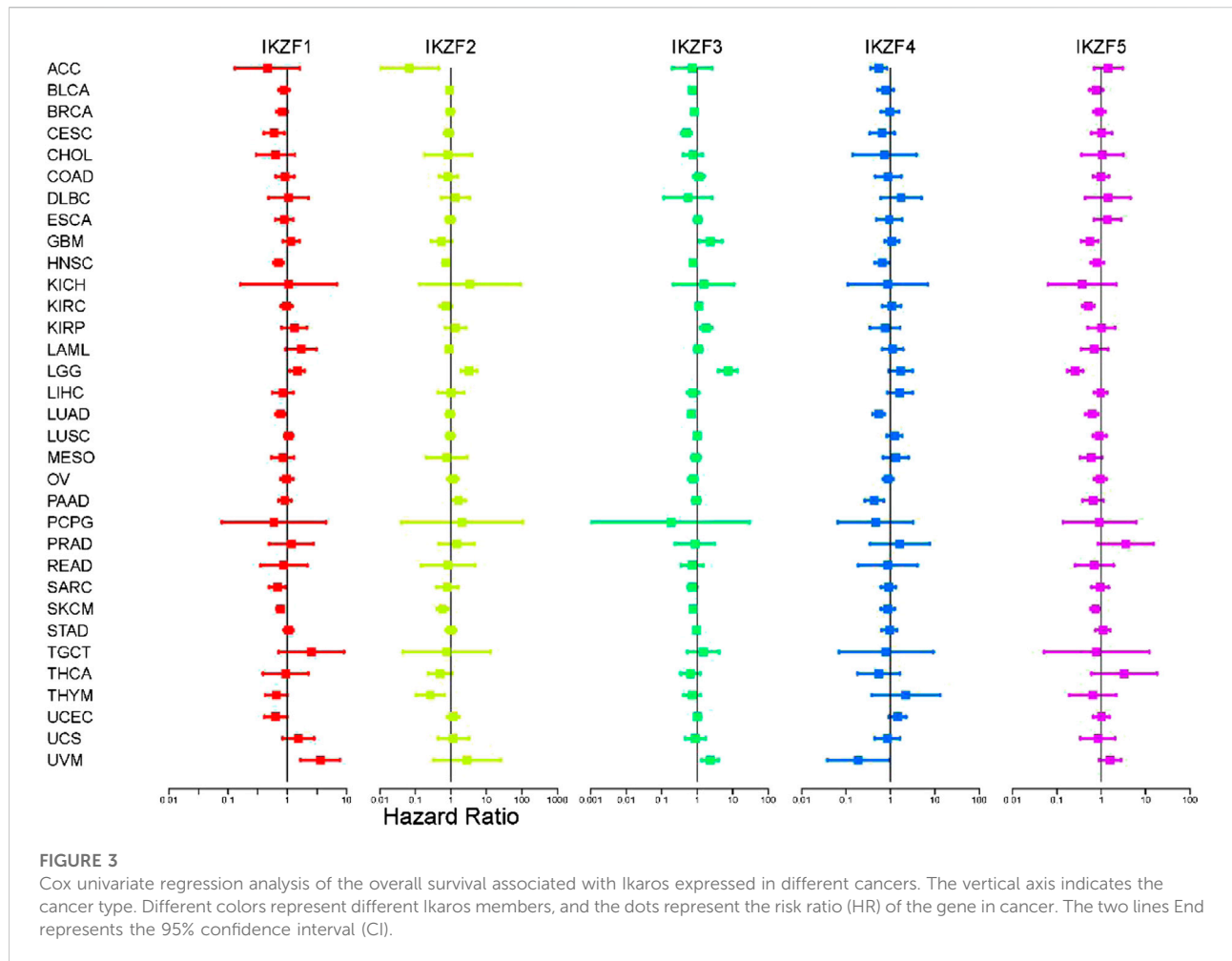
The Pearson coefficient index of every pair of Ikaros genes indicated that *IKZF1* and *IKZF3* were closely associated in various cancers, while *IKZF3* and *IKZF5* showed the weakest correlations (Figure 2A). We next screened 21 tumor datasets, each including at least three normal samples, using hierarchical cluster analysis of differentially expressed genes. The results confirmed the transcriptional relationship between *IKZF1* and *IKZF3*, because they ranked in the same cluster with a transcriptional pattern similar to that of *IKZF1* and *IKZF3* in different cancer types (Figure 2B).

To further investigate the role of *IKZF1–5* in predicting clinical outcomes, we conducted univariate Cox regression analysis of gene samples with the corresponding clinical data. We found that most patients with cancer that highly expressed *IKZF1–3* had a significantly improved prognosis. For example, high levels of *IKZF1–3* in patients with CESC,

HNSC, LUAD, SARC, and SKCM indicated longer survival but implied poor prognosis of patients with LGG and UVM with the same transcriptional profile (Figure 3). Although there was no significant association between *IKZF4* and *IKZF5*, their high expression levels indicate better outcomes of certain types of cancers. The high expression of *IKZF1*, *IKZF2*, *IKZF3*, and *IKZF5* in SKCM will indicate a better prognosis.

IKZF1–3 expression negatively correlates with the progression of SKCM

Compared with normal tissue, *IKZF2* and *IKZF5* mRNAs were expressed at lower levels in SKCM (Figure 4A). Considering that Ikaros genes may participate in the progression of SKCM, we investigated their transcriptional levels associated with different TNM stages. The results show that the expression of *IKZF1–3* mRNAs in SKCM significantly decreased with higher T-stage and significantly decreased when SKCM involved local lymph node metastasis that penetrated the original tissue boundary (Figure 4B). However, the levels of mRNAs of other members of the Ikaros family did not vary with higher T-stage. Moreover, among different tumor locations, Ikaros genes were expressed at the highest levels in regional lymph nodes, *IKZF1* and *IKZF3* were expressed at the lowest levels in distant



metastases, and *IKZF2*, *IKZF4*, and *IKZF5* were expressed at the lowest levels in the primary tumor.

IKZF3 can be used as an independent prognostic factor (Supplementary Figure S1).

Highly expressed *IKZF1–3* are associated with favorable clinical outcomes of patients with SKCM

The results of Kaplan-Meier analysis suggested that patients with SKCM with highly expressed *IKZF1–3* experienced significantly longer OS, DFS, and PFI than patients with lower levels ($p < 0.05$) (Figures 5A–C). However, the associations of the OS, DSS, and PFI rates of patients with higher expression levels of *IKZF4–5* were not significantly different (Figures 5D,E), and patients who expressed high levels of *IKZF1–3* experienced significantly longer DSS and longer tumor progression-free survival. These findings indicate that high expression of *IKZF1–3* mRNAs was significantly associated with good prognosis of patients with SKCM. Further multivariate-cox analysis indicates that

High expression of *IKZF1–3* activates an antitumor immune response

To further investigate Ikaros-associated signaling pathways, transcriptional data of SKCM samples were ranked by the relative levels of *IKZF1–5* expression in the upper 10th percentile (*IKZFhi*) vs. the lower 10th percentile (*IKZFlo*) and subjected to GSEA. Figures 6A–C shows that high expression of *IKZF1–3* was associated with several activated KEGG pathways, while *IKZF4* and *IKZF5* were not clustered in known pathways (data not shown). Moreover, the GSEA results demonstrate that high expression of *IKZF1–3* plays a critical role in the regulation of the activation of the immune response through the B-cell receptor signaling pathway, the T-cell receptor signaling pathway, leukocyte transendothelial migration, and natural killer cell-mediated cytotoxicity. Furthermore, *IKZF1-3hi* was closely

