## Review Article

# Association between *Faecalibacterium prausnitzii* Reduction and Inflammatory Bowel Disease: A Meta-Analysis and Systematic Review of the Literature

## Yuan Cao, Jun Shen, and Zhi Hua Ran

Renji Hospital, School of Medicine, Shanghai Jiao Tong University, Shanghai Institution of Digestive Disease, Shanghai Inflammatory Bowel Disease Research Center, Shanghai 200127, China

Correspondence should be addressed to Zhi Hua Ran; zhihuaran@vip.163.com

Received 31 December 2013; Accepted 17 February 2014; Published 27 March 2014

Academic Editor: Paolo Gionchetti

Copyright © 2014 Yuan Cao et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

*Background*. Laboratory data suggests a reduction of *Faecalibacterium prausnitzii* (*F. prausnitzii*) is confirmed both in fecal samples in inflammatory bowel disease (IBD) patients. Numerous observational studies have suspected dysbiosis, an imbalance between protective and harmful bacteria to be relevant to the etiology and pathogenesis of IBD. *Methods*. Medline, EMBASE, Pubmed, and others. were searched by 2 independent reviewers. Of 48 abstracts reviewed, 11 studies met our inclusion criteria (subject N = 1180). Meta-analysis was performed with Review Manager 5.2. *Results*. The bacterial count of *F. prausnitzii* in IBD patients was significantly lower (6.7888 ± 1.8875) log10 CFU/g feces than healthy controls (7.5791 ± 1.5812) log10 CFU/g feces; P < 0.0001. The Standardization Mean Difference of *F. prausnitzii* in IBD patients was -0.94 (95% confidence interval [CI]: -1.07-0.80). Subgroup analyses revealed a trend toward a greater effect for CD (SMD: -1.13, 95% CI: -1.32--0.94) when compared to UC (SMD: -0.78, 95% CI: -0.97--0.60). *Conclusions*. The abundance of *F. prausnitzii* was decreased in IBD patients compared with healthy controls. Furthermore, the reduction of *F. prausnitzii* and misbalance of the intestinal microbiota are particularly higher in CD patients with ileal involvement.

## 1. Introduction

IBD is suspected to arise from the interaction between the host's genetic background, mucosal immunity, and the resident bacterial flora [1]. Genome-wide association studies (GWAS) have identified more than 160 host genetic variants. Many are related to human gut microbiota [2]. In patients with inflammatory bowel diseases (IBD), the composition and diversity of the microbiota are always altered [3]. The imbalance between potentially "beneficial" and potentially "harmful" bacteria, also called dysbiosis, plays a role in the pathogenesis of chronic mucosal inflammatory lesions of IBD [4].

*F. prausnitzii* belongs to the phylum of Firmicutes and is the major bacterium of the *Clostridium leptum* group. The Meta-analysis of the Human Intestinal Tract project have shown that F. prausnitzii is one of the most abundant anaerobic bacteria in the human gut microbiota, with a proportion of around 5% of total bacteria in faeces [5]. F. prausnitzii plays an important role in providing energy to the colonocytes and maintaining the intestinal health [6]. Furthermore, there is emerging laboratory evidence illustrating a strong antiinflammatory effect of *F. prausnitzii* both in vitro and in vivo. And deficiency of *F. prausnitzii* might provoke and enhance inflammation [7]. Specially, a significant inverse correlation between disease activity and the count of F. prausnitzii was found in UC patients, even with quiescent disease [8]. The depletion of F. prausnitzii was observed in patients with untreated CD but not in the patients with chronic diarrhea, suggesting a relationship in the pathomechanisms of CD [9]. Finally, a diminished prevalence and abundance of F. prausnitzii are revealed in the fecal samples of patients with IBD. The *F. prausnitzii* level was much lower when the disease activity increased [10].

To further investigate the possible association between *F. prausnitzii* reduction and IBD, we conducted a metaanalysis and systematic review to estimate the relative risk of *F. prausnitzii* reduction in patients with and without IBD. Given the laboratory data previously cited, we hypothesized a relationship between *F. prausnitzii* reduction and IBD.

## 2. Materials and Methods

2.1. Search Strategy. This review was performed according to the standard guidelines for meta-analyses and systematic reviews of observational studies [11]. To find relevant articles for this review, we searched the following databases (from inception to November 2013): EMBASE, MEDLINE, Google Scholar, Pubmed, ACP Journal Club, the Cochrane Central Register of Controlled Trials, CMR, DARE, and HTA. The search strategy used free-text words to increase the sensitivity of the search. The following search terms were used: "inflammatory bowel disease," "Crohn's disease," "ulcerative colitis," "IBD," "UC," "CD," "Faecalibacterium prausnitzii," "F. prausnitzii," and "FP." Boolean operators (AND, OR, NOT) were used to widen and narrow the search results. The titles and abstracts from the search results were examined for potential inclusion. Also, the references from selected articles were examined as further search tools.

