Original Russian text www.bionet.nsc.ru/vogis/

# Comparative analysis of allele frequencies for DNA polymorphisms associated with disease and economically important traits in the genomes of Russian and foreign cattle breeds

A.V. Igoshin<sup>1</sup>, G.A. Romashov<sup>1</sup>, E.N. Chernyaeva<sup>2</sup>, N.P. Elatkin<sup>2</sup>, N.S. Yudin<sup>1</sup>, D.M. Larkin<sup>3</sup>

<sup>1</sup> Institute of Cytology and Genetics of the Siberian Branch of the Russian Academy of Sciences, Novosibirsk, Russia

<sup>2</sup> LLC "Miratorg-Genetika", Moscow, Russia

<sup>3</sup> Royal Veterinary College, London, United Kingdom

🖾 dmlarkin@gmail.com

Abstract. The genetic makeup of a breed including its genetic differences from other breeds determines its appearance and characteristics, including economically important traits and resistance to pathologies. To date, many loci controlling significant phenotypes have been identified, which is successfully used in the world practice of marker-assisted selection to improve breed properties. The aim of this study was a comparative analysis of frequencies for known causative nucleotide substitutions, insertions and deletions associated with disease and economically important traits in Russian and foreign cattle breeds. As a result, we identified frequencies of these DNA polymorphisms in the populations of Russian cattle breeds, compared them with those of foreign populations of the same breed, as well as other foreign breeds. Our results indicate similarities in frequencies for most of such alleles within breeds (populations of Russian and foreign breeding), as well as the relationship between the causative allele prevalence and the presence of phenotypic traits under the effect. We also found an excess of some undesirable alleles in the Russian cattle populations, which should be paid attention to when designing breeding programs. We found that the alleles increasing fertility in the Hereford breed have a higher frequency in the Russian Hereford population compared to the foreign counterpart. Interestingly, unlike for the European breeds, for Asian Turano-Mongolian Wagyu and Yakut cattle, there was a less clear link between phenotypic traits and frequencies of known causative alleles. Our work points to specific genetic variants that could be used to improve and/or maintain the performance of certain cattle breeds bred in the Russian Federation. Key words: cattle; selection; breed; Russian Federation; genetic variants; SNP, insertion; deletion.

**For citation:** Igoshin A.V., Romashov G.A., Chernyaeva E.N., Elatkin N.P., Yudin N.S., Larkin D.M. Comparative analysis of allele frequencies for DNA polymorphisms associated with disease and economically important traits in the genomes of Russian and foreign cattle breeds. *Vavilovskii Zhurnal Genetiki i Selektsii = Vavilov Journal of Genetics and Breeding*. 2022;26(3):298-307. DOI 10.18699/VJGB-22-28

# Сравнительный анализ частот ДНК-полиморфизмов, ассоциированных с заболеваниями и хозяйственно важными признаками, в геномах российских и зарубежных пород крупного рогатого скота

А.В. Игошин<sup>1</sup>, Г.А. Ромашов<sup>1</sup>, Е.Н. Черняева<sup>2</sup>, Н.П. Елаткин<sup>2</sup>, Н.С. Юдин<sup>1</sup>, Д.М. Ларкин<sup>3</sup>

Федеральный исследовательский центр Институт цитологии и генетики Сибирского отделения Российской академии наук, Новосибирск, Россия

<sup>2</sup> ООО «Мираторг-Генетика», Москва, Россия

<sup>3</sup> Королевский ветеринарный колледж, Лондон, Великобритания

🖾 dmlarkin@gmail.com

Аннотация. Генетический состав породы и ее генетические отличия от других пород определяют ее облик и характерные особенности, включая экономически важные признаки и встречаемость патологий. К настоящему времени выявлено множество локусов, контролирующих наиболее значимые фенотипы, что успешно используется в мировой практике для маркер-ассоциированной селекции в целях улучшения свойств пород. В настоящей работе проведен сравнительный анализ частот известных каузативных нуклеотидных замен, вставок и делеций, связанных с заболеваниями и хозяйственно ценными признаками, в российских и зарубежных породах крупного рогатого скота. Выявлены частоты вышеуказанных ДНК-полиморфизмов в популяциях российских пород крупного рогатого скота, выполнено их сравнение с частотами в зарубежных популяциях для пород, разводимых в Российской Федерации, а также с другими зарубежными породами. Наши результаты показывают схожесть частот большинства аллелей внутри пород (российского или зарубежного разведения), а также связь между представленностью аллелей исследуемых полиморфизмов и наличием определяемых ими фенотипических признаков. Были найдены и превышения по частотам ряда нежелательных аллелей в российских популяциях крупного рогатого скота, на которые стоит обратить внимание при селекционной работе с породами. Обнаружено, что аллели, отвечающие за повышенную фертильность породы герефорд, имеют повышенную частоту в популяциях российского разведения по сравнению с зарубежными популяциями. Интересно, что для азиатских турано-монгольских вагю и якутского скота наблюдалась меньшая связь между фенотипическими признаками и частотами известных каузативных аллелей по сравнению с европейскими породами. Наша работа указывает на конкретные генетические варианты, которые могут быть использованы для улучшения и/или поддержания качеств ряда пород крупного рогатого скота, разводимых в Российской Федерации.

Ключевые слова: крупный рогатый скот; селекция; порода; Российская Федерация; генетические варианты; SNP; инсерция; делеция.

#### Introduction

Common types of genetic variations, such as single nucleotide polymorphisms, nucleotide insertions and deletions, among others, can have "beneficial" or "harmful" effects on animal health and productivity (Liu, Bickhart, 2012; Bourque et al., 2018). That is why the sequencing of the Bos taurus genome caused a surge in research on the genetic diversity of cattle breeds and its relationship with economically important traits, adaptations and diseases, which opened up opportunities to use the knowledge gained for creating breeds with the necessary qualities and improving existing breeds (Larkin, Yudin, 2016; Yudin, Larkin, 2019). Now, according to the OMIA database (www.omia.org; Lenffer et al., 2006), 272 bovine traits are known to be genetically controlled, including a number of diseases. For 175 of them, causative mutations in the coding and non-coding regions of DNA have already been identified, the effect of which is related to various mechanisms, including changes in the protein sequence, in the stability, expression or processing of RNA (Ibeagha-Awemu et al., 2008; Yudin, Voevoda, 2015; Ciepłoch et al., 2017). Using this information, tests were developed for genotyping pathological mutations and removing carrier animals from the breeding herd (Romanenkova et al., 2015; Fornara et al., 2019; Sabetova et al., 2021). With this approach, it is possible to identify mutations at an early age for the timely culling of animals or embryos (Terletskiy et al., 2016). At the same time, it is worth considering that a "harmful" mutation may be "useful" for another economically important trait (Fasquelle et al., 2009). Identification of gene alleles associated with economically important traits allowed using them for marker-assisted selection (Pighetti, Elliott, 2011; Abd El-Hack et al., 2018). Marker-assisted selection is particularly important for traits that become evident with age or only in animals of the same sex, such as productivity or fertility (Zinovieva, 2016; Raina et al., 2020).