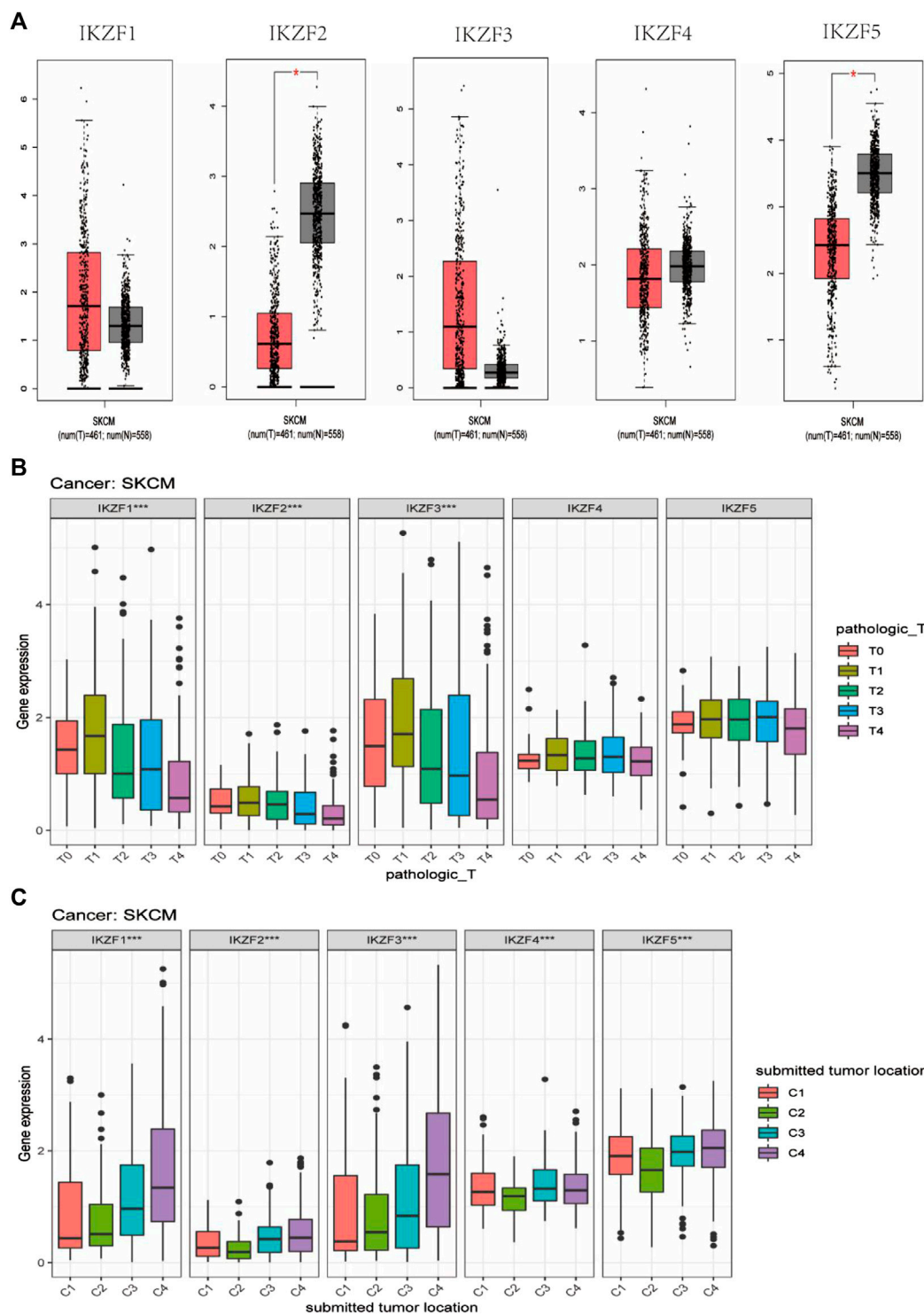


FIGURE 4

(A) GEPIA box diagram shows the expression levels of Ikaros genes in SKCM and healthy skin. T: tumor tissue, N: normal tissue; (B) Ikaros gene expression in patients with different pathological T-stages; (C) Ikaros gene expression at different tumor locations, C1 (Distant Metastasis), C2 (Primary Tumor), C3 (Regional Cutaneous or Subcutaneous Tissue [includes satellite and in-transit metastasis]), C4 (Regional Lymph Node) (* $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$).