2.2. Study Selection. For inclusion in the systematic review, a study had to meet the following criteria established by the study team: (1) *F. prausnitzii* counts measured by polymerase chain reaction-denaturing gradient gel electrophoresis (PCR-DGGE), terminal restriction fragment length polymorphism (T-RFLP), or fluorescence in situ hybridization (FISH), (2) studies of human, (3) inclusion of a control group, (4) IBD and control groups were similar in age and sex and from the same catchment area, and (5) data were reported that were sufficient to calculate *F. prausnitzii* reduction in both the IBD and control groups. Studies were excluded if they used data from a previously published study.

2.3. Data Extraction. To reduce reporting error and bias in data collection, 2 independent reviewers extracted data from selected studies using standardized data extraction forms. These forms, created by the study team, included the (a) title, (b) authors, (c) journal, (d) year of publication, (e) study design, (f) inclusion and exclusion criteria, (g) methods by which IBD was diagnosed, (h) methods by which F. prausnitzii reduction was diagnosed, (i) number of patients with ulcerative colitis (UC), (j) number of patients with Crohn's disease (CD), (k) number of patients in the control group, (l) reported previous use of antibiotics, probiotics, or prebiotics in the IBD and control groups, and (m) reported previous use of steroids, 5-aminosalicylates (5-ASAs), and tumor necrosis factor-alpha (TNF- $\alpha$ ) antibody medications in the IBD group. Studies were excluded if participants had used steroids, 5-ASAs, TNF- $\alpha$  antibody antibiotics, probiotics, or prebiotics in the last month preceding fecal

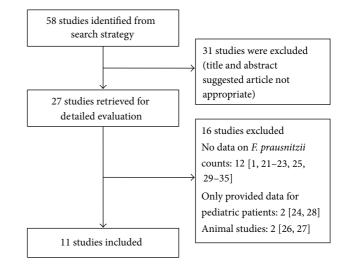


FIGURE 1: Flow diagram of studies identified in the systematic review.

sampling as this could influence the intestinal microbiota. If needed, authors were contacted regarding specific questions relating to their study. The independent reviewers conferred after data extraction was complete, discrepancies were identified, and review of the relevant article led to consensus.

2.4. Statistical Analysis. The primary outcome of this analysis was the Standardization Mean Difference (SMD) of *F. prausnitzii* counts in IBD versus controls. Std. mean difference was used to describe the counts of the *F. prausnitzii* in IBD patients versus the controls. We calculated the SMD with a 95% confidence interval (CI) based on a random-effects model as described by Mantel-Haenszel. Meta-analysis was performed with the Review Manager 5.2. Analysis with a funnel plot used to assess publication bias. An  $I^2$  statistic was used to measure the proportion of inconsistency in individual studies that could not be explained by chance. Any heterogeneity identified would prompt subgroup analysis in an attempt to explain these findings.

2.5. Assessment of Study Quality. Each study chosen for review was carefully assessed for study quality by the study team. Study quality was assessed using the following criteria: (1) study design, (2) method of IBD diagnosis, (3) method of patient enrollment (consecutive versus selected), (4) method of *F. prausnitzii* counts measurement, and (5) whether *F. prausnitzii* reduction was the primary or secondary outcome of the study.

## 3. Results

*3.1. Search Results.* Our initial search strategy yielded 58 potential articles for inclusion. After detailed analysis of selected articles, 27 articles were reviewed in detail. Subsequently, 16 articles did not meet inclusion criteria [1, 21–35]. The reasons for exclusion included: 12 studies did not provide data on *F. prausnitzii* counts [1, 21–23, 25, 29–35]. 2 studies

