

## Recent Advances in Indian Herbal Drug Research

Guest Editor: Thomas Paul Asir Devasagayam

# Radioprotective Potential of Plants and Herbs against the Effects of Ionizing Radiation

Ganesh C. Jagetia\*

Department of Radiobiology, Kasturba Medical College, Manipal-576 104, India

Received 19 September, 2006; Accepted 6 December, 2006

**Summary** Ionizing radiations produce deleterious effects in the living organisms and the rapid technological advancement has increased human exposure to ionizing radiations enormously. There is a need to protect humans against such effects of ionizing radiation. Attempts to protect against the deleterious effects of ionizing radiations by pharmacological intervention were made as early as 1949 and efforts are continued to search radioprotectors, which may be of great help for human application. This review mainly dwells on the radioprotective potential of plant and herbal extracts. The results obtained from *in vitro* and *in vivo* studies indicate that several botanicals such as *Gingko biloba*, *Centella asiatica*, *Hippophae rhamnoides*, *Ocimum sanctum*, *Panax ginseng*, *Podophyllum hexandrum*, *Amaranthus paniculatus*, *Emblica officinalis*, *Phyllanthus amarus*, *Piper longum*, *Tinospora cordifolia*, *Mentha arvensis*, *Mentha piperita*, *Syzygium cumini*, *Zingiber officinale*, *Ageratum conyzoides*, *Aegle marmelos* and *Aphanamixis polystachya* protect against radiation-induced lethality, lipid peroxidation and DNA damage. The fractionation-guided evaluation may help to develop new radioprotectors of desired activities.

**Key Words:** radioprotection, antioxidant, survival, micronuclei

## Introduction

### *Need for chemical radioprotection*

The discovery of X-rays by Roentgen in the year 1895 and radioactivity by Becquerel in the year 1896 can be considered as the turning point in human health care as the X-rays allowed to peep inside the human body [1, 2]. Although harmful effects of ionizing radiations were reported within a few months of discovery of X-rays, the real magnitude was not known. Study of occupational workers like physicians

and scientists handling radioactivity gave a clear picture of the harmful effects of ionizing radiations, which was further strengthened after the study of Japanese atomic bomb survivors of 1945. It is now fairly well established that radiation produces deleterious effects on the organisms and widespread use of radiation in diagnosis therapy, industry, energy sector and inadvertent exposure during air and space travel, nuclear accidents and nuclear terror attacks requires safeguard against human exposures. Lead shielding and other physical measures are cumbersome to use in such situations, therefore pharmacological intervention could be the most prudent strategy to protect humans against the harmful effect of ionizing radiations.

\*To whom correspondence should be addressed.

Tel: +91-820-2922122 Fax: +91-820-2571919

E-mail: gc.jagetia@gmail.com

### *Chemical radioprotection*

The use of chemicals to protect against the harmful effects of radiation was attempted after World War II with the realization of the need to safeguard humans against the military use of atomic weapons. Patt and his co-workers (1949) were the first to investigate the effect of amino-acid cysteine in rats exposed to lethal doses of X-rays [3]. They found that pretreatment of rats protected them against the radiation-induced lethality. Thereafter, several chemical compounds and their analogues have been screened for their radioprotective ability however, their high toxicity at optimum protective doses precluded their clinical use [4, 5]. The other major drawback of these compounds was that they were unable to provide post-irradiation protection. With the recognition that normal tissue protection during radiotherapy is as important as the destruction of cancer cells, the focus of protection research became more therapy oriented. Recent terror attacks throughout the world has strengthened the idea that it is necessary to devise appropriate measures against the nuclear terror attacks by using pharmacological agents that can protect against the ill effects of radiation.

The high toxicity of thiol compounds necessitated search for alternative agents, which could be less toxic and highly effective at non-toxic dose levels. It was also thought that products/compounds isolated from natural sources could be of substantial use as non-toxic radioprotectors. Therefore, investigators diverted their attention towards the plant and natural products during the last two decades. Plants have been reported to play an important role in the discovery of new drugs for the treatment of human diseases, which indicated that natural products play a highly significant role in the drug discovery and development process [6]. This was particularly evident in the areas of cancer and infectious diseases, where over 60% and 75% of these drugs, respectively, were shown to be of natural origin. A good chemical protector should be able to protect against the deleterious effect of ionizing radiation during therapeutic procedures as well as during nuclear accidents, space flight and background irradiation etc. An ideal radioprotector should be cheap, does not have toxic implications in a wide dose range, orally administered, rapidly absorbed, possesses a reasonably good dose reduction factor and can act through multiple mechanisms. The plant and natural products have all these qualities. They are usually non-toxic, relatively cheap, can be orally administered and could act through multiple mechanisms due to the presence of many chemicals. Therefore, screening of plants and natural products is a useful paradigm for radioprotection. The advantage of plants and natural products is that they are used in several traditional systems of medicines. They are usually considered non-toxic and widely accepted by humans. Their use as radioprotectors needs scientific evaluation and validation. Once this is done their use, as radioprotectors could be more successful than synthetic chemicals.