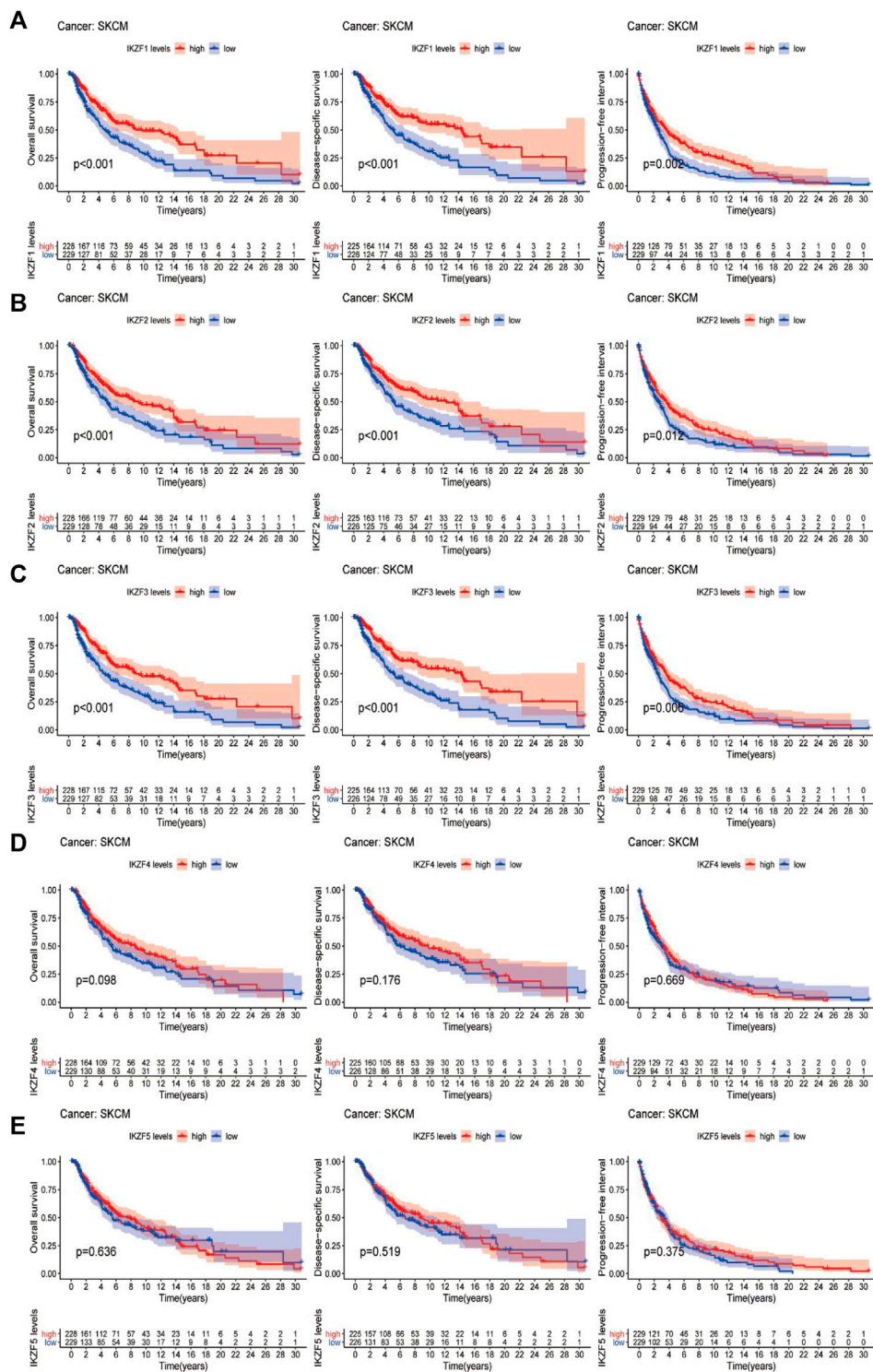


FIGURE 5 Comparison of Kaplan–Meier survival curves of patients with high or low expression of Ikaros. OS, DSS, and PFI rates associated with the expression of Ikaros genes in the SKCM cohort. (A–E), *IKZF1–5*, respectively.

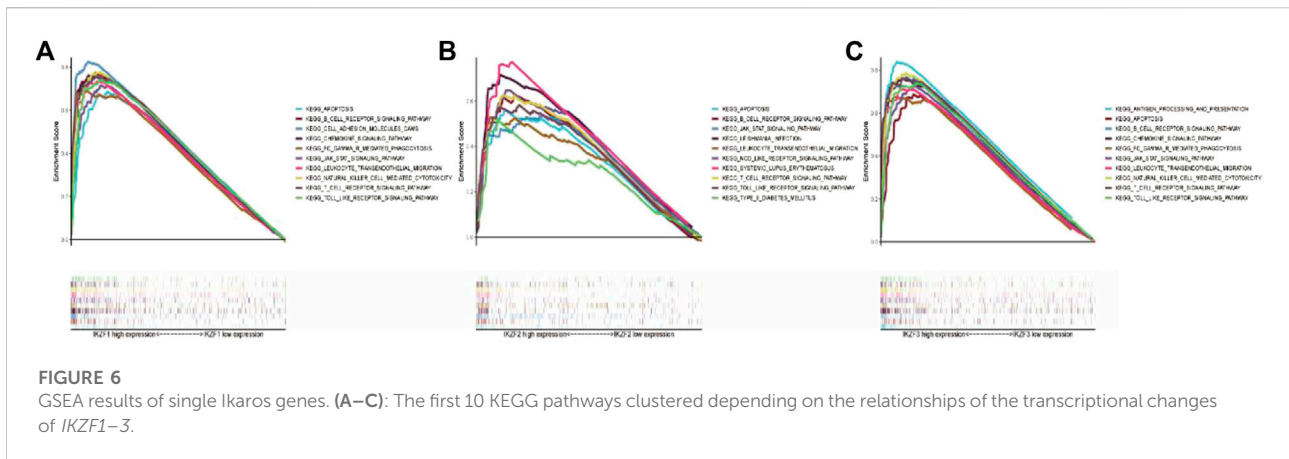


FIGURE 6
GSEA results of single Ikaros genes. (A–C): The first 10 KEGG pathways clustered depending on the relationships of the transcriptional changes of *IKZF1–3*.

associated with apoptosis and chemokine signaling pathways in the SKCM cohort, which indicates the anticancer effect of *IKZF1–3* through its regulation of the immune response.

Increased *IKZF1–3* expression enhances immunological infiltration of SKCM

Tumor infiltration analysis using CIBESORT and Estimate, shows that the stromal and immune scores significantly increased as the expression of *IKZF1–3* was enhanced in SKCM samples. Moreover, there was a positive correlation between increased *IKZF1–3* expression and the abundance of immunocytes such as B cells, CD4-positive T cells and CD8-positive T cells, although there was a negative correlation between increased *IKZF1–3* expression and tumor infiltration with M0/M2-polarized macrophages. Furthermore, increased infiltration of M0/M2 macrophages was a poor prognostic factor for patients. (Supplementary Figure S2). However, there was no significant relationship between tumor infiltration and expression of *IKZF4–5*.

Single-cell sequence analysis of Ikaros expression

Single-cell sequence analysis was applied to specifically determine the transcriptomic signature of Ikaros members. For this purpose, we employed melanoma tissue of the GSE72056 dataset. Using the Louvain clustering algorithm and a KNN graph, melanoma tissue was divided into the major cell types as follows: B cells, CD4-positive T cells, CD8-positive exhausted T cells, endothelial cells, fibroblasts, SKCM cells, monocytes, follicular helper T cells, Th1 helper cells, and proliferating T cells (8A, G). Consistent with the results mentioned above, the levels of *IKZF1* and *IKZF3* were highest in immunocytes and lowest in SKCM cells, which further

indicates the function of *IKZF1* and *IKZF3* in the immune response to tumor cells and indicates the status of immunocyte infiltration in the TME (Figures 7B,D). However, *IKZF2*, *IKZF4*, and *IKZF5* were basically not detectably expressed by immunocytes, malignant cells, and stromal cells (Figures 7C,E,F).

Correlation of *IKZF3* expression and immune factors

Relationship of *IKZF3* expression with immune factors was evaluated using TISIDB and TCGA databases. we calculated the correlation of *IKZF3* expression and Gene signatures of TILs, including three immunomodulators, chemokines and receptors (Figures 8A–E). Figure 8A shows correlations between *IKZF3* and immunostimulators, *IKZF3* strongly correlated with CD27, TNFRSF9 and ICOS, and weakly correlated with TNFSF9 and ULBP1. Figure 8B shows correlations between *IKZF3* and Immunoinhibitors, *IKZF3* strongly correlated with CD96, PDCD1 and TIGIT, and weakly correlated with KDR and VTCN1. Figure 8C shows correlations between *IKZF3* and chemokines, *IKZF3* strongly correlated with CXCL13, XCL2 and CCL4, and weakly correlated with CXCL1 and CCL27. Figure 8D shows correlations between *IKZF3* and receptors, *IKZF3* strongly correlated with CXCR3, CCR5 and CCR4, and weakly correlated with CXCR1 and CXCR2. Figure 8E shows *IKZF3* expression moderately to strongly correlated with MHC.