So far, Russian cattle breeds have been investigated for the presence of only a few, the most common mutations associated with economically important traits and health (Romanenkova et al., 2016, 2018; Usova et al., 2017; Surzhikova et al., 2019). The purpose of our work was to analyze the spectrum and frequencies of known causative DNA polymorphisms in nine Russian cattle breeds using genome sequencing data and to compare the frequencies of these polymorphisms with those

in worldwide breeds or foreign populations of the same breeds to determine the options for which the selection in Russian cattle could be conducted.

### Materials and methods

The list of single-nucleotide polymorphisms (SNPs), insertions and deletions, clinically and economically important for cattle, was compiled based on the information from the OMIA database (www.omia.org; Lenffer et al., 2006) and practical guidance of the Irish Cattle Breeding Federation (McClure M., McClure J., 2016). The genomic positions of polymorphisms specified in the Bos taurus UMD3.1 assembly coordinates were converted to the ARS-UCD1.2 assembly coordinates using liftOver (Kuhn et al., 2013). For polymorphisms present in the sample of Russian breeds, reference and alternative alleles were verified for matching those specified in the publications. For four substitutions of the twelve possible (T $\leftrightarrow$ A and G $\leftrightarrow$ C), such a verification is complicated, since: (1) there may be a change in the reference allele during the transition to a new genome assembly; (2) in the publication, the allele can be specified for a chain which is complementary to reference sequence. In such cases, we verified the alleles of polymorphisms in the context of codons (for substitutions in the coding sequence) or proximate sequences. For example, according to Hirano and colleagues (2013) and the OMIA database, replacing the nucleotide G with C at the BTA8:83909754 position, leading to the replacement of valine with leucine, results in perinatal weak calf syndrome. However, apparently, this replacement was indicated by the authors for the messenger RNA sequence since in the assembly ARS-UCD1.2 C stands for the reference nucleotide, being a part of the "AAC" triplet, which, in turn, corresponds to "GUU" mRNA codon, encoding valine. Thus, in the reference assembly of ARS-UCD1.2, the G allele will be "harmful".

In this paper, we used data on SNPs, insertions and deletions in the worldwide breeds from the "1000 Bull Genomes" Project (Hayes, Daetwyler, 2019), including the resequencing data of eight Russian breeds obtained earlier, as well as the resequencing data (".fastq"-files) for the Russian population of the Aberdeen Angus breed (hereinafter simply Angus), provided by LLC "Miratorg-Genetika". Of note, some of these animals were imported from the USA and Australia

#### Table 1. Breed analyzed

Breed	Geographic origin	Sample size
Altai	Russia	20
Buryat	Russia	19
Kalmyk	Russia	13
Kholmogory	Russia	32
Yakut	Russia	30
Yaroslavl	Russia	22
Aberdeen Angus (foreign)	Australia, Canada, New Zealand, USA etc.	401
Aberdeen Angus (Russian)	Russia (partially imported from USA and Australia)	46
Hereford (foreign)	Australia, Canada, New Zealand, USA etc.	123
Hereford (Russian)	Russia	18
Wagyu (foreign)	Australia	9
Wagyu (Russian)	Russia	20
Northern Finncattle	Finland	34
Western Finncattle	Finland	25
Eastern Finncattle	Finland	25
The rest (>180 populations/breeds)	-	4409

(Table 1). Additionally, we also used data on three native Finnish breeds provided by the Natural Resources Institute Finland (Luke). Finland borders with Russia and has a largely similar (although milder) climate, so the inclusion of Finnish breeds in the study could shed light on features of the selective breeding manifesting in the close natural conditions of the two countries.

Removal of adapter sequences from raw paired reads was performed using Trimmomatic-0.39. Clean reads were aligned to the ARS-UCD1.2 reference sequence using BWA-MEM v.0.7.17 (Li, Durbin, 2009). Files containing aligned sequences (".sam"-files) were then converted to the ".bam" format and sorted using the SAMtools v.1.8 software (Li et al., 2009). Further, libraries belonging to the same animal were pooled using the 'MergeSamFiles' module of the Picard v.2.18.2 package (http://broadinstitute.github.io/ picard). Duplicates were marked using the 'MarkDuplicates' module of the above-mentioned software. The OPTICAL DUPLICATE PIXEL DISTANCE parameter equaling 2500 was chosen according to the recommendations of the "1000 Bull Genomes" protocol. Base quality score recalibration was performed using the 'BaseRecalibrator' and 'PrintReads' modules of the GATK v.3.8 package (McKenna et al., 2010) using data provided by the "1000 Bull Genomes" Project (Hayes, Daetwyler, 2019). The variant calling and the merging of the resulting gVCF files were performed using the 'HaplotypeCaller' and 'GenotypeGVCFs' modules of the GATK v.3.8 program, respectively.

Extraction of SNPs, insertions, and deletions from genomewide VCF files was performed with the Tabix utility (Li, 2011), using the coordinates of polymorphisms from a previously generated list. The resulting VCF files containing the selected polymorphisms were used to calculate the frequencies of alternative alleles in the samples using the PLINK 2.0 program (Purcell et al., 2007) with the following parameters: --vcf --chrset 30 --freq --pheno --loop-cats. The count has been carried out for (1) breeds bred in Russia (Kholmogory, Yaroslavl, Altai, Yakut, Buryat, Kalmyk, Angus, Wagyu and Hereford), (2) foreign populations of those breeds (if present), (3) three Finnish breeds (Northern Finncattle, Western Finncattle and Eastern Finncattle), and (4) a combined sample of all other worldwide cattle breeds (see Table 1).

The presence of allele frequency differences between the abovementioned samples was tested using Fisher's exact test implemented in the 'fisher.test()' R function. Contingency tables  $2\times2$  were composed by counting the number of reference and alternative alleles in the chromosomal pool of each of the two groups studied. Three types of comparisons were made: (1) between a breed bred in Russia (or a foreign population of the same breed, if present) and a combined sample of other world breeds; (2) between a breed bred in Finland and the combined sample of the world's breeds using the polymorphisms identified in the first type of comparisons; (3) only between the Russian population and the foreign population of the same breed. To correct for multiple comparisons, we used the Storey and Tibshirani method

(Storey, Tibshirani, 2003) implemented in the 'qvalue()' R function (Storey et al., 2020).

