

Supplemental Table 1

Gene symbol	Gene name	Forward primer sequence	Reverse primer sequence
<i>GAPDH</i>	Glyceraldehyde-3-Phosphate Dehydrogenase	CTGCACCACCAACTGCTTAG	ACAGTCTTCTGGGTGGCAGT
<i>PDCD1</i>	Programmed Cell Death 1	GGACAATAGGAGCCAGGCG	CCCATAGTCCACAGAGAACACA
<i>NFATC1</i>	Nuclear Factor Of Activated T Cells 1	GGAGATGGAAGCGAAACTG	GGGGCTGGTTATCCTCTGAT
<i>CTSK</i>	Cathepsin K	TTGGCAGTGGGATATGGAAT	GCCACAGGCGTTGTTCTTAT
<i>MMP9</i>	Matrix Metalloproteinase 9	TCTTCCCTGGAGACCTGAGA	ATTTGACTCTCCACGCATC
<i>ACP5</i>	Acid Phosphatase 5	CGTGTGGTCCATAGCCGAG	CCACGCCATTCTCATCTTGC
<i>CRLR</i>	Calcitonin Receptor Like Receptor	GCCTAAGTTGCCAAAGGATTACC	TGAACTGGGACACTTTGCAAC
<i>RAMP1</i>	Receptor Activity Modifying Protein 1	AGGTAGACATGGAGGCCGT	ACCTGTCCACCTCTGCATTG
<i>CALCR</i>	Calcitonin Receptor	CAGCAGTTACCCGCATACCA	GGACAATACTCCAGCCGGTG
<i>RANK</i>	TNF Receptor Superfamily Member 11a	TCCACGGACAAATGCAGACC	GCATCGGATTTCTCTGTCCCA
<i>RANKL</i>	TNF Superfamily Member 11	GGCTGGGCCAGGTTGTC	ATTCTATTAGGATCCATCTGCGCT

Supplemental Table 2: Upregulated and downregulated common genes after PD-L1 stimulation in comparison to both control group and Nivolumab treatment.

Common upregulated genes	Common downregulated genes
LINC02416	CYP1B1
SLC45A2	EPS8L1
SPACA3	NCR3LG1
BEST3	FAHD2B
ENSG00000214144	CFAP61
RPSAP3	C3
COL1A2	ASPHD2
ANGPTL1	HLA-H
NPPC	ENTPD1
CAMK2A	ENSG00000259238
TNN	ENSG00000197320
AFAP1-AS1	RPS4XP16
ENSG00000284999	LINC02972
NPHS1	SIGLEC16
ENSG00000228566	CPAMD8
LINC01091	PVALB
JCHAIN	GLB1L
UOX	ANKRD35
F2R	GAL3ST4
KIRREL2	PSME2P3
SLC9B2	CXCL10
SATB1-AS1	KCNMA1
OXCT1	LRRC4C
ITGA2	SRPX
SH3GL2	NDUFB1P1
CTNND2	HLA-DQA2
MYO1B	
NYAP2	
PTPRK	
REM1	
TAFA4	
DUSP4	
ENSG00000233397	
FSD2	
DGKI	
ENSG00000225885	
GSDMA	
CTSK	
SEMA3D	
SLC6A17	

SLC22A1	
CLDN14	
COL5A1	
PKNOX2	
CLIC6	
EXT1	
SEMA3E	
PKIA	
ITGB3	
SSC5D	
AKAP6	
DPP4	
FHL2	
ENSG00000253227	
STRIP2	
KIAA0040	
ENSG00000230387	
ZNF462	
SHC4	
ATP1B4	
RUFY4	
RPS16P2	
ZEB1	
FOLR3	
RND1	
LRRC8E	
DYRK3	
NEURL3	
SH3D19	
BAMBI	
GRAP2	
SLC9B1	
DNM1	
SYNM	
TPGS2	
LIPH	
CALCR	
PIP5K1B	
PARVA	
OMG	
ADGRB1	
ADAM19	
ATOH8	
LRP6	

<i>N4BP2</i>	
<i>C12orf75</i>	
<i>ENSG00000291236</i>	
<i>SPRY4</i>	
<i>DMPK</i>	
<i>GLI3</i>	
<i>MAP1A</i>	
<i>STAP2</i>	
<i>EGR1</i>	
<i>MYO1D</i>	
<i>PKIG</i>	
<i>GCNT3</i>	
<i>ENSG00000289070</i>	
<i>UST</i>	
<i>STAMBPL1</i>	
<i>OCIAD2</i>	
<i>DNAJA4</i>	
<i>KIAA1614</i>	
<i>EOGT</i>	
<i>LRG1</i>	
<i>ARHGAP23</i>	
<i>HLA-DRB5</i>	
<i>KIAA1328</i>	
<i>ACP5</i>	
<i>AFAP1</i>	
<i>WSCD1</i>	
<i>THOP1</i>	
<i>GOLM1</i>	
<i>LTBP2</i>	
<i>AKR1C1</i>	
<i>RBFOX2</i>	
<i>SDC1</i>	
<i>XPR1</i>	
<i>GHRL</i>	
<i>PPFIBP1</i>	
<i>NAT14</i>	
<i>PHETA1</i>	
<i>SNAPC1</i>	
<i>NFATC1</i>	
<i>PACSIN3</i>	
<i>RCAN1</i>	
<i>RGS10</i>	
<i>STX1A</i>	
<i>LHFPL6</i>	