Correlation of *IKZF3* expression and drug responses

In order to understand the potential relationship between *IKZF3* expression and various types of drug response, we performed a correlation analysis to use CellMiner to identify

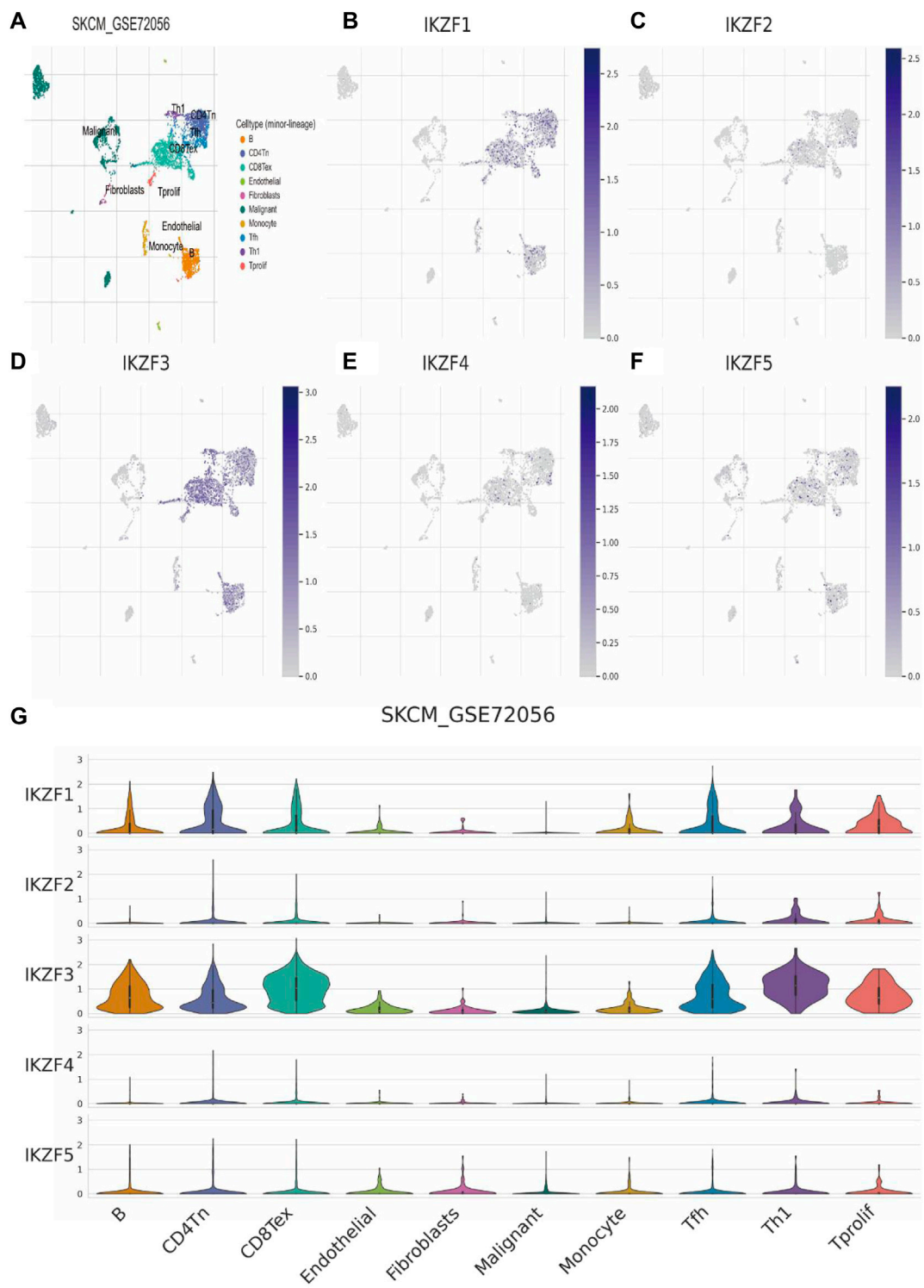


FIGURE 7

Transcriptomic signatures of different cell types in the melanoma microenvironment. **(A)**: There were 10 major cell types in the SKCM microenvironment as follows: B cells, CD4-positive T cells, CD8-positive T cells, endothelial cells, fibroblasts, malignant melanoma cells, monocytes, follicular helper T cells, Th1 helper cells, and proliferating T cells. **(B–E)**: Expression levels of *IKZF1–5* among different cell types in the melanoma.

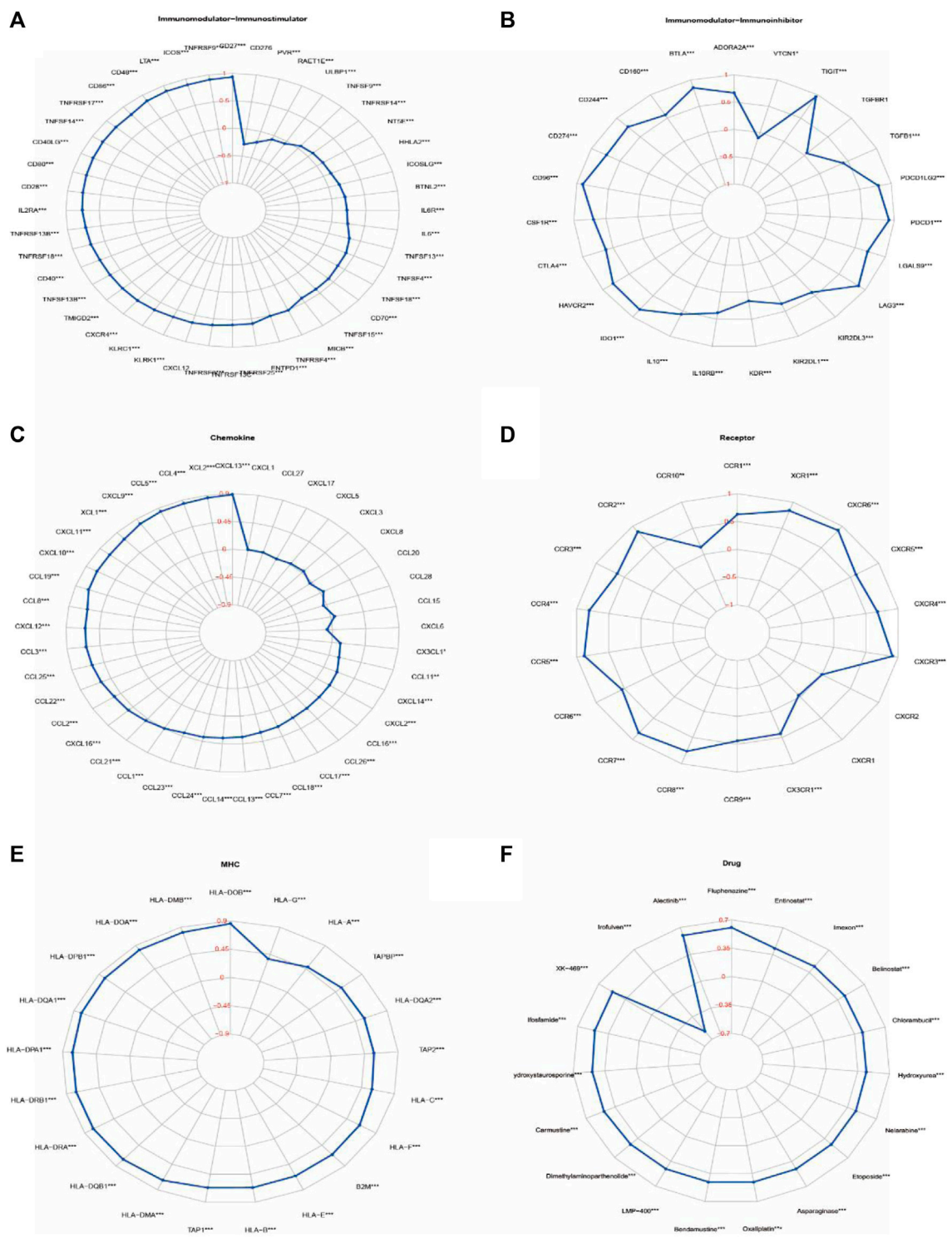


FIGURE 8 Correlations of *IKZF3* expression with immunomodulatory genes, drugs. (A–E) Correlation between immunomodulators and chemokines (or receptors) with *IKZF3* expression. (F) Correlation between *IKZF3* expression and drug responses. The red numbers of radar chart represent Spearman's correlation coefficient (* $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$).

