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Review

Molecular mechanisms in Alzheimer's disease and the impact of physical exercise with advancements in therapeutic approaches

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Abstract: Alzheimer's disease (AD) is one of the most common, severe neurodegenerative brain disorder characterized by the accumulation of amyloid-beta plaques, neurofibrillary tangles in the brain causing neural disintegration, synaptic dysfunction, and neuronal death leading to dementia. Although many US-FDA-approved drugs like Donepezil, Rivastigmine, Galantamine are available in the market, their consumption reduces only the symptoms of the disease but fails in potency to cure the disease. This disease affects many individuals with aging. Combating the disease tends to be very expensive. This review focuses on biochemical mechanisms in the neuron both at normal and AD state with relevance to the tau hypothesis, amyloid hypothesis, the risk factors influencing dementia, oxidative stress, and neuroinflammation altogether integrated with neurodegeneration. A brief survey is carried out on available biomarkers in the diagnosis of the disease, drugs used for the treatment, and the challenges in approaching therapeutic targets in inhibiting the disease pathologies. This review conjointly assesses the demerits with the inefficiency of drugs to reach targets, their side effects, and toxicity. Optimistically, this review directs on the advantageous strategies in using nanotechnology-based drug delivery systems to cross the blood-brain barrier for improving the efficacy of drugs combined with a novel neuronal stem cell therapy approach. Determinately, this review aims at the natural, non-therapeutic healing impact of physical exercise on different model organisms and the effect of safe neuromodulation treatments using repetitive Transcranial Magnetic Stimulation (rTMS), transcranial Electrical Stimulation (tES) in humans to control the disease pathologies prominent in enhancing the synaptic function.

Keywords: Alzheimer's disease; amyloid-beta plaques; dementia; neurofibrillary tangles; neuroinflammation; neurodegeneration; physical exercise

Abbreviations: AD: Alzheimer's disease; AB: Amyloid-beta; NFTs: Neurofibrillary Tangles; APP: Amyloid Precursor Protein; sAPPa: soluble Amyloid Precursor Protein alpha; AICD: Amyloid Precursor Protein Intra Cellular Domain; NMDAR: N-Methyl-D-Aspartate Receptor; AMPAR: α-Amino-3-hydroxy-5-Methyl-4-isoxazole Propionic acid Receptor; nAChRs: nicotinic Acetylcholine Receptors; mAChRs: muscarinic Acetylcholine Receptors; sAPPß: soluble Amyloid Precursor Protein beta; ROS: Reactive Oxygen Species; PKC: Protein Kinase C; PKA: Protein Kinase A; ERK2: Extracellular signal-Regulated Kinases2; CDK5: Cyclin-Dependent Kinases 5; AchE: AcetylcholinEsterase; CSF: Cerebro Spinal Fluid; PET: Positron Emission Tomography; BBB: Blood-Brain Barrier; PE: Physical Exercise; DLB: Dementia with Lewy Bodies; FTD: FrontoTemporal Dementia; MRI: Magnetic Resonance Imaging; fMRI: functional Magnetic Resonance Imaging; rTMS: repetitive Transcranial Magnetic Stimulation; tES: transcranial Electrical Stimulation; tDCS: transcranial Direct Current Stimulation; tACS: transcranial Alternating Current Stimulation; tRNS: transcranial Random Noise Stimulation; tPCS: transcranial Pulsed Current Stimulation, MCI: Mild Cognitive Impairment; Drp1: Dynamin-related protein-1; PSEN 1: Presenilin-1; PSEN 2: Presenilin-2; IDE: Insulin Degrading Enzyme; PD: Parkinson's Disease; ApoE: Apolipoprotein-E; PKB: Protein Kinase B; $GSK3\alpha/\beta$: Glycogen synthase kinase $3\alpha/\beta$; ETC: Electron Transport Chain; IGF-1: Insulin-like Growth Factor-1; IL-6: Interleukin-6; NOS-2: Nitrous Oxide Synthase-2; NF-kB: Nuclear Factor Kappa B; LRP-1: Lipoprotein Receptor-related Protein-1; OR-A: Orexin-A; OR-B: Orexin-B; GPCRs: G-Protein-Coupled Receptors; OX1R: Orexin Receptor-type-1; OX2R: Orexin Receptor-type-2; BDNF: Brain-Derived Neurotrophic Factor; OSA: Obstructive Sleep Apnea (OSA); CDH: Central Disorders of Hypersomnolence; REM: Rapid Eye Movement; EEG: Electroencephalogram; TBI: Traumatic Brain Injury; 123I-MIBG: 123I-Meta-IodoBenzylGuanidine; [11C]PiB: [11C]-Pittsburgh compound B; ¹⁸F: Fluorine 18-labeled; MoCA: Montreal Cognitive Assessment; T-tau: Total tau; P-tau: phosphorylated-tau; VLP-1: Visinin-Like Protein; NFL: Neuro Filament Light Protein; HFABP: Heart Fatty Acid Binding Protein; NSE: Neuron-Specific Enolase; CHI3L1: Chitinase-3-like protein-1; TNFSF10: Tumor Necrosis Factor Ligand Superfamily member-10; ACE: Angiotensin-Converting Enzyme; 3xTg-AD: triple-transgenic-AD; VBM: Voxel-Based Morphometry

1. Introduction

More than a hundred billion long branching extensions of neurons form complex molecular connections between inter-neuron circuits through trillions of synapses in the adult human brain [1]. Electrochemical nerve impulses travel rapidly between neural circuits regulating sensations, language, memories, thinking, skills, feelings, emotions with control, and co-ordinated body movements [2]. Neurodegenerative brain disorders are characterized by damaged brain cells with loss of neuron connections, drastically affecting a person's ability to carry out daily activities, medically termed as, Dementia [3]. Globally, dementia is the fifth leading cause of death with a new case every three seconds. Currently, about 23 million in the Asia Pacific [4], 8.8 million in Europe [5], 5.8 million in the USA [6], 5.3 million in India [7], 50 million people worldwide are living with dementia and it would reach 152 million by 2050 [8]. According to the global survey from world Alzheimer's reports in 2020, the total estimated worldwide cost of dementia is one-trillion US \$ and would reach two-trillion US \$ by 2030 [9].

AD is a type of progressive neurodegenerative disorder and the most common cause of dementia in aging groups [10]. The first case was reported in 1906 by Dr. Alois Alzheimer, a German physician

who identified the typical disease of the brain in a 51 years old woman patient; Auguste Deter with series of devastating symptoms like trouble in understanding, planning, execution of the task, changes in mood, psychological worsening, and profound memory loss [11]. Autopsy of the brain revealed pervasive atrophy of the cerebral cortex, brain tissues stained with silver salts showed distinct dense deposits of neuritic plaques [12]. Accumulation of abnormal protein aggregates or amyloid-beta (A β) plaques in the synapse region between communicating neurons interferes with nerve impulse transmission and leads to neuron death [13]. The internal supporting structure of a neuron is made up of integrated tubular microtubules, bridged by tau-proteins that stabilize the neuron [14]. Tau is an important cytoskeletal protein that maintains the structural stability of neurons associated with microtubule assembly, helps in nutrient transport [15].

Clinical aspects of Dementia are categorized into three stages, the early or mild stage with deterioration in learning and memory, language difficulties, declined vocabulary fluency, execution of functions, changes in personality and behavior, problems in olfactory and other sensory perception, loss in episodic, semantic, and implicit memories, depression with Mild Cognitive Impairment (MCI). Moderate or middle stage with progressive neuropsychiatric loss of reading, writing skills, difficulty in speech with incorrect word substitution, wandering, anxiety, irritability, aggression, agitation, delusion, sundowning, and loss of long-term memory. Severe or late-stage characterized by loss of speech with language reduced to phrases or only words, lack of emotion, fatigue, and decline in muscle mass with complete dependency on caregivers [16,17]. Dementia is caused by multiple etiological conditions like morphological modification in the brain with narrowed small blood vessels and declined vascular density followed by endothelial dysfunction which leads to vascular dementia. Small blood vessel narrowing in the brain is termed arteriosclerotic dementia. Decreased cerebral blood flow in the brain leads to the oxygen-depleted hypoxic condition. Inflammation of vessels in the brain causes vasculitis with ischemic hemorrhage and stroke. The molecular events of vascular dementia are closely associated with dementia caused by AD pathologies—A β plaques and neurofibrillary tangles. In turn, the accumulation of abnormal α -synuclein protein aggregates to form Lewy bodies in the neurons with brain atrophy causes dementia in Parkinson's Disease (PD). The atrophy of the frontal and temporal lobe in the brain causes frontotemporal dementia [18]. Neural loss, gliosis, spongiosis, neuroinflammation, impairment in language, physical inactivity, cognitive impairment, stress, hypertension, confusion, irritation, depression, and severe memory loss are the overall complex characteristics of dementia [19].

This review focus on the different molecular hypothesis of tau forming neurofibrillary tangles (NFTs), amyloidogenic processing of amyloid precursor protein in forming amyloid plaques, interconnected molecular events involved in neuroinflammation and neurodegeneration. This review sheds light on the factors influencing dementia, types of biomarkers available for the diagnosis of AD, activity, and toxicity of various drugs. As a measure of habitual curative, the vital effects of physical therapies and the outbreaks of research in divergent model organisms successful in decreasing the $A\beta$ plaques, tau phosphorylation, NFTs, neuroinflammation in the brain put forward ideas with multiple strategies on controlling disease pathologies and future therapeutic perspectives in dealing with AD [20].

2. Molecular mechanisms in AD

2.1. Tau hypothesis

Disaggregation of tau proteins forms NFTs inside the neurons and blocks the transport of nutrients lead to neurodegeneration, termed tauopathy [21,22]. In the AD state, microtubules dissociate from tau proteins by chemical changes due to hyperphosphorylation and pairs with another tau protein threads forming NFTs involving the mechanism of the tau hypothesis (Figure 1) [23].

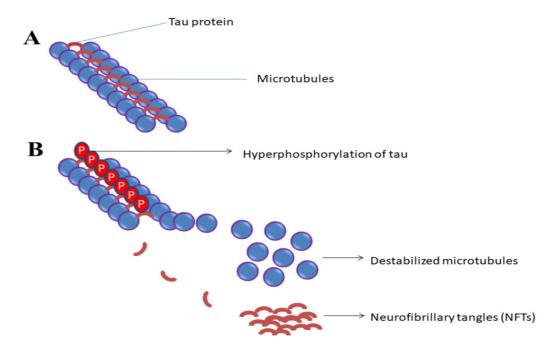


Figure 1. Role of Microtubules, tau proteins in neuron stability, and hyperphosphorylation in neural disintegration. (A) Microtubules and tau protein together stabilize structural integrity in a healthy neuron. (B) Hyperphosphorylation of tau leads to dissociation of tau proteins and destabilization of microtubules from the neuron. The dissociated tau filaments pairs to form NFTs, structural integrity of the neuron are lost in the disease state.

A β neuronal plaques and NFTs are the gold standard hallmarks of AD that develop gradually and spreads in the hippocampus and cerebral cortex regions of the brain [24]. Pathophysiology of the AD brain characterizes degeneration of neurons in the frontal, temporal, parietal lobes of the cerebral cortex, with cingulate gyrus and certain subcortical regions influencing synaptic dysfunction [25]. Due to cortical atrophies in AD, the gyri-folds of the brain diminish and the sulci-spaces in the folds of the brain are grossly enlarged. In AD state, the area of the brain shrinks in invariably in size different from that of a normal brain [26].

2.2. Molecular mechanism in the brain at normal state

A larger transmembrane protein Amyloid Precursor Protein (APP) present on the neuronal membrane is involved in the growth, development, survival, and repair of neurons [27]. APP has two

terminal ends spanning the neuronal membrane; N-terminal and C-terminal. An enzyme, α -secretase recognizes and cleaves the C-terminal end of APP near the cell surface to generate soluble Amyloid Precursor Protein alpha (sAPP α), which helps for neuronal plasticity and outgrowth of neurons [28]. sAPP α is further cleaved by an enzyme, γ -secretase to generate a secreted fragment called AICD (Amyloid Precursor Protein Intra Cellular Domain), which helps for neuronal transcription regulation and translocation [29]. Non-amyloidogenic APP processing (Figure 2) by α -secretase and γ -secretase sequentially is the scenario of events in the brain at normal conditions [30].

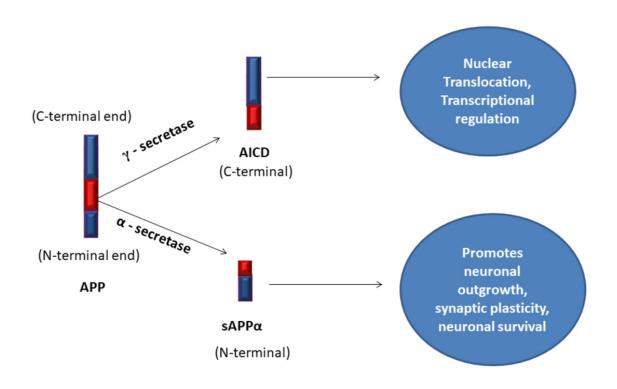


Figure 2. Non-amyloidogenic-Amyloid Precursor Protein (APP) processing by α -secretase and γ -secretase in a healthy neuron.

 α - secretase is the first enzyme that initiates the pathway of events, called the α -secretase pathway. In the normal state, the receptors on neurons are free to accept ligands for normal nerve impulse transmission. The glucose transporters on the neuron transport glucose, N-Methyl-D-Aspartate Receptor (NMDAR), α -Amino-3-hydroxy-5-Methyl-4-isoxazole Propionic acid Receptor (AMPAR) transport Na⁺, Ca²⁺ for maintaining homeostasis of synaptic transmission [31]. Nicotinic Acetylcholine Receptors (nAChRs) and muscarinic Acetylcholine Receptors (mAChRs) binds to acetylcholine neurotransmitters helping neurotransmission between communicating neurons [32,33].

