

ORIGINAL RESEARCH

RFI Gene Mutation in Familial Hemophagocytic Lymphohistiocytosis 2: A Family Report and Literature Review

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Objective: Gene mutation analysis was performed on a family with familial hemophagocytic lymphohistiocytosis (FHL) so as to provide an accurate etiological diagnosis, leading to genetic counseling for the family members.

Methods: The clinical data of two probands (siblings) with FHL in one family were analyzed, and eight genes related to the onset of the primary hemophagocytic lymphohistiocytosis (pHLH) (*PRF1*, *UNC13D*, *STX11*, *STXBP2*, *SH2D1A*, *BIRC4/XIAP*, *Rab27a*, *LYST*) were detected and analyzed in the probands and their parents with whole exome sequencing. **Results:** Proband 1 was a two-year-old male with the clinical manifestations of fever, hepatosplenomegaly, and a decreased peripheral blood cell count, sCD25: 12504pg/mL. The results of genetic testing showed that there was a c.1349C>T heterozygous missense mutation and a c.853_855del heterozygous mutation in the *PRF1* in proband 1. Proband 2 was an eight-year-old female with the clinical manifestations of convulsions and disturbance of consciousness with fever. The genetic test results were the same as those of proband 1. There was a single heterozygous mutation in the parents of the probands, and both probands had compound heterozygous mutations.

Conclusion: According to the clinical manifestations, laboratory tests, and results of the family molecular genetic testing, the probands could be clinically diagnosed as FHL2. The results of gene sequencing revealed that this was an autosomal recessive family with familial hemophagocytic syndrome. A rare pathogenic mutation (c.853_855del) in the *PRF1* was discovered in the two patients with HLH.

Keywords: FHL, PRF1, novel mutation

Hemophagocytic lymphohistiocytosis (HLH) is also referred to as hemophagocytic syndrome (HPS). HLH is a disease caused by the dysfunction of the immune system. The etiologies include molecular genetic abnormalities, infections, tumors, and autoimmune diseases. The main clinical manifestations are repeated fever, a decreased blood cell count to varying degrees, hepatosplenomegaly, and hepatic function abnormalities, together with manifestations of the central nervous system in many cases. Studies have found that children with HLH have cytotoxic cell dysfunction, meaning that antigens cannot be cleared in the normal manner. In turn, the monocytes and macrophages are constantly stimulated by antigens and are abnormally activated to produce a large number of cytokines, causing tissue damage. HLH progresses quickly with potentially dangerous consequences. If it is not treated in time, or treated ineffectively, survival duration is short and the

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prognosis is poor.⁴ HLH is traditionally divided into two subtypes, according to whether there are genetic defects: primary (hereditary) HLH and secondary (acquired) HLH. Hereditary HLH comprises the familial HLH (FHL) types 1–5 and HLH caused by primary immunodeficiency syndrome, and mainly occurs in infants or during early childhood. Acquired HLH is secondary to infections, rheumatic diseases and tumors.⁵

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FHL2 is the most common familial form of HLH with immune deficiency. FHL2 patients have mutations in genes affecting the expression of perforin which contribute to excessive activation and hyperplasia of CD8+ cytotoxic T lymphocytes (CTL) and natural killer (NK) cells. Perforin, a cytolytic protein, is a major factor responsible for the cytolytic activities of CTL and NK cells. Perforin, a pore-forming protein, creates pores in the target cytomembrane, generating apoptotic death through granzymes. PRF1 mutations cause decreased or absent expression of perforin protein, resulting in cytotoxic immunodeficiency. The loss of cytotoxic immune function contribute to an uncontrolled and persistent immune stimulus response in FHL2 patients, accompanied by high levels of cytokine release. If the immune system and excessive cytotoxic T-cell and NK-cell stimulation incontrollable. They can cause systemic inflammation and multiorgan failure.⁶

In 2004, the International Society of Tissue Cells revised the diagnostic criteria for familial and reactive HLH.⁷ In the present study, gene mutation analyses were performed on one family, consisting of two children with FHL and their parents, with a view to providing an accurate etiological diagnosis, followed by etiological diagnosis and genetic counseling for the family members.

Case Report

Proband 1, a 2-year-old male, reported "fever and hepatosplenomegaly" at the time of the hospital visit. There was no abnormality in the personal history and past history of the proband. The parents were not consanguineous and had no similar medical history. We perfected the laboratory examination and found: He had decreased peripheral blood cells, bone marrow cell morphology shows: lymphocytes increase, hemophagocytic cells are more common, and rised sCD25: 12504pg/mL. The ferritin level is normal.

Proband 2 (the sibling of proband 1), 8 years old, was first diagnosed with acute disseminated encephalomyelitis (ADEM) in a hospital in Beijing after presenting with "convulsions, disturbance of consciousness and fever for

two days" more than a year ago. Proband 2's symptoms recovered after immunotherapy. Magnetic resonance imaging (MRI) of the head during the hormone reduction period indicated that there were lesions that were aggravated, and this anomaly was followed by neurological symptoms. With the re-application of hormone therapy, proband 2's condition improved, but the intracranial lesions still showed progressive aggravation. Transient cytopenia, hepatosplenomegaly, and decreased NKC activity were observed during the course of the disease with normal hepatic function, levels of triacylglyceride and coagulation function in multiple detections. Eventually, after disease analysis and genetic examinations, proband 2 was given the diagnosis of "familial haemophagocytosis syndrome (FHPS) with the onset of the central nervous system". And the proband 2's bone marrow cell morphology shows: lymphocytes increase, hemophagocytic cells are more common.

