

Evaluation of the Phenolic Content, Antioxidant and Antimicrobial Activities of Oil and Non-Oil Extracts of *Citrus sinensis* (L.) Osbeck Seeds

Ehigbai I. Oikeh^{1,2}, Merit Ayevbunwan³, Francis Irabor⁴, Aiemere O. Oikeh¹, Faith E. Oviasogie⁵, and Ehimwenma S. Omoregie¹

¹Department of Biochemistry, Faculty of Life Sciences, ³Department of Medical Biochemistry, College of Basic Medical Sciences, and

⁵Department of Microbiology, Faculty of Life Sciences, University of Benin, Benin City, Edo State 301110, Nigeria

²Department of Chemistry, College of Arts and Sciences, University of Kentucky, Lexington, KY 40506, USA

⁴Department of Biological Sciences, Faculty of Natural and Applied Sciences, Michael and Cecilia Ibru University, Agbarha-Otor, Delta State 333106, Nigeria

ABSTRACT: The seeds of *Citrus sinensis* (L.) Osbeck (sweet orange) are waste products usually discarded. They may however contain phytochemicals that have potent bioactivities. In this study, the phenolic content, and antioxidant and antimicrobial activities of oil and non-oil (solid) extracts of *C. sinensis* seeds were evaluated using standard protocols. The seed oil contained significantly ($P>0.05$) higher contents of total phenol and total flavonoid when compared to the solid extract. However, the non-oil extract contained significantly ($P<0.05$) higher tannin contents than the seed oil. Ferric reducing antioxidant potential was not significantly different between both extracts. The antimicrobial activities of both extracts revealed that the seed oil possesses better antibacterial activities compared to the non-oil extract. The antifungal test revealed that the seed oil significantly inhibited the growth of *Candida albicans* (20 mm zone of inhibition at a concentration of 200 $\mu\text{g/mL}$), however, it did not inhibit the growth of *Aspergillus niger* and *Penicillium* sp. The minimum inhibitory concentration values against the bacterial and fungal strains were similar for both extracts in the range of 50~100 $\mu\text{g/mL}$. Minimum bactericidal concentration and minimum fungicidal concentration values ranged from 100~200 $\mu\text{g/mL}$ for both extracts. The results in this study indicate that *C. sinensis* seed oil and non-oil extracts possess antioxidant, and antibacterial and antifungal properties that may be differentially exploited in the development of antimicrobial agents.

Keywords: antimicrobial, *Citrus sinensis*, minimum bactericidal concentration, minimum fungicidal concentration, minimum inhibitory concentration

INTRODUCTION

More than eight thousand phenolic compounds are known to exist in nature. These compounds chemically contain at least one aromatic ring with one or more attached hydroxyl (OH^-) groups as well as other substituents. Phenolic derivatives are the largest group of secondary plant metabolites synthesized by higher plants and are believed to be essential for plant survival by providing defense against microbial attack and by making the plant unpalatable to herbivores (Apak et al., 2007; Tungmunnithum et al., 2018)

Several plant phenols have been identified as the bioactive principles of folk herbal medicine with reported

antioxidant, antitumor, antiviral, and antibiotic activities. Flavonoids and tannins are among the 15 major structural classes of plant phenolics believed to be responsible for these health benefits (Mahmud et al., 2009). They have also been implicated as being useful in managing ailments such as cardiovascular disease, inflammatory conditions, liver disease, and macular degeneration (Apak et al., 2007; Ayoola et al., 2008). Tannins are polyphenolic metabolites of plants that possess the ability to precipitate gelatin and other proteins from solution (Rahman et al., 2011). Flavonoids are a special class of more than 4,000 members in which the 2 aromatic rings are linked by 3 carbons cyclized with oxygen. Flavonoids are known to be potent antioxidants with diverse mechanisms of ac-

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Correspondence to Ehigbai I. Oikeh, Tel: +1-334-314-2011, E-mail: ehigbai.oikeh@uniben.edu

Author information: Ehigbai I. Oikeh (Researcher), Merit Ayevbunwan (Researcher), Francis Irabor (Researcher), Aiemere O. Oikeh (Graduate Student), Faith E. Oviasogie (Professor), Ehimwenma S. Omoregie (Professor)

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tion (Silva and Fernandes Júnior, 2010).