<i>ENSG00000273341</i>	
<i>RAB11FIP5</i>	
<i>PLXNA2</i>	
<i>BMAL2</i>	
<i>CBLB</i>	
<i>SMARCA1</i>	
<i>RARG</i>	
<i>VWA5A</i>	
<i>GREB1</i>	
<i>JDP2</i>	
<i>STRADB</i>	
<i>C1orf198</i>	
<i>OAF</i>	
<i>BDH1</i>	
<i>PACC1</i>	
<i>NDFIP2</i>	
<i>ZSWIM4</i>	
<i>ZFTA</i>	
<i>DHTKD1</i>	

Supplemental Table 3: Upregulated and downregulated common pathways after PD-L1 stimulation in comparison to both control group and Nivolumab treatment.

Common upregulated pathways	Common downregulated pathways
3-phosphoinositide Biosynthesis	PTEN Signaling
3-phosphoinositide Degradation	Protein Kinase A Signaling
ABRA Signaling Pathway	Adenosine Nucleotides Degradation II
Cell Cycle: G1/S Checkpoint Regulation	Role of BRCA1 in DNA Damage Response
Colorectal Cancer Metastasis Signaling	Leptin Signaling in Obesity
D-myo-inositol (1,4,5,6)-Tetrakisphosphate Biosynthesis	Th1 Pathway
D-myo-inositol (3,4,5,6)-tetrakisphosphate Biosynthesis	Th2 Pathway
D-myo-inositol-5-phosphate Metabolism	Cyclins and Cell Cycle Regulation
Estrogen Receptor Signaling	LXR/RXR Activation
Ferroptosis Signaling Pathway	BER (Base Excision Repair) Pathway
Gustation Pathway	CLEAR Signaling Pathway
Hepatic Fibrosis Signaling Pathway	Mismatch Repair in Eukaryotes
IL-1 Signaling	Ribonucleotide Reductase Signaling Pathway
Insulin Secretion Signaling Pathway	Estrogen-mediated S-phase Entry
Macrophage Alternative Activation Signaling Pathway	Role of Hypercytokinemia/hyperchemokineemia in the Pathogenesis of Influenza
Neuregulin Signaling	Macrophage Classical Activation Signaling Pathway
Neutrophil Extracellular Trap Signaling Pathway	Cell Cycle Control of Chromosomal Replication
NF- κ B Activation by Viruses	
p53 Signaling	
PAK Signaling	
Phagosome Formation	
RAR Activation	
Regulation of Cellular Mechanics by Calpain Protease	
RHOA Signaling	
Role of CHK Proteins in Cell Cycle Checkpoint Control	
Role Of Chondrocytes In Rheumatoid Arthritis Signaling Pathway	
Role of NFAT in Cardiac Hypertrophy	
Role Of Osteoclasts In Rheumatoid Arthritis Signaling Pathway	
Role of p14/p19ARF in Tumor Suppression	
Role of Tissue Factor in Cancer	
S100 Family Signaling Pathway	
Semaphorin Neuronal Repulsive Signaling Pathway	

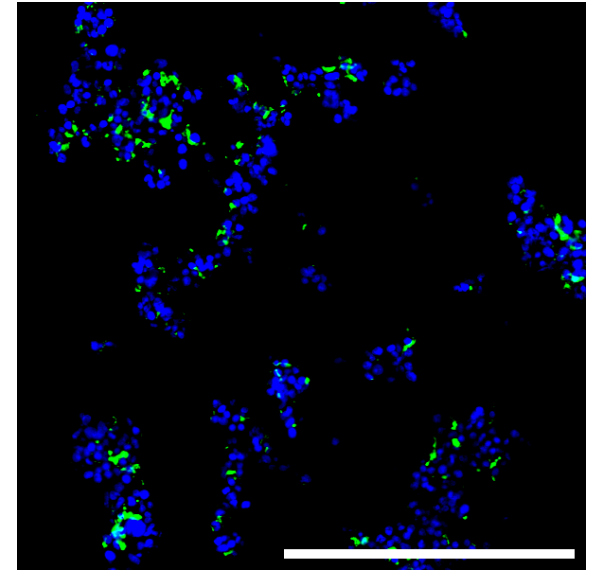
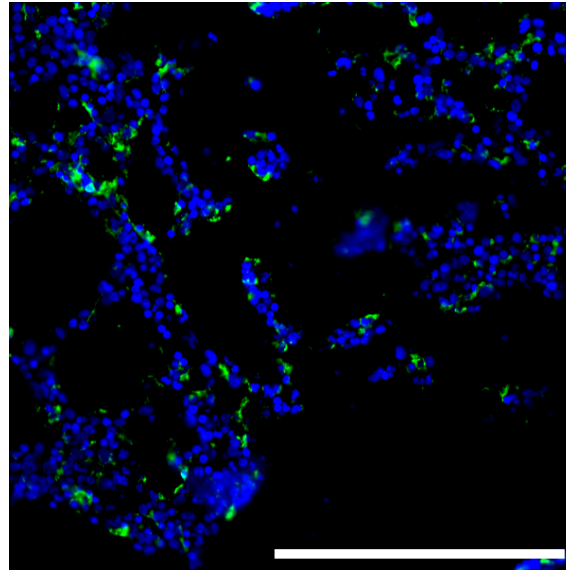
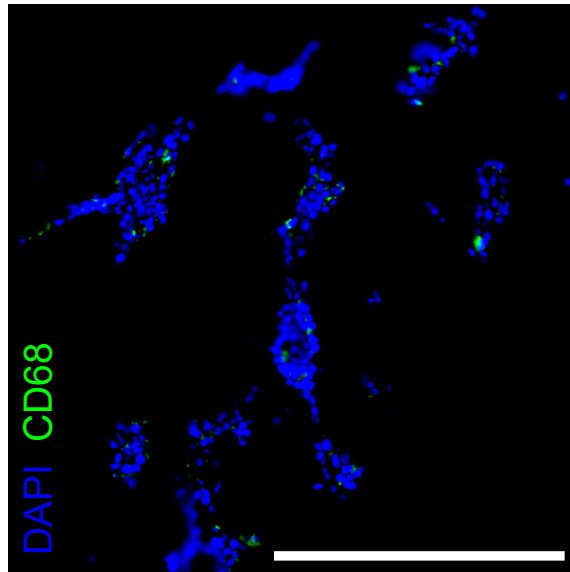
Senescence Pathway	
Superpathway of Inositol Phosphate Compounds	
WNT/Ca ⁺ pathway	