The proband 1 met the following five, out of a total of eight, clinical diagnostic criteria of HLH-2004: fever, splenomegaly, cytopenia, hemophagocytosis sCD25:12504pg/mL. While the proband 2 also met five: fever, splenomegaly, cytopenia, hemophagocytosis and decreased NKC activity. Since the two probands were strongly suspected of being HLH, genetic testing of 12 subjects in three generations of the family was carried out, and the blood was drawn and sent to the Beijing Kangxu Medical Laboratory. The proband 1 did full exon detection, using the method of next-generation sequencing, while the proband 2 did site verification, using sanger sequencing. The genotypes of these two sites can be determined by sanger sequencing. The results are shown in Figure 1. There was a heterozygous missense mutation in exon 3 c.1349C>T (nucleotide No. 1349 in the coding region was changed from C to T), which was detected in the probands, their father, grandfather and father's sister, and c.853 855del (nucleotide No. 853 855 in the coding region) hybrid combined frameshift mutation, which was detected in the probands, their mother, grandfather, and mother's brother. The probands's genes sequencing results in Figure 2. The c.1349C>T heterozygous missense mutation caused amino acid N450 to change from Thr to Met; while the c.853 855del heterozygous frameshift mutation caused the deletion of amino acid N285. The protein damage prediction results were analyzed by SIFT, Polyphen2 and Mutation Taster. The c.1349C>T heterozygous missense mutation is c. 1349c > t heterozygous missense mutation is harmful and pathogenic. While

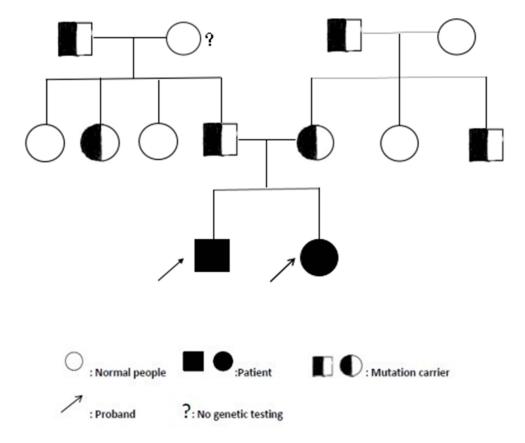


Figure I HLH pedigree map.

c.853 855del heterozygous frameshift mutation is suspected to be pathogenic. The above-mentioned mutations might have caused the protein function to be affected, which in turn would lead to the occurrence of HLH. The mutation site c.1349C>T (p.Thr450Met) has been included in the dbSNP database with the ID of rsl8965890, and literature reports related to HLH disease can be retrieved.⁸ While the pathogenicity of the mutation c.853 855del has not been reported in the Chinese literature, nor has it been included in the dbSNP database. Both the maternal and fraternal grandfathers, the maternal grandmother, the parents, the second aunt, and an uncle had single heterozygous mutations. None of the above-mentioned mutations were polymorphic changes, which have an extremely low frequency of occurrence in the population. The compound heterozygous variants found in the PRF1 gene of the probands were inherited from their parents, and the above variants might be pathogenic variants that caused the onset of the disease in the probands.

The proband 1 was Proband 1 received bone marrow transplantation successfully in January 2020. And proband 2 is scheduled to receive bone marrow transplantation this year.

Informed written consent was obtained from patients and their guardians for the publication of the case details and any accompanying images.

Literature Review

The CNKI and Wanfang databases were searched using the keywords of "familial hemophagocytic syndrome", "PRF1" and "novel mutation", and the PubMed database was searched using the keywords of "FHL", "PRF1" and "novel mutation" from January 1st, 2009 to June 30th, 2020.

From a comprehensive database of gene mutations related to the disease, the four genes for FHL, namely, *PRF1*, *UNC13D*, *STXBP2*, and *STX11*, were investigated. There were 433 different mutations of these genes to be found in the literature review, of which *UNC13D* had the maximum amount of mutations (189), followed by *PRF1* (157), *STXBP2* (66), and then *STX11* (21).