The existence of antimicrobial resistance of several pathogenic organisms is one of the issues that has continued to be of concern to scientists and the abuse of antimicrobial drugs is one factor that has contributed to this problem (Ekhaïse et al., 2012). Scientists have sustained the search for new and effective medicines targeted against various microbial strains. Plant materials continue to be the preferred starting material for discovery of potential antimicrobial drugs (Akinsulire et al., 2007; Ekhaïse et al., 2012) due to the presence of compounds such as phenolics, tannins, and essential oils synthesized in the secondary metabolism of medicinal plants (Nascimento et al., 2000).

The genus *Citrus* belong to the Rutaceae family and comprises trees, shrubs, and herbs of diverse sizes and uses. Citrus plants are among the world's most important crops (Mahmud et al., 2009). Citrus fruits and juices are known to contain several bioactive compounds that are important to human health. They include ascorbic acid, pectin, flavonoids, and phenolic compounds (Ghasemi et al., 2009; Oikeh et al., 2015; Jannat et al., 2016; Sir Elkhatim et al., 2018). We have in earlier studies shown that wastes generated from citrus fruits possess useful phytochemicals with antioxidant properties (Oikeh et al., 2013; Oikeh et al., 2014). This study therefore aims to contribute to the drug discovery process by providing information on the phenolic content and antimicrobial activities of oil and non-oil extracts of sweet orange seeds.

MATERIALS AND METHODS

Raw materials

Fresh oranges were purchased from New Benin Market in Benin City, Nigeria. The fruits were washed with distilled water and the peels were removed with the aid of a sharp knife. A juice extractor was used to extract the juice and then the seeds collected. The seeds were pulverized immediately after collection using a sterile mortar and pestle.

Preparation of extract

The pulverized seeds were subjected to Soxhlet extraction for a period of 12 h with 500 mL of ethanol and then concentrated using a rotary evaporator at reduced pressure. The extracts were stored in the refrigerator till required for use.

Determination of total phenolic content

Total phenolic content was determined according to Folin-Ciocalteu reagent method (Cicco et al., 2009). Different concentrations (0.2~1.0 mg/mL) of gallic acid were prepared in methanol. Concentrations of the extracts

were also prepared in methanol. Distilled water (4.5 mL) was added to 0.5 mL of the extract and mixed with 0.5 mL of a 10-fold diluted Folin-Ciocalteu reagent. Subsequently, 5 mL of 7% sodium carbonate and 2 mL of distilled water were added. The mixture stood for 90 min at room temperature before the absorbance was read at 760 nm. All determinations were performed in triplicates with gallic acid utilized as the positive control. The total phenolic content was expressed as gallic acid equivalents (GAE).

Determination of total flavonoid content

Total flavonoid contents were estimated using the method described in Ebrahimzadeh et al. (2008). The extract (0.5 mL of 1 mg/mL) was mixed with 1.5 mL of methanol. To this mixture, 0.1 mL of 10% aluminum chloride was added, followed by 0.1 mL of 1 M potassium acetate and 2.8 mL of distilled water. The mixture was incubated at room temperature for 30 min. The absorbance was measured by a spectrophotometer at 420 nm. The results were expressed as milligrams quercetin equivalents (QE) per gram of extract (mg QE/g extract).

Determination of total tannin content

The total tannin content was determined by the modified method of Polshettiwar et al. (2007). The sample (0.1 mL) was mixed with 0.5 mL of Folin-Denis reagent and 1 mL of 0.5% sodium carbonate (w/v) solution. The mixture was then made up to 5 mL with distilled water. The absorbance was measured at 755 nm within 30 min of reaction. The total tannin in the extract was expressed as the equivalent to tannic acid.