#### Gastroenterology Research and Practice

Author	Year	Location	Single versus multicenter	<i>n</i> , total	n, IBD (CD/UC)	<i>n</i> , control	Control composition	Mean age, IBD (CD/UC)	Mean age, control
Machiels et al. [8]	2013	Belgium	Single	214	0/127	87	Healthy controls	43	41.5
Varela et al. [12]	2013	Spain	Single	176	0/116	31	Healthy controls	40	32
Swidsinski et al. [13]	2008	Germany	Single	422	82/105	32	Healthy controls	35/41	40
Dörffel et al. [9]	2012	Germany	Single	171	50/0	25	Healthy controls	39	48
Vermeiren et al. [14]	2012	Belgium	Single	12	0/6	6	Healthy controls	Not reported	Not reported
Joossens et al. [15]	2011	Belgium	Single	207	68/0	55	Healthy controls	Not reported	Not reported
Wang et al. [16]	2013	China	Single	76	21/34	21	Healthy controls	Not reported	Not reported
Jia et al. [17]	2010	UK	Single	73	20/14	18	Healthy controls	Not reported	Not reported
Sokol et al. [18]	2009	France	Single	133	22/13	27	Healthy controls	37/40	36
Andoh et al. [19]	2012	Japan	Multicenter	188	67/0	121	Healthy controls	30	32
Willing et al. [20]	2009	Sweden	Single	20	6/0	6	Healthy controls	Not reported	Not reported

TABLE 1: Characteristics of the included studies.

IBD: inflammatory bowel disease; CD: Crohn's disease; UC: ulcerative colitis.

TABLE 2: Quality assessment of the included studies.

Author	F. prausnitzii counts	IBD diagnosis	Study type	Patient enrollment	Outcome	Samples
Machiels et al. [8]	RT-PCR	Not reported	Retrospective	Not reported	Primary	Stools
Varela et al. [12]	RT-PCR	Colonoscopy and histology	Retrospective	Not reported	Primary	Stools
Swidsinski et al. [13]	FISH	Colonoscopy	Retrospective	Not reported	Primary	Stools
Dörffel et al. [9]	FISH	Colonoscopy	Retrospective	Not reported	Primary	Stools
Vermeiren et al. [14]	RT-PCR	Not reported	Retrospective	Not reported	Primary	Stools
Joossens et al. [15]	RT-PCR	Not reported	Retrospective	Not reported	Primary	Stools
Wang et al. [16]	RT-PCR	Not reported	Retrospective	Not reported	Primary	Biopsies
Jia et al. [17]	RT-PCR	Not reported	Retrospective	Not reported	Primary	Stools
Sokol et al. [18]	RT-PCR	Not reported	Retrospective	Not reported	Primary	Stools
Andoh et al. [19]	T-RFLP	Not reported	Retrospective	Not reported	Primary	Stools
Willing et al. [20]	RT-PCR	Colonoscopy	Retrospective	Not reported	Primary	Biopsies

only provided data for pediatric patients [24, 28]. 2 studies were animal studies [26, 27]. Therefore, 11 studies [8, 9, 12–20] with 1180 patients fulfilled the inclusion criteria for the review (Figure 1).

3.2. Study Characteristics. The characteristics of the included studies are summarized in Tables 1 and 2. The results of each study are in Table 3. *F. prausnitzii* counts were expressed as log10 values per gram feces. The largest and earliest study examining the relationship between *F. prausnitzii* reduction and IBD was conducted in Germany by Swidsinski et al. [13]. The authors investigated sections of paraffinembedded punched fecal cylinders using fluorescence in situ hybridization (FISH). *F. prausnitzii* with high concentration was counted within a 10 \*  $10 \,\mu$ m area of the microscopic field representative of the region of interest. *F. prausnitzii* 

with uneven distribution or overall low concentrations was enumerated within larger areas of  $100 * 100 \,\mu\text{m}$ .

8 of the included studies confirmed the differences in the presence or intensity of *F. prausnitzii* counts after denaturing gradient gel electrophoresis (DGGE) by real-time PCR (RT-PCR) [8, 12, 14–18, 20]. *F. prausnitzii* cannot be cultured owing to its requirement for a complex anaerobic environment [19]. By RT-PCR, they were able to amplify, clone, and sequence the bacterial 16S ribosomal RNA genes and analyze the fecal samples individually to avoid the possible error [36].

Three of the included studies commented on the activity of IBD and *F. prausnitzii* counts. Wang et al. found sharply decreased *F. prausnitzii* in the feces of active CD and UC patients [16]. Sokol et al. and Andoh et al. reported lower counts of *F. prausnitzii* in active CD patients compared to CD patients in remission [18, 19].