A.

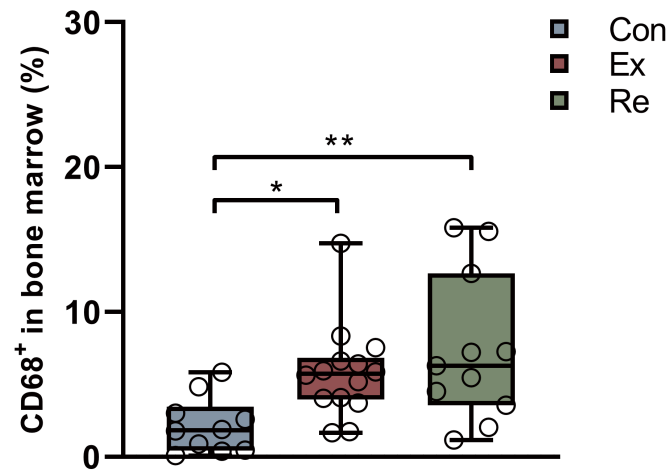
Control

PJI Explantation (Ex)

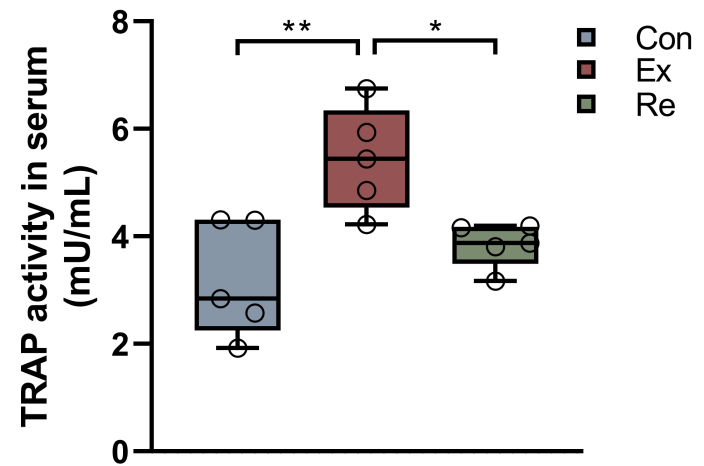
PJI Reimplantation (Re)