### Results

Our list of clinically and economically important polymorphisms contained 193 SNPs and 63 insertions/ deletions. A search in the VCF files revealed in Russian breeds the presence of 38 SNPs and one insertion from the above-mentioned list (Supplementary Table 1)<sup>1</sup>, which corresponded to at least 21 phenotypic traits.

When comparing 15 populations for 39 polymorphisms (585 comparisons in total) with a global sample, in 229 cases statistically significant (q < 0.05) differences in allele frequencies were found (see the Figure). The most significant differences with the total sample of worldwide breeds were observed for foreign populations of Angus and Hereford breeds (29 and 27 loci, respectively). Of the Russian populations, the Yakut breed had the largest number (16 loci) of differences from the worldwide sample. Of the Finnish breeds, the Northern Finncattle had the largest number (20 loci) of such differences.

The most significant (q = 4.24E-286) allele frequency difference from the global sample was observed for the foreign Angus population for SNP rs109688013 in the melanocortin-1 receptor gene MCIR, carriers of the alternative allele C of which have a black coat color (Klungland et al., 1995). The difference from the worldwide sample for this locus was also statistically significant for most of other populations as well, with the exception of the Northern Finncattle, as well as Russian and foreign Wagyu populations. In particular, the difference at this SNP was the highest among 39 loci for the Russian population of Angus (q = 6.01E-35), both populations of Herefords (q = 6.22E-37 for foreign and 7.34E-07 for Russian), for Altai (q = 1.99E-06), Kholmogory (q = 9.27E-12) and Yaroslavl (q = 2.76E-06) breeds. In foreign and Russian Angus populations, the frequency of the C allele coding for black color reaches 0.973 and 0.989, while in other worldwide breeds it is 0.339. In the populations of Altai, Kholmogory, Yaroslavl breeds, Russian and foreign Herefords, it has a frequency of 0.026, 0.828, 0.772, 0 and 0.019, respectively. In Finnish breeds, the frequency of the C allele varies from zero in Western Finncattle to 0.052 in Eastern Finncattle and 0.258 in Northern Finncattle.

Of the remaining loci, the greatest difference in the studied breeds from the global cattle population was observed for polymorphisms associated with milk traits, coat color and bleeding disorders. Thus, the Russian Wagyu population had the most significant (q = 6.44E-21) allele frequency difference from the worldwide sample for 15 bp insertion located at BTA27:16305660, which disrupts the *F11* gene function and, as a result, leads to a deficiency of blood coagulation factor XI, encoded by this gene (Kunieda et al., 2005). In Russian Wagyu population, the frequency of this insertion reaches 0.25, while in the global cattle population it is close to zero. The most significant differences from the worldwide sample for the foreign Wagyu population (q = 2.60E-05) and the Yakut breed (q = 2.21E-18) were observed for SNP rs210634530 in the gene of microphthalmia-associated transcription factor MITF, which is associated with the 'white spotting' phenotype (Fontanesi et al., 2012). The frequencies of the 'white spotting'-associated allele T in the Yakut breed and foreign Wagyu population are 0.083 and 0.111, respectively, while in the worldwide sample it reaches 0.65. In the Buryat and Kalmyk breeds, the most significant difference (q = 6.81E-10 and 2.33E-06, respectively) had SNP rs109191047 in the growth hormone gene *GH1*, associated with the composition of milk (Mullen et al., 2010). The frequency of G allele increasing the milk fat and protein content is 0.100 in the worldwide population, while in the above-mentioned breeds it reaches 0.526 and 0.500, respectively.

Comparisons between the Russian and corresponding foreign populations, made for the Angus, Hereford and Wagyu breeds, revealed four loci, statistically significantly (q < 0.05) differing in allele frequencies. Of these, three SNPs (rs43703017, rs43703015 and rs110014544) had differing frequencies in the Russian and foreign Angus populations and specified the alleles of the kappa-casein gene CSN3. One SNP located in the CAPN1 gene (rs17872050) differed between the Hereford populations and was associated with meat tenderness. Taking into account the frequency differences at the nominal significance level (p < 0.05), eight additional loci can be noted (Table 2), among which the V311A missense substitution (BTA26:34340886T>C) in the NHLRC2 gene differing between the Angus populations and in homozygotes leading to notomelia, a type of polymelia in which the additional limb is located along or near the midline of the back (Beever et al., 2014).

#### Discussion

#### **Breed-specific genetic features**

The gene pool of farm animals is formed under the influence of factors such as selection for productive traits, adaptation to environmental conditions, hybridization, *de novo* mutations, the founder effect and genetic drift (Notter, 1999; Xu et al., 2015).

As we showed above, a significant part of the polymorphisms taken into the analysis in the studied breeds differs in frequencies from the "worldwide average", reflecting the gene pool features of particular populations. For example, the Yakut cattle shows the highest divergence in allele frequencies among Russian breeds, expressed both in a greater number of differing loci and in a greater significance of these differences, which is consistent with the data on phylogeny of this breed and the analysis of its population structure (Yurchenko et al., 2018; Buggiotti et al., 2021).

Some of the polymorphisms studied make a definitive contribution to characteristic features of the breeds. For example, the content of the *MC1R* gene allele rs109688013-C in the breeds coincides well with the typical color of their representatives. Thus, in Angus having a black coat color, the frequency of this allele is close to one. In Yaroslavl and Kholmogory cattle, rs109688013-C also predominates, apparently defining black and black-mottled coats, mainly characteristic of these animals. At the same time, in Herefords, which are not characterized by a black color, the frequency of the C allele is close to zero. Similarly, there is a link between color and the frequency of the C allele in populations of Finnish breeds. In breeds that have mainly lighter coats (fawn,

<sup>&</sup>lt;sup>1</sup> Supplementary Tables 1 and 2 are available in the online version of the paper: http://vavilov.elpub.ru/jour/manager/files/Suppl\_Igoshin\_engl.pdf