Cancer: SKCM

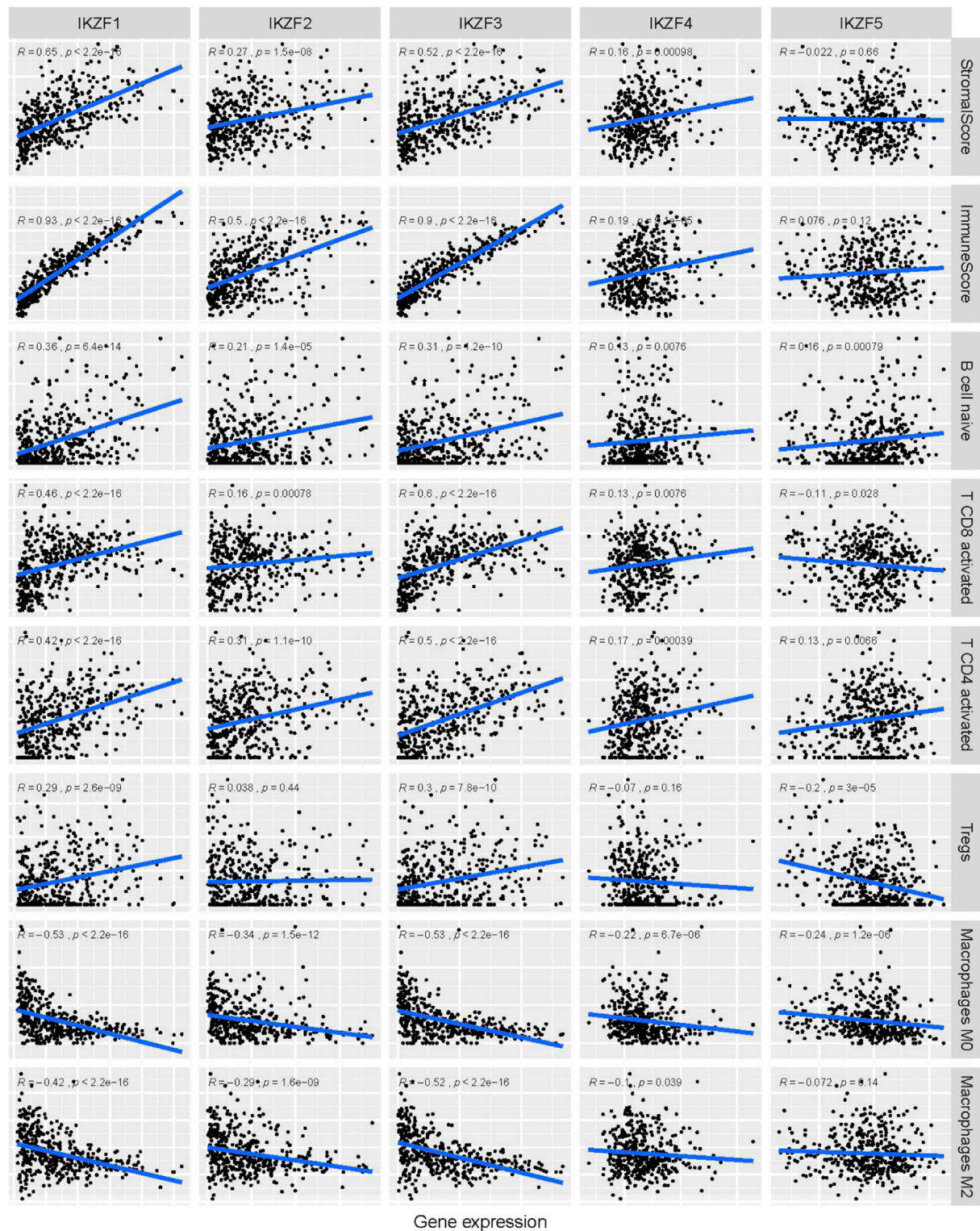


FIGURE 9

Correlation analysis of Ikaros family genes and the abundance of immune-infiltrating cells in the tumor microenvironment. Horizontal labels represent different levels of Ikaros gene expression, vertical labels represent different infiltrating immune cells, and the line represents the correlation between them. The correlation value is represented by the index R.

potential drug candidates. IKZF3 expression in tumor cells was positively associated with increased drug sensitivity to Fluphenazine, Alectinib, and negatively associated with Irofulven. We have screened a total of 20 drugs that are most related to the expression of IKZF3, which are also shown in the radar chart (Figure 8F).

Discussion

With an extremely low survival rate and high potential for metastatic growth, SKCM causes heavy economic losses and medical burdens annually worldwide. Except for conventional surgical removal of the carcinoma and regular radiation therapy, immunotherapy is one of the most promising treatments for SKCM (Besser et al., 2010; Hodi et al., 2010).

The recent research have determined the effect of immunotherapy in the treatment of tumors. As the development of cancer is highly associated with immune micro-environment, determination of the pivotal role of immune response in the tumor is of importance in developing new immunotherapeutic strategies (Roma-Rodrigues et al., 2019). IMiD (Immunomodulatory drugs) covers thalidomide, Len and pomalidomide, and is clinically approved for the treatment of MM (multiple myeloma) and other malignant tumors. Among them, MM and MDS (myelodysplastic syndrome) are widely used and studied. IMiD can achieve the therapeutic effect by inducing selective ubiquitination and proteasome degradation of Ikaros (Gao et al., 2020). In addition, IMiD has the ability to directly inhibit the growth of tumor cells and strong immune stimulation characteristics, so it has multiple effects on the existence of different cell components in the tumor microenvironment. The efficacy and safety of IMiD have been verified in a wide range of clinical trials. It is increasingly clear that IMiDs are promising in the treatment of MM (Gao et al., 2020), MDS and CLL (chronic lymphocytic leukemia) (Vitale et al., 2016). At the same time, Ikaros can be used as prognostic factors for MM patients treated by Len (Tachita et al., 2020). Previous Research has shown that patients with lower IKZF3 expression level have a poorer therapeutic effect with Len, which will also lead to shorter progression-free survival and overall survival (Pourabdollah et al., 2016). Moreover, inhibiting RUNX to protect Ikaros from degradation can significantly improve the drug resistance of IMiDs in MM (Zhou et al., 2019). In addition, the combined use of imatinib in children with B-ALL can enhance the therapeutic effect of IKZF1-deletion patients. Imatinib, as an intensive treatment for B-ALL in IKZF1- deletion children, significantly reduces the risk of recurrence and improves the 5-year overall survival of patients (Yeoh et al., 2018). CX4945, an inhibitor of CK2, can restore Ikaros function and play an anti-leukemia role *in vitro* or preclinical leukemia model (Borgo et al., 2021). In breast cancer, the application of ginseng polysaccharides can inhibit the proliferation of MDA-MB-231 cells by activating IKZF1 (Zhou et al., 2020). Through the inhibition of the CTLA4 *via*

ipilimumab and the application of adoptive cellular immunotherapy with tumor-infiltrated lymphocytes, it enhanced the anti-tumor effects of T cells (Besser et al., 2010; Hodi et al., 2010).