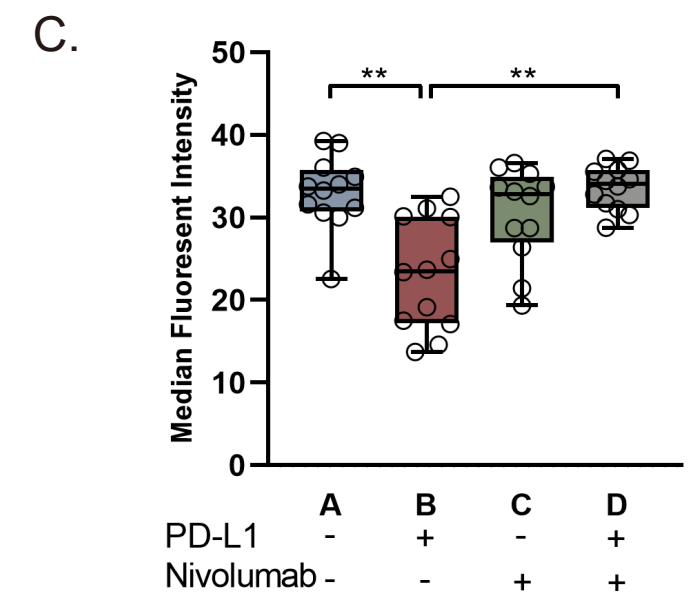
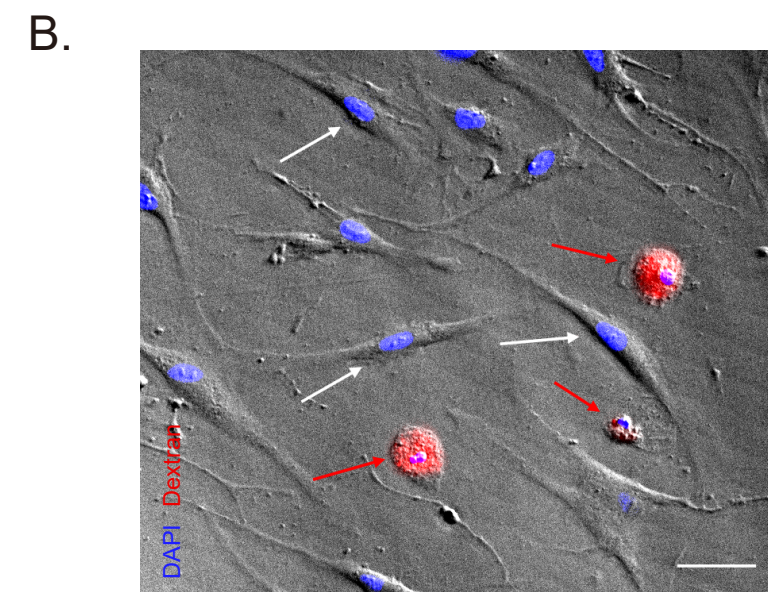
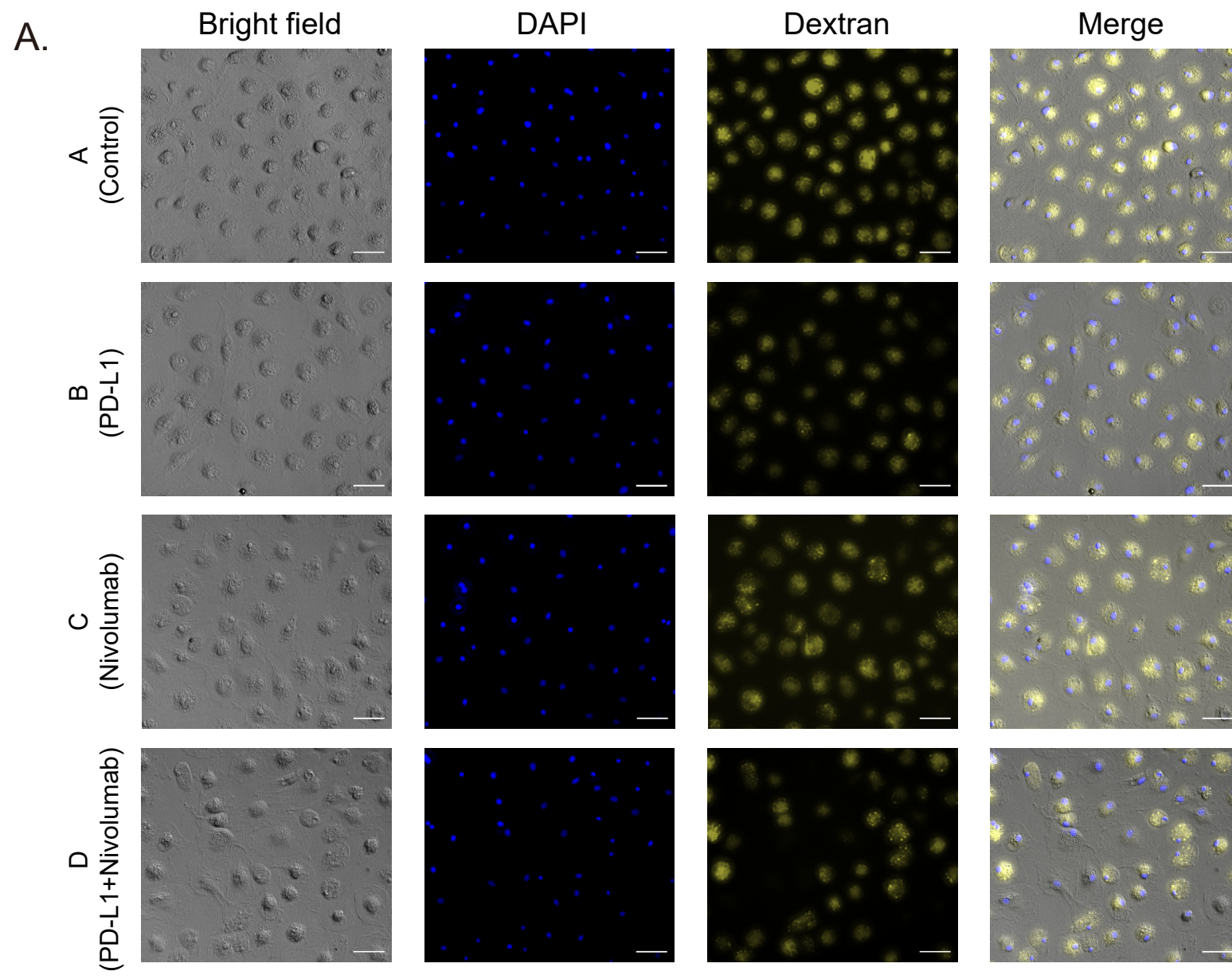