	Gene						Allele	frequ	uencie	25							Phenotype
F11										****	*						Factor XI deficiency
IARS										***	*						Perinatal weak calf syndrome
NHLRC2								***	****								Developmental duplications
ROR2		*	*			*		**	*****				***	*	*	*	Interdigital hyperplasia
LRP4		*		*	*				*								Syndactyly (mule foot)
TLR4			*			**	*		****	**	*	*	***	**	**	**	Sysceptibility to infections
STAT1								*	*				****	**	**		Decreased embryo survival rate
STAT3					*	**	*	**	****	*	*	*	*	*	*		Decreased embryo survival rate
STAT5A					****					*	*	*	***		***		Reduced fertility
STAT5A		*			****		*	*	****				**		*		Reduced fertility
STAT5A					****					*	*		***	*	****		Reduced fertility
STAT3					****	*			*		*	*	***		***	*	Reduced fertility
CAST			**	**	**	*			****								More tender meat
CAST				*	****	*			****								More tender meat
CAST									****	*		*	*	*		*	More tender meat
CAPN1			*	**			*		**				**		*	**	More tender meat
CAPN1			*						***				**				More tender meat
CAPN1		*	**		*				**		*		***				More tender meat
CSN2									*								CSN2*F allele
CSN2		***	*		*	*		*	****				***				Milk more favourable for cheese making
CSN2									*				*				Decreased milk protein vield
CSN2		***	**					***	****	**			****		*		A2-milk
CSN3		**															CSN3*I allele
CSN3							*										CSN3*G1 and CSN3*H alleles
CSN3				*					****				*	*	***	*	Milk more favourable for cheese making
CSN3				*					****				*	*	***	*	Decreased kappa case in concentration
CSN3								****	****				**				Less favourable milk coagulation properties
CSN3				*					*****				*	*	***	*	Milk more favourable for cheese making
GH1						***	***	**	*****	*			***		*		Increased milk fat and protein content
GH1		*				*				***				*	***		Decreased milk protein vield
GHR			**		*							*	***	*			Decreased milk fat vield
LGB		*			***		**	***	*****			**	***	**		**	Milk more favourable for cheese making
LGB		Ŷ			***	Ĵ		***	*****	Ŷ		**	+++	•••	Ŷ	**	Milk more favourable for cheese making
LGB								***	*****			**	+++	Ĵ.	- ++	***	Milk more favourable for cheese making
LGB					***			***	*****			**	***		**	**	Milk more favourable for cheese making
KRT27		î		+++						Ŷ		**		<u></u>			
MC1R		****	***	***	****	**		*****	*****			***	*****	**		***	Dominant black: rat tail syndrome
MITF		*		*	****	**			**	***	**	***	****		***	**	White spotting
	5	^ >	<b>ب</b>		t t	÷.	×	-	-	-	-		~	a	۵ ۵	e U	white spotting
	Norlc	uobc	oslav	Alta	Yaku	lurya	almy	ssian	reign	ssian	reign	ssian	'eign	cattl	cattl	cattle	
	_	olme	Yar			È	¥	s (Ru.	s (foi	ı (Ru	n (foi	ł (Ru:	d (for	Finn	Finn	Finn	
		수						ngu	Ngu	ʻagyı	/agyı	eforc	eforc	tern	Jern	tern	
								Ä	4	\$	S	Her	Her	Eas	Nort	Wes	

Frequencies of clinically and economically significant polymorphisms in Russian and foreign cattle populations.

On the left are the genes containing the polymorphic variants under study. A darker tone corresponds to a higher frequency of a reference (green) or alternative (red) allele. On the right is the phenotype associated with this allele. The polymorphism designations and their frequencies are given in Suppl. Table 2. The asterisks indicate loci that have significant frequency differences between the specified breed and the global cattle population: \*\*\*\*\*\* q < 1.0E–25, \*\*\*\* q < 1.0E–10, \*\*\* q < 1.0E–5, \*\* q < 1.0E–3, \* q < 0.05. The frame indicates loci that differ (p < 0.05) between the Russian and foreign populations of the same breed: red color means that the "harmful" allele has a large content in the Russian population, blue – in the foreign, black – the significance of the allele for beef breeds is not established.

			5 1 1					
Locus	Allele		Frequency of associated we phenotype	of allele vith the specified	Breed	Phenotype	Statistical significance	
	Reference	Alternative	Russian population	Foreign population			<i>p</i> -value	q-value
rs43705173	G*	A	0.853	0.971	Hereford	Decreased embryo survival rate	0.0086	0.1465
rs43703015	T*	С	0.304	0.157	Angus	Decreased rennet coagula- tion time, decreased lactose concentration	0.0011	0.0353
rs43703016	C	А*	0.696	0.833	Angus	Decreased kappa casein concentration	0.0024	0.0563
rs43703017	A	G*	0.283	0.113	Angus	Less favourable coagulation properties and increased milk fat content	4.88E–05	0.0057
rs110014544	G*	A	0.304	0.159	Angus	Decreased rennet coagulation time	0.0012	0.0353
rs41255587	G*	A	0.620	0.725	Angus	More tender meat	0.0385	0.3753
rs109221039	A*	G	0.793	0.894	Angus	More tender meat	0.0088	0.1465
rs208753173	G*	A	0.917	0.988	Hereford	Reduced fertility	0.0298	0.349
rs110942700	Т	C*	0.083	0.267	Hereford	Decreased embryo survival rate	0.0201	0.262
BTA26:34340886	Т	C*	0.065	0.024	Angus	Developmental duplications	0.037	0.3753
rs17871051	G*	A	0.722	0.894	Hereford	More tender meat	0.0127	0.1854
rs17872050	C*	Т	0.500	0.799	Hereford	More tender meat	0.0003	0.0154

Table 2. Differences between Russian and foreign populations within the same breed

\* The allele associated with the phenotype specified.

light brown and red, often white muzzle, belly and back), it is low (0.053 in Eastern Finncattle) or zero (in Western Finncattle). In Northern Finncattle, which has a predominantly white coat color (some individuals are black-mottled), the frequency of rs109688013-C is 0.258. Breeds for which the red (Kalmyk) or brown (Altai, Buryat) colors are typical have rs109688013-C in low frequency (0.03-0.08). However, in Wagyu populations, which are usually characterized by black color, the frequency of this allele is far from one and has values of 0.42 in Russian population and 0.67 in foreign population, probably reflecting the genetic characteristics of Turano-Mongolian breeds. This discrepancy is also observed in Yakut cattle, in which a black-and-white color is common, but the frequency of rs109688013-C is vanishingly small. Given the genetic divergence of Turano-Mongolian breeds from other breeds, it can be assumed that other loci are involved in the control of body coloration.