Author	log 10 copies/g IBD patients (CD/UC)	log 10 copies/g healthy controls	Р
Machiels et al. [8]	$0/(10.95 \pm 1.41)$	$11.72 \pm 1.08$	< 0.0001
Varela et al. [12]	$0/(8.02 \pm 0.57)$	$8.90 \pm 0.37$	< 0.0001
Swidsinski et al. [13]	$(9.75 \pm 9.77)/(10.14 \pm 10.02)$	$10.17 \pm 9.65$	< 0.0001
Dörffel et al. [9]	$(9.06 \pm 9.33)/0$	$10.21 \pm 9.94$	< 0.001
Vermeiren et al. [14]	$0/(5.56 \pm 0.83)$	$6.63 \pm 0.95$	0.07
Joossens et al. [15]	$(9.44 \pm 1.85)/0$	$10.97 \pm 1.25$	< 0.0001
Wang et al. [16]	$(0.03 \pm 0.06)/(0.23 \pm 0.51)$	$1.40 \pm 1.06$	< 0.0001
Jia et al. [17]	$(5.71 \pm 5.34)/(5.93 \pm 5.87)$	$5.93 \pm 5.83$	< 0.05
Sokol et al. [18]	$(8.81 \pm 0.52)/(8.70 \pm 0.63)$	$10.4 \pm 0.2$	0.0004
Andoh et al [19]	$(0.40 \pm 0.09)/0$	$0.81 \pm 0.04$	< 0.0001
Willing et al. [20]	$(0.40 \pm 0.89)/0$	$8.72 \pm 2.49$	< 0.001

TABLE 3: Study results.

1 of the included studies examined the *F. prausnitzii* counts before and after treatment by an element diet [17]. It suggests recovery following elemental diet is attributed to lower levels of gut flora.

1 of the included studies reported the relationship between the maintenance of clinical remission and the recovery of the *F. prausnitzii* population after relapse Varela et al. found low counts of *F. prausnitzii* were associated with less than 12 months of remission and more than 1 relapse/year [12].

3.3. Meta-Analysis of SMD. Overall, the bacterial count of *F. prausnitzii* in IBD patients was significantly lower (6.7888 ± 1.8875) log10 CFU/g feces than healthy controls (7.5791 ± 1.5812) log10 CFU/g feces; P < 0.0001. The SMD of *F. prausnitzii* in IBD patients was -0.94 (95% confidence interval [CI]: -1.07--0.80) (Figure 2). Subgroup analyses revealed a trend toward a greater effect for CD (SMD: -1.13, 95% CI: -1.32--0.94) when compared to UC (SMD: -0.78, 95% CI: -0.97--0.60). There was significant heterogeneity in the included studies ( $I^2 = 96\%$ ). Furthermore, analysis of the funnel plots for publication bias suggested a possible bias against small studies demonstrating high SMD (Figure 3).

#### 4. Discussion

Our systematic review and meta-analysis of the literature has identified recent studies examining the relationship between *F. prausnitzii* reduction and IBD. The majority of recent studies find a higher rate of *F. prausnitzii* reduction in IBD patients as compared to controls. All of the 11 included studies found significantly lower *F. prausnitzii* counts in IBD patients versus controls. Our meta-analysis suggests a possible link with the reduction of *F. prausnitzii* andmisbalance of the intestinal microbiota and IBD patients, especially CD patients with ileal involvement. The levels of *F. prausnitzii* were extremely low in two studies [19, 28]. Wang et al.

took biopsies samples from active CD patients and found extremely lower F. prausnitzii counts compared to stools [28]. Andoh et al. demonstrated a difference in gut microbiota of the Japanese population, suggesting that environmental factors such as sanitation, diet, hygiene, and ethnicity were important for shaping the gut microbiota [19]. However, significant heterogeneity and the possibility of publication bias limit our certainty in this association. Furthermore, Hansen et al. challenged the current model of a protective role for F. prausnitzii in CD [24]. They reported an increasein mucosal Faecalibacterium in pediatric CD patients compared with controls, which suggested a more dynamic role for this organism than previously described in adult IBD. It is possible that the microbial signature of pediatric IBD is distinct from adult disease. Furthermore, the early host and microbiota response to IBD may induce proliferation of F. prausnitzii to reverse the inflammatory change, which still remains to be explained.