2.3. Molecular mechanism in the brain at AD state

In the AD state, instead of α -secretase, an enzyme β -secretase or β -Amyloid Cleaving Enzyme-1 (BACE-1) performs amyloidogenic-APP processing from the C-terminal end to form

soluble Amyloid Precursor Protein beta (sAPP β). Eventually, sAPP β enters death receptor-6 on the neural membrane and activates caspases (caspase-6). The activated caspases initiate apoptotic pathways and induce neural death (Figure 3) [34].

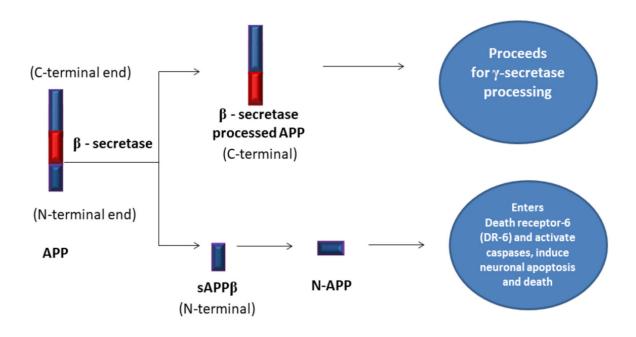


Figure 3. Amyloidogenic-Amyloid Precursor Protein (APP) processing by β -secretase in AD state.

The remaining membrane-bound APP is further recognized by an enzyme, γ -secretase generates A β monomer fragments of (A β 40/A β 42) 40–42 amino acids length [35]. Out of which, A β 40 is majorly formed, leaving only around 10% A β 42. Many monomers of A β peptide aggregates form dense, insoluble oligomers or senile plaques [36]. Amyloid hypothesis comprises the cleavage of APP from BACE-1 to form A β -peptides. The misfolded peptides formed are different in conformation and are released from donor neurons to the extracellular space either as naked protein or vesicles as exosomes taken up by recipient neurons through receptor-mediated endocytosis [37]. The formed A β 40/A β 42 accumulates on glucose transporter receptor [38], NMDAR [39], AMPAR [40], nAChR [41], mAChRs [42] causing impairment in synaptic transmission by blocking ion channels and neurotransmitters through calcium dysregulation [43]. The formation of A β plaques could be linked to preceding cortical tau pathology, but A β independent regulators like Apolipoprotein-E (ApoE), cholesterol metabolism, receptor-mediated endocytosis, and microglial activation may induce tau pathology [44].

A β plaques formed in the neurons activate microglia and astrocytes that produce chemokines and cytokines involved in the formation of Reactive Oxygen Species (ROS) [45]. This creates mitochondrial oxidative stress and activates series of apoptotic caspases by the production of p53, Bad, and Bax production leading to lipid peroxidation, membrane damage, and neuronal death [46,47]. ROS formed by A β activates—protein kinase C (PKC), protein kinase A (PKA), and Extracellular Signal-Regulated Kinases2 (ERK2), induces hyperphosphorylation of tau and destabilizing microtubules forming NFTs [48]. Hyperphosphorylation of tau is also mediated by activation of protein

kinase B (PKB) or Akt to activate Glycogen Synthase Kinase $3\alpha/\beta$ or GSK $3\alpha/\beta$ [49]. P35-Calpain mediated activation of Cyclin-Dependent Kinases 5 (CDK5) and P25 also induces hyperphosphorylation of tau to form NFTs and neuronal death (Figure 4) [50].

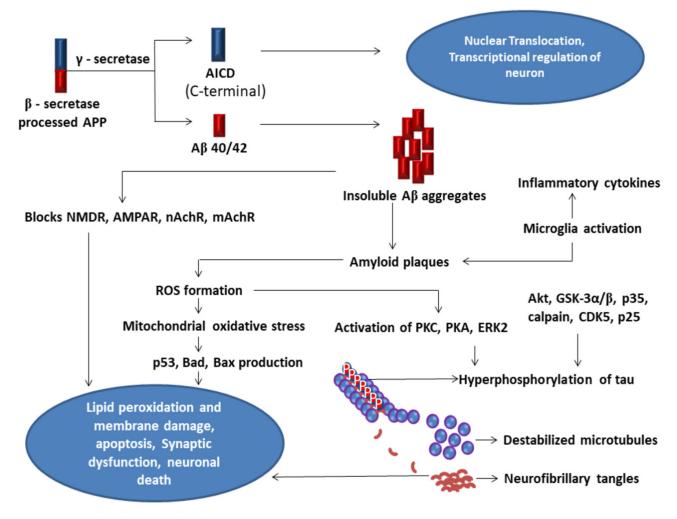


Figure 4. Multiple molecular factors involved in neural death—Post to β-secretase processing the remained C-terminal APP-transmembrane protein is recognized and cleaved by γ -secretase to form AICD which helps neuronal regulation by transcription and translocation. Cleavage by γ -secretase also generates Aβ short peptide of 40–42 amino acids length (Aβ40/Aβ42). Monomers of Aβ peptide form Aβ clumps of amyloid plaques. The formed amyloid plaques block NMDAR, AMPAR, nAChR, mAChRs on the neuronal membrane and impair synaptic transmission. Aβ plaques induce mitochondrial oxidative stress by the generation of ROS and activate series of caspases with the production of p53, Bad, Bax inducing apoptosis with neuron death. Amyloid plaques also activate inflammatory cells like astrocytes and microglia to produce chemokines and cytokines to induce inflammatory reactions. ROS formed by Aβ activatesPKC, PKA, ERK2, and hyperphosphorylates tau to disintegrate leads to disassociation of microtubules, forms clumps of NFTs. Activation of PKB or Akt to activate GSK3α/β, P35-Calpain mediated activation of CDK5 and P25 also induces hyperphosphorylation of tau to form NFTs, all together ultimately leads to neural death. Regulation of memory and learning in the brain is controlled by the cholinergic neurotransmitter system with AcetylcholinEsterase (AchE) enzyme found in neuromuscular junction acts through hydrolytic activity [51]. Acetylcholine is an important neurotransmitter required for regular synaptic transmission [52]. AchE degrades acetylcholine into choline and acetate terminating a synaptic transmission. Degeneration of the cholinergic system is an observed mechanism in AD, where the levels of acetylcholine are drastically reduced by the activity of AchE [53].

2.4. Integrated connection of mitochondrial oxidative stress and inflammation with $A\beta$

Aging cell degenerates by free radicals generated through oxidative imbalance forming ROS [54]. Enzymes like catalase, superoxide dismutase, glutathione peroxidase combat ROS by antioxidant defense mechanism [55]. If anti-oxidant enzyme levels decrease or levels of ROS exceed the toxic threshold due to overproduction, it induces oxidative stress in a cell [56] leading to mitochondrial dysfunction which is a common phenomenon in aging diseases like AD, PD, and other neurodegenerative disorders [57]. Mitochondria, being the powerhouse of the cell produce a majority of ATPs by coupling electron transfer to the pumping of protons across the inner mitochondrial membrane [58]. In certain cases, electrons escape the Electron Transport Chain (ETC) and cut oxygen to ROS like \cdot O₂, \cdot OH, and H₂O₂ causing oxidative damage [59]. Brain cells use more than 20% of the oxygen consumed by the body and form a high rate of ROS during oxidative phosphorylation [60].

Aging-associated AD causes early cellular changes in the mitochondria causing dysfunction. Aß enters mitochondria, induces oxidative stress with free radicals, damages mitochondrial membrane, decreases cytochrome oxidase activity, blocks mitochondrial protein transport, alters ETC, and inhibits ATP production. A β aggregates in the synaptic terminals degenerate neurons, blocks neural circuits, and lead to cognitive dysfunction [61]. Employing mitochondrial therapeutic approaches on using natural and targeted anti-oxidants for scavenging free radicals maintains mitochondrial functioning with enhanced ATP production, decreases lipid peroxidation, and protects neurons from oxidative stress [62]. Studies using cDNA microarray in APP-Tg2576 transgenic mice model for gene expression related to mitochondrial energy metabolism and apoptosis showed up-regulation in 2-months aged-Tg2576 mice, 5 & 18-months aged-Tg2576 mice. In situ hybridization confirmed the mitochondrial genes-ATPase-6, heat shock protein-86 (HSP-86), programmed cell death gene-8 (PCD-8) are up-regulated in Tg2576 mice compared to wild type mice, which correlates with mutated APP/A β -induced mitochondrial energy metabolism impairment, oxidative damage in the neurons of hippocampal regions, and cerebral cortex [63]. Removal of damaged mitochondria from the cell is a normal process called Mitophagy. Excessive A β , p-tau and ROS production induces impaired autophagy and mitophagy in aging and AD [64]. Dynamin-related protein1 (Drp1) are natural enhancers that regulate mitochondrial fission and induce mitophagy. Drp1 is synergistically associated with A β and p-tau inducing abnormal mitophagy in AD, inhibited Drp1 levels up-regulate normal mitophagy in AD [65]. Formation of NFTs due to hyperphosphorylation and destabilization of tau from microtubule assembly is associated with oxidative stress, lipid peroxidation, decreased Insulin-like Growth Factor-1 (IGF-1), $A\beta$ oligomers mediated by astrocytes, intraneural $A\beta$ accumulation, impaired axonal transport, caspases activated mitochondrial apoptosis, reduced ATP synthesis and synaptic dysfunction [66].

The brain is rich in lipids with unsaturated fatty acids, being easy targets for lipid peroxidation. It has a weak antioxidant defense mechanism compared to other organs with fewer ROS detoxifying enzymes compared to the liver and kidney [67]. A higher level of iron catalyzes the formation of ROS in certain areas of the brain [68]. Glutathione, tri-peptide functions in defense against reactive oxygen species by detoxifying ROS and reduction of peroxides [69]. Oxidative damage caused by A β induces toxicity in the cerebral region of the brain by inhibition of mitochondrial enzymes like α -ketoglutarate dehydrogenase, cytochrome c oxidase, and pyruvate dehydrogenase leading to reduced levels of mitochondria [70]. The formation of Aβ plaques and NFTs induces activation of inflammatory cells such as astrocytes & microglia to secrete pro-inflammatory cytokines like Tumour Necrosis Factor-α, Interleukin-6 (IL-6), and anti-inflammatory intermediates [71]. Astrocytes activated by $A\beta$ also release chemokines and cytokines including IL- β , Nitrous Oxide Synthase-2 (NOS-2) [72]. Between neurons and glial cells, chemokines act as messengers and attract microglia to further secrete pro-inflammatory mediators which collectively induce neuronal damage [73]. A nuclear transcription factor called Nuclear Factor Kappa B (NF-kB) regulates many mechanisms including cell growth and development, cell survival, inflammatory responses, and apoptosis [74]. A β has an impact to increase NF- κ B activity in brain cells with degenerating neurons [75]. Death of neurons may take place either by inflammation of neurons called necrosis or by hyperactivation of apoptosis induced by death caspases [76]. Unfortunately, the loss of neurons in the adult brain cannot be compensated by the generation of new neurons [77].

3. Risk factors influencing dementia in AD

3.1. Impact of obesity, hyperinsulininsm, diabetes & cardiovascular diseases in dementia associated with AD

Obesity and dementia are closely associated, the fat cells in the brain release adipose secreted proteins with inflammatory cytokines induces increased blood supply, damages white matter in the brain, and causes vascular dementia with cognitive impairment, increases the risk of AD [78]. Diet rich in carbohydrates and saturated lipids is one of the major causes of obesity which affects insulin secretion and glucose metabolism in the cerebral region of the brain. Oxidative stress, impaired insulin secretion, activation of inflammatory cytokines is the common intracellular mechanisms in Type-2 Diabetes and AD [79]. Insulin is a hormonal protein regulating blood sugar level which is also transported from the peripheral nervous system to the brain, especially essential for cognitive development of the brain. In insulin-deficient conditions, the risk for obesity increases, and insulin uptake by the brain declines which leads to dementia [80]. The expression of the level of insulin and its receptors decline by normal aging and still diminish in AD [81].

Combined factors with age and obesity, elevated insulin levels are associated with activation of neuro-inflammatory signaling pathways and co-laterally increase amyloid-beta deposits in the brain causing AD [82]. As a common strategy, routine physical activity and a disciplined dietary lifestyle with control in intake of fat and carbohydrates may be the key to reduce the risk of complications associated with insulin resistance, diabetes, and AD. Obesity and stress with high blood pressure induced hypertension are interconnected in causing endothelial dysfunction, increasing cardiovascular morbidity with atherosclerosis and ischemic white and grey matter atrophy with inadequate blood flow to the brain, accumulation of amyloid plaques leading to cognitive dysfunction with encephalopathy and dementia [83,84]. The dietary lifestyle and smoking habits impact metabolic function and increase the risk of association between diabetes and dementia [85].

Brain MRI studies in patients with diabetes are two-fold more prone to develop brain atrophy with a high risk of developing AD as compared to normal without diabetes [86,87]. Studies on rat model (BBZDR/Wor) with diabetes revealed loss of neurons, neural dystrophy, elevated amyloid levels, and tau hyperphosphorylation with decreased expression of insulin and IGF-1 receptors [88]. Zinc-binding metalloprotease Insulin Degrading Enzyme (IDE) binds to substrates insulin and amyloid- β with more affinity for insulin, predictable to be associated with the interconnection between diabetes and AD. Studies in the transgenic AD mice models fed with a high-fat diet reported impaired insulin signaling with diminished IDE levels and increased amyloid- β levels [89]. The loss of IDE function may induce hyperinsulinemia correlated with insulin resistance and altered glucose tolerance, but the exact underlying mechanisms remain unclear. IDE inhibition by drugs showed a reduction in the degradation of insulin and amyloid-beta [90]. Hyperinsulinemia in diabetes resulted in suppressed degradation of amyloid-beta by IDE which further induces AB deposition in the brain [91]. Chronic hyperinsulinemia alters the insulin signaling pathway and increases insulin resistance in the brain with reduced uptake of insulin transport across the blood-brain barrier (BBB) induces cognitive dysfunction with dementia [92]. Diabetes and AD are closely related to many characteristic risk factors which include vascular parameters like stress, hypertension, anxiety, high blood pressure, cerebrovascular diseases, endothelial function, hypercholesterolemia, lipoprotein receptors, and oxidative stress. Lipoprotein receptor-related protein-1 (LRP-1) is a key signaling protein involved in metabolic defects and neurodegenerative diseases including AD. Studies in rats have shown improvement in A\beta-induced learning and memory impairments with functional LRP-1 regulating signaling pathways for AB clearance from the brain [93,94]. The study reported an inverse in dementia by regulation of amyloid beta-processing and synaptic plasticity in the hippocampus of the brain monitored by a unique Leptin hormone secreted by adipose cells and enterocytes in the small intestine. Leptin acts as a neuroprotective hormone, inhibits the activity of the BACE enzyme, activates A β degradation, inhibits GSK3 β , inhibits oxidative stress, and inhibits long-term potentiation and depression. Obesity in middle age groups with Leptin resistance increases the risk for developing AD [95].