Also, coat color is associated with the SNP rs210634530 in the *MITF* gene, the T allele of which defines the 'white spotting' phenotype. The highest frequency of rs210634530-T is observed in populations of Hereford cattle (fixed in the

Russian sample and 0.92 in the foreign population), which is characterized by a white head and belly. In addition, this allele prevails in the populations of Kholmogory, Yaroslavl, Altai and Kalmyk breeds, which have white spotting in color, as well as in Angus. In other populations, the frequency of the T allele varies from low (Yakut breed) to moderate (Buryat, Wagyu). The link between the content of rs210634530-T and coat color can be demonstrated for Finnish breeds. As mentioned above, many individuals of Western Finncattle and Eastern Finncattle have a white muzzle, back, and belly. Northern Finncattle has either a white or, less often, black-and-white coat. It should be noted that in addition to SNP rs210634530, additional loci appear to be involved in the control of the 'white spotting' phenotype (Fontanesi et al., 2012), therefore, the link between the frequency of rs210634530-T and coat color may not be so straightforward.

Some of the genetic features of the breeds are not quite obvious at first glance. For example, both Russian and foreign populations of Angus and Wagyu have a high (0.89–0.95) frequency of rs43703011-G allele of the beta-casein gene *CSN2*. Variations of the *CSN2* gene at several non-synonymous

positions determine its alleles – A1, A2, A3, B, C, etc. The above-mentioned allele G of rs43703011 is shared by several alleles of the CNS2 gene, the most common of which is A2. The so-called A2-milk is considered more preferable for consumption, due to better absorption and fewer undesirable effects from the human digestive system (Jianqin et al., 2016). In recent years, breeding programs in many countries have aimed to increase the frequency of the A2 allele in dairy cattle (Sebastiani et al., 2020). Given that Angus and Wagyu are beef breeds and are not used for milk production, the increased G allele content they have can hardly be explained by selection to improve milk quality. The most plausible explanation is selection for meat productivity. Thus, according to Hohmann et al. (2020), the carriage of the A2 allele increases average daily weight gain and weaning weight in German Angus and Simmentals. Therefore, increasing the frequency of the rs43703011-G allele, and consequently the A2 allele of the CSN2 gene, can be useful for improving not only dairy but also beef breeds.

Some of the variants found are specific to one breed and virtually absent in others. The most breed-specific are clinically important polymorphisms in the F11, IARS and NHLRC2 genes. The previously mentioned insertion in the F11 gene, leading to a deficiency of blood coagulation factor XI, is almost exclusively observed in foreign and Russian Wagyu populations. At the same time, among more than 5 thousand other animals from the "1000 Bull Genomes" Project, this mutation is harbored by only two animals. Association of factor XI activity with ATATGTGCAGAATAT insertion has been initially demonstrated for Wagyu (Kunieda et al., 2005). The homozygous genotype for this mutation is associated with a blood clotting disorder and an increase in the duration of bleeding. In the Russian population of Wagyu, its frequency equals 0.25, which is consistent with the data of early publications on its prevalence in the Japanese black breed (Watanabe et al., 2006; Ohba et al., 2008). At the same time, in the foreign Wagyu population, here represented by a sample from Australia, this insertion has a frequency of 0.11.

Other examples of breed-specific variants are single nucleotide substitutions in the IARS (BTA8:83909754C>G) and NHLRC2(BTA26:34340886T>C)genes.BTA8:83909754C>G mutation in the IARS gene in homozygote leads to perinatal weak calf syndrome and increased prenatal mortality (Hirano et al., 2013, 2016). This variant is specific to Wagyu, and besides, it was found only in one animal from the "1000 Bull Genomes" Project. In the Russian and Australian samples of this breed, its frequencies are 0.075 and 0.056, respectively. The BTA26:34340886T>C mutation in the NHLRC2 gene mentioned earlier, in homozygote leading to notomelia, is breed-specific for Angus, and was first discovered in this breed (Beever et al., 2014). Besides Angus, in the sample of "1000 Bull Genomes", the mutant allele is found only in one animal of an unknown ('crossbreed') breed. In the Russian and foreign populations of this breed, it has frequencies of 0.065 and 0.024, respectively.

# Differences between Russian and foreign populations of the same breed

The presence in our analysis of foreign Angus, Herefords and Wagyu populations can shed light on the features of the selection and adaptation of Russian populations of these breeds. Overall, Russian and foreign samples of the same breed demonstrate similar allele frequency profiles, with statistically confirmed differences present only in a small number of loci. The differences observed can be explained by many factors or their combinations. For example, an almost threefold excess of the BTA26:34340886-C allele (leading to the appearance of additional limbs) content in the Russian Angus population compared to the foreign one (see Table 2) may be a consequence of the founder effect or genetic drift in general, as well as less intensive efforts for elimination of this variant in the Russian herd.

Interpopulation differences in the loci associated with reproduction may result from an adaptation to environmental conditions. In the Russian population of Herefords, alleles of several polymorphisms that negatively affect the survival of embryos (rs43705173-G and rs110942700-C) and fertility (rs208753173-G) have a lower frequency compared to the foreign sample of this breed. It can be assumed that the Russian sample of Herefords, in this work represented by a population bred in Western Siberia since the 1960s (Vsyakikh, Kurinsky, 1976), was subject to selection for reproductive performance. This assumption is supported by the data of Afanasyeva and co-authors, according to which in the conditions of the Altai Territory, the population of Herefords of Siberian selection shows a much lower stillbirth rate (1.4%) compared to animals of Finnish selection (6.6%) imported in 2011 (Afanasyeva et al., 2015). Low temperatures are known to negatively affect the reproduction of cattle, reducing fertility and increasing perinatal mortality (Gwazdauskas, 1985; Mee, 2020). Therefore, population differences at these loci may reflect the process of genetic adaptation aimed at compensating a decrease in reproductive functions caused by cold.

Of particular interest are single nucleotide polymorphisms associated with meat traits and differing in samples of Angus (rs41255587 and rs109221039 in the *CAST* gene) and Herefords (rs17871051 and rs17872050 in the *CAPN1* gene). For all four SNPs, the foreign populations of these breeds demonstrate a higher content of alleles increasing meat tenderness. This is an important gastronomic feature and its improvement is included in the breeding programs of foreign beef breeds (Tatum, 2006). At the same time, we are not aware of extensive breeding attempts of this kind in Russia, which is probably the reason for the observed differences between the samples. Therefore, the Russian populations of Angus and Herefords have the potential for the improvement of meat quality by selection for *CAST* and *CAPN1* alleles.