### *Assessment of radioprotective potential of plants and herbs*

The most pragmatic approach to select the possible candidate to evaluate radioprotective effect is to look into the available properties of the substance. Whether a substance has anti-inflammatory, antioxidant, antimicrobial, immunomodulatory, free radical scavenging or anti-stress properties, if so, it may act as a potential radioprotector and could be the right candidate for evaluation of its radioprotective activity.

Short-term *in vitro* tests can provide a basis for detailed evaluation of radioprotective activity. The simplest tests could be the evaluation of lipid peroxidation *in vitro*. Assay of free radicals and antioxidant status of a pharmacological agent can also provide some leads regarding the radioprotective potential of such agents. If a plant or a natural product is found to inhibit lipid peroxidation and scavenge free radicals, it may act as a possible radioprotector. The next step is to evaluate its radioprotective potential *in vitro* using cell survival and micronuclei assays. If it is found to elevate cell survival and reduce radiation-induced micronuclei formation, it certainly has a potential as a radioprotector.

There are other short-term tests like DNA strand breaks, apoptosis and estimation of glutathione (GSH) and enzymes like catalase, glutathione peroxidase etc. that can also provide an inkling of the radioprotective activity of any pharmacological agent. However, the gold standard for radioprotective activity is the evaluation of 30-day survival in rodents, since the animal studies with death as the end point are the most confirmatory, because the 30-day survival after lethal whole body irradiation clearly indicates the capacity of the pharmacological agent in test to modulate the recovery and regeneration of the gastrointestinal epithelium and the hemopoietic progenitor cells in the bone marrow, the two most radiosensitive organs that are essential for sustenance of the life [7]. The most reliable procedures involve determination of a dose reduction factor (DRF). In animal studies, DRFs are typically determined by irradiating mice with or without administering radioprotective agent at a range of radiation doses and then comparing the endpoint of interest. For example, the DRF for 30-day survival ( $LD_{50/30}$  drug-treated divided by  $LD_{50/30}$  vehicle-treated) quantifies protection of the hemopoietic system [8, 9]. With sufficient loss of hemopoietic stem cells, death follows due to infection, hemorrhage, and anemia. The GI syndrome in mice can be assessed by determining survival up to ten days (measure of GI death) after exposure to comparatively high doses of whole-body radiation, whereas hemopoietic syndrome can be assessed by monitoring the survival of irradiated animals up to 30 days post-irradiation [7-11]. The intestinal crypt cell assay or functional changes also serve as indicators of GI damage [12]. The most informative and useful preclinical studies relate protective effects to the drug's toxicity in the same animal model.

The efficacy of radioprotectors in clinical practice requires

different end points. Among other endpoints amenable to the determination of beneficial effects of radioprotectors, the most readily evaluable is protection against mucositis and xerostomia resulting from head and neck radiotherapy and various side effects when the GI tract is in the radiation field [13].

#### Plants and herbs as radioprotectors

Several botanicals have been screened for their radioprotective activity (Table 1). An intravenous infusion of an ethanol extract of *Gingko biloba* leaves, at a dose of 100 mg/person was found to be effective in patients with vasogenic edema observed after irradiation of the brain [14]. It has been reported to protect against the clastogenic factors from plasma of human subjects exposed to irradiation [15]. Treatment of recovery workers from the Chernobyl accident site was found to be effective when an oral dose of 40 mg/day of *G. biloba* was given 3 times daily for 2 months [16].

Aqueous extract of *Centella asiatica* reduced the adverse effect of low dose irradiation in Sprague Dawley rats by

inhibiting radiation-induced body weight loss and conditioned taste aversion [17]. Similarly, it has been found to protect against the radiation-induced weight loss in mice exposed to 8 Gy  $\gamma$ -radiation [18].

Oral administration of a *Hippophae rhamnoides* fruit juice concentrate to rats before or after irradiation increased life span, restored the 11-oxycorticosteroid level in the blood and weight of isolated adrenals, and also normalized their basal activity and response to (ACTH) (corticotropin) under *in vitro* conditions [19]. Hydroalcoholic extract of berries of *H. rhamnoides* also protected mice against  $\gamma$ -radiation-induced mortality, decline in endogenous colony forming unit (CFU), micronuclei formation and various other hematological parameters [20–22].

The radioprotective property of *Osimum sanctum* was first reported by Jagetia *et al.* [23] against the radiation-induced mortality, thereafter studies by Uma Devi and her coworkers established its radioprotective efficacy by evaluating mouse survival, spleen colony assay, and chromosome aberrations in mouse bone marrow cells. Apart from these osmium has