The previous studies have revealed that SKCM patients with a high abundance of T and B cells or lowly infiltrated M0 and M2 macrophages have a significant prognosis, while the metastatic SKCM patients with low infiltration of B and CD8⁺ T cells have a worse outcome than the previous condition (Iglesia et al., 2016; Wang et al., 2020). The present study suggests that expressed *IKZF1-3* could activate the B cell receptor signaling pathway, T cell receptor signaling pathway, leukocyte trans-endothelial migration, antigen processing and presentation and several other immune regulation processes. Obviously, *IKZF1-3* is positively correlated with T and B cells infiltration, but negatively correlated with the M0 and M2 infiltration. Moreover, Ikaros is specifically expressed in the immunocytes. It seems that *IKZF1-3* participate the regulation of immunocytes' differentiation, thus making it could be considered as an indicator for the status of TME in SKCM patients. Besides the regulation of hematopoietic process, Ikaros also regulates the balance of autoimmunity and suppresses the growth of tumor (Fan and Lu, 2016). Ikaros genes mediate lymphocyte proliferation and differentiation (Fan and Lu, 2016). *IKZF1-3* play an important role in the regulation of lymphatic system differentiation. The disorder of its expression has the directive relationship with the hematological malignancies and primary immunodeficiency. Lack of Ikaros family may lead to a variety of immune related diseases, including immune thrombocytopenia (Sriaroon et al., 2019), systemic lupus erythematosus (Chen et al., 2020), rheumatoid arthritis (Yang et al., 2019). For example, mutations of *IKZF1-3* in leukemias are associated with poor prognosis, mainly caused by the disruption of lymphocyte fates (Rebollo and Schmitt, 2003; Payne and Dovat, 2011). In addition, Some solid tumors are related to the abnormal expression of Ikaros family proteins. Higher levels of Ikaros have been proved to be related to poor differentiation and late stage of ovarian cancer (He et al., 2012). Moreover, hypermethylation of Ikaros levels can be regarded as a marker of colorectal cancer progression and poor prognosis (Javierre et al., 2011). As a nuclear transcription factor, IKZF1 could inhibit the proliferation of HCC through suppressing the promotor of ANXA4D (Liu et al., 2017). In lung cancer, overexpressed IKZF3 upregulates the expression of TWIST and matrix metalloproteinase-16, which promotes the epithelial-mesenchymal-transition and the transformation of cancer stem cells and leads to poor prognosis (Hung et al., 2020). Moreover, the highly expressed Ikaros are closely associated with recurrence and metastasis lung adenocarcinoma (Zhao et al., 2020). But, in several tumors that normally lack IKZF1 expression, overexpression of Ikaros leads to enhanced immune recruitment infiltration and tumor sensitivity to CTLA4 and PD1 inhibitors (Chen et al., 2018). However, no research, to our knowledge, has investigated the correlation

between Ikaros expression and SKCM with the aim of developing markers to predict prognosis and tumor progression.

In the present study, we first determined the associations between patients' overall characteristics and clinical prognosis with the expression of Ikaros across cancer types. We employed different analytical methods to identify the relationship between the transcriptional properties of *IKZF1-5* and different cancers (Figures 1–3). For patients with SKCM, the expression of *IKZF1-3* significantly decreased with increased T-stage or metastasis (Figure 4). Moreover, highly expressed *IKZF1-3*, but not *IKZF4-5*, were positively associated with a favorable prognostic outcome (Figure 5) of patients with SKCM through regulating immunocyte infiltration (Figure 9) and the immune response (Figure 6, 7). Together, these results indicate that highly expressed *IKZF1-3* gain clinical significance for predicting the prognosis of patients with SKCM, *IKZF3* is an independent prognostic factor of SKCM, thus suggesting that *IKZF3* are prognostic biomarkers and immunotherapeutic targets of melanoma. To the best of our knowledge, there is no prior study that comprehensively investigated the relationship between Ikaros gene expression and SKCM.

A limitation of the present study is the lack of experimental evidence that activation of *IKZF1-3*-related pathways predict the prognosis of SKCM. Therefore, detailed epigenetic regulation of Ikaros during the pathogenesis of melanoma should be further investigated.

In conclusion, our present study systematically describes the distributions and functions of Ikaros family genes across cancer types. Although the roles of other Ikaros family members such as *IKZF4-5* in melanoma remain unclear, *IKZF3* may serve as biomarkers for the outcomes of treatments for SKCM that achieve a positive clinical prognosis.

Data availability statement

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found in the article/Supplementary Material.

Ethics statement

Written informed consent was obtained from the individual(s) for the publication of any potentially identifiable images or data included in this article.

Author contributions

L-KY: Conception and design, Provision of study material, Collection and/or assembly of data, Data analysis and

interpretation, Manuscript writing. C-XL: Manuscript writing. S-HL: Provision of study material, Collection and/or assembly of data. J-JL: Provision of study material, Collection and/or assembly of data. L-LX: Provision of study material, Collection and/or assembly of data. G-HX: Provision of study material, Collection and/or assembly of data, Data analysis and interpretation. H-WL: Conception and design, Administrative support, Data analysis and interpretation, Final approval of manuscript. XL: Conception and design, Financial support, Administrative support, Data analysis and interpretation, Final approval of manuscript.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Supplementary material

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fgene.2022.1036402/full#supplementary-material>