3.2. Genetic aspects of dementia in AD

Mutations in multiple genes cause autosomal early-onset and late-onset familial AD in more than 1% of cases of AD. APP-coding gene located on chromosome 21q21 exhibit majority of missense mutations and comprises 10–15% early-onset familial AD between 40–50 years of age [96]. APP is degraded into different product lengths of amino acids APP563, APP695, APP714, APP751, and APP770 isoforms [97]. APP695 is the predominant isoform in neurons is confined to the central nervous system and a larger part of early-onset familial AD mutations are responsible for elevated A β 42 comparatively to A β 40 in the brain [98,99].

Presenilin-1 (PSEN 1) gene with the chromosomal location on 14q24.2 codes for membrane protein γ -secretase complex [100]. 18–50% autosomal dominant early-onset familial AD in 25–65 years age group is most commonly caused by PSEN1 missense mutation suppressing γ -secretase activity with an increased ratio of A β 42 over A β 40 accelerates dementia [101,102]. PSEN1-L166P mutation induces a high rise in A β 42 production associated with defective intracellular notch signaling domain [103].

Presenilin-2 (PSEN 2) gene is located on chromosome 1q42.13 codes for aspartyl-protease γ -secretase [104]. Rare PSEN2 missense, point, and substitution mutations cause early-onset familial AD affecting high variable age groups between 45–88 years [105]. Studies in human and mice models reported PSEN2 mutations producing fewer rates of A β 42/A β 40 in the neurons in contrast to PSEN1 [106].

APOE gene with chromosomal location 19q.13.2 includes gene clusters APOC1, APOC2, APOC4 with three allelic genotypes APOE \in 2, APOE \in 3, and APOE \in 4. APO \in 4 genotype mediates A β deposition, hyperphosphorylation of tau linked with high risk for developing early-onset AD, even associated with trauma-stroke reported in humans and transgenic mice [107–110]. APOE is involved in cholesterol metabolism, APOE ϵ 4 enhances amyloid aggregation by receptor-mediated endocytosis, induces cholesterol efflux from neurons and astrocytes. APOE ϵ 3 isoform has a high affinity for binding to A β compared to APOE ϵ 4 [111,112]. Patients with AD most commonly display APO ϵ 4 based mitochondrial deterioration, amyloid plaques, and NFTs pathologies compared to other APOE allelic forms. APOE ϵ 4 allelic form has a major prevalence for late-onset AD above 65 years [113,114].

3.3. Role of orexin receptors, anxiety, depression, sleep-deprivation, & traumatic brain injury associated with dementia in AD

Orexins—Orexin-A (OR-A), Orexin-B (OR-B) are hypocretin neuropeptide ligands secreted majorly from the lateral hypothalamus, also secreted from locus coeruleus, tuberomammillary nucleus, paraventricular nucleus, and raphe nuclei regions of the brain that binds and activates G-protein-coupled receptors (GPCRs), Orexin receptor-type-1 (OX1R) and Orexin receptor-type-2 (OX2R). Orexin receptors mediate multiple molecular signaling mechanisms connected to feeding behavior, circadian rhythm, energy homeostasis, and drug addiction, sleep disorders, depression, dementia, ischemic stroke, and associated with the pathogenesis of AD [115–119]. In AD, the impaired cholinergic pathway alters the sleep-wake cycle with insomnia at night and excess sleep in the day, OR-A increases Aβ42, P-Tau levels in the Cerebro Spinal Fluid (CSF) causes rapid eye movement sleep, orexinergic neurons in the hypothalamus degenerate, and cognitive function deteriorates [120-122]. Neuropeptide 26RFa (QRFP), an endogenous ligand of the human orphan receptor GPR103 and GPCRs expressed in the hypothalamic nuclei of the brain executes the same function as orexins. GPR103 correlates with 48% protein sequence homology with OX1R, 47% homology with OX2R. GPR103 forms a functional heterodimer with OXRs signaling cascade function, closely associated with potential neuroprotective effects in AD [123]. The expression and signaling pathways of QRFP/GPR103 are yet to be understood.

Alterations in sleep with deprived sleep quality less than 6 hours and excessive sleep above 9 hours are associated with impairment in cognition [124,125]. Globally American Academy of Sleep Medicine classifies sleep disorders into six major categories which are linked to having a high risk of causing AD [126]. The first group is insomnia, a depressive condition with difficulty falling asleep associated with the pathogenesis of AD by inducing tau aggregation, amyloid-beta accumulation, neuroinflammation and decreases brain-derived neurotrophic factors (BDNFs) [127]. Secondly, Sleep-related breathing disorders with obstructive sleep apnea (OSA) is a potential risk factor in AD characterized by oxygen-deprived hypoxic repetitive paused breathing, anxiety, fragmentation of sleep, depression, excess day time sleep with naps, the prevalence for OSA is over 70% in AD [128]. Sleep-related hypoventilation and hypoxemia disorders are other sleep-related breathing disorders that

altogether promote AD pathogenesis [129]. Thirdly, Central Disorders of Hypersomnolence (CDH) with narcolepsy type 1 & 2, a condition with heavy drowsiness with daytime naps and Idiopathic hypersomnia, a chronic neurological disorder with unsatisfying sleep even after a full night sleep. Kleine-Levin syndrome, a type of rare CDH syndrome with repeating episodes of excessive sleep up to 20hrs a day with behavioral and cognitive anomalies is associated with AD [130]. The fourth group of sleep disorders includes disturbances in circadian rhythm inducing sleep-wake disorders with stress, depression, anxiety, and modification in the melatonin hormonal secretion linked with AD [131]. Parasomnias are the fifth type characterized by abnormal behavior, anxiety, agitation, emotional breakdown, dream with hallucination, falling asleep, sleep disturbances, rapid eye movement (REM) with undesirable body movements, walking or talking during sleep. Lastly, Sleep-Related Movement Disorders with symptoms include drowsiness, clumsiness, confusion, irritability, instability with imbalance; tremor, ataxia are related to cognitive dysfunction, dementia, and PD [132].

Healthy subjective sleep duration is optimal between 6–8 hrs, reported in normal adults [133]. Altered short sleep less than 6hr, prolonged sleep more than 9 hr, and excess daytime sleep with naps are sensitive and early signs of sleep disruption in turn are associated with cognitive decline and dementia [134,135]. The assessment of subjective sleep in AD patients or caregivers reported anxiety, elevated sleep disturbances, and the abnormal circadian rhythm of sleep-wake patterns, high rated daytime short sleep with naps [136]. Wrist actigraphy analysis of objective sleep on human rest and activity cycles confirmed altered sleep disruption with less total sleep time and a high number of awakenings in AD patients [137]. Studies using Polysomnography (PSG) which reported impaired brain waves signified a high rate of sleep awakening and reduction in total sleep time in AD patients [138].

Electroencephalogram (EEG) detects electrical activity of the brain about communication between neurons via electrical impulses that are active all time, even during sleep. EEG studies in AD patients revealed impaired functional connectivity between frontoparietal and frontotemporal regions of the brain with the reduced slow-wave activity associated with REM sleep anomalies correlated with damaged cholinergic circuit and cognitive impairment [139]. Melatonin hormone released by the pineal gland in the brain at night is associated with the biological rhythm of the sleep and wake cycle, the hormonal release is altered in early AD stages. The CSF-melatonin levels were reported to be declined in preclinical stages of AD associated with sleep deprivation, anxiety, neural degeneration, and cognitive dysfunction [140].

Traumatic brain injury (TBI) causes neurovascular injury with cerebrovascular damage of capillaries, arterial stiffness, perivascular accumulation, damage of blood-brain-barrier, endothelial and mitochondrial dysfunction [141]. TBI is associated with AD-like pathologies inducing A β aggregation; hyperphosphorylation of tau with amyloid plaques, NFTs mediated cognitive impairment, and encephalopathy [142,143]. Clinical biomarkers with tau, p-tau, A β were reported in the CSF caused by TBI, indicative of AD. Neuroimaging diagnostic analysis using MRI revealed cerebrovascular damage with micro bleeding, impairment of blood flow in the BBB, and hypo-perfusion produced by TBI. Positron Emission Tomography (PET) scan also exposed accumulation of A β plaques, NFTs in the brain accelerated by TBI signifying AD pathologies with cognitive impairment [144].

3.4. Challenges in the differential diagnosis of dementia and functional connectivity of the brain associated with AD, DLB & FTD

In some cases of Dementia with Lewy Body (DLB), Aβ-plaques and NFTs co-exist along with α -synuclein oligometric and fibrils with more synaptic loss; the challenge is in discriminating Lewy body dementia from AD [145]. Some of the morphological and clinical studies of the brain help in diagnosing patients with DLB and AD, respectively. AD patients exhibited decreased α -synuclein levels in the CSF and elevated Aβ-plaques, NFTs, and increased neural loss [146]. MRI studies of the brain in patients with DLB exhibited small gray matter atrophy; cortical thickness modification in the posterior parietal lobe of the brain and regional thinning was restricted to lateral frontal, the superior temporal occipital region with less severe hippocampal atrophy [147,148]. MRI of patients with AD displayed cortical thinning in the subgenual cingulate region, para-hippocampal, and tempo-parietal cortices of the brain with severe hippocampal atrophy affecting the subiculum region, CA1, entorhinal cortex, or hippocampus [149,150]. Patients with DLB are clinically diagnosed with increased cholinergic dysfunction by gray matter atrophy in the substantia innominate and dorsal mesopontine region of the brain distinguishing from patients with AD [151]. In the early stages of DLB, the damage was diagnosed in the white matter parietal-occipital regions of the brain which is not observed in AD [152]. Studies by functional Magnetic Resonance Imaging (fMRI) in patients with DLB showed enhanced neural connection in the putamen and inferior parietal cortical regions of the brain and declined functionality in the frontal-parietal operculum, the medial prefrontal cortex. Comparatively, the fMRI examination in AD patients exhibited many complex brain network mechanisms with decreased connectivity in the hippocampus, increased prefrontal activity, decreased mesial temporal lobe activation, impairment in memory coding default network between the lateral parietal, temporal, prefrontal, precuneus, posterior cingulate, and medial, and cortical regions of the brain [153,154].

Imaging a Dopamine transporter with 123I-FP-CIT radiotracer-based single-photon emission tomography (FP-CIT-SPECT) is one of the sensitive and specific diagnostic tools in assessing the dopaminergic function [155]. The dopamine transporter uptake declines in the basal ganglia of the brain cause DLB with dopaminergic dysfunction which is differentiable in contrast to AD [156,157]. The radiopharmaceutical 123I-meta-iodobenzylguanidine (123I-MIBG) made from Iobenguane, an aralkyl guanidine analog of adrenergic neurotransmitter nor-epinephrine, acts as an antagonist in blocking adrenergic neurons. 123I-MIBG cardia scintigraphy is one more diagnostic tool used in assessing the cardiac postganglionic sympathetic degeneration in DLB cases, but DLB patients additionally diagnosed with cardiovascular heart diseases and diabetes have unveiled fallacious results [158,159].

Patients with DLB have displayed additional pathologies of amyloid deposits, NFTs, along with α-synuclein protein aggregates. The radioligand biomarker [11C]-Pittsburgh compound B ([11C]PiB) in PET is employed for examining amyloid plaques. Hypo-retention of ([11C]PiB) is observed in patients with DLB in contrast to AD patients, as the amyloid deposition load is excess in AD state [160]. PET imaging with radioligand Fluorine 18-labeled (¹⁸F) AV-1451 biomarker exhibited excessive uptake in assessing tau and NFTs deposits in the precuneus and temporal gyrus regions of the brain in cognitively impaired AD patients. Meanwhile, (¹⁸F) AV-1451 uptake in the temporal lobe of the brain was decreased in DLB patients [161]. Montreal Cognitive Assessment (MoCA) was reported to be one of the efficient diagnostic methods to examine cognitive

impairment. Observation on visual perception, spatial relationships of objects, task orientation, language fluency, long-term semantic memory, and short-term memory showed a substantial decline in frontotemporal dementia (FTD) patient groups compared to AD [162].

4. Biomarkers and therapeutic approaches in AD

Evaluation of A β 42-biomarker in the CSF using ELISA and Mass Spectrometry has been reported to decrease A\beta42 levels due to sequestration of senile plaques in AD patients [163]. In vivo amyloid investigation has also been conducted using PET with diverse amyloid tracers like ¹¹C-PiB, ¹⁸F-AV1451, ¹⁸F-florbetapir (Amyvid), ¹⁸F-flutemetamol (Vizamyl), and ¹⁸F-florbetaben (Neuraceq) [164,165]. Other CSF biomarkers like—Total tau (T-tau), phosphorylated-tau (P-tau), Chitinase-3-like protein-1 (CHI3L1), Visinin-Like Protein (VLP-1), Neuro Filament Light Protein (NFL), Heart Fatty Acid Binding Protein (HFABP), Neuron-Specific Enolase (NSE), and blood biomarker-Plasma T-tau are shown to be elevated in patients with AD [166]. Tau PET, neurofilament light, and neurogranin are used as new biomarkers in clinical trials to study tau pathology in AD patients [167]. To date, there is no cure for AD and only a few medications can control depression and symptoms that may occur as the disease progresses [168]. US-FDA-approved drugs such as donepezil, galantamine, rivastigmine and tacrine act as cholinesterase inhibitors slowing down the metabolic breakdown of acetylcholine by improving communication between the neural cells and reducing the progression of cognitive impairment proved effective for some patients in the early to middle stages [169–171]. Memantine has shown efficacy by acting as a non-competitive NMDA receptor antagonist protecting neurons against excess amounts of glutamate, a messenger chemical released in large amounts to cell-surface NMDA receptors which cause neurodegeneration [172,173]. Antipsychotic drugs are moderately useful in reducing aggression and psychosis with dementia [174]. These drugs are also associated with adverse metabolic side effects such as cerebrovascular events movement difficulties, cognitive decline that does not permit their routine use [175,176]. When used in the long-term, they have been shown to associate with increased mortality [177]. γ -secretase inhibitors such as semagacestat, avagacestat, tarenflurbil showed low brain penetration, exhibited side effects associated with cognitive decline in daily activities, increased rates of infections, and skin cancer in AD patients [178-180].