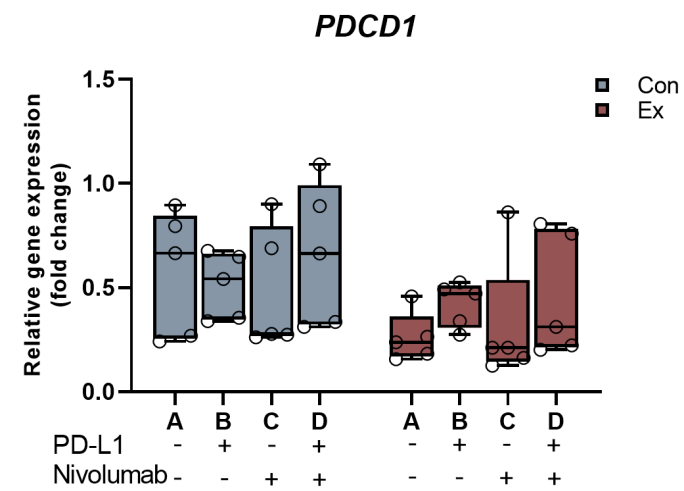
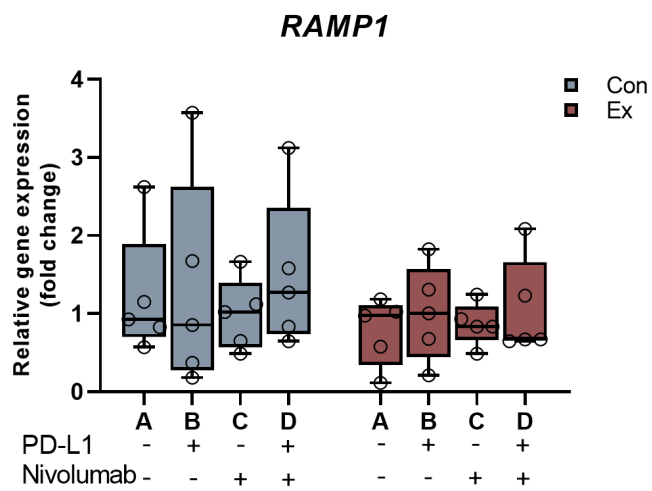
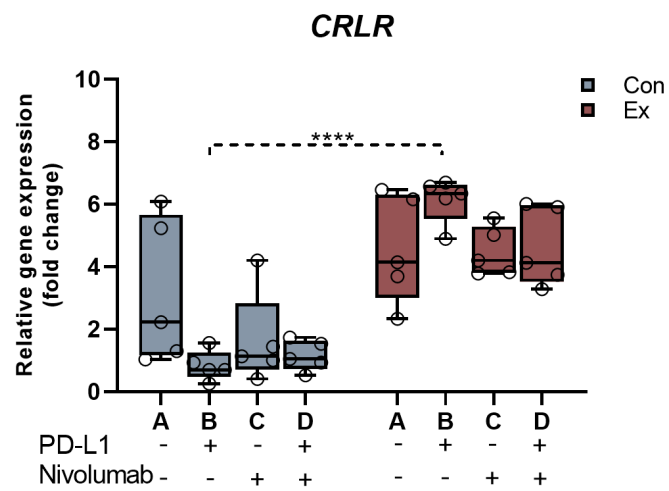
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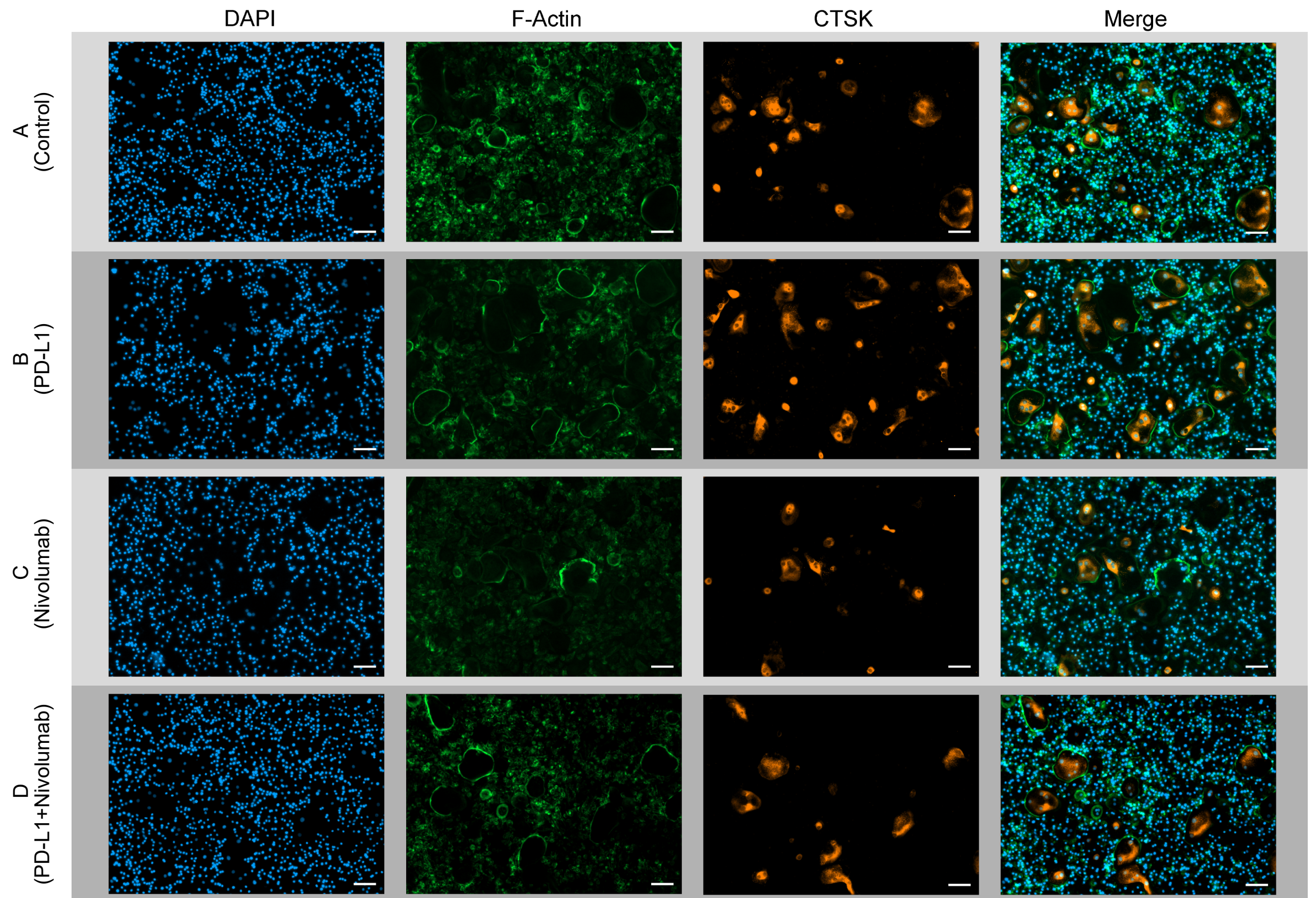
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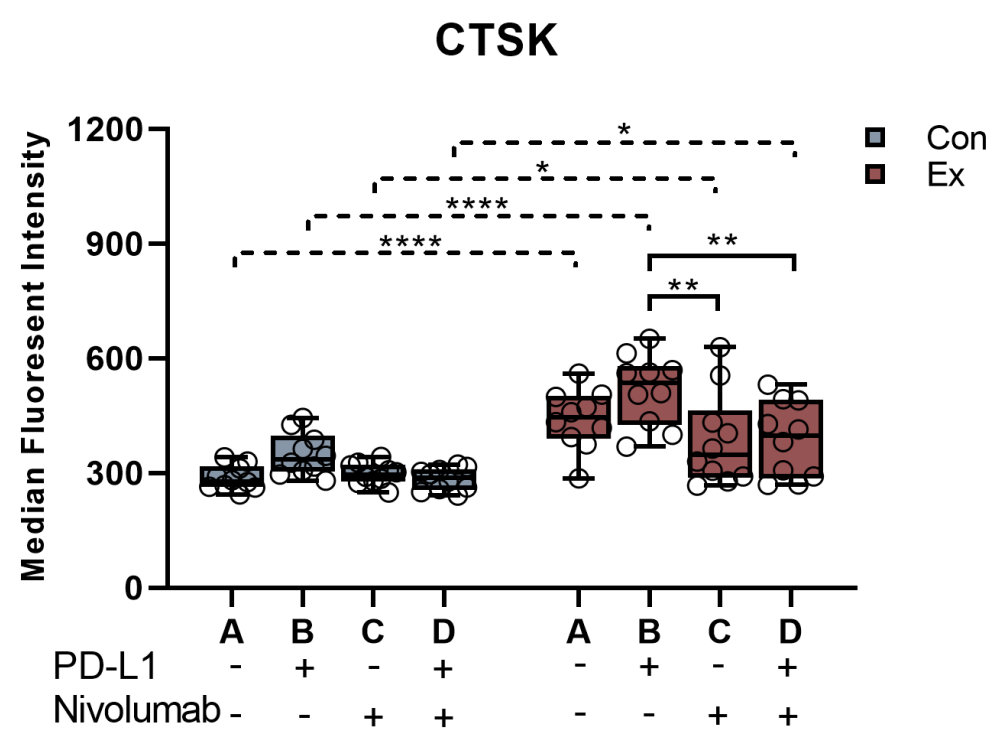