Ferric reducing antioxidant power (FRAP) assay

The Benzie and Strain (1996) protocol with slight modification was employed for this assay. Different concentrations (100~500 mg/mL) of the concentrates and the standard were serially diluted with distilled water. Then 1 mL of FRAP reagent [200 mL of 300 mM sodium acetate buffer at pH 3.6, 20 mL of 10.0 mM 2,4,6-tripyridyl-S-triazine (TPTZ) solution, 20 mL of 20.0 mM $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ solution and 24 mL of distilled water] was added to each test tube. The resulting mixture was vigorously shaken and then incubated at 37°C for 4 min. The increase in absorbance at 593 nm was measured and compared with the standard ascorbic acid.

Antimicrobial assay

Test microorganisms: Seven microorganisms were used in this study: consisting of 4 bacterial and 3 fungal strains. Two of the 4 bacterial strains were Gram-positive (*Staphylococcus aureus* and *Enterococcus faecalis*) and the other 2 were Gram-negative (*Escherichia coli* and *Salmonella* sp). The 3 fungal strains used were *Candida albicans*, *Aspergil-*

lus niger, and *Penicillium* sp. All microorganisms were obtained from Lahor Research and Diagnostic Laboratories, Benin City, Nigeria.

Antimicrobial susceptibility assay: This was carried out using the protocol described by Owoseni and Ajayi (2010). Test organisms were sub-cultured onto fresh suitable broth medium. Broth cultures were then incubated at 37°C till the turbidity of 0.5 McFarland's standard (1.5×10^8 colony forming unit/mL) was obtained. Mueller-Hinton agar was used as bacterial medium and Sabouraud agar as fungal medium. All were incubated appropriately as specified for each test organism. The turbidity of the actively growing broth culture was then adjusted with sterile saline to obtain 0.5 McFarland's standard turbidity. This was used to flood and then drain dry the surface of solid Mueller-Hinton agar plates. Wells of 5 mm in diameter and about 2 cm apart were punched in the culture media with a sterile cork borer. The extracts were thereafter used to fill the boreholes. Each plate was kept in the refrigerator at 4°C for 1 h before incubating at 37°C for 24 h (bacteria) and 72 h (fungi). Zones of inhibition around the wells, measured in millimeters, were used as positive bioactivity.

Minimum inhibitory concentration (MIC)

The organisms that showed susceptibility to the different solvent extracts were introduced into the broths containing different concentrations of each extract (serial dilutions of the extracts corresponding to 200, 100, 50, 25, and 12.5 µg/mL). The tubes were then incubated for 24 h at 37°C. The MIC was taken as the lowest concentration of the extracts that did not permit any visible growth (Owoseni and Ajayi, 2010).

Minimum bactericidal concentration (MBC) and minimum fungicidal concentration (MFC)

The tubes that showed no turbidity in the MIC test were taken and a loopfull from each tube was streaked onto Mueller-Hinton agar. The plates were incubated for 24 h at 37°C and the absence of growth was observed. The concentration of the extracts that showed no growth was recorded as the MBC/MFC (CLSI, 2008a; CLSI, 2008b; CLSI, 2012).

Statistical analysis

The data were expressed as mean ± standard error of mean for three replicates. The data were subjected to one-way analysis of variance and differences between means were determined by Duncan's multiple range test using SPSS Statistics version 17.0 (SPSS Inc., Chicago, IL, USA) where applicable. *P*-values <0.05 were regarded as significant.

RESULTS AND DISCUSSION

Phytochemicals possessing antioxidant activities are important for different reasons. They are known to be useful against diseases linked to free radicals, DNA damage and carcinogenesis as well as in the prevention of oxidative reactions in foods. They are also believed to possess anti-inflammatory and antimicrobial properties (Zhang et al., 2015; Lee et al., 2017; Mintie et al., 2020). Several plants and plant parts hold promise as rich reservoirs of antimicrobial compounds and thus, require sustained pharmacological investigations. *Citrus sinensis* seeds contain phytoconstituents such as flavonoids, saponins, tannins, cardiac glycoside alkaloids, and steroids (Oikeh et al., 2013). Some of these phytoconstituents are known to contain diverse antimicrobial properties (Owoseni and Ajayi, 2010; Dhiman et al., 2012).

