



# Draft Genome Sequence of *Clostridium cochlearium* Strain AGROS13, Isolated from a Sheep Dairy Farm in New Zealand

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**ABSTRACT** We report the draft genome sequence of a new *Clostridium cochlearium* strain, AGROS13, which was isolated from a sheep dairy farm environment in New Zealand. The genome is 2.7 Mbp, with a GC content of 28.2%. The genome sequence was found to be closely related to that of *Clostridium cochlearium* ATCC 17787. The new strain harbors a biosynthetic gene cluster coding for an unknown sactipeptide.

*Clostridium* species are obligate or facultative anaerobic bacteria, producing endospores that are highly resistant to heat and other environmental factors (1, 2). Some *Clostridium* species are well-known pathogens (1, 3–6), whereas some are detrimental to milk and dairy product quality (7, 8). Of all the important *Clostridium* species known, not much has been stated about *Clostridium cochlearium*, a species that has been found to spoil dairy products and also has been isolated from infant formula milk (9, 10). Here, we report the whole-genome sequence of a new *Clostridium cochlearium* strain, AGROS13, which was isolated from a New Zealand sheep dairy farm silage sample.

Bacteria were isolated using a previous methodology, with slight modifications (11). Briefly, 20 g of silage was weighed in a stomacher bag, suspended in 50 ml of phosphate buffer to blend the sample, and centrifuged at  $3,466 \times g$  for 1 h. The pellet was resuspended in 10 ml of phosphate buffer and heated at 80°C for 10 min. One milliliter of the heated sample was added to cooked meat-glucose starch medium (12) and incubated anaerobically at 35°C for 48 h. The growth suspension was serially diluted, plated on Shahidi-Ferguson agar, and incubated anaerobically for 24 h (13). Proteolytic activity was preliminarily investigated by visualizing a clear zone around the bacterial growth on a skimmed milk agar plate (14). The presumptive *C. cochlearium* strain AGROS13 was found to be proteolytic, indicating potentially a dairy spoilage bacterium. Genomic DNA was extracted from pure cultures grown in tryptic soy broth (Fort Richard, New Zealand) by using the phenol-chloroform extraction method (15). The quality and concentration of DNA were determined using a Qubit 2.0 fluorometer (Thermo Fisher Scientific, USA).

The whole genome of *Clostridium* strain AGROS13 was prepared with the NuGEN Celero enzymatic fragmentation DNA library kit and sequenced using the Illumina MiSeq sequencing platform version 3 (Massey Genome Services, Palmerston North, New Zealand), producing 484,293 read pairs of 300 nucleotides and 291,544,386 bp and giving a coverage of  $\sim 109$ -fold. The reads were quality trimmed, filtered, and assembled via the A5-miseq pipeline version 20160825 with default settings (16). The assembly produced 75 contigs, with a total genome size of 2.7 Mb, an  $N_{50}$  value of 78 kb, and a GC content of 28.2%. A BUSCO version 3.0.2 (17) test using the bacterial reference produced a completeness score of 93.9%.

A two-way average nucleotide identity test (<http://enve-omics.ce.gatech.edu/ani>) of

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the new *Clostridium* strain AGROS13 produced a 98.96% match with *Clostridium cochlearium* NCTC 13027 (GenBank accession number [NZ\\_LT906477.1](https://ncbi.nlm.nih.gov/nucl/NZ_LT906477.1)) (18). A comparative genomic analysis of these two genomes using *in silico* digital DNA-DNA hybridization (dDDH) via the Type (strain) Genome Server (TYGS) (<https://tygs.dsmz.de>) (19) resulted in a dDDH ( $d_g$ ) value of 80%, indicating the same species but with probable differences at the strain level. We investigated the presence of biosynthetic gene clusters (BGCs) in strain AGROS13 using antiSMASH version 5.1.2 (<https://antismash.secondarymetabolites.org>) (20). The software predicted the presence of a BGC encoding an unknown sactipeptide, a ribosomally synthesized and posttranslationally modified peptide (RiPP) (21). RiPPs have been recognized as a predominant group of natural antimicrobial compounds, of which sactipeptides and lanthipeptides are the dominant ones identified in some *Clostridium* species (22, 23). Further studies are required to identify the sactipeptide and to investigate its properties. As part of the submission process, NCBI annotated the genomic scaffolds with PGAP version 4.11 (24), resulting in 2,692 genes being annotated in total.