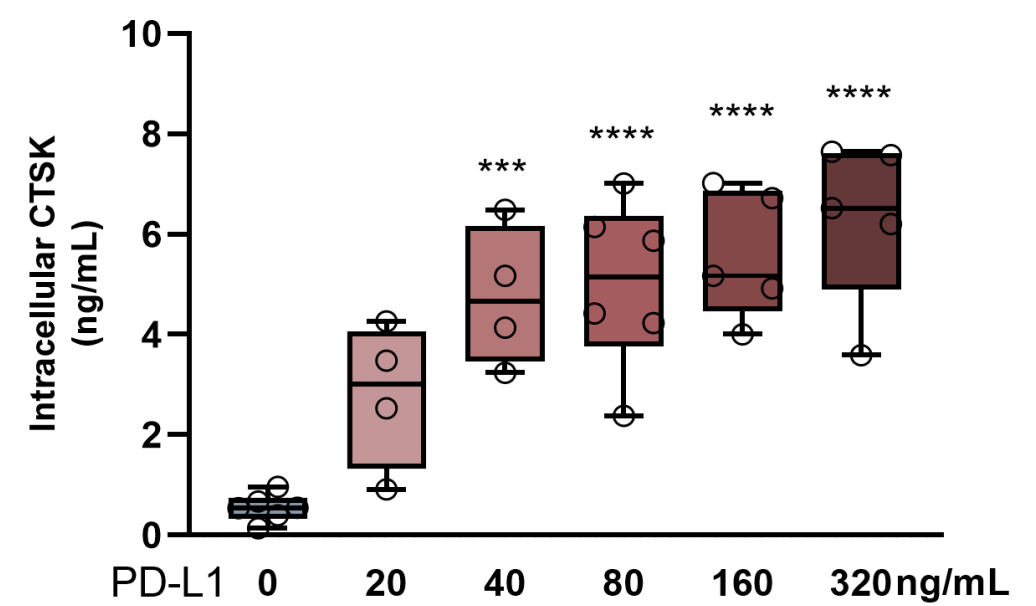
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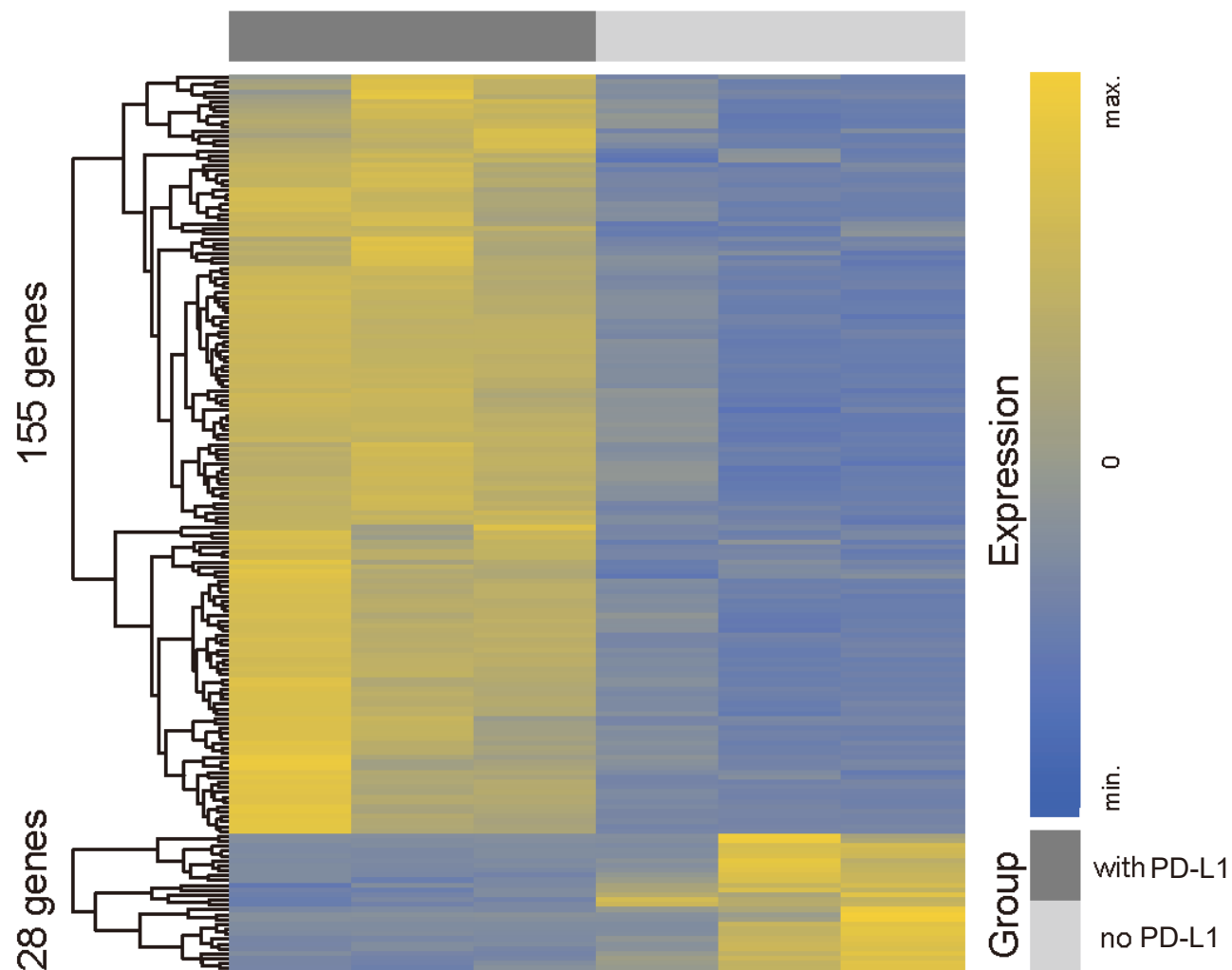