Of the studied loci differing between populations of the same breed, four SNPs (rs43703015, rs43703016, rs43703017 and rs110014544) determining the kappa-casein gene *CSN3* alleles deserve to be noticed. Their allele frequencies differ between Russian and foreign Angus populations. These polymorphisms are associated with milk traits, in particular, with the concentration of kappa-casein in milk and milk coagulation properties, which is important for cheesemaking. At the same time, the effect of *CSN3* alleles on the productivity of beef cattle is poorly understood. Investigations of Tambasco et al. (2003)  $\mu$  Curi et al. (2005) found no association between *CSN3* alleles and meat traits. Thus, the observed differences can be attributed to the founder effect, or selection for

economically important traits whose associations with *CSN3* polymorphisms have not yet been identified.

# Polymorphisms of clinical significance present in Russian breeds

In Russian breeds, there is a number of polymorphic variants, in homozygous state causing hereditary diseases, some of which (mutations in the genes F11, IARS and NHLRC2) have already been discussed above due to their breed specificity. Also, the variants in the ROR2 and LRP4 genes should be mentioned that are associated with the manifestation of interdigital hyperplasia (proliferation of tissue between the hooves) and syndactyly (fusion of the fingers, also called 'mule foot'), respectively. Unlike the F11, IARS and NHLRC2 genes, the "harmful alleles" in ROR2 (rs377953295-A) and LRP4 (rs453049317-T) are not breed-specific, and are widespread both in Russian breeds and in the rest of the worldwide cattle population. Of the Russian populations, the Kalmyk (0.192) and Altai (0.15) breeds have the highest content of the rs37795322-A allele of the ROR2 gene. In the worldwide sample, its frequency reaches 0.13. The rs453049317-T variant in the LRP4 gene has the highest frequency in the Altai breed (0.2) and in the Russian Angus (0.12), while in the rest of the worldwide population it is 0.076.

Currently, testing for genetic defects is widely used in the practice of animal husbandry in many countries (Terletskiy et al., 2016). For example, testing for mutations in the *F11* and *IARS* genes is included in the genetic screening programs recommended by the Australian Wagyu Association (https://www.wagyu.org.au/content/uploads/2020/08/ Genetic-Conditions-in-Wagyu-FactSheet-2020.pdf). At the same time, the elimination of undesirable alleles should be approached with caution. For example, there is an assumption that the carriage of mutations associated with syndactyly improves the milk productivity of cows, which can partially explain the spread of this pathology in cattle (Johnson et al., 2006).

## Conclusion

Our analysis showed the allele frequency distribution for the most clinically and economically important DNA polymorphisms present in Russian cattle breeds. A number of variants leading to common hereditary disorders in cattle have significant representation in Russian populations, and probably need to be eliminated. Also, the differences between Russian and foreign cattle populations at several loci are presumably of adaptive importance. The data of this study may be useful in cattle breeding programs aimed at improving the existing cattle breeds, and creating new ones.

## References

- Abd El-Hack M.E., Abdelnour S.A., Swelum A.A., Arif M. The application of gene marker-assisted selection and proteomics for the best meat quality criteria and body measurements in Qinchuan cattle breed. *Mol. Biol. Rep.* 2018;45(5):1445-1456. DOI 10.1007/s11033-018-4211-y.
- Afanasyeva A.I., Knyazev S.S., Lotz K.N. Reproductive capacity of Hereford beef cattle of Siberian and Finnish breeding under the conditions of the Altai region. Vestnik Altayskogo Gosudarstvennogo Agrarnogo Universiteta = Bulletin of the Altai State Agricultural University. 2015;8(130):86-89. (in Russian)

- Beever J.E., Marron B.M., Parnell P.F., Teseling C.F., Steffen D.J., Denholm L.J. Developmental Duplications (DD): 1. Elucidation of the underlying molecular genetic basis of polymelia phenotypes in Angus cattle. In: Proc. XXVIII World Buiatrics Congress. Cairns, 2014.
- Bourque G., Burns K.H., Gehring M., Gorbunova V., Seluanov A., Hammell M., Imbeault M., Izsvák Z., Levin H.L., Macfarlan T.S., Mager D.L., Feschotte C. Ten things you should know about transposable elements. *Genome Biol.* 2018;19:199. DOI 10.1186/ s13059-018-1577-z.
- Buggiotti L., Yurchenko A.A., Yudin N.S., Vander Jagt C.J., Vorobieva N.V., Kusliy M.A., Vasiliev S.K., Rodionov A.N., Boronetskaya O.I., Zinovieva N.A., Graphodatsky A.S., Daetwyler H.D., Larkin D.M. Demographic history, adaptation, and NRAP convergent evolution at amino acid residue 100 in the world northernmost cattle from Siberia. *Mol. Biol. Evol.* 2021;38(8):3093-3110. DOI 10.1093/molbev/msab078.
- Ciepłoch A., Rutkowska K., Oprzkadek J., Poławska E. Genetic disorders in beef cattle: a review. *Genes Genomics*. 2017;39(5):461-471. DOI 10.1007/s13258-017-0525-8.
- Curi R.A., de Oliveira H.N., Gimenes M.A., Silveira A.C., Lopes C.R. Effects of CSN3 and LGB gene polymorphisms on production traits in beef cattle. *Genet. Mol. Biol.* 2005;28(2):262-266. DOI 10.1590/S1415-47572005000200015.
- Fasquelle C., Sartelet A., Li W., Dive M., Tamma N., Michaux C., Druet T., Huijbers I.J., Isacke C.M., Coppieters W., Georges M., Charlier C. Balancing selection of a frame-shift mutation in the *MRC2* gene accounts for the outbreak of the Crooked Tail Syndrome in Belgian Blue Cattle. *PLoS Genet.* 2009;5(9):e1000666. DOI 10.1371/journal.pgen.1000666.
- Fontanesi L., Scotti E., Russo V. Haplotype variability in the bovine *MITF* gene and association with piebaldism in Holstein and Simmental cattle breeds. *Anim. Genet.* 2012;43(3):250-256. DOI 10.1111/j.1365-2052.2011.02242.x.
- Fornara M.S., Kostyunina O.V., Filipchenko A.A., Sermyagin A.A., Zinovyeva N.A. Polymorphism determination system of gene SUGT1 associated with Fleckvieh fertility haplotype FH4. *Veterinariya, Zootekhniya i Biotekhnologiya = Veterinary Medicine, Zootechnics and Biotechnology.* 2019;3:92-97. DOI 10.26155/vet. zoo.bio.201903015. (in Russian)
- Gwazdauskas F.C. Effects of climate on reproduction in cattle. *J. Dairy Sci.* 1985;68(6):1568-1578. DOI 10.3168/ jds.S0022-0302(85)80995-4.
- Hayes B.J., Daetwyler H.D. 1000 Bull Genomes Project to map simple and complex genetic traits in cattle: applications and outcomes. *Annu. Rev. Anim. Biosci.* 2019;7:89-102. DOI 10.1146/annurevanimal-020518-115024.
- Hirano T., Kobayashi N., Matsuhashi T., Watanabe D., Watanabe T., Takasuga A., Sugimoto M., Sugimoto Y. Mapping and exome sequencing identifies a mutation in the *IARS* gene as the cause of hereditary perinatal weak calf syndrome. *PLoS One*. 2013;8(5):e64036. DOI 10.1371/journal.pone.0064036.
- Hirano T., Matsuhashi T., Takeda K., Hara H., Kobayashi N., Kita K., Sugimoto Y., Hanzawa K. *IARS* mutation causes prenatal death in Japanese Black cattle. *Anim. Sci. J.* 2016;87(9):1178-1181. DOI 10.1111/asj.12639.
- Hohmann L.G., Weimann C., Scheper C., Erhardt G., König S. Associations between maternal milk protein genotypes with preweaning calf growth traits in beef cattle. J. Anim. Sci. 2020;98(10):skaa280. DOI 10.1093/jas/skaa280.
- Ibeagha-Awemu E.M., Kgwatalala P., Ibeagha A.E., Zhao X. A critical analysis of disease-associated DNA polymorphisms in the genes of cattle, goat, sheep, and pig. *Mamm. Genome.* 2008;19(4):226-245. DOI 10.1007/s00335-008-9101-5.
- Jianqin S., Leiming X., Lu X., Yelland G.W., Ni J., Clarke A.J. Effects of milk containing only A2 beta casein versus milk containing both A1 and A2 beta casein proteins on gastrointestinal physiology, symptoms of discomfort, and cognitive behavior of people with self-