References

- Atkins, M. B., Kunkel, L., Sznol, M., and Rosenberg, S. A. (2000). High-dose recombinant interleukin-2 therapy in patients with metastatic melanoma: Long-term survival update. *Cancer J. Sci. Am.* 6 (1), S11–S14.
- Balch, C. M., Buzaid, A. C., Soong, S. J., Atkins, M. B., Cascinelli, N., Coit, D. G., et al. (2001). Final version of the American Joint Committee on Cancer staging system for cutaneous melanoma. *J. Clin. Oncol.* 19 (16), 3635–3648. doi:10.1200/JCO.2001.19.16.3635
- Besser, M. J., Shapira-Frommer, R., Treves, A. J., Zippel, D., Itzhaki, O., Hershkovitz, L., et al. (2010). Clinical responses in a phase II study using adoptive transfer of short-term cultured tumor infiltration lymphocytes in metastatic melanoma patients. *Clin. Cancer Res.* 16 (9), 2646–2655. doi:10.1158/1078-0432.CCR-10-0041
- Borgo, C., D'Amore, C., Sarno, S., Salvi, M., and Ruzzene, M. (2021). Protein kinase CK2: A potential therapeutic target for diverse human diseases. *Signal Transduct. Target. Ther.* 6 (1), 183. doi:10.1038/s41392-021-00567-7
- Chen, J. C., Perez-Lorenzo, R., Saenger, Y. M., Drake, C. G., and Christiano, A. M. (2018). IKZF1 enhances immune infiltrate recruitment in solid tumors and susceptibility to immunotherapy. *Cell Syst.* 7 (1), 92–103. doi:10.1016/j.cels.2018.05.020
- Chen, L., Niu, Q., Huang, Z., Yang, B., Wu, Y., and Zhang, J. (2020). IKZF1 polymorphisms are associated with susceptibility, cytokine levels, and clinical features in systemic lupus erythematosus. *Med. Baltim.* 99 (41), e22607. doi:10.1097/MD.00000000000022607
- Fan, Y., and Lu, D. (2016). The Ikaros family of zinc-finger proteins. *Acta Pharm. Sin. B* 6 (6), 513–521. doi:10.1016/j.apsb.2016.06.002
- Ferlay, J., Soerjomataram, I., Dikshit, R., Eser, S., Mathers, C., Rebelo, M., et al. (2015). Cancer incidence and mortality worldwide: Sources, methods and major patterns in GLOBOCAN 2012. *Int. J. Cancer* 136 (5), E359–E386. doi:10.1002/ijc.29210
- Fu, Q., Chen, N., Ge, C., Li, R., Li, Z., Zeng, B., et al. (2019). Prognostic value of tumor-infiltrating lymphocytes in melanoma: A systematic review and meta-analysis. *Oncotarget* 8 (7), 1593806. doi:10.1080/2162402X.2019.1593806
- Gao, S., Wang, S., and Song, Y. (2020). Novel immunomodulatory drugs and neo-substrates. *Biomark. Res.* 8, 2. doi:10.1186/s40364-020-0182-y
- He, L. C., Gao, F. H., Xu, H. Z., Zhao, S., Ma, C. M., Li, J., et al. (2012). Ikaros inhibits proliferation and, through upregulation of Slug, increases metastatic ability of ovarian serous adenocarcinoma cells. *Oncol. Rep.* 28 (4), 1399–1405. doi:10.3892/or.2012.1946
- Hodi, F. S., O'Day, S. J., McDermott, D. F., Weber, R. W., Sosman, J. A., Haanen, J. B., et al. (2010). Improved survival with ipilimumab in patients with metastatic melanoma. *N. Engl. J. Med.* 363 (8), 711–723. doi:10.1056/NEJMoa1003466
- Hung, J. J., Kao, Y. S., Huang, C. H., and Hsu, W. H. (2020). Author Correction: Overexpression of Aiolos promotes epithelial-mesenchymal transition and cancer stem cell-like properties in lung cancer cells. *Sci. Rep.* 10 (1), 1309. doi:10.1038/s41598-020-57957-0
- Iglesia, M. D., Parker, J. S., Hoadley, K. A., Serody, J. S., Perou, C. M., and Vincent, B. G. (2016). Genomic analysis of immune cell infiltrates across 11 tumor types. *J. Natl. Cancer Inst.* 108 (11), djw144. doi:10.1093/jnci/djw144
- Javierre, B. M., Rodriguez-Ubrea, J., Al-Shahrour, F., Corominas, M., Grana, O., Ciudad, L., et al. (2011). Long-range epigenetic silencing associates with deregulation of Ikaros targets in colorectal cancer cells. *Mol. Cancer Res.* 9 (8), 1139–1151. doi:10.1158/1541-7786.MCR-10-0515
- John, L. B., and Ward, A. C. (2011). The Ikaros gene family: Transcriptional regulators of hematopoiesis and immunity. *Mol. Immunol.* 48 (9–10), 1272–1278. doi:10.1016/j.molimm.2011.03.006
- Lee, N., Zakka, L. R., Mihm, M. C., Jr., and Schatton, T. (2016). Tumour-infiltrating lymphocytes in melanoma prognosis and cancer immunotherapy. *Pathology* 48 (2), 177–187. doi:10.1016/j.pathol.2015.12.006
- Liu, Y. Y., Ge, C., Tian, H., Jiang, J. Y., Zhao, F. Y., Li, H., et al. (2017). The transcription factor Ikaros inhibits cell proliferation by downregulating ANXA4 expression in hepatocellular carcinoma. *Am. J. Cancer Res.* 7 (6), 1285–1297.
- Locy, H., de Mey, S., de Mey, W., De Ridder, M., Thielemans, K., and Maenhout, S. K. (2018). Immunomodulation of the tumor microenvironment: Turn foe into friend. *Front. Immunol.* 9, 2909. doi:10.3389/fimmu.2018.02909
- Megahed, M., Schön, M., Selimovic, D., and Schön, M. P. (2002). Reliability of diagnosis of melanoma *in situ*. *Lancet (London, Engl.)* 359 (9321), 1921–1922. doi:10.1016/S0140-6736(02)08741-X
- Molnár, A., Georgopoulos, K., and Molnar, A. (1994). The Ikaros gene encodes a family of functionally diverse zinc finger DNA-binding proteins. *Mol. Cell. Biol.* 14 (12), 8292–8303. doi:10.1128/mcb.14.12.8292
- Nazarian, R., Shi, H., Wang, Q., Kong, X., Koya, R. C., Lee, H., et al. (2010). Melanomas acquire resistance to B-RAF(V600E) inhibition by RTK or N-RAS upregulation. *Nature* 468 (7326), 973–977. doi:10.1038/nature09626
- Newman, A. M., Liu, C. L., Green, M. R., Gentles, A. J., Feng, W., Xu, Y., et al. (2015). Robust enumeration of cell subsets from tissue expression profiles. *Nat. Methods* 12 (5), 453–457. doi:10.1038/nmeth.3337
- Payne, K. J., and Dovat, S. (2011). Ikaros and tumor suppression in acute lymphoblastic leukemia. *Crit. Rev. Oncog.* 16 (1–2), 3–12. doi:10.1615/critrevoncog.v16.i1-2.20
- Perdomo, J., Holmes, M., Chong, B., and Crossley, M. (2000). Eos and pegasus, two members of the Ikaros family of proteins with distinct DNA binding activities. *J. Biol. Chem.* 275 (49), 38347–38354. doi:10.1074/jbc.M005457200
- Pourabdollah, M., Bahmanyar, M., Atenafu, E. G., Reece, D., Hou, J., and Chang, H. (2016). High IKZF1/3 protein expression is a favorable prognostic factor for survival of relapsed/refractory multiple myeloma patients treated with lenalidomide. *J. Hematol. Oncol.* 9 (1), 123. doi:10.1186/s13045-016-0354-2
- Rebollo, A., and Schmitt, C. (2003). Ikaros, Aiolos and Helios: Transcription regulators and lymphoid malignancies. *Immunol. Cell Biol.* 81 (3), 171–175. doi:10.1046/j.1440-1711.2003.01159.x
- Roma-Rodrigues, C., Mendes, R., Baptista, P. V., and Fernandes, A. R. (2019). Targeting tumor microenvironment for cancer therapy. *Int. J. Mol. Sci.* 20 (4), E840. doi:10.3390/ijms20040840
- Siegel, R. L., Miller, K. D., and Jemal, A. (2016). Cancer statistics. *Ca. Cancer J. Clin.* 66 (1), 7–30. doi:10.3322/caac.21332
- Soengas, M. S., and Lowe, S. W. (2003). Apoptosis and melanoma chemoresistance. *Oncogene* 22 (20), 3138–3151. doi:10.1038/sj.onc.1206454
- Sriaroon, P., Chang, Y., Ujhazi, B., Csomos, K., Joshi, H. R., Zhou, Q., et al. (2019). Familial immune thrombocytopenia associated with a novel variant in IKZF1. *Front. Pediatr.* 7, 139. doi:10.3389/fped.2019.00139
- Sun, D., Wang, J., Han, Y., Dong, X., Ge, J., Zheng, R., et al. (2021). Tisch: A comprehensive web resource enabling interactive single-cell transcriptome visualization of tumor microenvironment. *Nucleic Acids Res.* 49 (D1), D1420–d1430. doi:10.1093/nar/gkaa1020
- Sun, L., Liu, A., and Georgopoulos, K. (1996). Zinc finger-mediated protein interactions modulate Ikaros activity, a molecular control of lymphocyte development. *EMBO J.* 15 (19), 5358–5369. doi:10.1002/j.1460-2075.1996.tb00920.x
- Tachita, T., Kinoshita, S., Ri, M., Aoki, S., Asano, A., Kanamori, T., et al. (2020). Expression, mutation, and methylation of cereblon-pathway genes at pre- and post-lenalidomide treatment in multiple myeloma. *Cancer Sci.* 111 (4), 1333–1343. doi:10.1111/cas.14352
- Tsai, K. K., Zarzoso, I., and Daud, A. I. (2014). PD-1 and PD-L1 antibodies for melanoma. *Hum. Vaccin. Immunother.* 10 (11), 3111–3116. doi:10.4161/21645515.2014.983409
- Vitale, C., Falchi, L., Ten Hacken, E., Gao, H., Shaim, H., Van Roosbroeck, K., et al. (2016). Ofatumumab and lenalidomide for patients with relapsed or refractory chronic lymphocytic leukemia: Correlation between responses and immune characteristics. *Clin. Cancer Res.* 22 (10), 2359–2367. doi:10.1158/1078-0432.CCR-15-2476
- Wang, X., Xiong, H., Liang, D., Chen, Z., Li, X., and Zhang, K. (2020). The role of SRG in the survival and immune infiltrates of skin cutaneous melanoma (SKCM) and SKCM-metastasis patients. *BMC cancer* 20 (1), 378. doi:10.1186/s12885-020-06849-7
- Weber, J. (2010). Immune checkpoint proteins: A new therapeutic paradigm for cancer—preclinical background: CTLA-4 and PD-1 blockade. *Semin. Oncol.* 37 (5), 430–439. doi:10.1053/j.seminoncol.2010.09.005
- Yang, M., Liu, Y., Mo, B., Xue, Y., Ye, C., Jiang, Y., et al. (2019). Helios but not CD226, TIM3 and Foxp3 is a potential marker for CD4+ treg cells in patients with rheumatoid arthritis. *Cell. Physiol. Biochem.* 52 (5), 1178–1192. doi:10.33594/0000000080
- Yeoh, A., Lu, Y., Chin, W., Chiew, E. K. H., Lim, E. H., Li, Z., et al. (2018). Intensifying treatment of childhood B-lymphoblastic leukemia with IKZF1 deletion reduces relapse and improves overall survival: Results of Malaysia-Singapore ALL 2010 study. *J. Clin. Oncol.* 36 (26), 2726–2735. doi:10.1200/JCO.2018.78.3050