Table 1. Radioprotective effect of various herbs and plants

S. No.	Name	Test system	Observation	Ref. No.
1	<i>Gingko biloba</i>	Human	Brain edema, clastogenic factors	14–16
2	<i>Centella asiatica</i>	Rat, mice	Weight loss, taste aversion	17, 18
3	<i>Hippophae rhamnoides</i>	Rat, Mice	Survival, ACTH Survival, CFU, micronuclei	19 20
4	<i>Osimum sanctum</i>	Mice	Survival, CFU, chromosome damage, lipid peroxidation, glutathione	23–25
5	<i>Panax ginseng</i>	Mice	Survival, CFU, apoptosis, testicular enzymes	26–32
6	<i>Podophyllum hexandrum</i>	Mice	Survival, GI damage, nervous system of developing mice, GST, SOD	20, 33–36
7	<i>Tinospora cordifolia</i>	Mice	Survival, CFU, Blood cells	37, 38
8	<i>Emblica officinalis</i>		Survival, weight loss	39
9	<i>Phyllanthus amarus</i>	Mice	WBC, SOD, catalase, GST, GSHpx, glutathione reductase	40
10	<i>Amaranthus paniculatus</i>	Mice	Survival, CFU, spleen weight, Lipid peroxidation, GSH	41
11	<i>Piper longum</i>	Mice	WBC, $\alpha$ -Esterase, glutathione pyruvate transaminase, alkaline phosphatase, lipid peroxidation	42
12	<i>Syzigium cumini</i>	HPBLS, Mice	Micronuclei Radiation-sickness, GI & BM deaths	7 44–45
13	<i>Mentha arvensis</i>	Mice	Radiation-sickness, GI & BM deaths	47
14	<i>Mentha piperita</i>	Mice	Hematological constituents, serum phosphatase, CFU, spleen weight, goblet cells/villus section and chromosomal damage	48–50
15	<i>Zingiber officinale</i> ,	Mice	Radiation-sickness, GI & BM deaths, free radicals, GSH lipid peroxidation	52–53
16	<i>Ageratum conyzoides</i>	Mice	Radiation-sickness, GI & BM deaths, DPPH radical	54
17	<i>Aegle marmelos</i>	HPBLS Mice	Micronuclei, free radicals Radiation-sickness, GI & BM deaths, Lipid peroxidation, GSH, CFU, villus height, crypt cells, goblet cells	56 57–59
18	<i>Aphanamixis polystachya</i>	Mice	aberrant cells, chromatid breaks, chromosome breaks, dicentrics, acentric fragments and total aberrations	60

GSH: glutathione, GST: glutathione-s-transferase; GSHpx: glutathione peroxidase; SOD: superoxide dismutase; GI: gastrointestinal; BM: bone marrow; CFU: colony forming units; HPBLS: Human peripheral blood lymphocytes; DPPH: diphenylpicryl hydrazyl radical

been reported to protect against radiation-induced lipid peroxidation and reduction in glutathione concentration [24, 25].

The radioprotective efficacy of ginseng (*Panax ginseng*) has been reported by several workers [26–30]. Ginseng treatment caused recovery of thrombocyte and erythrocyte counts in blood after irradiation [31]. The whole extract of ginseng and the relative protective effects of various fractions (carbohydrate, protein and saponins) have been evaluated. The results showed that the water-soluble whole extract of ginseng provided best protection against radiation induced damage in C3H mice, whereas isolated protein and carbohydrate fractions were less effective, the saponin fraction was ineffective [32]. Similar results were obtained by Kim and coworkers, who found that whole ginseng extract and its fractions increased endogenous spleen colony formation in irradiated mice and also reduced apoptosis in jejunal crypt cells [27]. The radioprotective effect of ginseng root extract on testicular enzymes (acid and alkaline phosphatases and lipid peroxidation) has also been reported [28].

*Podophyllum hexandrum* has been reported to protect against radiation-induced mortality, gastrointestinal damage and embryonic nervous system of developing mice [20, 33–35]. It has also been reported to protect against radiation-induced decline in glutathione-S-transferase, superoxide dismutase in the liver and intestine of irradiated mice [36].

Oral administration of an aqueous extract of guduchi, *Tinospora cordifolia* has been reported to increase the survival of mice exposed to radiation [37]. Treatment of mice with hydroalcoholic extract of *Tinospora cordifolia* has been found to protect against the radiation-induced micronuclei formation and oxidative stress and decline in the mouse survival, spleen CFU and hematological parameters [38].

The fruit pulp of Amala, *Embllica officinalis* (EO) has been reported to increase the survival and inhibit radiation-induced weight loss in mice [39]. *Phyllanthus amarus* has been reported to protect against the radiation-induced decline in white blood cells (WBC), superoxide dismutase, catalase, glutathione-S-transferase, glutathione peroxidase, and glutathione reductase [40].

Daily oral administration of 800 mg/kg body weight (b. wt.) of Rajgira (*Amaranthus paniculatus*) leaf extract for 15 consecutive days before whole body exposure to  $\gamma$ -radiation protected mice against the radiation-induced lethality with a dose reduction factor of 1.36. It increased endogenous spleen colony forming units and spleen weight without any side effects or toxicity. Rajgara extract also arrested radiation-induced lipid peroxidation and the decline in reduced glutathione in the liver and blood of mice [41].

The ethanolic extract of *Piper longum* (pippali) fruits was found to protect mice against the radiation-induced decline in WBC, bone marrow cells  $\alpha$ -esterase positive

cells and GSH. Pippali extract also reduced the elevated levels of glutathione pyruvate transaminase (GPT), alkaline phosphatase (ALP), lipid peroxidation (LPO) in liver and serum of irradiated animals [42].