Verubecestat, the first small molecule BACE1 inhibitor after long-term treatment in animals reported efficient BBB permeability effectively decreased A β 40, A β 42, sAPP β in CSF, and the brain. Preclinical treatment studies of verubecestat in rats and monkeys did not report adverse effects, such as reduced nerve myelination, neurodegeneration, altered glucose homeostasis, or hepatotoxicity, seen in BACE1-null mice [181]. Unexpectedly administration of verubecestat in patients with early-onset AD caused slight shrinkage in hippocampal and total brain volume which resulted in worsening of cognitive symptoms and other side effects of changes in hair color due to inhibition of BACE2 for its control of hair pigmentation, thus verubecestat was discontinued due to lack of efficacy [182]. Upon oral administration of small-molecule BACE1 inhibitors like lanabecestat, atabecestat in AD patients displayed reduced A β , total tau, and phosphorylated tau levels in the CSF, but the usage of these drugs is terminated due to side effects of skin rashes, liver toxicity, and neuropsychiatric symptoms [183,184].

Due to the inefficacy of drugs in crossing the BBB, different nanotechnology-based drug delivery systems with metal-based silver nanoparticles, gold nanoparticles, polymeric nanoparticles

such as poly lactic-co-glycolic acid, polylactic acid, poly butyl cyanoacrylate, polysorbate-80, Cholesterol, DPPC, Methylcellulose, dimethyl-β-CD, sodium taurocholate mediated liposomes are explored in the trial. Using these versatile encapsulated delivery systems; galantamine, curcumin, dexibuprofen, rivastigmine drugs are tested on model systems like Neuro2a cells, HeLa cells, SH-SY5Y cells, GI-1 glioma cells, *in vivo* mice, PC12 cells, bEnd3 cells, glial cells, APPswe/PS1dE Mice, Tg2576 mice, and Balb-C type mice. Conclusively, the advantage of drugs to cross the BBB was easily achieved in vivo with uninterrupted drug delivery to the brain exhibiting inhibition of AchE, cleared amyloid plaques, restricted neurodegeneration, and established neuroprotective effect by improving memory and cognition [185]. A potent pro-apoptotic member; Tumor Necrosis Factor Ligand Superfamily member-10 (TNFSF10) mediates neuroinflammation and Aβ-induced neuronal death. Therapeutics using the anti-TNFSF10 antibody in triple-transgenic-AD (3xTg-AD) mice showed declined neuroinflammation and neurodegeneration in the brain hippocampus [186].

Reports evidenced for higher levels of Angiotensin-Converting Enzyme (ACE) protein expression in the brain with deposition of amyloid-beta and degeneration of hippocampal neurons. Treatment with an ACE inhibitor like captopril has revealed successful regeneration of hippocampal CA1 neurons and reduction of amyloid plaques in the hippocampus of Tg2576 AD mice [187]. Bilateral transplantation of neuronal stem cells in an aged 3xTg-AD restored cognitive and synaptic deficits producing high levels of BDNF without modifying both amyloid plaques and tangle pathologies [188]. Neuronal stem cells that deliver disease-modifying proteins survive for a long period by secreting Aβ-degrading enzyme, neprilysin (NEP) leads to declined Aβ pathology and enhanced the synaptic connectivity in 3xTgAD and Thy1-APP transgenic AD mice models [189]. The Human CNS stem cell line derived from fetal brain tissue has been shown to recover cognitive function by improving the synaptic connectivity in both 3xTg-AD and CaM/Tet-DTA models via growth-associated proteins without affecting A β and tau pathology [190]. The therapeutic approach demonstrated in transgenic AD mice models by neuronal stem cells in enhancing neurotrophic factors for improving synaptic activity and delivery of disease-modifying proteins for reducing $A\beta$ pathology could become one of the promising future aspects to combat AD [191]. Early diagnosis of AD using several CSF biomarkers is useful but the pharmacological approach to treat AD is beneficial only in mild to moderate conditions. FDA-approved drugs do help for reducing the symptoms in moderate to severe conditions up to a certain extent; there is a gap in disease-modifying clinical and therapeutic approaches as the disease remains incurable as it advances [192]. The drugs, suvorexant-first dual orexin blocker to treat insomnia and Lemborexant (in phase-3 trials), almorexant, daridorexant, filorexant are some promising dual antagonists of orexin-OX1 and OX2 receptors associated with relieving neuropsychiatric symptoms of sleep-wake disorders and insomnia in AD [193–196]. In silico evaluation revealed five ligands for site-specific targets on blocking serine protein kinase p-O ester scaffolds of tau with high pharmacokinetics and revealed to be one of the promising therapeutic approaches as p-tau inhibitors for tauopathies in AD [197].

5. Healing impact of Physical exercise along with neuromodulative treatments in AD

5.1. Molecular role of different forms of Physical exercise in AD

Physical exercise (PE) has a beneficial effect in decreasing blood pressure, raised endothelial function, improves memory with cognitive psychological function, decreases anxiety and depression

enhancing sleep quality by expressing neurotrophic factors inducing neurogenesis [198,199]. PE has a positive impact on molecules such as PKC, MAPK, Akt, NF-KB, calmodulin kinase, and calcineurin which are involved in antioxidant defense mechanism and cytoprotection through signaling pathways [200]. Animal studies using intra-hippocampal β-Amyloid infusion in Male Wistar rats subjected to one-time treadmill aerobic running and anaerobic strength exercise session exhibited consolidated object recognition learning when examined at different time interval memory tests [201]. Performing physical activity release excess IGF-1 to the brain cells to prevent neurodegeneration. Treadmill running exercise has reported reduced neurodegeneration with enhanced uptake of circulating IGF-1 in different experiments involving excitotoxin domoic acid injected C57BL/6 male mice which produce partial neural loss in hippocampus, adult male Wistar rats injected with neurotoxin 3-acetylpyridine which damages neurons in the brain stem, and pcd mouse inheriting degeneration in the Purkinje cells of the cerebellum [202]. Active PE reduces AB induced neuroinflammation in the functional tissue of the brain by clearing A β deposits through interstitial fluid drainage by upregulated A β transporters [203]. PE in adults with MCI has reported a reduction in tau levels in the CSF [204]. Physical activity controls oxidative stress-induced by vascular risk factors induce cerebrovascular neuroinflammation and neurodegeneration with improved neuron function [205]. PE helps in anti-inflammatory mechanisms by upregulating the expression of IL-10 anti-inflammatory cytokine and downregulating TNF- α , IL-1 β pro-inflammatory cytokines [206]. Treadmill exercise in Swiss mice, Balb/cJ, Balb/c-IL4^{tm2Nnt}/J knockout mice has revealed reduced microglial activation and an increase in IL-4 cytokines and M2-macrophages which secretes anti-inflammatory cytokines with a decrease in pro-inflammatory cytokines secreting M1-macrophages count reducing the risk of neuroinflammation [207]. The decreasing impact of Aß, tau phosphorylation, tau levels, NFTs, pro-inflammatory cytokines, microglia, astrocytes, and the increasing impact of anti-inflammatory cytokines (IL-1a, IL-4, IL-6), BDNF, and other factors by PE to enhance cognition is explained in Figure 5.

Physical exercise		
Aβ plaques↓	IL-1α, IL-4, IL-6个	
tau phosphorylation $~ igstarrow$	Microglia activation↓	
ptau levels↓	Astrocytes↓	
total tau ↓	BDNF个 in the brain	
tau acetylation $igvee$	anti-inflammatory cytokines个	
NFT formation↓	pro-inflammatory cytokines↓	
Enhanced memory and cognitive function		

Figure 5. Molecular effects of Physical Exercise in AD for improving memory and cognition.

Author (Year)	Model	Test	Molecular alterations by PE
Adlard et al. (2005)	TgCRND8 mice	Morris water maze experiment	$A\beta\downarrow$ in frontal cortex, hippocampus and
[211]			improved spatial learning
Um et al. (2011)	Tg-NSE/PS2m mice	Treadmill exercise	$A\beta\downarrow$ in the brain and improved learning
[212]			
Leem et al. (2009)	Tg-NSE/htau23 mice	Mouse tread mill	ptau levels \downarrow in hippocampus, phospho-PKCa \uparrow
[213]			phospho-AKT \uparrow phospho-PI3K \uparrow phospho-PKA \downarrow
			phospho-ERK↓,GSK3β↑
Belarbi et al. (2011)	THY-Tau22 transgenic	Running wheel test	NFT formation \downarrow in the hippocampus and
[214]	model		enhanced spatial learning
Pajonk et al. (2010)	Human	Aerobic exercise training	MRI revealed increased hippocampal volume
[210]		(cycling)	correlating with neurons and synapses↑,
			improved neural plasticity
Liu et al. (2020)	3xTg-AD mice (B6;	Resistance training, open field	expression of pan tau \downarrow A β deposition \downarrow
[208]	129-sen1tm1 Mpm Tg	test, Novel object recognition	tau hyperphosphorylation \downarrow total tau \downarrow
	(APPSwe, tauP301L)	test, Y-maze test	improved cognition, decreased
	1L fa/MmJax)		neuroinflammation in the frontal cortex and
			hippocampus
Alkadhi et al.	Adult male Wistar rats	Rodent treadmill	Increase in APP, BACE-1 and A β prevented in
(2017) [215]	(infusion of A β 1–42)		the hippocampus, BDNF \uparrow in thebrain
Bobinski et al.	Swiss mice, Balb/cJ,	Treadmill	Microglia activation ↓ anti-inflammatory
(2018) [207]	$Balb/c\text{-}IL4^{tm2Nnt}\!/J$		cytokine IL-4 ↑ anti-inflammatory cytokine
	knockout mice		secreting M2-macrophages ↑pro-inflammatory
			cytokine secreting M1-macrophages \downarrow
Marquez et al.	Human	Cycling	BDNF↑ in the serum
(2015) [209]			
Hashiguchi et	APP/PS1	Resistance exercise	microglia↑ Aβ plaques↓ IL-1α↑ IL-4↑ IL-6↑
al.(2019) [216]	double transgenic mice	OF test	Improved neural function
He et al. (2017)	C57BL/6J mice	Voluntary Wheel Training	Astrocytes Microglia Glymphatic clearance of
[203]	Thy1-GFP transgenic	Morris Water Maze	A β ↑ postsynaptic density protein (PSD95) ↑
	mice		dendrites↑ unaltered BBB permeability
Mankhong et al.	Rats	The Single Pellet Reaching	tau↓ inhibited tau modification and tau
(2020) [217]		(SPR) Test. Rotarod Test	acetylation, phosphorylated glycogen synthase
		The Radial Maze Test	kinase 3-beta Tyr 216 (p-GSK3β Y216)↓
		Treadmill Aerobic Exercise	SIRT1↓ Improved memory and cognitive
		Training	function

Table 1. Impact of Physical Exercise (PE) on modulating Alzheimer' disease pathologies in different model organisms.

Meanwhile, studies using 3xTg-AD mice by resistance training, an examination by novel object recognition, open field-test reported decreased hyperphosphorylation of tau, total tau, and declined A β deposits in the hippocampus and frontal cortex of the brain with improved cognition [208]. Trials using humans conducting cycling aerobic exercise showed increased hippocampal volume determined by MRI studies correlated by an increase in BDNFs [209] and elevated density of neurons, with

ameliorated synaptic plasticity [210]. A detailed effect of physical exercise in different model organisms and the molecular changes with relevance to AD is explained in Table 1.

PE influences the pharmacokinetics of the drugs suitable for absorption, metabolism, and distribution through oral, subcutaneous, intramuscular, and transdermal ways by altering physicochemical characteristics like pH of enzymes in the gastrointestinal tract, pH of the blood and muscle. PE also induces total body clearance of drugs and increases renal and biliary excretion, avoiding drug toxicity [218].

WHO recommends health guidelines on performing the physical activity for aged people above 65 years at least 150 mins-moderate/75 mins-heavy or equivalent physical activity daily with short-10mins intermediate breaks. Considering added health benefits, daily 300 mins-moderate/150 mins-heavy or an equivalent physical activity is recommended. Aged individuals with poor mobility are suggested to do mild balance exercises thrice a week and any kind of muscle-strengthening physical activity, twice a week. The elderly adults with any health complications and incapable of performing the recommended activities are advised at least to remain active physically, depending on the individual capabilities [219]. 2020 WHO physical activity guidelines describe that adult age groups between 18–64 years are recommended with 150–300 mins-moderate/75–150 mins-heavy physical activity and categorizes different forms of physical activities as i) aerobic physical activity including yoga, meditation, walking, running, swimming, and bicycling, ii) balance training exercises with different postures, iii) muscle and bone-strengthening exercises with treadmill running, gymnasium, weight lifting, iv) Household-domain physical activity, v) Leisure-domain physical activity with sports, dancing and gardening, vi) Light-intensity physical activity (LPA) with slow walking, bathing, or other incidental light activities at a normal breathing rate. [220].