reported intolerance to traditional cows' milk. *Nutr. J.* 2016;15:35. DOI 10.1186/s12937-016-0147-z.

- Johnson E.B., Steffen D.J., Lynch K.W., Herz J. Defective splicing of *Megf7/Lrp4*, a regulator of distal limb development, in autosomal recessive mulefoot disease. *Genomics*. 2006;88(5):600-609. DOI 10.1016/j.ygeno.2006.08.005.
- Klungland H., Våge D.I., Gomez-Raya L., Adalsteinsson S., Lien S. The role of melanocyte-stimulating hormone (MSH) receptor in bovine coat color determination. *Mamm. Genome.* 1995;6(9):636-639. DOI 10.1007/BF00352371.
- Kuhn R.M., Haussler D., Kent W.J. The UCSC genome browser and associated tools. *Brief. Bioinform.* 2013;14(2):144-161. DOI 10.1093/bib/bbs038.
- Kunieda M., Tsuji T., Abbasi A.R., Khalaj M., Ikeda M., Miyadera K., Ogawa H., Kunieda T. An insertion mutation of the bovine *F11* gene is responsible for factor XI deficiency in Japanese black cattle. *Mamm. Genome.* 2005;16(5):383-389. DOI 10.1007/ s00335-004-2462-5.
- Larkin D.M., Yudin N.S. The genomes and history of domestic animals. Molecular Genetics, Microbiology and Virology. 2016;31(4):197-202. DOI 10.3103/S0891416816040054.
- Lenffer J., Nicholas F.W., Castle K., Rao A., Gregory S., Poidinger M., Mailman M.D., Ranganathan S. OMIA (Online Mendelian Inheritance in Animals): an enhanced platform and integration into the Entrez search interface at NCBI. *Nucleic Acids Res.* 2006;34(1):D599-D601. DOI 10.1093/nar/gkj152.
- Li H. Tabix: fast retrieval of sequence features from generic TABdelimited files. *Bioinformatics*. 2011;27(5):718-719. DOI 10.1093/ bioinformatics/btq671.
- Li H., Durbin R. Fast and accurate short read alignment with Burrows– Wheeler transform. *Bioinformatics*. 2009;25(14):1754-1760. DOI 10.1093/bioinformatics/btp324.
- Li H., Handsaker B., Wysoker A., Fennell T., Ruan J., Homer N., Marth G., Abecasis G., Durbin R. The Sequence Alignment/Map format and SAMtools. *Bioinformatics*. 2009;25(16):2078-2079. DOI 10.1093/bioinformatics/btp352.
- Liu G.E., Bickhart D.M. Copy number variation in the cattle genome. *Funct. Integr. Genomics.* 2012;12(4):609-624. DOI 10.1007/ s10142-012-0289-9.
- McClure M., McClure J. Genetic Disease and Trait Information for IDB Genotyped Animals in Ireland. Bandon: Irish Cattle Breeding Federation, 2016.
- McKenna A., Hanna M., Banks E., Sivachenko A., Cibulskis K., Kernytsky A., Garimella K., Altshuler D., Gabriel S., Daly M., DePristo M.A. The Genome Analysis Toolkit: a MapReduce framework for analyzing next-generation DNA sequencing data. *Genome Res.* 2010;20(9):1297-1303. DOI 10.1101/gr.107524.110.
- Mee J.F. Investigation of bovine abortion and stillbirth/perinatal mortality similar diagnostic challenges, different approaches. *Ir. Vet. J.* 2020;73:20. DOI 10.1186/s13620-020-00172-0.
- Mullen M.P., Berry D.P., Howard D.J., Diskin M.G., Lynch C.O., Berkowicz E.W., Magee D.A., MacHugh D.E., Waters S.M. Associations between novel single nucleotide polymorphisms in the *Bos taurus* growth hormone gene and performance traits in Holstein-Friesian dairy cattle. *J. Dairy Sci.* 2010;93(12):5959-5969. DOI 10.3168/jds.2010-3385.
- Notter D.R. The importance of genetic diversity in livestock populations of the future. J. Anim. Sci. 1999;77(1):61-69. DOI 10.2527/1999.77161x.
- Ohba Y., Takasu M., Nishii N., Takeda E., Maeda S., Kunieda T., Kitagawa H. Pedigree analysis of factor XI deficiency in Japanese black cattle. J. Vet. Med. Sci. 2008;70(3):297-299. DOI 10.1292/ jvms.70.297.
- Pighetti G.M., Elliott A.A. Gene polymorphisms: the keys for marker assisted selection and unraveling core regulatory pathways for mastitis resistance. J. Mammary Gland Biol. Neoplasia. 2011;16(4):421-432. DOI 10.1007/s10911-011-9238-9.