Several plant and herbal products form the supplements of daily human diet. The potential of dietary ingredients for radioprotection has remained unexplored area until now. The dietary supplements, if found radioprotective may be of crucial importance, as they are in daily human use, nontoxic and have wide acceptability. Jamun, *Syzygium cumini* Linn. Skeels also known as *Eugenia cumini* (family Myrtaceae), and has been reported to possess several medicinal properties in the folklore system of medicine [43]. The micronucleus study of radioprotective effect of dichloromethane and methanol (1:1) extract of jamun (SCE) in human peripheral blood lymphocytes (HPBLs) ascertained its radioprotective potential, where 12.5  $\mu\text{g/ml}$  SCE was found to reduce the micronuclei up to a maximum extent. *In vivo* evaluation further established its radioprotective activity where it was found to reduce radiation-induced sickness, gastrointestinal and bone marrow deaths [7, 44]. Not only leaf but the hydroalcoholic extract of jamun seeds (JSE) also exhibited a greatest protective effect at 80 mg/kg JSE. The JSE was more effective when administered through the intraperitoneal route at equimolar doses than the oral. The JSE treatment protected mice against the gastrointestinal as well as bone marrow deaths with a DRF of 1.24 [45].

Pudina or Mint (*Mentha arvensis* Linn., Family Lamiaceae), a plant, native of Japan, is used as a food seasoner, household remedy, and for industrial purposes [46]. Treatment of mice with 10 mg/kg b. wt. of chloroform extract of mint (*Mentha arvensis* Linn) protected against the radiation-induced sickness, gastrointestinal and bone marrow deaths with a DRF of 1.2. Further it was non-toxic up to a dose of 1000 mg/kg b. wt., the highest drug dose that could be tested for acute toxicity [47]. Pre-treatment of mice with leaf extract of another species of pudina, i.e. *Mentha piperita* has been reported to protect mice against the radiation-induced decline in hematological constituents, serum phosphatase, endogenous spleen colonies formation, spleen weight, goblet cells/villus section and chromosomal damage [48–50].

The rhizome of *Zingiber officinale*, commonly known as ginger, is consumed daily worldwide as a spice and flavoring agent. The rhizome of ginger has been reported to possess diverse medicinal properties in the traditional Indian system of medicine, the Ayurveda, and it is widely used in several medicinal preparations [51]. Administration of 10 mg/kg (i.p) or 250 mg/kg (orally) hydroalcoholic extract once daily, consecutively for 5 days was found to protect mice against the radiation-sickness, gastrointestinal as well as bone marrow deaths with a DRF of 1.15. Ginger has been reported to increase glutathione, reduce lipid peroxidation *in vivo* and scavenging of various free radicals *in vitro* [52, 53].

*Ageratum conyzoides*, (family: Asteraceae) is commonly known as Billy Goat Weed. It has been used in various parts of Africa, Asia and South America for curing various diseases. The study of various doses of alcoholic extract of *Ageratum conyzoides*, Linn. revealed that the best protective dose was 75 mg/kg and it reduced radiation-induced, sickness gastrointestinal as well as bone marrow deaths. A DRF was found to be 1.3. The radioprotective effect was due to scavenging of DPPH (1,1-diphenyl-2-picrylhydrazyl), free radical [54].

*Aegle marmelos* Correa, commonly known as bael, is a spinous tree belonging to family Rutaceae. It is grown throughout the sub-continent as well as Bangladesh, Burma and Srilanka [55]. The hydroalcoholic extract of *Aegle marmelos* (AME) protected cultured HPBLs against the radiation-induced micronuclei at a concentration of 5 µg/ml. It was also reported to scavenge  $\cdot\text{OH}$ ,  $\text{O}_2^-$ , DPPH, ABTS<sup>+</sup> and NO (nitric oxide) radicals *in vitro* in a concentration dependent manner [56]. The radioprotective efficacy of 15 or 250 mg/kg AME was further confirmed in animal studies where its intraperitoneal as well as oral administration has been found to protect mice against the radiation-induced sickness, gastrointestinal and bone marrow deaths and mortality giving a DRF of 1.2. It also protected mice against the radiation-induced lipid peroxidation and elevated GSH concentration in the liver, kidney, stomach and intestine at 31 days post-irradiation. Oral administration also protected mice against the gamma radiation-induced decline in erythrocytes, leukocytes, lymphocytes and clonogenicity of hemopoietic progenitor cells assessed by exogenous spleen colony forming assay. Pretreatment of mice with AME elevated the villus height and the crypt number accompanied by a decline in goblet and dead cell number [57, 58]. Not only leaf but also the hydroalcoholic extract of *Aegle marmelos* fruit administered intraperitoneally at a dose of 20 mg/kg once daily, consecutively for five days found to protect mice against the radiation-induced sickness, gastrointestinal as well as bone marrow deaths with a DRF of 1.1 [59].