Mechanistic theories of microbial etiopathogenesis between the possible protective benefit of F. prausnitzii against IBD have been proposed. Duncan et al. demonstrated that the major end products of glucose fermentation by F. prausnitzii strains are substantial quantities of butyrate [37]. Butyrate plays a major role in gut physiology, protection against pathogen invasion, and modulation of immune system [38]. Butyrate is the primary energy source for intestinal epithelial cells, which are fundamental elements for the maintenance of barrier integrity [26]. Therefore, butyrate may contribute to the anti-inflammatory effect. Additionally, butyrate may inhibit inflammatory response through inhibition of histone deacetylase activity, resulting in suppression of NF- $\kappa$ B activity and hyperacetylation of histones [12]. Furthermore, Himmel et al. found that F. prausnitzii could induce relatively low amounts of IL-12 and large amounts of IL-10 and Tregs in epithelial and PBMC models to restrain the progression of inflammation [39]. While Sokol et al. reported that F. prausnitzii led to significantly lower IL-12 and IFN-y production levels

Study or subgroup	IBD (CD/UC)			Healthy controls				Std. mean difference	Std. mean difference
Study of subgroup	Mean	SD	Total	Mean	SD	Total	Weight	IV, fixed, 95% CI	IV, fixed, 95% CI
Andoh et al. 2012	0.4	0.09	67	0.81	0.04	121	3.4%	-6.53 [-7.26, -5.80]	$\leftarrow$
Dörffel et al. 2012	9.06	9.33	50	10.21	9.94	25	7.9%	-0.12 [-0.60, 0.36]	
Jia et al. 2010	5.71	5.34	20	5.93	5.83	18	4.5%	$-0.04 \ [-0.68, 0.60]$	
Jia et al. 2010	5.93	5.87	14	5.93	5.83	18	3.7%	0.00 [-0.70, 0.70]	
Joossens et al. 2011	9.44	1.85	68	10.97	1.25	55	12.9%	-0.94 [-1.32, -0.57]	
Machiels et al. 2013	10.95	1.4	127	11.72	1.08	87	23.3%	-0.60 [-0.88, -0.32]	
Sokol 2009	8.8	0.5	22	10.4	0.2	27	1.6%	-4.30 [-5.36, -3.25]	$\leftarrow$
Sokol 2009	8.7	0.6	13	10.4	0.2	27	1.2%	-4.44 [-5.66, -3.22]	<i>←</i>
Swidsinski 2008	9.75	9.77	82	10.17	9.65	32	10.9%	-0.04 [-0.45, 0.37]	
Swidsinski 2008	10.14	10	105	10.17	9.65	32	11.6%	-0.00 [-0.40, 0.39]	
Varela 2013	8.02	0.57	116	8.9	0.37	31	9.4%	-1.64 [-2.08, -1.20]	
Vermeiren 2011	5.56	0.83	6	6.63	0.95	6	1.2%	-1.11 [-2.36, 0.15]	
Wang 2013	0.026	0.058	21	1.402	1.059	21	3.4%	-1.80 [-2.53, -1.07]	
Wang 2013	0.225	0.512	34	1.402	1.059	21	4.7%	-1.52 [-2.13, -0.90]	
Willing 2009	0.4	0.89	6	8.7	2.49	6	0.3%	-4.10 [-6.40, -1.80]	<
Total (95% CI)			75	1		527	100.0%	-0.94 [-1.07, -0.80]	•
Heterogeneity: $\chi^2$ =	= 393.06	, df = 1	4 ( <i>P</i> <	0.00001)	; $I^2 = 9$	6%			
Test for overall effec	zt: Z = 1	13.65 ()	P < 0.00	0001)					-2 $-1$ 0 1 2

FIGURE 2: Forest plot of rate of *F. prausnitzii* reduction in patients with IBD versus controls.

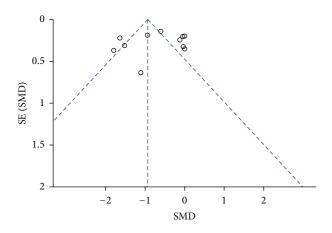


FIGURE 3: Funnel plot analysis.

and higher secretion of IL-10 in vitro peripheral blood mononuclear cells [26].

The data on the incidence of *F. prausnitzii* reduction and IBD found in the literature has several limitations. Most of the studies did not comment on the participants' previous history of treatment such as antibiotics, probiotics, or prebiotics which may influence the intestinal microbiota. It is therefore possible that study participants had been treated for dysbiosis prior to entering the study, thereby producing a false *F. prausnitzii* reduction. Additionally, we did not evaluate confounding factors including diet and smoking in our study, even though these factors might reduce *F. prausnitzii* levels and faecal butyrate values [40]. Furthermore, some of the

included studies did not clearly identify the criteria for the IBD diagnosis. Few commented on personal review of the endoscopic findings or histology. Also we did not relate the differences in microbiota to geography and ethnicity. Lastly, most studies were performed at a single medical center.