Physical activities directly or indirectly have a major impact on relieving obesity, stress, sleep cardiovascular diseases, and improve hippocampal neurogenesis, deprivation, induce synaptogenesis, enhances BDNF with cognition reported both in human and rat models [221-223]. Aerobic exercise promotes beneficiary structural and functional changes in hippocampal size, reduced loss of grey, white matter in the frontal and temporal regions of the brain observed by voxel-based morphometry (VBM), MRI significantly improved neuroplasticity with spatial and learning memory in humans [224,225]. A scheduled 3–12 months physical aerobic exercise training in aged individuals showed enhanced neural connectivity with improved cognition, memory, reduced anxiety, stress, and depression [226,227]. A study using analyzing physical exercise in 198 subjects with MCI and 1,126 with normal cognition reported conservative activity in the mid-age and 39% lower risk of developing MCI by aging [228]. Physical activity with aerobic exercise enhanced cognition, independent functioning, learning, and memory in older adults with MCI and dementia [229]. PE with aerobic multicomponent training in AD patients displayed reduced neuroinflammation, ROS, improved cognition analyzed by neuropsychological battery test [230,231]. Physical activity with walking, stretching balance exercise for 3-sessions a week till 3-months decreased depression, better mood, increased behavioral response, and cognitive function in early to moderate-AD patients [232,233]. Stationary bike aerobic arms cycling for 20 mins thrice a week for 3-months at 70% maximal heart rate showed enhanced attention, response to verbal communication with improved cognitive thinking [234]. Synergistically, multiple antioxidant-rich healthy diets with regular physical activity in aged peoples could decrease the risk for dementia by reducing ROS, mitochondrial dysfunction, and other disease pathologies associated with MCI, AD and improve neurocognitive function [235].

5.2. Combined role of Neuromodulatory treatment-with physical exercise in AD

Repetitive Transcranial Magnetic Stimulation (rTMS) is a neuromodulation instrument treatment technique that involves electro-magnetic impulses delivered through coils applied on the forehead as helmet helps stimulate synaptic transmission in the focal cortical regions of the brain. rTMS values with a repetitive minimum frequency below 1 Hz with the continuous form of theta-burst stimulation (cTBS) indicate debased low synaptic transmission, high-frequency values between 5-50 Hz intermittent forms of theta-burst stimulation (iTBS) correlates with enhanced cortical excitability in synaptic transmission. rTMS is an FDA-approved treatment procedure for major depression, treatment-resistant depression, anxiety, AD, and psychotic disorders [236–238]. rTMS with 20 Hz stimulation on the dorsolateral prefrontal cortex (DFLPC) and precuneus of the brain reported improved episodic memory, language function, identifying objects, comprehensive auditory learning in patients with mild to moderate AD [239,240]. Performing various cognitive tasks in mild AD patients with mild depression during rTMS procedure with 20 Hz stimulation, 40 pulses per burst in 5-second intervals on the right and left DFLPC region of the brain showed improvement in retained cognition for certain weeks post-treatment [241]. Patients with early AD diagnosed by brain CSF protein levels, shown beneficiary results on delayed recall and enhancement in memory after two weeks of rTMS 20 Hz stimulation on the precuneus region of the brain [242].

rTMS with cognitive training in 30 mild-to-moderate AD patients involving grammar task, comprehensive meaning, attentive spatial memory tasks recognition-categorizing objects, shapes, places, colors, naming letters focusing the Broca area, Wernicke area, left to right DLPFCs and PSAC areas of the brain, with the schedule of 1 h daily, weekly 5-days, for 6 weeks reported safe, effective, improved cognition up to a year in 80% of AD patients [243]. Combined application on performing rTMS with PE in stroke patients has shown a positive impact on improving voluntary functioning with reduced long-term depression, modulates neurotransmitters, induces BDNF generation, and improves neural plasticity [244]. rTMS (18 Hz, 2-Sec on, 18-Sec off) combined with stationary aerobic cycling in patients with depression on continuous assessment of alertness, response, cycling capability reported that all patients were comfortable during rTMS-physical activity proved effective, improved mood with reduced depressive symptoms [245].

Transcranial electrical stimulation (tES) is a safer and reliable technique that noninvasively stimulates the brain by passing electrical current via electrodes through the soft tissue and skull into the brain cortex and alters brain function, also includes multiple types of tES-transcranial direct current stimulation (tDCS), transcranial alternating current stimulation (tACS), transcranial random noise stimulation (tRNS), transcranial pulsed current stimulation (tPCS), have proven beneficiary in inducing cortical excitability and neuromodulator effects [246]. Comparative evaluation on the effects tES types- tDCS, tACS, tRNS, and tPCS in a beta-amyloid-induced AD rat model (Sprague-Dawley male rats bilaterally microinjected with A β 25–35 dose of 5 µg/2.5 ml/day into brain hippocampus), physical activity(swimming) training with behavior assessment using Morris water maze task and tES induction (20 min-session, current intensity-200 µA, ramp-10 secs in multiple sessions) proved that both tDCS and tACS was most significant in improving learning and memory behavior induced by $A\beta$ [247]. Aerobic exercise with tDCS synergistically enhances BDNFs and other growth factors; releases neurochemicals in the brain improves synaptic activity and motor function with cognition [248].

6. Conclusions

Aβ plaques and NFTs are the pathologic hallmarks of AD linked with multi-factorial integrated molecular events. Since the disease progression in the early stages is asymptomatic; fluid biomarkers are boon for early diagnosis of AD. Unfortunately, FDA-approved drugs used in AD treatment could only reduce the symptoms but not the disease progression. Successful use of neuronal stem cells on transgenic AD mice models has shown promising results in both reducing AB by secreting A β -degrading enzyme, NEP and enhancing BDNF. Treatment with γ -secretase inhibitors, BACE1 inhibitors can strongly cut down A β 40, A β 42, total tau, and phosphorylated tau pathologies. Experiments on cycling, treadmill PE in human trials, transgenic AD mice, and rat models appear as the best informative sources in being a preventive measure to avoid developing AD and prove efficient in modulating A β , tau, and neuroinflammation in developed AD. The pharmacokinetics in assimilation and absorption of the drugs could be more effective by PE, conjointly the ability of drugs to cross the BBB could be achieved by nanotechnology-based drug delivery systems for degrading amyloid plaques, inhibiting acetylcholine esterase, and preventing neurodegeneration by enhancing neuron function. Conclusively, the joint approaches on using neuronal stem cells with nanotechnology-based multi approached drug delivery systems along with a healthy diet and physically active stress-free lifestyle with neuromodulatory treatments like rTMS, tES may prevent AD pathologies and improve the synaptic transmission of the brain that would become one of the future therapeutic research advancements in tackling AD.

Conflict of Interest

The authors declare no conflict of interest.

References

- 1. Drachman DA (2005) Do we have brain to spare? *Neurology* 64: 2004–2005.
- 2. Beul SF, Hilgetag CC (2019) Neuron density fundamentally relates to architecture and connectivity of the primate cerebral cortex. *Neuroimage* 189: 777–792.
- 3. Cunningham EL, McGuinness B, Herron B, et al. (2015) Dementia. Ulster Med J 84: 79-87.
- 4. Alzheimer's Disease International (2014) Dementia in the Asia Pacific Region. Alzheimer's Disease International Report, London, Available from: https://www.alzint.org/resource/dementia-in-the-asia-pacific-region/.
- 5. Europe A (2019) Dementia in Europe yearbook 2019: Estimating the prevalence of dementia in Europe, Luxembourg: Alzheimer Europe, 1–102.
- 6. Alzheimer's disease facts and figures (2020) Alzheimer's Dement. Alzheimer's Association Report 2020. Available from: https://doi.org/10.1002/alz.12068.
- 7. Kumar CTS, Shaji KS, Varghese M, et al. (2019) Dementia in India 2020. Cochin: Alzheimer's and Related Disorders Society of India (ARDSI), 1–96.
- 8. Barnes DE, Yaffe K (2011) The projected effect of risk factor reduction on Alzheimer's disease prevalence. *Lancet Neurol* 10: 819–828.

- Fleming R, Zeisel J, Bennett K (2020) World Alzheimer Report 2020: Design Dignity Dementia: dementia-related design and the built environment Volume 1: Case Studies. London, England: Alzheimer's Disease International, 1–248.
- 10. Trevisan K, Cristina-Pereira R, Silva-Amaral D, et al. (2019) Theories of Aging and the Prevalence of Alzheimer's Disease. *Biomed Res Int* 2019: 1–9.
- 11. Alzheimer A (1987) About a peculiar disease of the cerebral cortex. By Alois Alzheimer, 1907 (Translated by L Jarvik and H Greenson). *Alzheimer Dis Assoc Disord* 1: 3–8.
- 12. Tiraboschi P, Hansen LA, Thal LJ, et al. (2004) The importance of neuritic plaques and tangles to the development and evolution of AD. *Neurology* 62: 1984–1989.
- 13. Carter J, Lippa CF (2001) Beta-Amyloid, Neuronal Death and Alzheimers Disease. *Curr Mol Med* 1: 733–737.
- 14. Medina M, Hernández F, Avila J (2016) New features about tau function and dysfunction. *Biomolecules* 6: 21.
- 15. Castellani RJ, Perry G (2019) Tau biology, tauopathy, traumatic brain injury, and diagnostic challenges. *J Alzheimers Dis* 67: 447–467.
- 16. Zvěřová M (2019) Clinical aspects of Alzheimer's disease. Clin Biochem 72: 3-6.
- 17. Yang Y, Kandiah N, Kim S, et al. (2020) Clinical aspects of neurobehavioral symptoms of dementia. *Dement Neurocogn Disord* 19: 54–64.
- Raz L, Knoefel J, Bhaskar K (2016) The neuropathology and cerebrovascular mechanisms of dementia. J Cereb Blood Flow Metab 36: 172–186.
- 19. Livingston G, Huntley J, Sommerlad A, et al. (2020) Dementia prevention, intervention, and care: report of the Lancet Commission. *Lancet* 396: 413–446.
- 20. Kelly AM (2018) Exercise-induced modulation of neuroinflammation in models of Alzheimer's disease. *Brain Plast* 4: 81–94.
- 21. Saha P, Sen N (2019) Tauopathy: A common mechanism for neurodegeneration and brain aging. *Mech Ageing Dev* 178: 72–79.
- 22. Hernandez F, Avila J (2007) Tauopathies. Cell Mol Life Sci 64: 2219-2233.
- 23. Šimić G, Babić Leko M, Wray S, et al. (2016) Tau protein hyperphosphorylation and aggregation in Alzheimer's disease and other tauopathies, and possible neuroprotective strategies. *Biomolecules* 6: 6.
- 24. Vickers JC, Dickson TC, Adlard PA, et al. (2000) The cause of neuronal degeneration in Alzheimer's disease. *Prog Neurobiol* 60: 139–165.
- 25. Serrano-Pozo A, Frosch MP, Masliah E, et al. (2011) Neuropathological alterations in Alzheimer disease. *Cold Spring Harb Perspect Med* 1: a006189.
- 26. Wenk GL (2003) Neuropathologic Changes in Alzheimer's Disease. J Clin Psychiatry 64: 7–10.
- 27. Priller C, Bauer T, Mitteregger G, et al. (2006) Synapse formation and function is modulated by the amyloid precursor protein. *J Neurosci* 26: 7212–7221.
- 28. Turner PR, O'connor K, Tate WP, et al. (2003) Roles of amyloid precursor protein and its fragments in regulating neural activity, plasticity and memory. *Prog Neurobiol* 70: 1–32.
- 29. Sastre M, Steiner H, Fuchs K, et al. (2001) Presentilin-dependent γ -secretase processing of β -amyloid precursor protein at a site corresponding to the S3 cleavage of Notch. *EMBO Rep* 2: 835–841.
- 30. Selkoe DJ (2002) Deciphering the genesis and fate of amyloid beta-protein yields novel therapies for Alzheimer disease. *J Clin Invest* 110: 1375–1381.

- 31. Dingledine R, Borges K, Bowie D, et al. (1999) The glutamate receptor ion channels. *Pharmacol Rev* 51: 7–62.
- 32. Hu M, Liu QS, Chang KT, et al. (2002) Nicotinic regulation of CREB activation in hippocampal neurons by glutamatergic and nonglutamatergic pathways. *Mol Cell Neurosci* 21: 616–625.
- 33. Yakel JL (2013) Cholinergic receptors: functional role of nicotinic ACh receptors in brain circuits and disease. *Pflügers Archi* 465: 441–450.
- 34. Hussain I, Powell D, Howlett DR, et al. (1999) Identification of a novel aspartic protease (Asp 2) as β-secretase. *Mol Cell Neurosci* 14: 419–427.
- 35. Seubert P, Oltersdorf T, Lee MG, et al. (1993) Secretion of β -amyloid precursor protein cleaved at the amino terminus of the β -amyloid peptide. *Nature* 361: 260–263.
- Hashimoto M, Rockenstein E, Crews L, et al. (2003) Role of protein aggregation in mitochondrial dysfunction and neurodegeneration in Alzheimer's and Parkinson's diseases. *Neuromolecular Med* 4: 21–36.
- 37. Peng C, Trojanowski JQ, Lee VM (2020) Protein transmission in neurodegenerative disease. *Nat Rev Neurol* 16: 199–212.
- Mattson MP, Guo ZH, Geiger JD (1999) Secreted form of amyloid precursor protein enhances basal glucose and glutamate transport and protects against oxidative impairment of glucose and glutamate transport in synaptosomes by a cyclic GMP-mediated mechanism. *J Neurochem* 73: 532–537.
- 39. Snyder EM, Nong Y, Almeida CG, et al. (2005) Regulation of NMDA receptor trafficking by amyloid-β. *Nat Neurosci* 8: 1051–1058.
- Gu Z, Liu W, Yan Z (2009) β-Amyloid impairs AMPA receptor trafficking and function by reducing Ca²⁺/calmodulin-dependent protein kinase II synaptic distribution. *J Biol Chem* 284: 10639–10649.
- 41. Tozaki H, Matsumoto A, Kanno T, et al. (2002) The inhibitory and facilitatory actions of amyloid-β peptides on nicotinic ACh receptors and AMPA receptors. *Biochem Biophys Rese Communn* 294: 42–45.
- 42. Liskowsky W, Schliebs R (2006) Muscarinic acetylcholine receptor inhibition in transgenic Alzheimer-like Tg2576 mice by scopolamine favours the amyloidogenic route of processing of amyloid precursor protein. *Int J Dev Neurosci* 24: 149–156.
- 43. Bojarski L, Herms J, Kuznicki J (2008) Calcium dysregulation in Alzheimer's disease. *Neurochem Int* 52: 621–633.
- 44. Van der Kant R, Goldstein LS, Ossenkoppele R (2020) Amyloid-β-independent regulators of tau pathology in Alzheimer disease. *Nat Rev Neurosci* 21: 21–35.
- 45. Ramesh G, MacLean AG, Philipp MT (2013) Cytokines and chemokines at the crossroads of neuroinflammation, neurodegeneration, and neuropathic pain. *Mediators Inflamm* 2013: 480739.
- 46. Alberghina L, Colangelo AM (2006) The modular systems biology approach to investigate the control of apoptosis in Alzheimer's disease neurodegeneration. *BMC Neurosci* 7: S2.
- 47. Gunn AP, Wong BX, Johanssen T, et al. (2016) Amyloid-β peptide Aβ3pE-42 induces lipid peroxidation, membrane permeabilization, and calcium influx in neurons. *J Biol Chem* 291: 6134–6145.
- 48. Gan X, Huang S, Wu L, et al. (2014) Inhibition of ERK-DLP1 signaling and mitochondrial division alleviates mitochondrial dysfunction in Alzheimer's disease cybrid cell. *Biochim Biophys Acta* 1842: 220–231.