Rohituka, *Aphanamixis polystachya* Wall. Parker [*Amoora rohituka*, *Amoora aphanamixis* (Roxb.) Wight & Arn.] is a member of the family Meliaceae. The ethyl acetate fraction of *Aphanamixis polystachya* at a dose of 7.5 mg/kg b. wt. before exposure to 1–5 Gy of whole body gamma-radiation significantly reduced the frequencies of aberrant cells and chromosomal aberrations like acentric fragments, chromatid and chromosome breaks, centric rings, dicentric, exchanges and total aberrations at all post-irradiation scoring times. It also showed a concentration dependent scavenging of hydroxyl, superoxide, 2,2'-diphenyl-1-picryl hydrazyl (DPPH) radicals and the 2,2'-azino-bis-3-ethyl benzothiazoline-6-sulphonic acid (ABTS) cation radicals *in vitro*. EAP treatment also reduced lipid peroxidation in bone marrow cells in a concentration dependent manner [60].

#### Mechanism of action

Ionizing radiations induce reactive oxygen species in the form of  $\cdot\text{OH}$ ,  $\text{H}$ , singlet oxygen and peroxy radicals that follows a cascade of events leading to DNA damage such as single- or double-strand breaks (DSB), base damage, and DNA-DNA or DNA-protein cross-links, and these lesions cluster as complex local multiply damaged sites. The DNA-DSBs are considered the most lethal events following ionizing radiation and has been found to be the main target of cell killing by radiation. The putative mechanisms of radioprotection by plant and herbal radioprotectors are shown in Fig. 1. The radioprotective activity of plant and herbs may be mediated through several mechanisms, since they are complex mixtures of many chemicals. The majority of plants and herbs contain polyphenols, scavenging of radiation-induced free radicals and elevation of cellular antioxidants by plants and herbs in irradiated systems could be leading mechanisms for radioprotection. The polyphenols present in the plants and herbs may upregulate mRNAs of antioxidant enzymes such as catalase, glutathione transferase, glutathione peroxidase, superoxide dismutase and thus may counteract the oxidative stress-induced by ionizing radiations. Upregulation of DNA repair genes may also protect against radiation-induced damage by bringing error free repair of DNA damage. Reduction in lipid peroxidation and elevation in non-protein sulphhydryl groups may also contribute to some extent to their radioprotective activity. The plants and herb may also inhibit activation of protein kinase C (PKC), mitogen activated protein kinase (MAPK), cytochrome P-450, nitric oxide and several other genes that may be

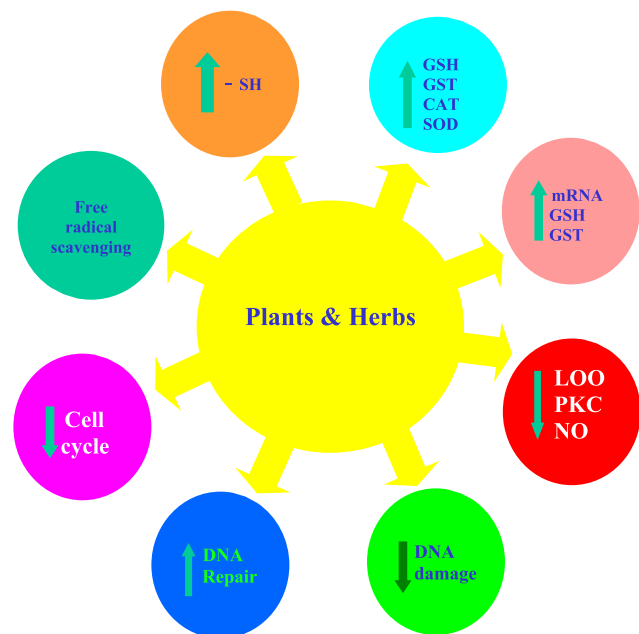


Fig. 1. The putative mechanisms of radioprotection by various plants and herbs.

responsible for inducing damage after irradiation.

## Conclusions

Humans are exposed to ionizing radiations during diagnostic, therapeutic and industrial purposes. Apart from these humans also get exposed to ionizing radiations during air and space travel, background radiation nuclear accidents, and use of electronic devices. Nuclear terror attacks are not distant possibility, therefore it is essential to protect humans from ionizing radiations by pharmacological intervention. Recently, focus of radiation protection has shifted to test the radioprotective potential of plants and herbs in the hope that one day it will be possible to find a suitable pharmacological agent/s that could protect humans against the deleterious effects of ionizing radiation in clinical and other conditions as well as during nuclear terror attack. Majority of plant and herbs described in this review have medicinal properties and are being used in traditional Ayurvedic or Chinese systems of medicine to treat various ailments in humans. Most of these plants and herbs certainly have potential as radioprotectors of future. They protect against the radiation-induced damage by scavenging of free radicals and increasing antioxidant status. Fractionation guided evaluation may result in the development of ideal radioprotector/s in the near future.