Future studies should address these limitations. After confirming the diagnosis of IBD through the endoscopic and histological findings, PCR-DGGE, T-RFLP, or FISH for F. prausnitzii counts would be initiated. In patients found to have F. prausnitzii reduction, probiotics or prebiotics may be used to restore the "ecological balance" of intestinal microbiota. Dörffel et al. reported rifaximin was associated with an increased level of F. prausnitzii [9]. Other specific treatments such as infliximab and a high-dose cortisol therapy were shown to reverse the depletion of F. prausnitzii [13]. The mechanism for the inverse association between F. prausnitzii reduction and the initiation and perpetuation of inflammatory bowel disease has yet to be defined. Healthy controls who are age- and sex-matched to the IBD group would be selected from the same area as the IBD group and tested for F. prausnitzii by the same method. In both groups, a thorough history examining previous treatment such as antibiotics, probiotics, or prebiotics, steroids, 5aminosalicylates (5-ASAs), and tumor necrosis factor alpha  $(TNF-\alpha)$  antibody would be obtained. Prideaux et al. reported that regardless of ethnicity or geography, Crohn's disease resulted in reduced bacterial diversity. However, in ulcerative colitis, diversity was reduced in Chinese subjects only. It suggested that ethnicity might also play an important role in the pathogenesis of IBD [41].

## 5. Conclusions

In summary, our meta-analysis and systematic review suggest a possible protective benefit of *F. prausnitzii* against the development of IBD. However, significant variation among the studies and the possibility of publication bias limit the certainty of this association. Therefore, further treatment such as probiotics or prebiotics to increase the levels of *F. prausnitzii* in IBD are lead to attempts. If *F. prausnitzii* is found to indeed protect against IBD, we can approach the treatment such as supplementing the microorganisms that produce butyric acid.

## **Conflict of Interests**

The authors declare that there is no conflict of interests regarding the publication of this paper.

#### **Authors' Contribution**

All of the authors contributed equally to the study.

## Acknowledgment

The study was supported by the National Science Foundation of China (no. 81170362 and 81370508).

## References

- M. Martinez-Medina, X. Aldeguer, F. Gonzalez-Huix, D. Acero, and L. J. Garcia-Gil, "Abnormal microbiota composition in the ileocolonic mucosa of Crohn's disease patients as revealed by polymerase chain reaction-denaturing gradient gel electrophoresis," *Inflammatory Bowel Diseases*, vol. 12, no. 12, pp. 1136–1145, 2006.
- [2] L. Jostins, S. Ripke, R. K. Weersma et al., "Host-microbe interactions have shaped the genetic architecture of inflammatory bowel disease," *Nature*, vol. 491, no. 7422, pp. 119–124, 2012.
- [3] B. Chassaing and A. Darfeuille-Michaud, "The commensal microbiota and enteropathogens in the pathogenesis of inflammatory bowel diseases," *Gastroenterology*, vol. 140, no. 6, pp. 1720–1728, 2011.
- [4] A. W. Walker, J. D. Sanderson, C. Churcher et al., "Highthroughput clone library analysis of the mucosa-associated microbiota reveals dysbiosis and differences between inflamed and non-inflamed regions of the intestine in inflammatory bowel disease," *BMC Microbiology*, vol. 11, article 7, 2011.
- [5] M. Arumugan, J. Raes, E. Pelletier et al., "Enterotypes of the human gut microbioma," *Nature*, vol. 473, no. 7346, pp. 174–180, 2011.
- [6] P. Louis and H. J. Flint, "Diversity, metabolism and microbial ecology of butyrate-producing bacteria from the human large intestine," *FEMS Microbiology Letters*, vol. 294, no. 1, pp. 1–8, 2009.
- [7] C. D. Packey and R. B. Sartor, "Commensal bacteria, traditional and opportunistic pathogens, dysbiosis and bacterial killing in inflammatory bowel diseases," *Current Opinion in Infectious Diseases*, vol. 22, no. 3, pp. 292–301, 2009.