- 49. Hooper C, Killick R, Lovestone S (2008) The GSK3 hypothesis of Alzheimer's disease. J Neurochem 104: 1433–1439.
- 50. Town T, Zolton J, Shaffner R, et al. (2002) p35/Cdk5 pathway mediates soluble amyloid-β peptide-induced tau phosphorylation in vitro. *J Neurosci Res* 69: 362–372.
- 51. Das A, Dikshit M, Nath C (2005) Role of molecular isoforms of acetylcholinesterase in learning and memory functions. *Pharmacol Biochem Behav* 81: 89–99.
- 52. Tiwari P, Dwivedi S, Singh MP, et al. (2013) Basic and modern concepts on cholinergic receptor: A review. *Asian Pac J Trop Dis* 3: 413–420.
- 53. Leonard BE (2004) Pharmacotherapy in the treatment of Alzheimer's disease: an update. *World Psychiatry* 3: 84–88.
- 54. Gutteridge JM, Halliwell B (2000) Free radicals and antioxidants in the year 2000: a historical look to the future. *Ann N Y Acad Sci* 899: 136–147.
- 55. Uttara B, Singh AV, Zamboni P, et al. (2009) Oxidative stress and neurodegenerative diseases: a review of upstream and downstream antioxidant therapeutic options. *Curr Neuropharmacol* 7: 65–74.
- 56. Mancuso C, Scapagini G, Curro D, et al. (2007) Mitochondrial dysfunction, free radical generation and cellular stress response in neurodegenerative disorders. *Front Biosci* 12: 1107–1123.
- 57. Beal MF (1998) Mitochondrial dysfunction in neurodegenerative diseases. *Biochim Biophys Acta* 1366: 211–223.
- 58. Fernie AR, Carrari F, Sweetlove LJ (2004) Respiratory metabolism: glycolysis, the TCA cycle and mitochondrial electron transport. *Curr Opin Plant Biol* 7: 254–261.
- 59. Turrens JF (2003) Mitochondrial formation of reactive oxygen species. J Physiol 552: 335-344.
- 60. Halliwell B (1992) Reactive oxygen species and the central nervous system. *J Neurochem* 59: 1609–1623.
- 61. Reddy PH, Beal MF (2008) Amyloid beta, mitochondrial dysfunction and synaptic damage: implications for cognitive decline in aging and Alzheimer's disease. *Trends Mol Med* 14: 45–53.
- 62. Reddy PH, Tripathi R, Troung Q, et al. (2012) Abnormal mitochondrial dynamics and synaptic degeneration as early events in Alzheimer's disease: implications to mitochondria-targeted antioxidant therapeutics. *Biochim Biophys Acta* 1822: 639–649.
- 63. Reddy PH, McWeeney S, Park BS, et al. (2004) Gene expression profiles of transcripts in amyloid precursor protein transgenic mice: up-regulation of mitochondrial metabolism and apoptotic genes is an early cellular change in Alzheimer's disease. *Hum Mol Genet* 13: 1225–1240.
- 64. Tran M, Reddy PH (2021) Defective Autophagy and Mitophagy in Aging and Alzheimer's Disease. *Front Neurosci* 14: 612757.
- 65. Pradeepkiran JA, Reddy PH (2020) Defective mitophagy in Alzheimer's disease. *Ageing Res Rev* 64: 101191.
- 66. Reddy PH (2011) Abnormal tau, mitochondrial dysfunction, impaired axonal transport of mitochondria, and synaptic deprivation in Alzheimer's disease. *Brain Res* 1415: 136–148.
- 67. Garbarino VR, Orr ME, Rodriguez KA, et al. (2015) Mechanisms of oxidative stress resistance in the brain: lessons learned from hypoxia tolerant extremophilic vertebrates. *Arch Biochem Biophys* 576: 8–16.
- 68. Gerlach M, Ben-Shachar D, Riederer P, et al. (1994) Altered brain metabolism of iron as a cause of neurodegenerative diseases? *J Neurochem* 63: 793–807.
- 69. Chance B, Sies H, Boveris A (1979) Hydroperoxide metabolism in mammalian organs. *Physiol Rev* 59: 527–605.

- 70. Hirai K, Aliev G, Nunomura A, et al. (2001) Mitochondrial abnormalities in Alzheimer's disease. *J Neurosci* 21: 3017–3023.
- 71. Tuppo EE, Arias HR (2005) The role of inflammation in Alzheimer's disease. *Int J Biochem Cell Biol* 37: 289–305.
- 72. Chao CC, Hu S, Sheng WS, et al. (1996) Cytokine-stimulated astrocytes damage human neurons via a nitric oxide mechanism. *Glia* 16: 276–284.
- 73. Biber K, Vinet J, Boddeke HW (2008) Neuron-microglia signaling: chemokines as versatile messengers. *J Neuroimmunol* 198: 69–74.
- 74. Perkins ND (2007) Integrating cell-signalling pathways with NF-κB and IKK function. *Nat Rev Mol Cell Biol* 8: 49–62.
- 75. O'Neill LA, Kaltschmidt C (1997) NF-kB: a crucial transcription factor for glial and neuronal cell function. *Trends Neurosci* 20: 252–258.
- 76. Friedlander RM (2003) Apoptosis and caspases in neurodegenerative diseases. *N Engl J Med* 348: 1365–1375.
- 77. Katsimpardi L, Lledo PM (2018) Regulation of neurogenesis in the adult and aging brain. *Curr Opin Neurobiol* 53: 131–138.
- 78. Anjum I, Fayyaz M, Wajid A, et al. (2018) Does obesity increase the risk of dementia: a literature review. *Cureus* 10: e2660.
- 79. Verdile G, Keane KN, Cruzat VF, et al. (2015) Inflammation and oxidative stress: the molecular connectivity between insulin resistance, obesity, and Alzheimer's disease. *Mediators Inflamm* 2015: 105828.
- 80. Nameni G, Farhangi MA, Hajiluian G, et al. (2017) Insulin deficiency: a possible link between obesity and cognitive function. *Int J Dev Neurosci* 59: 15–20.
- 81. Steen E, Terry BM, Rivera EJ, et al. (2005) Impaired insulin and insulin-like growth factor expression and signaling mechanisms in Alzheimer's disease—is this type 3 diabetes? J Alzheimers Dis 7: 63–80.
- 82. Craft S (2005) Insulin resistance syndrome and Alzheimer's disease: age-and obesity-related effects on memory, amyloid, and inflammation. *Neurobiol Aging* 26: 65–69.
- 83. Dye L, Boyle NB, Champ C, et al. (2017) The relationship between obesity and cognitive health and decline. *Proc Nutr Soc* 76: 443–454.
- 84. Skoog I, Gustafson D (2003) Hypertension, hypertension-clustering factors and Alzheimer's disease. *Neurol Res* 25: 675–680.
- 85. Luchsinger JA, Reitz C, Patel B, et al. (2007) Relation of diabetes to mild cognitive impairment. *Arch Neurol* 64:570–575.
- 86. Luchsinger JA, Tang MX, Shea S, et al. (2004) Hyperinsulinemia and risk of Alzheimer disease. *Neurology* 63: 1187–1192.
- 87. Tiehuis AM, Van der Graaf Y, Visseren FL, et al. (2008) Diabetes increases atrophy and vascular lesions on brain MRI in patients with symptomatic arterial disease. *Stroke* 39: 1600–1603.
- 88. Li ZG, Zhang W, Sima AA (2007) Alzheimer-like changes in rat models of spontaneous diabetes. *Diabetes* 56: 1817–1824.
- 89. Ho L, Qin W, Pompl PN, et al. (2004) Diet-induced insulin resistance promotes amyloidosis in a transgenic mouse model of Alzheimer's disease. *FASEB J* 18: 902–904.

- 90. Bennett RG, Hamel FG, Duckworth WC (2003) An insulin-degrading enzyme inhibitor decreases amylin degradation, increases amylin-induced cytotoxicity, and increases amyloid formation in insulinoma cell cultures. *Diabetes* 52: 2315–2320.
- 91. Shiiki T, Ohtsuki S, Kurihara A, et al. (2004) Brain insulin impairs amyloid-β (1–40) clearance from the brain. *J Neurosci* 24: 9632–9637.
- 92. Lee HJ, Seo HI, Cha HY, et al. (2018) Diabetes and Alzheimer's disease: mechanisms and nutritional aspects. *Clin Nutr Res* 7: 229–240.
- 93. Xi YD, Li XY, Ding J, et al. (2013) Soy isoflavone alleviates Aβ1-42-induced impairment of learning and memory ability through the regulation of RAGE/LRP-1 in neuronal and vascular tissue. *Curr Neurovasc Res* 10: 144–156.
- 94. Deane R, Wu Z, Sagare A, et al. (2004) LRP/amyloid β-peptide interaction mediates differential brain efflux of Aβ isoforms. *Neuron* 43: 333–344.
- 95. Lloret A, Monllor P, Esteve D, et al. (2019) Obesity as a risk factor for Alzheimer's disease: implication of leptin and glutamate. *Front Neurosci* 13: 508.
- 96. Bird TD (2008) Genetic aspects of Alzheimer disease. Genet Med 10: 231-239.
- 97. Kang J, Lemaire HG, Unterbeck A, et al. (1987) The precursor of Alzheimer's disease amyloid A4 protein resembles a cell-surface receptor. *Nature* 325: 733–736.
- 98. Weidemann A, König G, Bunke D, et al. (1989) Identification, biogenesis, and localization of precursors of Alzheimer's disease A4 amyloid protein. *Cell* 57: 115–126.
- 99. Scheuner D, Eckman C, Jensen M, et al. (1996) Secreted amyloid beta-protein similar to that in the senile plaques of Alzheimer's disease is increased in vivo by the presenilin 1 and 2 and APP mutations linked to familial Alzheimer's disease. *Nat Med* 2: 864–870.
- 100. Sherrington R, Rogaev EI, Liang Y, et al. (1995) Cloning of a gene bearing missense mutations in early-onset familial Alzheimer's disease. *Nature* 375: 754–760.
- 101. Theuns J, Del-Favero J, Dermaut B, et al. (2000) Genetic variability in the regulatory region of presenilin 1 associated with risk for Alzheimer's disease and variable expression. *Hum Mol Genet* 9: 325–331.
- 102. Citron M, Westaway D, Xia W, et al. (1997) Mutant presenilins of Alzheimer's disease increase production of 42-residue amyloid beta-protein in both transfected cells and transgenic mice. *Nat Med* 3: 67–72.
- 103. Moehlmann T, Winkler E, Xia X, et al. (2002) Presenilin-1 mutations of leucine 166 equally affect the generation of the Notch and APP intracellular domains independent of their effect on Abeta 42 production. *Proc Natl Acad Sci USA* 99: 8025–8030.
- 104. Levy-Lahad E, Wasco W, Poorkaj P, et al. (1995) Candidate gene for the chromosome 1 familial Alzheimer's disease locus. *Science* 269: 973–977.
- 105. Sherrington R, Froelich S, Sorbi S, et al. (1996) Alzheimer's disease associated with mutations in presenilin 2 is rare and variably penetrant. *Hum Mol Genet* 5: 985–988.
- 106. Kovacs DM, Fausett HJ, Page KJ, et al. (1996) Alzheimer-associated presenilins 1 and 2: neuronal expression in brain and localization to intracellular membranes in mammalian cells. *Nat Med* 2: 224–229.
- 107. Corder EH, Saunders AM, Strittmatter WJ, et al. (1993) Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families. *Science* 261: 921–923.
- 108. Nicoll JA, Roberts GW, Graham DI (1995) Apolipoprotein E epsilon 4 allele is associated with deposition of amyloid beta-protein following head injury. *Nat Med* 1: 135–137.