- Purcell S., Neale B., Todd-Brown K., Thomas L., Ferreira M.A.R., Bender D., Maller J., Sklar P., de Bakker P.I.W., Daly M.J., Sham P.C. PLINK: a tool set for whole-genome association and populationbased linkage analyses. *Am. J. Hum. Genet.* 2007;81(3):559-575. DOI 10.1086/519795.
- Raina V.S., Kour A., Chakravarty A.K., Vohra V. Marker-assisted selection vis-à-vis bull fertility: coming full circle – a review. *Mol. Biol. Rep.* 2020;47(11):9123-9133. DOI 10.1007/ s11033-020-05919-0.
- Romanenkova O.V., Gladyr E.A., Kostyunina O.V., Zinovieva N.A. Development of test system for diagnostics of cattle fertility haplotype HH3 associated with early embryonic mortality. *Dostizheniya Nauki i Tekhniki APK = Achievements of Science and Technology of AIC*. 2015;29(11):91-94. (in Russian)
- Romanenkova O.V., Gladyr E.A., Kostyunina O.V., Zinovieva N.A. Screening of cattle for the presence of mutation in *APAF1* gene, which is associated with fertility haplotype HH1. *Dostizheniya Nauki i Tekhniki APK = Achievements of Science and Technology of AIC*. 2016;30(2):94-97. (in Russian)
- Romanenkova O.S., Volkova V.V., Kostyunina O.V., Zinovieva N.A. Diagnostics of HH5 haplotype for Russian Holstein and Blackand-White cattle population. *Molochnoe i Myasnoe Skotovodstvo = Dairy and Beef Cattle Breeding*. 2018;6:13-15. DOI 10.25632/ MMS.2018.2018.20295. (in Russian)
- Sabetova K.D., Podrechneva I.Yu., Belokurov S.G., Schegolev P.O., Kofiadi I.A. Test system for *BLAD* mutation diagnosis in cattle populations. *Russ. J. Genet.* 2021;57(8):936-941. DOI 10.1134/ S1022795421080135.
- Sebastiani C., Arcangeli C., Ciullo M., Torricelli M., Cinti G., Fisichella S., Biagetti M. Frequencies evaluation of β-casein gene polymorphisms in dairy cows reared in Central Italy. *Animals* (*Basel*). 2020;10(2):252. DOI 10.3390/ani10020252.
- Storey J.D., Bass A.J., Dabney A., Robinson D. qvalue: Q-value estimation for false discovery rate control. R Packag. version 2.24.0. 2020. DOI 10.18129/B9.bioc.qvalue.
- Storey J.D., Tibshirani R. Statistical significance for genome-wide experiments. *Proc. Natl. Acad. Sci. USA*. 2003;100(16):9440-9445. DOI 10.1073/pnas.1530509100.
- Surzhikova E.S., Sharko G.N., Mikhailenko T.N. Allelic spectrum of CSN3, PIT-1, PRL genes in horned cattle of Black-and-White breed. *Novosti Nauki v APK = Science News of AIC*. 2019;3:136-139. DOI 10.25930/2218-855X/032.3.12.2019. (in Russian)
- Tambasco D.D., Paz C.C.P., Tambasco-Studart M., Pereira A.P., Alencar M.M., Freitas A.R., Coutinho L.L., Packer I.U., Regitano L.C.A. Candidate genes for growth traits in beef cattle crosses *Bos taurus* × *Bos indicus. J. Anim. Breed. Genet.* 2003;120(1):51-56. DOI 10.1046/j.1439-0388.2003.00371.x.
- Tatum J.D. Pre-Harvest Cattle Management Practices for Enhancing Beef Tenderness. Colorado State Univ., 2006.
- Terletskiy V.P., Buralkhiyev B.A., Usenbekov Y.S., Yelubayeva M., Tyshchenko V.I., Beyshova I.S. Screening for mutations that determine the development of hereditary diseases in breeding cattle. *Aktual'nye Voprosy Veterinarnoi Biologii = Actual Questions of Veterinary Biology*. 2016;3:3-7. (in Russian)
- Usova T.P., Usmanova N.N., Litvina N.I., Usov N.V. The spread of BLAD-syndrome of breeding bulls of Holstein breed of Russian and import selection. *Vestnik Rossiyskogo Gosudarstvennogo Agrarnogo* Zaochnogo Universiteta = Bulletin of the Russian State Agricultural Correspondence University. 2017;25:20-24. (in Russian)
- Vsyakikh A.S., Kurinsky M.S. Imported Cattle in the USSR. Moscow: Kolos Publ., 1976. (in Russian)
- Watanabe D., Hirano T., Sugimoto Y., Ogata Y., Abe S., Ando T., Ohtsuka H., Kunieda T., Kawamura S. Carrier rate of Factor XI deficiency in stunted Japanese black cattle. J. Vet. Med. Sci. 2006;68(12):1251-1255. DOI 10.1292/jvms.68.1251.
- Xu L., Bickhart D.M., Cole J.B., Schroeder S.G., Song J., Tassell C.P., Sonstegard T.S., Liu G.E. Genomic signatures reveal new evidences

for selection of important traits in domestic cattle. *Mol. Biol. Evol.* 2015;32(3):711-725. DOI 10.1093/molbev/msu333.

- Yudin N.S., Larkin D.M. Whole genome studies of origin, selection and adaptation of the Russian cattle breeds. Vavilovskii Zhurnal Genetiki i Selektsii = Vavilov Journal of Genetics and Breeding. 2019;23(5):559-568. DOI 10.18699/VJ19.525. (in Russian)
- Yudin N.S., Voevoda M.I. Molecular genetic markers of economically important traits in dairy cattle. *Russ. J. Genet.* 2015;51(5):506-517. DOI 10.1134/S1022795415050087.
- Yurchenko A., Yudin N., Aitnazarov R., Plyusnina A., Brukhin V., Soloshenko V., Lhasaranov B., Popov R., Paronyan I.A., Plemyashov K.V., Larkin D.M. Genome-wide genotyping uncovers genetic profiles and history of the Russian cattle breeds. *Heredity (Edinb.)*. 2018;120(2):125-137. DOI 10.1038/s41437-017-0024-3.
- Zinovieva N.A. Haplotypes affecting fertility in Holstein cattle. *Sel'skokhozyaistvennaya Biologiya* = *Agricultural Biology*. 2016;51(4):423-435. DOI 10.15389/agrobiology.2016.4.423eng.

ORCID ID

N.S. Yudin orcid.org/0000-0002-1947-5554 D.M. Larkin orcid.org/0000-0001-7859-6201

Acknowledgements. The work was supported by the Russian Ministry of Education and Science grant No. 075-15-2021-1004. Conflict of interest. The authors declare no conflict of interest. Received November 30, 2021. Revised December 20, 2021. Accepted December 30, 2021.