- [8] K. Machiels, M. Joossens, J. Sabino et al., "A decrease of the butyrate-producing species Roseburia hominis and Faecalibacterium prausnitzii defines dysbiosis in patients with ulcerative colitis," *Gut*, 2013.
- [9] Y. Dörffel, A. Swidsinski, V. Loening-Baucke, B. Wiedenmann, and M. Pavel, "Common biostructure of the colonic microbiota in neuroendocrine tumors and Crohn's disease and the effect of therapy," *Inflammatory Bowel Diseases*, vol. 18, no. 9, pp. 1663– 1671, 2012.
- [10] K. Machiels, M. Joossens, V. de Preter et al., "Association of Faecalibacterium prausnitzii and disease activity in ulcerative colitis," *Gastroenterology*, vol. 140, supplement 1, article S142, no. 5, 2011.
- [11] J. Luther, M. Dave, P. D. R. Higgins, and J. Y. Kao, "Association between Helicobacter pylori infection and inflammatory bowel disease: a meta-analysis and systematic review of the literature," *Inflammatory Bowel Diseases*, vol. 16, no. 6, pp. 1077–1084, 2010.
- [12] E. Varela, C. Manichanh, M. Gallart et al., "Colonisation by Faecalibacterium prausnitzii and maintenance of clinical remission in patients with ulcerative colitis," *Alimentary Pharmacology & Therapeutics*, vol. 38, no. 2, pp. 151–161, 2013.
- [13] A. Swidsinski, V. Loening-Baucke, M. Vaneechoutte, and Y. Doerffel, "Active Crohn's disease and ulcerative colitis can be specifically diagnosed and monitored based on the biostructure of the fecal flora," *Inflammatory Bowel Diseases*, vol. 14, no. 2, pp. 147–161, 2008.
- [14] J. Vermeiren, P. van den Abbeele, D. Laukens et al., "Decreased colonization of fecal Clostridium coccoides/Eubacterium rectale species from ulcerative colitis patients in an *in vitro* dynamic gut model with mucin environment," *FEMS Microbiology Ecology*, vol. 79, no. 3, pp. 685–696, 2012.
- [15] M. Joossens, G. Huys, M. Cnockaert et al., "Dysbiosis of the faecal microbiota in patients with Crohn's disease and their unaffected relatives," *Gut*, vol. 60, no. 5, pp. 631–637, 2011.
- [16] W. Wang, L. Chen, R. Zhou et al., "Increased proportion of Bifidobacterium and the Lactobacillus group and loss of butyrate-producing bacteria in inflammatory bowel disease," *Journal of Clinical Microbiology*, 2013.
- [17] W. Jia, R. N. Whitehead, L. Griffiths et al., "Is the abundance of Faecalibacterium prausnitzii relevant to Crohn's disease?" *FEMS Microbiology Letters*, vol. 310, no. 2, pp. 138–144, 2010.
- [18] H. Sokol, P. Seksik, J. P. Furet et al., "Low counts of Faecalibacterium prausnitzii in colitis microbiota," *Inflammatory Bowel Diseases*, vol. 15, no. 8, pp. 1183–1189, 2009.
- [19] A. Andoh, H. Kuzuoka, T. Tsujikawa et al., "Multicenter analysis of fecal microbiota profiles in Japanese patients with Crohn's disease," *Journal of Gastroenterology*, vol. 47, no. 12, pp. 1298– 1307, 2012.
- [20] B. Willing, J. Halfvarson, J. Dicksved et al., "Twin studies reveal specific imbalances in the mucosa-associated microbiota of patients with ileal Crohn's disease," *Inflammatory Bowel Diseases*, vol. 15, no. 5, pp. 653–660, 2009.
- [21] T. Fujimoto, H. Imaeda, K. Takahashi et al., "Decreased abundance of Faecalibacterium prausnitzii in the gut microbiota of Crohn's disease," *Journal of Gastroenterology and Hepatology*, vol. 28, no. 4, pp. 613–619, 2013.
- [22] M. Galecka, P. Szachta, A. Bartnicka, L. Łykowska-Szuber, P. Eder, and A. Schwiertz, "Faecalibacterium prausnitzii and Crohn's disease—is there any connection?" *Polish Journal of Microbiology*, vol. 62, no. 1, pp. 91–95, 2013.