- 109. Liu Y, Laakso MP, Karonen JO, et al. (2002) Apolipoprotein E polymorphism and acute ischemic stroke: a diffusion- and perfusion-weighted magnetic resonance imaging study. *J Cereb Blood Flow Metab* 22: 1336–1342.
- 110. Horsburgh K, McCulloch J, Nilsen M, et al. (2000) Increased neuronal damage and apoE immunoreactivity in human apolipoprotein E, E4 isoform-specific, transgenic mice after global cerebral ischaemia. *Eur J Neurosci* 12: 4309–4317.
- 111. Huang Y (2006) Molecular and cellular mechanisms of apolipoprotein E4 neurotoxicity and potential therapeutic strategies. *Curr Opin Drug Discov Devel* 9: 627–641.
- 112. Mahley RW, Weisgraber KH, Huang Y (2006) Apolipoprotein E4: a causative factor and therapeutic target in neuropathology, including Alzheimer's disease. *Proc Natl Acad Sci USA* 103: 5644–5651.
- 113. Nagy Z, Esiri MM, Jobst KA, et al. (1995) Influence of the apolipoprotein E genotype on amyloid deposition and neurofibrillary tangle formation in Alzheimer's disease. *Neurosci* 69: 757–761.
- 114. Gibson GE, Haroutunian V, Zhang H, et al. (2000) Mitochondrial damage in Alzheimer's disease varies with apolipoprotein E genotype. *Ann Neurol* 48: 297–303.
- 115. Yamanaka A, Sakurai T, Katsumoto T, et al. (1999) Chronic intracerebroventricular administration of orexin—A to rats increases food intake in daytime, but has no effect on body weight. *Brain Res* 849: 248–252.
- 116. Chow M, Cao M (2016) The hypocretin/orexin system in sleep disorders: preclinical insights and clinical progress. *Nat Sci Sleep* 8: 81–86.
- 117. Tsuneki H, Wada T, Sasaoka T (2012) Role of orexin in the central regulation of glucose and energy homeostasis. *Endocr J* 59: 365–374.
- 118. Ben J, Soares FM, Cechetti F, et al. (2009) Exercise effects on activities of Na⁺, K⁺-ATPase, acetylcholinesterase and adenine nucleotides hydrolysis in ovariectomized rats. *Brain Res* 1302: 248–255.
- 119. Feng P, Vurbic D, Wu Z, et al. (2008) Changes in brain orexin levels in a rat model of depression induced by neonatal administration of clomipramine. *J Psychopharmacol* 22: 784–791.
- 120. Liguori C, Romigi A, Nuccetelli M, et al. (2014) Orexinergic system dysregulation, sleep impairment, and cognitive decline in Alzheimer disease. *JAMA Neurol* 71: 1498–1505.
- 121. Osorio RS, Ducca EL, Wohlleber ME, et al. (2016) Orexin—A is associated with increases in cerebrospinal fluid phosphorylated-tau in cognitively normal elderly subjects. *Sleep* 39: 1253–1260.
- 122. Slats D, AHR Claassen J, Jan Lammers G, et al. (2012) Association between hypocretin-1 and amyloid-β42 cerebrospinal fluid levels in Alzheimer's disease and healthy controls. *Curr Alzheimer Res* 9: 1119–1125.
- 123. Davies J, Chen J, Pink R, et al. (2015) Orexin receptors exert a neuroprotective effect in Alzheimer's disease (AD) via heterodimerization with GPR103. *Sci Rep* 5: 1–2.
- 124. Jaussent I, Bouyer J, Ancelin ML, et al. (2012) Excessive sleepiness is predictive of cognitive decline in the elderly. *Sleep* 35: 1201–1207.
- 125. Suh SW, Han JW, Lee JR, et al. (2018) Sleep and cognitive decline: a prospective nondemented elderly cohort study. *Ann Neurol* 83: 472–482.
- 126. American Academy of Sleep Medicine (2014) International Classification of Sleep Disorders, third Eds., Illinois: Darien, 1–383.

- 127. Sadeghmousavi S, Eskian M, Rahmani F, et al. (2020) The effect of insomnia on development of Alzheimer's disease. *J Neuroinflammation* 17: 289.
- 128. Gehrman PR, Martin JL, Shochat T, et al. (2003) Sleep-disordered breathing and agitation in institutionalized adults with Alzheimer disease. *Am J Geriatr Psychiatry* 11: 426–433.
- 129. Andrade AG, Bubu OM, Varga AW, et al. (2018) The Relationship between Obstructive Sleep Apnea and Alzheimer's Disease. *J Alzheimers Dis* 64: S255–S270.
- 130. Pan W, Kastin AJ (2014) Can sleep apnea cause Alzheimer's disease? *Neurosci Biobehav Rev* 47: 656–669.
- 131. Weldemichael DA, Grossberg GT (2010) Circadian rhythm disturbances in patients with Alzheimer's disease: a review. *Int J Alzheimers Dis* 2010: 716453.
- 132. Bhidayasiri R, Sringean J, Rattanachaisit W, et al. (2017) The sleeping brain in Parkinson's disease: A focus on REM sleep behaviour disorder and related parasomnias for practicing neurologists. *J Neurol Sci* 374: 32–37.
- 133. Panel CC, Watson NF, Badr MS, Belenky G, et al. (2015) Joint consensus statement of the American Academy of Sleep Medicine and Sleep Research Society on the recommended amount of sleep for a healthy adult: methodology and discussion. *Sleep* 38: 1161–1183.
- 134. Matricciani L, Bin YS, Lallukka T, et al. (2017) Past, present, and future: trends in sleep duration and implications for public health. *Sleep Health* 3: 317–323.
- 135. Lo JC, Groeger JA, Cheng GH, et al. (2016) Self-reported sleep duration and cognitive performance in older adults: a systematic review and meta-analysis. *Sleep Med* 17: 87–98.
- 136. Peter-Derex L, Yammine P, Bastuji H, et al. (2015) Sleep and Alzheimer's disease. *Sleep Med Rev* 19: 29–38.
- 137. Camargos EF, Louzada FM, Nóbrega OT (2013) Wrist actigraphy for measuring sleep in intervention studies with Alzheimer's disease patients: application, usefulness, and challenges. *Sleep Med Rev* 17: 475–488.
- 138. Brzecka A, Leszek J, Ashraf GM, et al. (2018) Sleep disorders associated with Alzheimer's disease: a perspective. *Front Neurosci* 12: 330.
- 139. Tsolaki A, Kazis D, Kompatsiaris I, et al. (2014) Electroencephalogram and Alzheimer's disease: clinical and research approaches. *Int J Alzheimers Dis* 2014: 349249.
- 140. Wu YH, Feenstra MG, Zhou JN, et al. (2003) Molecular changes underlying reduced pineal melatonin levels in Alzheimer disease: alterations in preclinical and clinical stages. *J Clin Endocrinol Metab* 88: 5898–5906.
- 141. Fossati S, Cam J, Meyerson J, et al. (2010) Differential activation of mitochondrial apoptotic pathways by vasculotropic amyloid-β variants in cells composing the cerebral vessel walls. *FASEB J* 24: 229–241.
- 142. Washington PM, Morffy N, Parsadanian M, et al. (2014) Experimental traumatic brain injury induces rapid aggregation and oligomerization of amyloid-beta in an Alzheimer's disease mouse model. *J Neurotrauma* 31: 125–134.
- 143. Washington PM, Villapol S, Burns MP (2016) Polypathology and dementia after brain trauma: does brain injury trigger distinct neurodegenerative diseases, or should they be classified together as traumatic encephalopathy? *Exp Neurol* 275: 381–388.
- 144. Ramos-Cejudo J, Wisniewski T, Marmar C, et al. (2018) Traumatic brain injury and Alzheimer's disease: the cerebrovascular link. *EBioMedicine* 28: 21–30.

- 145. Elahi FM, Miller BL (2017) A clinicopathological approach to the diagnosis of dementia. *Nat Rev Neurol* 13: 457–476.
- 146. Lim X, Yeo JM, Green A, et al. (2013) The diagnostic utility of cerebrospinal fluid alpha-synuclein analysis in dementia with Lewy bodies: a systematic review and meta-analysis. *Parkinsonism Relat d* 19: 851–858.
- 147. Watson R, Colloby SJ, Blamire AM, et al. (2015) Assessment of regional gray matter loss in dementia with Lewy bodies: a surface-based MRI analysis. *Am J Geriatr Psychiatry* 23: 38–46.
- 148. Chow N, Aarsland D, Honarpisheh H, et al. (2012) Comparing hippocampal atrophy in Alzheimer's dementia and dementia with Lewy bodies. *Dement Geriatr Cogn Disord* 34: 44–50.
- 149. Lebedev AV, Westman E, Beyer MK, et al. (2013) Multivariate classification of patients with Alzheimer's and dementia with Lewy bodies using high-dimensional cortical thickness measurements: an MRI surface-based morphometric study. *J Neurol* 260: 1104–1115.
- 150. Watson R, O'Brien JT, Barber R, et al. (2012) Patterns of gray matter atrophy in dementia with Lewy bodies: a voxel-based morphometry study. *Int Psychogeriatr* 24: 532–540.
- 151. Whitwell JL, Weigand SD, Shiung MM, et al. (2007) Focal atrophy in dementia with Lewy bodies on MRI: a distinct pattern from Alzheimer's disease. *Brain* 130: 708–719.
- 152. Watson R, Blamire AM, Colloby SJ, et al. (2012) Characterizing dementia with Lewy bodies by means of diffusion tensor imaging. *Neurology* 79: 906–914.
- 153. Galvin JE, Price JL, Yan Z, et al. (2011) Resting bold fMRI differentiates dementia with Lewy bodies vs Alzheimer disease. *Neurology* 76: 1797–1803.
- 154. Johnson KA, Fox NC, Sperling RA, et al. (2012) Brain imaging in Alzheimer disease. *Cold Spring Harb Perspect Med* 2: a006213.
- 155. McKeith I, O'Brien J, Walker Z, et al. (2007) Sensitivity and specificity of dopamine transporter imaging with 123I-FP-CIT SPECT in dementia with Lewy bodies: a phase III, multicentre study. *Lancet Neurol* 6: 305–313.
- 156. Brigo F, Turri G, Tinazzi M (2015) 123I-FP-CIT SPECT in the differential diagnosis between dementia with Lewy bodies and other dementias. *J Neurol Sci* 359: 161–171.
- 157. Shimizu S, Hanyu H, Kanetaka H, et al. (2005) Differentiation of dementia with Lewy bodies from Alzheimer's disease using brain SPECT. *Dement Geriatr Cogn Disord* 20: 25–30.
- 158. Hanyu H, Shimizu S, Hirao K, et al. (2006) Comparative value of brain perfusion SPECT and [(123) I] MIBG myocardial scintigraphy in distinguishing between dementia with Lewy bodies and Alzheimer's disease. *Eur J Nucl Med Mol Imaging* 33: 248–253.
- 159. Yoshita M, Arai H, Arai H, et al. (2015) Diagnostic accuracy of 123 I-meta-iodobenzylguanidine myocardial scintigraphy in dementia with Lewy bodies: a multicenter study. *PloS One* 10: e0120540.
- 160. Edison P, Rowe CC, Rinne JO, et al. (2008) Amyloid load in Parkinson's disease dementia and Lewy body dementia measured with [11C] PIB positron emission tomography. J Neurol Neurosurg Psychiatry 79: 1331–1338.
- 161. Kantarci K, Lowe VJ, Boeve BF, et al. (2017) AV-1451 tau and β-amyloid positron emission tomography imaging in dementia with Lewy bodies. *Ann Neurol* 81: 58–67.
- 162. Coleman KK, Coleman BL, MacKinley JD, et al. (2016) Detection and differentiation of frontotemporal dementia and related disorders from alzheimer disease using the Montreal Cognitive Assessment. *Alzheimer Dis Assoc Disord* 30: 258–263.

- 163. Kuhlmann J, Andreasson U, Pannee J, et al. (2017) CSF Aβ1–42—an excellent but complicated Alzheimer's biomarker—a route to standardisation. *Clinica Chimica Acta* 467: 27–33.
- 164. Lashley T, Schott JM, Weston P, et al. (2018) Molecular biomarkers of Alzheimer's disease: progress and prospects. *Dis Models Mech* 11: dmm031781.
- 165. Morbelli S, Bauckneht M (2018) Amyloid PET imaging: standardization and integration with other Alzheimer's disease biomarkers. *In Biomarkers for Alzheimer's Disease Drug Development*. Humana Press, New York, NY, 203–212.
- 166. Olsson B, Lautner R, Andreasson U, et al. (2016) CSF and blood biomarkers for the diagnosis of Alzheimer's disease: a systematic review and meta-analysis. *Lancet Neurol* 15: 673–684.
- 167. Guzman-Martinez L, Maccioni RB, Farías GA, et al. (2019) Biomarkers for Alzheimer's Disease. *Curr Alzheimer Res* 16: 518–528.
- 168. Gill SS, Rochon PA, Herrmann N, et al. (2005) Atypical antipsychotic drugs and risk of ischaemic stroke: population based retrospective cohort study. *BMJ* 330: 445.
- 169. Giacobini E, Pepeu G (2006) Cholinesterases in human brain: the effect of cholinesterase inhibitors on Alzheimer's disease and related disorders. *Brain Cholinergic Syst Health Dis* 235–264.
- 170. Li WM, Pi RB, Chan HH, et al. (2005) Novel dimeric acetylcholinesterase inhibitor bis7-tacrine, but not donepezil, prevents glutamate-induced neuronal apoptosis by blocking N-methyl-D-aspartate receptors. *J Biol Chem* 280: 18179–18188.
- 171. Hyde C, Peters J, Bond M, et al. (2013) Evolution of the evidence on the effectiveness and cost-effectiveness of acetylcholinesterase inhibitors and memantine for Alzheimer's disease: systematic review and economic model. *Age Ageing* 42: 14–20.
- 172. Lipton SA (2005) The molecular basis of memantine action in Alzheimer's disease and other neurologic disorders: low-affinity, uncompetitive antagonism. *Curr Alzheimer Res* 2: 155–165.
- 173. Hynd MR, Scott HL, Dodd PR (2004) Glutamate-mediated excitotoxicity and neurodegeneration in Alzheimer's disease. *Neurochem Int* 45: 583–595.
- 174. Liperoti R, Pedone C, Corsonello A (2008) Antipsychotics for the treatment of behavioral and psychological symptoms of dementia (BPSD). *Curr Neuropharmacol* 6: 117–124.
- 175. Reynolds GP, Kirk SL (2010) Metabolic side effects of antipsychotic drug treatment—pharmacological mechanisms. *Pharmacol Ther* 125: 169–179.
- 176. Vigen CL, Mack WJ, Keefe RS, et al. (2011) Cognitive effects of atypical antipsychotic medications in patients with Alzheimer's disease: outcomes from CATIE-AD. *Am J Psychiatry* 168: 831–839.
- 177. Ralph SJ, Espinet AJ (2018) Increased all-cause mortality by antipsychotic drugs: updated review and meta-analysis in dementia and general mental health care. *J Alzheimers Dis Rep* 2: 1–26.
- 178. Doody RS, Raman R, Farlow M, et al. (2013) A phase 3 trial of semagacestat for treatment of Alzheimer's disease. *N Engl J Med* 369: 341–350.
- 179. Coric V, Salloway S, van Dyck CH, et al. (2015) Targeting prodromal Alzheimer disease with avagacestat: a randomized clinical trial. *JAMA neurology* 72: 1324–1333.
- 180. Muntimadugu E, Dhommati R, Jain A, et al. (2016) Intranasal delivery of nanoparticle encapsulated tarenflurbil: A potential brain targeting strategy for Alzheimer's disease. *Eur J Pharm Sci* 92: 224–234.
- 181. Das B, Yan R (2019) A Close Look at BACE1 Inhibitors for Alzheimer's Disease Treatment. *CNS Drugs* 33: 251–263.