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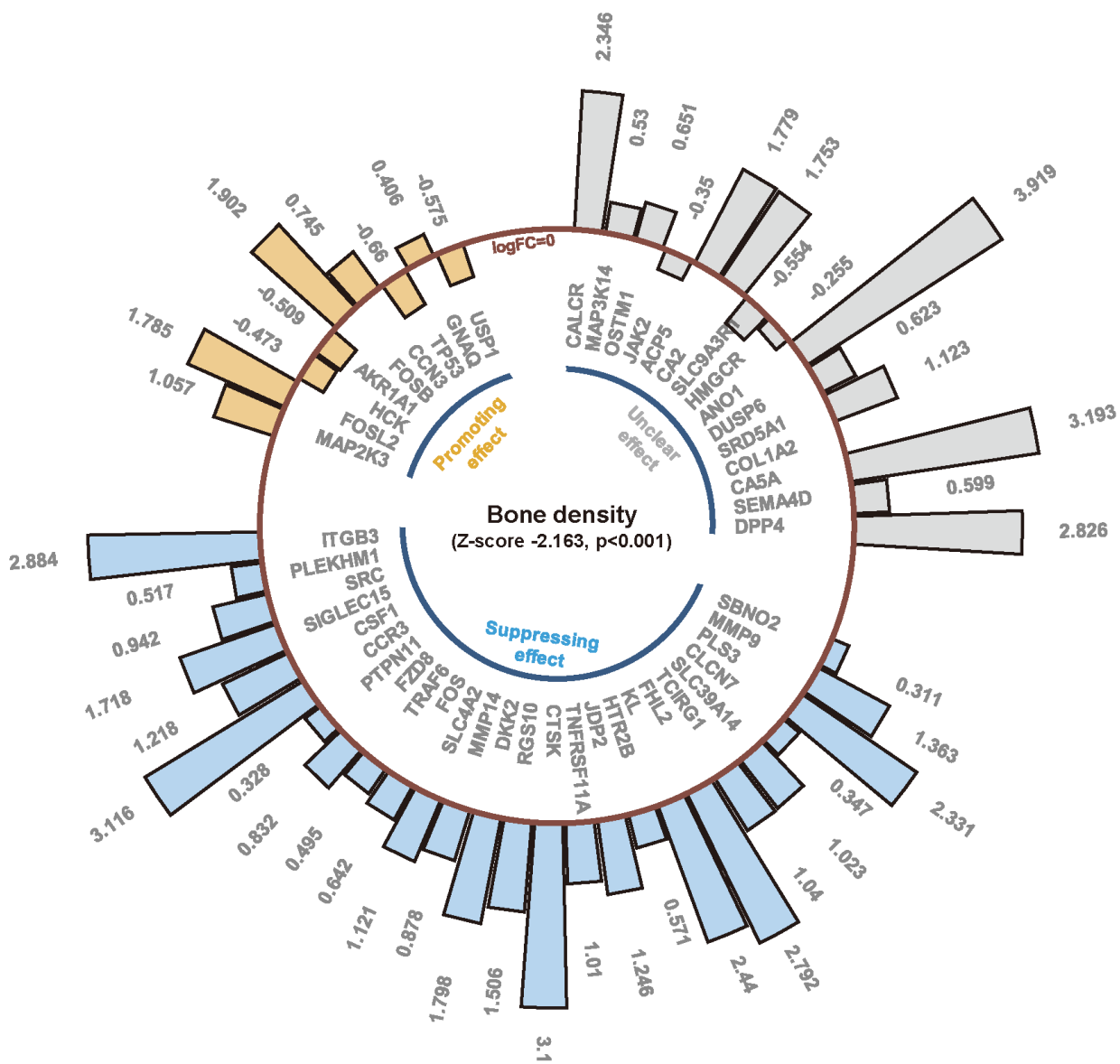
C.