- [23] S. Miquel, R. Marín, O. Rossi et al., "Faecalibacterium prausnitzii and human intestinal health," *Current Opinion in Microbiology*, vol. 16, no. 3, pp. 255–261, 2013.
- [24] R. Hansen, R. K. Russell, C. Reiff et al., "Microbiota of Pediatric IBD: increased Faecalibacterium prausnitzii and reduced bacterial diversity in Crohn's but not in ulcerative colitis," *The American Journal of Gastroenterology*, vol. 107, no. 12, pp. 1913– 1922, 2012.
- [25] C. L. O'Brien, G. E. Allison, and P. Pavli, "The more the merrier: Faecalibacterium prausnitzii in Crohn's disease," *Journal of Gastroenterology and Hepatology*, vol. 28, no. 5, pp. 757–759, 2013.
- [26] H. Sokol, B. Pigneur, L. Watterlot et al., "Faecalibacterium prausnitzii is an anti-inflammatory commensal bacterium identified by gut microbiota analysis of Crohn disease patients," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 105, no. 43, pp. 16731–16736, 2008.
- [27] X. Qiu, M. Zhang, X. Yang, N. Hong, and C. Yu, "Faecalibacterium prausnitzii upregulates regulatory T cells and anti-inflammatory cytokines in treating TNBS-induced colitis," *Journal of Crohn's and Colitis*, vol. 7, no. 11, pp. e558–e568, 2013.
- [28] M. Wang, G. Molin, S. Ahrné, D. Adawi, and B. Jeppsson, "High proportions of proinflammatory bacteria on the colonic mucosa in a young patient with ulcerative colitis as revealed by cloning and sequencing of 16S rRNA genes," *Digestive Diseases and Sciences*, vol. 52, no. 3, pp. 620–627, 2007.
- [29] S. Mondot, S. Kang, J. P. Furet et al., "Highlighting new phylogenetic specificities of Crohn's disease microbiota," *Inflammatory Bowel Diseases*, vol. 17, no. 1, pp. 185–192, 2011.
- [30] Y. H. Siaw and A. Hart, "Commentary: is Faecalibacterium prausnitzii a potential treatment for maintaining remission in ulcerative colitis?" *Alimentary Pharmacology & Therapeutics*, vol. 38, no. 5, p. 551, 2013.
- [31] S. Maccaferri, B. Vitali, A. Klinder et al., "Rifaximin modulates the colonic microbiota of patients with Crohn's disease: an *in vitro* approach using a continuous culture colonic model system," *Journal of Antimicrobial Chemotherapy*, vol. 65, no. 12, pp. 2556–2565, 2010.
- [32] J. L. Benjamin, C. R. Hedin, A. Koutsoumpas et al., "Smokers with active Crohn's disease have a clinically relevant dysbiosis of the gastrointestinal microbiota," *Inflammatory Bowel Diseases*, vol. 18, no. 6, pp. 1092–1100, 2012.
- [33] S. Angelberger, W. Reinisch, A. Makristathis et al., "Temporal bacterial community dynamics vary among ulcerative colitis patients after fecal microbiota transplantation," *The American Journal of Gastroenterology*, vol. 108, no. 10, pp. 1620–1630, 2013.
- [34] S. D. McLaughlin, A. W. Walker, C. Churcher et al., "The bacteriology of pouchitis: a molecular phylogenetic analysis using 16s rRNA gene cloning and sequencing," *Annals of Surgery*, vol. 252, no. 1, pp. 90–98, 2010.
- [35] S. Cucchiara, V. Iebba, M. P. Conte, and S. Schippa, "The microbiota in inflammatory bowel disease in different age groups," *Digestive Diseases*, vol. 27, no. 3, pp. 252–258, 2009.
- [36] K. E. Ashelford, N. A. Chuzhanova, J. C. Fry, A. J. Jones, and A. J. Weightman, "At least 1 in 20 16S rRNA sequence records currently held in public repositories is estimated to contain substantial anomalies," *Applied and Environmental Microbiology*, vol. 71, no. 12, pp. 7724–7736, 2005.
- [37] S. H. Duncan, P. Louis, and H. J. Flint, "Lactate-utilizing bacteria, isolated from human feces, that produce butyrate as a major fermentation product," *Applied and Environmental Microbiology*, vol. 70, no. 10, pp. 5810–5817, 2004.

- [38] G. T. Macfarlane and S. Macfarlane, "Fermentation in the human large intestine: its physiologic consequences and the potential contribution of prebiotics," *Journal of Clinical Gastroenterology*, vol. 45, supplemment, pp. S120–S127, 2011.
- [39] M. E. Himmel, Y. Yao, P. C. Orban, T. S. Steiner, and M. K. Levings, "Regulatory T-cell therapy for inflammatory bowel disease: more questions than answers," *Immunology*, vol. 136, no. 2, pp. 115–122, 2012.
- [40] R. F. J. Benus, T. S. van der Werf, G. W. Welling et al., "Association between Faecalibacterium prausnitzii and dietary fibre in colonic fermentation in healthy human subjects," *British Journal of Nutrition*, vol. 104, no. 5, pp. 693–700, 2010.
- [41] L. Prideaux, S. Kang, J. Wagner et al., "Impact of ethnicity, geography, and disease on the microbiota in health and inflammatory bowel disease," *Inflammatory Bowel Diseases*, vol. 19, no. 13, pp. 2906–2918, 2013.