- 182. Egan MF, Kost J, Voss T, et al. (2019) Randomized trial of verubecestat for prodromal Alzheimer's disease. *N Engl J Med* 380: 1408–1420.
- 183. Burki T (2018) Alzheimer's disease research: the future of BACE inhibitors. Lancet 391: 2486.
- 184. Henley D, Raghavan N, Sperling R, et al. (2019) Preliminary results of a trial of atabecestat in preclinical Alzheimer's disease. *N Engl J Med* 380: 1483–1485.
- 185. Wong KH, Riaz MK, Xie Y, et al. (2019) Review of current strategies for delivering Alzheimer's disease drugs across the blood-brain barrier. *Int J Mol Sci* 20: 381.
- 186. Di Benedetto G, Burgaletto C, Carta AR, et al. (2019) Beneficial effects of curtailing immune susceptibility in an Alzheimer's disease model. *J Neuroinflammation* 16: 166.
- 187. Quitterer U, AbdAlla SN (2020) Improvements of symptoms of Alzheimers disease by inhibition of the angiotensin system. *Pharmacol Res* 154: 104230.
- 188. Blurton-Jones M, Kitazawa M, Martinez-Coria H, et al. (2009) Neural stem cells improve cognition via BDNF in a transgenic model of Alzheimer disease. *Proc Natl Acad Sci USA* 106: 13594–13599.
- 189. Blurton-Jones M, Spencer B, Michael S, et al. (2014) Neural stem cells genetically-modified to express neprilysin reduce pathology in Alzheimer transgenic models. *Stem Cell Res Ther* 5: 46.
- 190. Ager RR, Davis JL, Agazaryan A, et al. (2015) Human neural stem cells improve cognition and promote synaptic growth in two complementary transgenic models of Alzheimer's disease and neuronal loss. *Hippocampus* 25: 813–826.
- 191. Martini AC, Forner S, Trujillo-Estrada L, et al. (2018) Past to future: what animal models have taught us about Alzheimer's disease. *J Alzheimers Dis* 64: S365–S378.
- 192. Briggs R, Kennelly SP, O'Neill D (2016) Drug treatments in Alzheimer's disease. *Clin Med* 16: 247–253.
- 193. Cummings J, Lee G, Ritter A, et al. (2020) Alzheimer's disease drug development pipeline: 2020. *Alzheimers Dement TRCI* 6: e12050.
- 194. Bennett T, Bray D, Neville MW (2014) Suvorexant, a dual orexin receptor antagonist for the management of insomnia. *Pharm Ther* 39: 264–266.
- 195. Equihua-Benítez AC, Guzmán-Vásquez K, Drucker-Colín R (2017) Understanding sleep-wake mechanisms and drug discovery. *Expert Opin Drug Discov* 12: 643–657.
- 196. Steiner MA, Winrow CJ, editors (2014) Insomnia and beyond-Exploring the therapeutic potential of orexin receptor antagonists. *Frontiers Research Topics E-book series*, Switzerland: Frontiers Media SA, 1–219.
- 197. Pradeepkiran JA, Reddy PH (2019) Structure based design and molecular docking studies for phosphorylated tau inhibitors in Alzheimer's disease. *Cells* 8: 260.
- 198. Rêgo ML, Cabral DA, Costa EC, et al. (2019) Physical exercise for individuals with hypertension: It is time to emphasize its benefits on the brain and cognition. *Clin Med Insights Cardiol* 13: 1179546819839411.
- 199. Archer T, Josefsson T, Lindwall M (2014) Effects of physical exercise on depressive symptoms and biomarkers in depression. *CNS Neurol Disord Drug Targets* 13: 1640–1653.
- 200. Vina J, Sanchis-Gomar F, Martinez-Bello V, et al. (2012) Exercise acts as a drug; the pharmacological benefits of exercise. *Br J Pharmacol* 167: 1–12.
- 201. Daré LR, Garcia A, Neves BH, et al. (2020) One Physical Exercise Session Promotes Recognition Learning In Rats With Cognitive Deficits Related To Amyloid Beta Neurotoxicity. *Brain Res* 1744: 146918.

- 202. Carro E, Trejo JL, Busiguina S, et al. (2001) Circulating insulin-like growth factor I mediates the protective effects of physical exercise against brain insults of different etiology and anatomy. J Neurosci 21: 5678–5684.
- 203. He XF, Liu DX, Zhang Q, et al. (2017) Voluntary exercise promotes glymphatic clearance of amyloid beta and reduces the activation of astrocytes and microglia in aged mice. *Front Mol Neurosci* 10: 144.
- 204. Baker LD, Frank LL, Foster-Schubert K, et al. (2010) Effects of aerobic exercise on mild cognitive impairment: a controlled trial. *Arch Neurol* 67: 71–79.
- 205. Lange-Asschenfeldt C, Kojda G (2008) Alzheimer's disease, cerebrovascular dysfunction and the benefits of exercise: from vessels to neurons. *Exp Gerontol* 43: 499–504.
- 206. Ziemann E, Zembroñ-Lacny A, Kasperska A, et al. (2013) Exercise training-induced changes in inflammatory mediators and heat shock proteins in young tennis players. J Sports Sci Med 12: 282–289.
- 207. Bobinski F, Teixeira JM, Sluka KA, et al. (2018) IL-4 mediates the analgesia produced by low-intensity exercise in mice with neuropathic pain. *Pain* 159: 437–450.
- 208. Liu Y, Chu JM, Yan T, et al. (2020) Short-term resistance exercise inhibits neuroinflammation and attenuates neuropathological changes in 3xTg Alzheimer's disease mice. J Neuroinflammation 17: 4.
- 209. Saucedo Marquez CM, Vanaudenaerde B, Troosters T, et al. (2015) High-intensity interval training evokes larger serum BDNF levels compared with intense continuous exercise. *J Appl Physiol* 119: 1363–1373.
- 210. Pajonk FG, Wobrock T, Gruber O, et al. (2010) Hippocampal plasticity in response to exercise in schizophrenia. *Arch Gen Psychiat* 67: 133–143.
- 211. Adlard PA, Perreau VM, Pop V, et al. (2005) Voluntary exercise decreases amyloid load in a transgenic model of Alzheimer's disease. *J Neurosci* 25: 4217–4221.
- 212. Um HS, Kang EB, Koo JH, et al. (2011) Treadmill exercise represses neuronal cell death in an aged transgenic mouse model of Alzheimer's disease. *Neurosci Res* 69: 161–173.
- 213. Leem YH, Lim HJ, Shim SB, et al. (2009) Repression of tau hyperphosphorylation by chronic endurance exercise in aged transgenic mouse model of tauopathies. J Neurosci Res 87: 2561–2570.
- 214. Belarbi K, Burnouf S, Fernandez-Gomez FJ, et al. (2011) Beneficial effects of exercise in a transgenic mouse model of Alzheimer's disease-like Tau pathology. *Neurobiol Dis* 43: 486–494.
- 215. Alkadhi KA, Dao AT (2018) Exercise decreases BACE and APP levels in the hippocampus of a rat model of Alzheimer's disease. *Mol Cell Neurosci* 86: 25–29.
- 216. Hashiguchi D, Campos HC, Wuo-Silva R, et al. (2020) Resistance Exercise Decreases Amyloid Load and Modulates Inflammatory Responses in the APP/PS1 Mouse Model for Alzheimer's Disease. *J Alzheimers Dis* 73:1525–1539.
- 217. Mankhong S, Kim S, Moon S, et al. (2020) Effects of Aerobic Exercise on Tau and Related Proteins in Rats with the Middle Cerebral Artery Occlusion. *Int J Mol Sci* 21: 5842.
- 218. Khazaeinia T, Ramsey AA, Tam YK (2020) The effects of exercise on the pharmacokinetics of drugs. *J Pharm Pharm Sci* 3: 292–302.
- 219. Taylor D (2014) Physical activity is medicine for older adults. Postgrad Med J 90: 26-32.

- 220. World Health Organization (2020) WHO guidelines on physical activity and sedentary behaviour: at a glance. Available from: World Health Organization. https://apps.who.int/iris/handle/10665/337001. Licence: CC BY-NC-SA 3.0 IGO, 1–17.
- 221. Spirduso WW, Francis K, MacRae P (1995) Physical dimensions of aging, xiii Eds., USA Champaign Illinois: Human kinetics, 1–432.
- 222. Van Praag H, Shubert T, Zhao C, et al. (2005) Exercise enhances learning and hippocampal neurogenesis in aged mice. *J Neurosci* 25: 8680–8685.
- 223. Lledo PM, Alonso M, Grubb MS (2006) Adult neurogenesis and functional plasticity in neuronal circuits. *Nat Rev Neurosci* 7: 179–193.
- 224. Erickson KI, Voss MW, Prakash RS, et al. (2011) Exercise training increases size of hippocampus and improves memory. *PNAS* 108: 3017–3022.
- 225. Erickson KI, Kramer AF (2009) Aerobic exercise effects on cognitive and neural plasticity in older adults. *Br J Sports Med* 43: 22–24.
- 226. Voss MW, Prakash RS, Erickson KI, et al. (2010) Plasticity of brain networks in a randomized intervention trial of exercise training in older adults. *Front Aging Neurosci* 2: 32.
- 227. Langlois F, Vu TT, Chassé K, et al. (2013) Benefits of physical exercise training on cognition and quality of life in frail older adults. *J Gerontol:Series B* 68: 400–404.
- 228. Geda YE, Roberts RO, Knopman DS, et al. (2010) Physical exercise, aging, and mild cognitive impairment: a population-based study. *Arch Neurol* 67: 80–86.
- 229. Nuzum H, Stickel A, Corona M, et al. (2020) Potential benefits of physical activity in MCI and dementia. *Behav Neurol* 2020: 7807856.
- 230. Burns JM, Cronk BB, Anderson HS, et al. (2008) Cardiorespiratory fitness and brain atrophy in early Alzheimer disease. *Neurology* 71: 210–216.
- 231. Chen WW, Zhang X, Huang WJ (2016) Role of physical exercise in Alzheimer's disease. *Biomed Rep* 4: 403–407.
- 232. Teri L, Gibbons LE, McCurry SM, et al. (2003) Exercise plus behavioral management in patients with Alzheimer disease: a randomized controlled trial. *JAMA* 290: 2015–2022.
- 233. Teri L, McCurry SM, Buchner DM, et al. (1998) Exercise and activity level in Alzheimer's disease: a potential treatment focus. *J Rehabil Res Dev* 35: 411–419.
- 234. Palleschi L, Vetta F, De Gennaro E, et al. (1996) Effect of aerobic training on the cognitive performance of elderly patients with senile dementia of Alzheimer type. *Arch Gerontol Geriatr* 22: 47–50.
- 235. George EK, Reddy PH (2019) Can healthy diets, regular exercise, and better lifestyle delay the progression of dementia in elderly individuals? *J Alzheimers Dis* 72: S37–S58.
- 236. Fitzgerald PB, Daskalakis ZJ (2013) Repetitive transcranial magnetic stimulation treatment for depressive disorders: a practical guide. *Springer Science & Business Media*, Germany: Springer-Verlag Berlin Heidelberg, 1–121.
- 237. Weiler M, Stieger KC, Long JM, et al. (2020) Transcranial Magnetic Stimulation in Alzheimer's disease: are we ready? *eNeuro* 7.
- 238. Abraham G, Milev R, Lazowski L, et al. (2007) Repetitive transcranial magnetic stimulation for treatment of elderly patients with depression—an open label trial. *Neuropsychiatr Dis Treat* 3: 919–924.

- 239. Cotelli M, Manenti R, Cappa SF, et al. (2008) Transcranial magnetic stimulation improves naming in Alzheimer disease patients at different stages of cognitive decline. *Eur J Neurol* 15: 1286–1292.
- 240. Cotelli M, Calabria M, Manenti R, et al. (2011) Improved language performance in Alzheimer disease following brain stimulation. *J Neurol Neurosurg Psychiatry* 82: 794–797.
- 241. Rutherford G, Lithgow B, Moussavi Z (2015) Short and long-term effects of rTMS treatment on Alzheimer's disease at different stages: a pilot study. *J Exp Neurosci* 9: 43–51.
- 242. Koch G, Bonnì S, Pellicciari MC, et al. (2018) Transcranial magnetic stimulation of the precuneus enhances memory and neural activity in prodromal Alzheimer's disease. *Neuroimage*. 169: 302–311.
- 243. Rabey JM, Dobronevsky E (2016) Repetitive transcranial magnetic stimulation (rTMS) combined with cognitive training is a safe and effective modality for the treatment of Alzheimer's disease: clinical experience. *J Neural Transm* 123: 1449–1455.
- 244. Yang YW, Pan WX, Xie Q (2020) Combined effect of repetitive transcranial magnetic stimulation and physical exercise on cortical plasticity. *Neural Regen Res* 15: 1986–1994.
- 245. Ross RE, VanDerwerker CJ, Newton JH, et al. (2018) Simultaneous aerobic exercise and rTMS: Feasibility of combining therapeutic modalities to treat depression. *Brain Stimul* 11: 245–246.
- 246. Reed T, Kadosh RC (2018) Transcranial electrical stimulation (tES) mechanisms and its effects on cortical excitability and connectivity. *J Inherit Metab Dis* 41: 1123–1130.
- 247. Zarifkar AH, Zarifkar A, Nami M, et al. (2018) The Comparison of the Effects of Different Transcranial Electrical Stimulation (tES) Paradigms on Beta-Amyloid (Aβ 25–35)-Induced Memory Impairment upon Morris Water Maze Task in Male Rats. *J Neurol Neurosci* 9: 265.
- 248. Steinberg F, Pixa NH, Fregni F (2019) A review of acute aerobic exercise and transcranial direct current stimulation effects on cognitive functions and their potential synergies. *Front Hum Neurosci* 12: 534.



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