Upon completion of the Soxhlet extraction process, a yield of 15.69% was obtained (Table 1). This consisted predominantly of oil (13.05%) and a denser non-oil portion (2.64%). Both components were separated by draining off the oil and then used for further analysis. The *C. sinensis* oil seed extract was observed to contain significantly higher amounts of total flavonoids and total phenols compared to the non-oil extract (Fig. 1A and 1B). Total tannins were however significantly higher in the non-oil extract compared to the oil extract (Fig. 1C). The result of the ferric reducing antioxidant potential assay revealed that there was no significant difference between the FRAP values of both seed extracts and the reference antioxidant, ascorbic acid (Fig. 1D). This result suggests that the seeds may possess good reductive properties comparable to ascorbic acid and this could be attributed to the presence of phenolics such as flavonoids and tannins present in both extracts.

Phenolic compounds are known to play important roles in plant defense against pathogens and herbivore predators. They are effective in the control of human pathogenic infections (Jimoh et al., 2008). The antioxidant properties of phenolic compounds are mainly attributed to their redox properties, allowing them to act as reducing agents, hydrogen donors, etc. (Firoozi et al., 2016; Filip, 2017). Flavonoids interfere with the activities of enzymes involved in reactive oxygen species generation and quenching of free radicals (Doughari, 2012). Tannins are astringent in nature. Their antioxidant capacity is linked to the presence of readily oxidizable hydroxyl groups and their

Table 1. Percentage yield of oil and non-oil extracts of dry *Citrus sinensis* seed

	Yield (%)
Seed oil	13.05
Non-oil extract	2.64

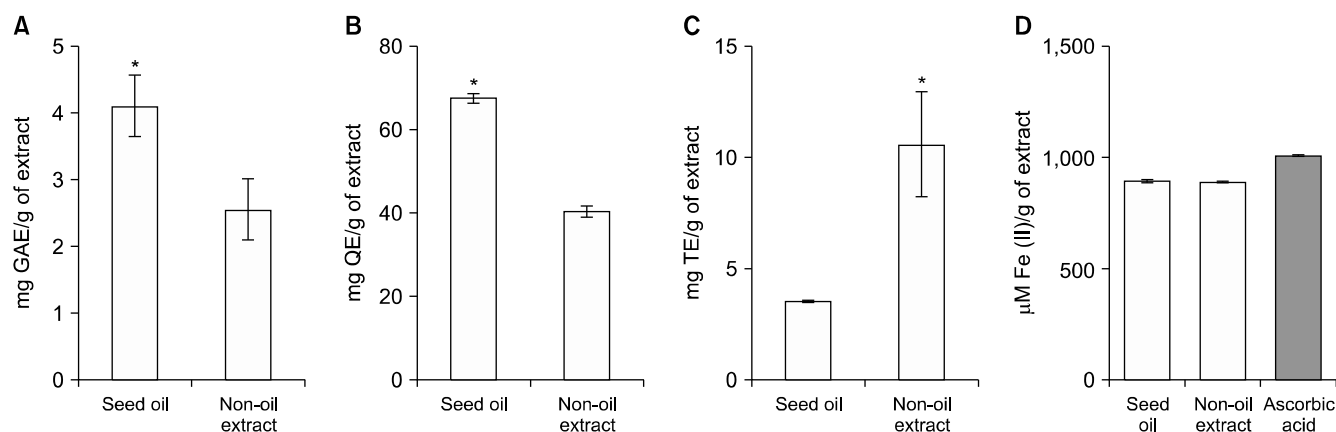


Fig. 1. (A) Total phenolic content, (B) flavonoid content, (C) tannin content, and (D) ferric reducing antioxidant potential of oil and non-oil extracts of dry *Citrus sinensis* seed. Values are mean \pm SEM (n=3). *Significantly different from the other group at $P<0.05$. GAE, gallic acid equivalents; QE, quercetin equivalents; TE, tannic equivalents.