## References

- [1] Roentgen, W.C.: Sitzungsberichte Würzburger Physik-med. Gesellschaft, **137**, 132–141, 1895; translation by Arthur Stanton as On a New Kind of Rays. *Nature*, **53**, 274–276, 1896.
- [2] Becquerel, A.H.: On the rays emitted by phosphorescence. *Comptes Rendus*, **122**, 420–421, 1896.
- [3] Patt, H.M., Tyree, E.B., Straube, R.L., and Smith, D.E.: Cysteine protection against X-irradiation. *Science*, **110**, 213–214, 1949.
- [4] Sweeney, T.R.: *Survey of compounds from the antiradiation drug development program of the U.S. Army Medical Research and Development Command*. Government Printing Office, Washington D.C. publication, pp. 308–318, 1979.
- [5] Maisin, J.R.: Bacq and Alexander Award lecture—chemical radioprotection: past, present, and future prospects. *Int. J. Radiat. Biol.*, **73**, 443–450, 1998.
- [6] Cragg, G.M., Newman, D.J., and Snader, K.M.: Natural products in drug discovery and development. *J. Nat. Prod.*, **60**, 52–60, 1997.
- [7] Jagetia, G.C. and Baliga, M.S.: The evaluation of the radioprotective effect of the leaf extract of *Syzygium cumini* (Jamun) in the mice exposed to lethal dose of radiation. *Nahrung/Food*, **47**, 181–185, 2003.
- [8] Yuhas, J.M. and Storer, J.B.: Chemoprotection against three modes of radiation death in the mouse. *Int. J. Radiat. Biol. Relat. Stud. Phys. Chem. Med.*, **15**, 233–237, 1969.
- [9] Brown, D.Q., Graham, W.J., Mackensie, L.J., Pittock, J.W., and Shaw, L.: Can WR-2721 be improved upon. *Pharmacol. Ther.*, **39**, 157–168, 1988.
- [10] Jagetia, G.C., Shrinath Baliga, M., Malagi, K.J., and Sethukumar Kamath, M.: The evaluation of the radioprotective effect of Triphala (an Ayurvedic rejuvenating drug) in the mice exposed to gamma-radiation. *Phytomedicine*, **9**, 99–108, 2002.
- [11] Jagetia, G.C., Baliga, M.S., Aruna, R., Rajanikant, G.K., and Jain, V.: Effect of abana (a herbal preparation) on the radiation-induced mortality in mice. *J. Ethnopharmacol.*, **86**, 159–165, 2003.
- [12] Weiss, J.F., Landauer, M.R., Gunter-Smith, P.J., and Hanson, W.R.: Effect of radioprotective agents on survival after acute intestinal radiation injury. in *Radiation and the Gastrointestinal Tract*, eds. By Dubois, A., King, G.L., and Livengood, D.R., CRC Press, Boca Raton, FL, pp. 183–199, 1995.
- [13] Malaker, K.: Clinical experience with radioprotectors. in *Radioprotectors: Chemical, Biological and Clinical Perspective*, eds. By Bump, E.A. and Malaker, K., CRC Press, Boca Raton, FL, pp. 373–410, 1998.
- [14] Hannequin, D., Thibert, A., and Vaschalde, Y.: Development of a model to study the anti-oedema properties of *Ginkgo biloba* extract. *Presse Med.*, **15**, 1575–1576, 1986.
- [15] Emerit, I., Arutyunyan, R., Oganessian, N., Levy, A., Cernjavsky, L., Sarkisian, T., Pogossian, A., and Asrian, K.: Radiation-induced clastogenic factors: Anticlastogenic effects of *Ginkgo biloba* extract. *Free Radic. Biol. Med.*, **18**, 985–991, 1995.
- [16] Emerit, I., Oganessian, N., Sarkisian, T., Arutyunyan, R., Pogossian, A., Asrian, K., Levy, A., and Cernjavski, L.: Clastogenic factors in the plasma of Chernobyl recovery workers: Anticlastogenic effects of *Ginkgo biloba* extract. *Radiat. Res.*, **144**, 198–205, 1995.
- [17] Shobi, V. and Goel, H.C.: Protection against radiation-induced conditioned taste aversion by *Centella asiatica*. *Physiol. Behavior*, **73**, 19–23, 2001.
- [18] Sharma, J. and Sharma, R.: Radioprotection of Swiss albino mouse by *Centella asiatica* extract. *Phytother. Res.*, **16**, 785–786, 2002.
- [19] Mizina, T.Y. and Sitnikova, S.G.: Antiradiation activity of juice concentrate from *Hippophae rhamnoides* L. fruits. *Rastitel'nye Resursy*, **35**, 85–92, 1999.
- [20] Goel, H.C., Prasad, J., Singh, S., Sagar, R.K., Kumar, I.P., and Sinha, A.K.: Radioprotection by a herbal preparation of *Hippophae rhamnoides*, RH-3, against whole body lethal irradiation in mice. *Phytomedicine*, **9**, 15–25, 2002.
- [21] Agrawal, P.K. and Goel, H.C.: Protective effect of RH-3 with special reference to radiation-induced micronuclei in mice bone marrow. *Ind. J. Exp. Biol.*, **40**, 525–530, 2002.
- [22] Goel, H.C., Agrawala, P.K., Pathania, V., and Malhotra, N.: Immunomodulatory and cytoprotective role of RP-1 in gamma-irradiated mice. *Mol. Cell. Biochem.*, **254**, 73–81, 2003.
- [23] Jagetia, G.C., Uma Devi, P., Shinghatgiri, M., Singh, N., and Kohli, R.: Radiation modifying effect of *Ocimum sanctum* on mouse survival. *Annals of National Academy of Sciences (India)*, 1986.