**Data availability.** The raw reads have been deposited in the NCBI SRA under the accession number [SRX8326676](https://ncbi.nlm.nih.gov/sra/SRX8326676). This whole-genome shotgun project has been deposited in DDBJ/ENA/GenBank under the accession number [JABFIF000000000](https://ncbi.nlm.nih.gov/nucl/JABFIF000000000). The version described in this paper is version [JABFIF010000000](https://ncbi.nlm.nih.gov/nucl/JABFIF010000000).

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## REFERENCES

- Wells CL, Wilkins TD. 1996. Clostridia: sporeforming anaerobic bacilli, p 279–296. In Baron S (ed), *Medical microbiology*, 4th ed. University of Texas Medical Branch at Galveston, Galveston, TX.
- Brown K. 2000. Control of bacterial spores. *Br Med Bull* 56:158–171. <https://doi.org/10.1258/0007142001902860>.
- Hatheway CL. 1993. *Clostridium botulinum* and other clostridia that produce botulinum neurotoxin, p 3–20. In Hauschild AHW, Dodds KL (ed), *Clostridium botulinum: ecology and control in foods*. CRC Press, Boca Raton, FL.
- Kelly CP, LaMont JT. 1998. *Clostridium difficile* infection. *Annu Rev Med* 49:375–390. <https://doi.org/10.1146/annurev.med.49.1.375>.
- Brook I. 2008. Current concepts in the management of *Clostridium tetani* infection. *Expert Rev Anti Infect Ther* 6:327–336. <https://doi.org/10.1586/14787210.6.3.327>.
- Uzal FA, Freedman JC, Shrestha A, Theoret JR, Garcia J, Awad MM, Adams V, Moore RJ, Rood JI, McClane BA. 2014. Towards an understanding of the role of *Clostridium perfringens* toxins in human and animal disease. *Future Microbiol* 9:361–377. <https://doi.org/10.2217/fmb.13.168>.
- Garde S, Ávila M, Gómez N, Nunez M, Garde S, Ávila M, Gómez N, Nunez M. 2013. *Clostridium* in late blowing defect of cheese: detection, prevalence, effects and control strategies, p 503–518. In Castelli H, du Vale L (ed), *Handbook on cheese: production, chemistry and sensory properties*. Nova Science Publishers, New York, NY.
- Reindl A, Dzielciel M, Hein I, Wagner M, Zangerl P. 2014. Enumeration of clostridia in goat milk using an optimized membrane filtration technique. *J Dairy Sci* 97:6036–6045. <https://doi.org/10.3168/jds.2014-8218>.
- Lycken L, Borch E. 2006. Characterization of *Clostridium* spp. isolated from spoiled processed cheese products. *J Food Prot* 69:1887–1891. <https://doi.org/10.4315/0362-028x-69.8.1887>.
- Barash JR, Hsia JK, Arnon SS. 2010. Presence of soil-dwelling clostridia in commercial powdered infant formulas. *J Pediatr* 156:402–408. <https://doi.org/10.1016/j.jpeds.2009.09.072>.
- Gupta TB, Brightwell G. 2017. Farm level survey of spore-forming bacteria on four dairy farms in the Waikato region of New Zealand. *Microbiologyopen* 6:e00457. <https://doi.org/10.1002/mbo3.457>.
- Murray PR, Baron EJ, Jorgensen JH, Pfaller MA, Tenover FC, Tenover FC (ed). 2003. *Manual of clinical microbiology*, 8th ed. ASM Press, Washington, DC.
- Shahidi SA, Ferguson AR. 1971. New quantitative, qualitative, and confirmatory media for rapid analysis of food for *Clostridium perfringens*. *Appl Environ Microbiol* 21:500–506. <https://doi.org/10.1128/AEM.21.3.500-506.1971>.