Table 2. Zones of inhibition for some bacterial strains at various concentrations of dry *Citrus sinensis* seed oil and non-oil extracts (unit: mm)

Concentration ($\mu\text{g/mL}$)	<i>Staphylococcus aureus</i>		<i>Enterococcus faecalis</i>		<i>Escherichia coli</i>		<i>Salmonella</i> sp.	
	Seed oil	Non-oil extract	Seed oil	Non-oil extract	Seed oil	Non-oil extract	Seed oil	Non-oil extract
200	6	6	8	2	4	2	18	4
100	4	4	6	1	2	1	16	2
50	2	2	—	—	—	—	—	—

degree of polymerization (Smeriglio et al., 2017; Huang et al., 2018).

The antibacterial susceptibility test for oil and non-oil extracts of dry *C. sinensis* seed against some bacterial strains is shown in Table 2. The observed zones of inhibition varied from one organism to another and at different concentrations of either extract. The Gram-positive strain *Staphylococcus aureus* showed concentration-dependent susceptibility to both extracts at 200, 100, and 50 $\mu\text{g/mL}$, the zone of microbial inhibition corresponded to 6, 4, and 2 mm, respectively. However, *Enterococcus faecalis* was insensitive to either extract at 50 $\mu\text{g/mL}$ but at 200 and 100 $\mu\text{g/mL}$, the degree of susceptibility of the organism to the seed oil was 8 mm and 6 mm, respectively. Susceptibility was however lower for the non-oil extract (2 and 1 mm, respectively). Inhibition of *Escherichia coli* and *Salmonella* sp. growth was observed at 100 and 200 $\mu\text{g/mL}$ for both extracts. Zones of inhibition observed for *Salmonella* sp. were 16 and 18 mm, respectively for the oil extract and 2 and 4 mm, respectively for the non-oil extract. *E. coli* inhibition zones were much smaller (2 and 4 mm, respectively for the oil extract; 1 and 2 mm, respectively for the non-oil extract).

Several studies have reported lower zones of inhibition in Gram-negative organisms compared to Gram-positive organisms. This is attributed to the Gram-negative outer membrane functioning as a barrier that prevents the penetration of antimicrobial compounds directed against the

bacteria. The presence of hydrolytic enzymes that break down foreign molecules in the periplasmic space is also another factor that limits the susceptibility of Gram-negative bacteria to antimicrobial substances (Holetz et al., 2002; Oikeh et al., 2015). Our results however show that the *C. sinensis* oil has good inhibitory action against *Salmonella* sp. The *C. sinensis* seed oil may therefore be a potent source of antimicrobials targeted against *Salmonella* sp. The effectiveness of the seed oil against *Salmonella* sp. is particularly important due to the prevalence of *Salmonella* sp. in food-borne infections.

Table 3 shows the minimum inhibitory and bacteriocidal concentrations of oil and non-oil extracts for dry *C. sinensis* seed against some bacterial strains. The MIC is the lowest concentration of a drug that is bacteriostatic, preventing the visible growth of bacteria. MICs are used by diagnostic laboratories mainly to confirm resistance, but mostly as a research tool to determine the activity of novel antimicrobial agents. On the other hand, MBC is the minimum concentration of an antimicrobial agent that can kill 99.9% of bacteria (Andrews, 2001). The MIC values ranged from 50 to 100 $\mu\text{g/mL}$, while MBC values ranged from 100 to 200 $\mu\text{g/mL}$ for both extracts studied. The results obtained showed that the MIC values for both the oil and non-oil extract were lower than their MBC values. This suggests that both extracts possess bacteriostatic effect at lower concentrations and bacteriocidal effect at higher concentrations.