- [24] Uma Devi, P. and Ganasoundari, A.: Radioprotective effect of leaf extract of Indian medicinal plant *Ocimum sanctum*. *Ind. J. Exp. Biol.*, **33**, 205–209, 1995.
- [25] Ganasoundari, A., Uma Devi, P., and Rao, M.N.A.: Protection against radiation-induced chromosome damage in mouse bone marrow by *Ocimum sanctum*. *Mutat. Res.*, **373**, 271–276, 1997.
- [26] Kim, S.H., Cho, C.K., Yoo, S.Y., Koh, K.H., Yun, H.G., and Ki, M.T.H.: In vivo radioprotective activity of *Panax ginseng* and diethylthiocarbamate. *In Vivo*, **7**, 467–470, 1993.
- [27] Kim, S.H., Son, C.H., Nah, S.Y., Jo, S.K., Byun, M.W., and Shin, D.H.: Modification of radiation response in mice by *Panax ginseng* and diethylthiocarbamate. *In Vivo*, **15**, 407–411, 2001.
- [28] Kumar, M., Sharma, M.K., Saxena, P.S., and Kumar, A.: Radioprotective effect of *Panax ginseng* on the phosphatases and lipid peroxidation level in testes of Swiss albino mice. *Biol. Pharm. Bull.*, **26**, 308–312, 2003.
- [29] Pande, S., Kumar, M., and Kumar, A.: Evaluation of radio-modifying effects of root extract of *Panax ginseng*. *Phytother. Res.*, **12**, 13–17, 1998a.
- [30] Takeda, A., Katoh, N., and Yonezawa, M.: Restoration of radiation injury by ginseng III. Radioprotective effect of thermostable fraction of ginseng extract on mice, rats and guinea pigs. *J. Radiat. Res.*, **23**, 150–167, 1982.
- [31] Yonezawa, M., Katoh, N., and Takeda, A.: Restoration of radiation injury by ginseng. IV. Stimulation of recoveries in CFU and megakaryocyte counts related to the prevention of occult blood appearance in x-irradiated mice. *J. Radiat. Res.*, **26**, 436–442, 1985.
- [32] Zhang, J.S., Sigdestad, C.P., Gemmell, M.A., and Grdina, D.J.: Modification of radiation response in mice by fractionated extracts of *Panax ginseng*. *Radiat. Res.*, **112**, 156–163, 1987.
- [33] Goel, H.C., Prasad, J., Singh, S., Sagar, R.K., Prem Kumar, I., and Sinha, A.K.: Radioprotection by a herbal preparation of *Hippophae rhamnoides*, RH-3, against whole body lethal irradiation in mice. *Phytomedicine*, **9**, 15–25, 2002.
- [34] Goel, H.C., Saji Kumar, S., and Sharma, A.K.: Effects of *Podophyllum hexandrum* on radiation-induced delay of postnatal appearance of reflexes and physiological markers in rats irradiated in utero. *Phytomedicine*, **9**, 447–454, 2002.
- [35] Salin, C.A., Samanta, N., and Goel, H.C.: Protection of mouse jejunum against lethal irradiation by *Podophyllum hexandrum*. *Phytomedicine*, **8**, 413–422, 2001.
- [36] Mittal, A., Pathania, V., Agarwala, P.K., Prasad, J., Singh, S., and Goel, H.C.: Influence of *Podophyllum hexandrum* on endogenous antioxidant defense system in mice: possible role in radioprotection. *J. Ethnopharmacol.*, **76**, 253–262, 2002.
- [37] Pahadiya, S. and Sharma, J.: Alteration of lethal effects of gamma rays in Swiss albino mice by *Tinospora cordifolia*. *Phytother. Res.*, **17**, 552–554, 2003.
- [38] Goel, H.C., Prasad, J., Singh, S., Sagar, R.K., Agrawala, P.K., Bala, M., Sinha, A.K., and Dogra R.: Radioprotective potential of a herbal extract of *Tinospora cordifolia*. *J. Radiat. Res.*, **45**, 61–68, 2004.
- [39] Singh, I., Sharma, A., Nunia, V., and Goyal, P.K.: Radio-protection of Swiss albino mice by *Embllica officinalis*. *Phytother. Res.*, **19**, 444–446, 2005.
- [40] Hari Kumar, K.B. and Kuttan, R.: Protective effect of an extract of *Phyllanthus amarus* against radiation-induced damage in mice. *J. Radiat. Res.*, **45**, 133–139, 2004.
- [41] Krishna, A. and Kumar, A.: Evaluation of radioprotective effects of rajgira (*Amaranthus paniculatus*) extract in swiss albino mice. *J. Radiat. Res.*, **46**, 233–239, 2005.
- [42] Sunila, E.S. and Kuttan, G.: Protective effect of *Piper longum* fruit ethanolic extract on radiation induced damages in mice: a preliminary study. *Fitoterapia*, **76**, 649–655, 2005.
- [43] Warriar, P.K., Nambiar, V.P.K., and Ramankutty, C.: *Indian medicinal plants*. Orient Longman Ltd., Hyderabad, India, 5, pp. 225–228, 1996.
- [44] Jagetia, G.C. and Baliga, M.S.: *Syzygium cumini* (Jamun) reduces the radiation-induced DNA damage in the cultured human peripheral blood lymphocytes: a preliminary study. *Toxicol Lett.*, **132**, 19–25, 2002.
- [45] Jagetia, G.C., Baliga, M.S., and Venkatesh, P.: Influence of seed extract of *Syzygium Cumini* (Jamun) on mice exposed to different doses of gamma-radiation. *J. Radiat. Res.*, **46**, 59–65, 2005.
- [46] CHEMEXCIL. *Selected medicinal plants of India*. Basic Chemicals, Pharmaceutical and Cosmetic Export Promotion Council, Bombay 400 039, India, pp. 205–207, 1992.
- [47] Jagetia, G.C. and Baliga, M.S.: Influence of the leaf extract of *Mentha arvensis* Linn. (mint) on the survival of mice exposed to different doses of gamma radiation. *Strahlenther Onkol.*, **178**, 91–98, 2002.
- [48] Samarth, R.M. and Kumar, A.: *Mentha piperita* (Linn.) leaf extract provides protection against radiation induced chromosomal damage in bone marrow of mice. *Indian. J. Exp. Biol.*, **41**, 229–237, 2003.
- [49] Samarth, R.M., Goyal, P.K., and Kumar, A.: Modulation of serum phosphatases activity in Swiss albino mice against gamma irradiation by *Mentha piperita* Linn. *Phytother. Res.*, **16**, 586–589, 2002.
- [50] Samarth, R.M., Goyal, P.K., and Kumar, A.: Modulatory effect of *Mentha piperita* (Linn.) on serum phosphatases activity in Swiss albino mice against gamma irradiation. *Indian. J. Exp. Biol.*, **39**, 479–482, 2001.
- [51] Warriar, P.K., Nambiar, V.P.K., and Ramankutty, C.: *Indian medicinal plants*. Orient Longman Ltd., Hyderabad, India, 5, pp. 225–228, 1996.
- [52] Jagetia, G.C., Baliga, M.S., Venkatesh, P., and Ulloor, J.N.: Influence of ginger rhizome (*Zingiber officinale* Rosc) on survival, glutathione and lipid peroxidation in mice after whole-body exposure to gamma radiation. *Radiat Res.*, **160**, 584–592, 2003.
- [53] Jagetia, G.C., Baliga, M.S., and Venkatesh, P.: Ginger (*Zingiber officinale* Rosc.), a dietary supplement, protects mice against radiation-induced lethality: mechanism of action. *Cancer Biother. Radiopharm.*, **19**, 422–435, 2004.
- [54] Jagetia, G.C., Shirwaikar, A., Rao, S.K., and Bhilegaonkar, P.M.: Evaluation of the radioprotective effect of *Ageratum conyzoides* linn. Extract in mice exposed to different doses of gamma radiation. *J. Pharm. Pharmacol.*, **55**, 1151–1158,