- Yoshida, T., and Georgopoulos, K. (2014). Ikaros fingers on lymphocyte differentiation. *Int. J. Hematol.* 100 (3), 220–229. doi:10.1007/s12185-014-1644-5
- Yoshihara, K., Shahmoradgoli, M., Martínez, E., Vegesna, R., Kim, H., Torres-García, W., et al. (2013). Inferring tumour purity and stromal and immune cell admixture from expression data. *Nat. Commun.* 4, 2612. doi:10.1038/ncomms3612
- Zhao, W., Chen, T. B., and Wang, H. (2020). Ikaros is heterogeneously expressed in lung adenocarcinoma and is involved in its progression. *J. Int. Med. Res.* 48 (8), 300060520945860. doi:10.1177/0300060520945860
- Zhou, H., Yan, Y., Zhang, X., Zhao, T., Xu, J., and Han, R. (2020). Ginseng polysaccharide inhibits MDA-MB-231 cell proliferation by activating the inflammatory response. *Exp. Ther. Med.* 20 (6), 229. doi:10.3892/etm.2020.9359
- Zhou, J., Dudley, M. E., Rosenberg, S. A., and Robbins, P. F. (2005). Persistence of multiple tumor-specific T-cell clones is associated with complete tumor regression in a melanoma patient receiving adoptive cell transfer therapy. *J. Immunother.* 28 (1), 53–62. doi:10.1097/00002371-200501000-00007
- Zhou, N., Gutierrez-Uzquiza, A., Zheng, X. Y., Chang, R., Vogl, D. T., Garfall, A. L., et al. (2019). RUNX proteins desensitize multiple myeloma to lenalidomide via protecting IKZFs from degradation. *Leukemia* 33 (8), 2006–2021. doi:10.1038/s41375-019-0403-2

Glossary

TME Tumor microenvironment	LGG Brain Lower Grade Glioma
TIL Tumor-infiltrating lymphocyte	LIHC Liver hepatocellular carcinoma
GSEA Gene Set Enrichment Analysis	LUAD Lung adenocarcinoma
HR Hazard ratio	LUSC Lung squamous cell carcinoma
OS Overall survival	MESO Mesothelioma
PFI Progression-free interval	OV Ovarian serous cystadenocarcinoma
DSS Disease-free survival	PAAD Pancreatic adenocarcinoma
ACC Adrenocortical carcinoma	PCPG Pheochromocytoma and Paranganglioma
BLCA Bladder Urothelial Carcinoma	PRAD Prostate adenocarcinoma
BRCA Breast invasive carcinoma	READ Rectum adenocarcinoma
CESC Cervical squamous cell carcinoma and endocervical adenocarcinoma	SARC Sarcoma
CHOL Cholangiocarcinoma	SKCM Skin Cutaneous Melanoma
COAD Colon adenocarcinoma	STAD Stomach adenocarcinoma
DLBC Lymphoid Neoplasm Diffuse Large B-cell Lymphoma	TGCT Testicular Germ Cell Tumors
ESCA Esophageal carcinoma	THCA Thyroid carcinoma
GBM Glioblastoma multiforme	THYM Thymoma
HNSC Head and Neck squamous cell carcinoma	UCEC Uterine Corpus Endometrial Carcinoma
KICH Kidney Chromophobe	UCS Uterine Carcinosarcoma
KIRC Kidney renal clear cell carcinoma	UVM Uveal Melanoma
KIRP Kidney renal papillary cell carcinoma	MDS Myelodysplastic syndrome
LAML Acute Myeloid Leukemia	MM Multiple myeloma
	IMiD Immunomodulatory drugs
	CLL Chronic lymphocytic leukemia