Table 3. Minimum inhibitory and bacteriostatic concentrations of dry *Citrus sinensis* seed oil and non-oil extract against microbial strains (unit: $\mu\text{g}/\text{mL}$)

	Gram-positive				Gram-negative			
	<i>Staphylococcus aureus</i>		<i>Enterococcus faecalis</i>		<i>Escherichia coli</i>		<i>Salmonella</i> sp.	
	MIC	MBC	MIC	MBC	MIC	MBC	MIC	MBC
Seed oil	50	100	100	200	100	200	100	200
Non-oil extract	50	100	100	200	100	200	100	200

MIC, minimum inhibitory concentration; MBC, minimum bactericidal concentration.

Table 4. Zones of inhibition of some fungal strains at various concentrations of dry *Citrus sinensis* seed oil and non-oil extracts (unit: mm)

Concentration ($\mu\text{g}/\text{mL}$)	<i>Candida albicans</i>		<i>Aspergillus niger</i>		<i>Penicillium</i> sp.	
	Seed oil	Non-oil extract	Seed oil	Non-oil extract	Seed oil	Non-oil extract
200	20	2	–	8	–	4
100	8	1	–	4	–	2
50	4	–	–	2	–	1

Table 5. Minimum inhibitory and fungicidal concentrations of fresh and dry *Citrus sinensis* seed oil and non-oil extracts against some fungal strains (unit: $\mu\text{g}/\text{mL}$)

	<i>Candida albicans</i>		<i>Aspergillus niger</i>		<i>Penicillium</i> sp.	
	MIC	MFC	MIC	MFC	MIC	MFC
Seed oil	50	100	–	–	–	–
Non-oil extract	100	200	50	100	50	100

MIC, minimum inhibitory concentration; MFC, minimum fungicidal concentration.

The antifungal activities of the extracts for dry *C. sinensis* seed revealed that *C. albicans* was more susceptible to the seed oil compared to non-oil extract with observed zones of inhibition ranging from 4 mm to 20 mm for seed oil extract at different concentrations while non-oil extract recorded 1 mm and 2 mm at 100 and 200 $\mu\text{g}/\text{mL}$, respectively (Table 4). However, *A. niger* and *Penicillium* sp. were not susceptible to seed oil at all concentrations studied. The bioactivity of the seed oil against *C. albicans* is important due to the disease-causing potential of *C. albicans*. Previous studies of the authors have also shown good susceptibility of *C. albicans* to some citrus fruit concentrates (Oikeh et al., 2015). Citrus fruits may hold promise as a starting material for the discovery of new anti-Candidiasis agents.

Table 5 shows the inhibitory and fungicidal concentrations of oil and non-oil extracts for dry *C. sinensis* seed against some fungi. MIC values ranged from 50 to 100 $\mu\text{g}/\text{mL}$, while MFC values ranged from 100 to 200 $\mu\text{g}/\text{mL}$ for both extracts studied against the different fungal strains. The seed oil was more effective in inhibiting the growth of *C. albicans* with MIC and MFC values of 50 $\mu\text{g}/\text{mL}$ and 100 $\mu\text{g}/\text{mL}$, respectively, compared to 100 $\mu\text{g}/\text{mL}$ and 200 $\mu\text{g}/\text{mL}$ observed for the non-oil extract. This result suggests that *C. sinensis* oil extract is a better fungicidal agent against *C. albicans* compared to the non-oil ex-

tract. MIC and MFC values for the non-oil extract against *A. niger* and *Penicillium* sp. were both 50 $\mu\text{g}/\text{mL}$ and 100 $\mu\text{g}/\text{mL}$, respectively. MIC and MFC values could not be obtained for the *C. sinensis* seed oil as these fungal species were not susceptible to the oil.

In conclusion, the results of this study show differences in the antimicrobial activities of the oil and non-oil extracts against the bacterial and fungal strains studied. This study has provided preliminary *in vitro* data on the antimicrobial potential of extracts obtained from seeds of *C. sinensis*. The results in this study provide information on potential antibacterial and antifungal leads. More extensive studies for the potential identification and isolation of lead compounds as well as confirmatory *in vivo* studies will be carried out.

AUTHOR DISCLOSURE STATEMENT

The authors declare no conflict of interest.

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