- 2003.
- [55] Sharma, R.K. and Dash, B.: *Carka Samhita* Volume II. Chowkamba Sanskrit Series Office, Varanasi, India, 1998.
- [56] Jagetia, G.C., Venkatesh, P., and Baliga, M.S.: Evaluation of radioprotective effect of *Aegle marmelos* (L.) Correa in the cultured human peripheral blood lymphocytes exposed to different doses of  $\gamma$ -radiation: a micronucleus study. *Mutagenesis*, **18**, 387–393, 2003.
- [57] Jagetia, G.C., Venkatesh, P., and Baliga, M.S.: Evaluation of the radioprotective effect of bael leaf (*Aegle marmelos*) extract in mice. *Int. J. Radiat. Biol.*, **80**, 281–290, 2004.
- [58] Jagetia, G.C. and Venkatesh, P.: Radioprotection by oral administration of *Aegle marmelos* (L.) Correa in vivo. *J. Environ. Pathol. Toxicol. Oncol.*, **24**, 315–32, 2005.
- [59] Jagetia, G.C., Venkatesh, P., and Baliga, M.S.: Fruit extract of *Aegle marmelos* protects mice against radiation-induced lethality. *Integrat. Cancer Ther.*, **3**, 323–332, 2004.
- [60] Jagetia, G.C. and Venkatesha, V.A.: Treatment of mice with stem bark extract of *Aphanamixis polystachya* reduces radiation-induced chromosome damage. *Int. J. Radiat. Biol.*, **82**, 197–209, 2006.