- Martley FG, Jayashankar SR, Lawrence RC. 1970. An improved agar medium for the detection of proteolytic organisms in total bacterial counts. *J Appl Bacteriol* 33:363–370. <https://doi.org/10.1111/j.1365-2672.1970.tb02208.x>.
- Gupta SK, Haigh BJ, Seyfert H-M, Griffin FJ, Wheeler TT. 2017. Bovine milk RNases modulate pro-inflammatory responses induced by nucleic acids in cultured immune and epithelial cells. *Dev Comp Immunol* 68:87–97. <https://doi.org/10.1016/j.dci.2016.11.015>.
- Coil D, Jospin G, Darling AE. 2015. A5-miseq: an updated pipeline to assemble microbial genomes from Illumina MiSeq data. *Bioinformatics* 31:587–589. <https://doi.org/10.1093/bioinformatics/btu661>.
- Simão FA, Waterhouse RM, Ioannidis P, Kriventseva EV, Zdobnov EM. 2015. BUSCO: assessing genome assembly and annotation completeness with single-copy orthologs. *Bioinformatics* 31:3210–3212. <https://doi.org/10.1093/bioinformatics/btv351>.
- Goris J, Konstantinidis KT, Klappenbach JA, Coenye T, Vandamme P, Tiedje JM. 2007. DNA-DNA hybridization values and their relationship to whole-genome sequence similarities. *Int J Syst Evol Microbiol* 57:81–91. <https://doi.org/10.1099/ijs.0.64483-0>.
- Meier-Kolthoff JP, Göker M. 2019. TYGS is an automated high-throughput platform for state-of-the-art genome-based taxonomy. *Nat Commun* 10:2182. <https://doi.org/10.1038/s41467-019-10210-3>.
- Blin K, Shaw S, Steinke K, Villebro R, Ziemert N, Lee SY, Medema MH, Weber T. 2019. antiSMASH 5.0: updates to the secondary metabolite genome mining pipeline. *Nucleic Acids Res* 47:W81–W87. <https://doi.org/10.1093/nar/gkz310>.
- Arnison PG, Bibb MJ, Bierbaum G, Bowers AA, Bugni TS, Bulaj G, Camarero JA, Campopiano DJ, Challis GL, Clardy J, Cotter PD, Craik DJ, Dawson M, Dittmann E, Donadio S, Dorrestein PC, Entian K-D, Fischbach MA, Garavelli JS, Göransson U, Gruber CW, Haft DH, Hemscheidt TK, Hertweck C, Hill C, Horswill AR, Jaspars M, Kelly WL, Klinman JP, Kuipers OP, Link AJ, Liu W, Marahiel MA, Mitchell DA, Moll GN, Moore BS, Müller R, Nair SK, Nes IF, Norris GE, Olivera BM, Onaka H, Patchett ML, Piel J, Reaney MJT, Rebuffat S, Ross RP, Sahl H-G, Schmidt EW, Selsted ME, Severinov K, Shen B, Sivonen K, Smith L, Stein T, Süßmuth RD, Tagg JR, Tang G-L, Truman AW, Vederas JC, Walsh CT, Walton JD, Wenzel SC, Willey JM, van der Donk WA. 2013. Ribosomally synthesized and post-translationally modified peptide natural products: overview and recom-

- mendations for a universal nomenclature. *Nat Prod Rep* 30:108–160. <https://doi.org/10.1039/c2np20085f>.
22. Letzel A-C, Pidot SJ, Hertweck C. 2014. Genome mining for ribosomally synthesized and post-translationally modified peptides (RiPPs) in anaerobic bacteria. *BMC Genomics* 15:983. <https://doi.org/10.1186/1471-2164-15-983>.
23. Pahalagedara A, Flint S, Palmer J, Brightwell G, Gupta TB. 2020. Antimicrobial production by strictly anaerobic *Clostridium* species. *Int J Antimicrob Agents* 55:105910. <https://doi.org/10.1016/j.ijantimicag.2020.105910>.
24. Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>.