A.



B.



Supplement figure 1: (A) Representative images of bone marrow specimens showing distribution of macrophages (CD68 positive cells). Scale bar: 200 μ m. (B) Quantification of macrophages in bone marrow. (C) TRAP activity test in plasma among three groups.

Supplement figure 2: (A, B) Representative fluorescent imaging of monocytes after treatment with FITC-labeled dextran at day 7. White arrows, BMSCs; red arrows, monocytes. (C) Median fluorescent intensity of FITC in monocytes acquired in cell culture plate at day 7. BMSCs, bone marrow mesenchymal stem cells.

Supplement figure 3: Quantification of expression of *RAMP1*, *CRLR*, and *PDCD1* in osteoclasts by RT-qPCR indicate osteoclast function.

Supplement figure 4: (A, B) Immune cytochemistry of CTSK in cultured osteoclasts from monocytes in patients at explantation surgery and quantification. Scale bar: 100 μ m. (C) Quantification of intracellular CTSK of osteoclasts stimulated with gradient concentrations of PD-L1.

Supplement figure 5: (A) Heatmap plot demonstrating distinct gene expression patterns between osteoclasts stimulated with PD-L1 and without stimulation. Yellow and blue indicates relative up- and downregulation, respectively. (B) Two downregulated disease modules consisting of a list of contributing genes, including those with promoting effect, suppressing effect and unclear effect. IPA, ingenuity pathway analysis.