Thus, it was shown that mutations in the *PRF1* accounted for the main proportion of the occurrences of FHL. Genotypic characteristics and immune indicators were evaluated in a study concerning 311 cases of HPS in China. It was reported that in the 128 cases where

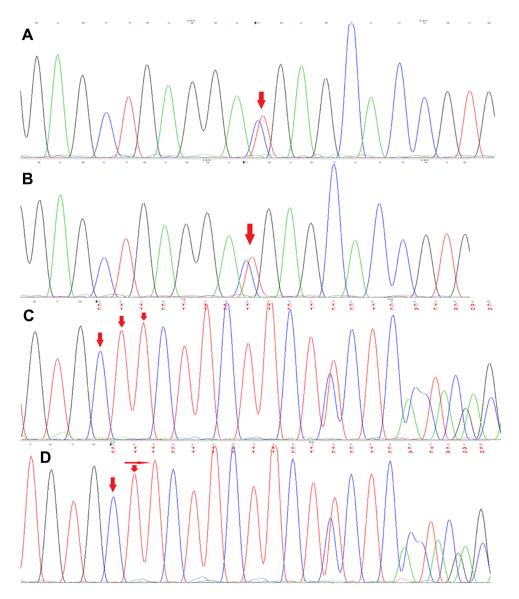


Figure 2 The probands's genes sequencing results (A). The sequencing peak of c.1349C>T of Proband I. (B) The sequencing peak of c.1349C>T of Proband 2. (C) The sequencing peak of c.853 855del of Proband I. (D) The sequencing peak of c.853 855del of Proband 2.

genetic testing was complete, the different types of genetic variation could be clarified. The mutation rate of the UNC13D gene was said to be the highest (29%), and then LYST (21%), PRF1 (17%) and STXBP2 (10%). The most common of the main pathogenic genes in 39 cases of primary HPS was PRF1 (16/39, 41%), follow by UNC13D (10/39, 26%) and BIRC4 (6/39, 15%). Also, there were differences in the genes related to primary HPS in children and adolescents, in that PRF1 was more prominent in children and adolescents while in grown-up UNC13D was the most common, followed by *PRF1*.¹⁰

There was a report in X of a new fatal case of congenital pyrimidine metabolic abnormality and rapid progression of multiple organ failure. It was found that the patient had two single gene defects at the same time, including the c.853 855del. At present, this gene defect has not been reported in China. The mixed phenotype of the two coexisting single gene defects might have been the main reason for the rapid deterioration of the condition in the patient, leading to multiple organ failure and early death.11

Discussion

HLH is a group of clinical syndromes in which lymphocytes and macrophages are excessively activated and then proliferate due to multiple etiologies. A large number of inflammatory cytokines are released causing a "cytokine storm", resulting in multiple organ dysfunction. Current

studies believe that the main pathogenesis of the disease is an immune regulation disorder or immune imbalance, causing the excessive secretion of lymphokines and monocyte factors by T lymphocytes and mononuclear macrophages. In addition, a cascade of inflammatory mediators will lead to corresponding clinical symptoms. The highly activated macrophages, which have strong cell activation, proliferation, and hemophagocytic abilities, together with the increased levels of inflammation, can inhibit blood cell production and reduce the amount of blood cells.¹

Classify FHL as five subtypes, types 1 to 5, all of which are autosomal recessive diseases. Except for FHL1, in which the gene defect is unknown, four different pathogenic genes have been identified: PFR1 (FHL2), UNC13D (FHL3), STX11 (FHL4), and STXBP2 (FHL5). 12 The incidence of gene defects in FHL, of which FHL2 and FHL3 are the most common, is largely correlated with race. 13 The PRF1 is located at 10q21-22 and contains three exons. The coding regions are the 2nd and 3rd exons. The perforin encoded in PRF1 contains 555 amino acids, including three transmembrane regions, the EGF-like region and three functional regions in the C2 region. The perforin can produce cytotoxic particles in the cytoplasm of target cells, which can lead to apoptosis. The NK cells and CTL cells mainly kill the target cells through the perforin/granzyme pathway. When this gene is mutated, the expression of perforin is down-regulated, and so the stability is reduced. When cytotoxic cells make contact with the target cell, the damaged perforin cannot smoothly form a channel on the membrane of the target cell. This will prevent the perforin from delivering the granzyme into the target cell to induce the apoptosis and osmotic dissolution, and, eventually, lead to impaired function in the killing of the target cells.

The *PRF1* mutation is the most frequency mutation, accounting for 20–40% of all FHL cases. The highest rate of *PRF1* mutation was reported in African-American families (>50%), and then the population of Turkey (43%), Japan (42%), and India (41%). According NCBI clinvar database (https://www.ncbi.nlm.nih.gov/clinvar), a total of 173 *PFR1* mutations have been reported now, among which 28 mutations were definitely pathogenic, all occurring in the second and third exons of the coding region. Some special mutation sites are more common in subjects from specific races and regions. The nonsense mutations

(c.1122G>A) of *PRF1* mutations are more common in the Turkish population, the frameshift mutations (c.1090_1091delCT and c.50delT) are more common in Japanese and African-American populations, while the missense mutations, p.Ala91Val and p.Trp129Ser, have appeared in the Italian and Indian populations, respectively. The missense mutations (c.445G>A) are common in Spanish patients while in Japanese patients with HLH, the most common mutation is c.1090_1091delCT (accounting for approximately 62.5% of the *PRF1* mutations), followed by c.207delC (about 37.5% of *PRF1* mutations). ^{15,16}

In addition, mutations and variants of the *PRF1* have also been reported in other diseases, including perforin deficiency syndrome, multiple sclerosis, type 1 diabetes, non-Hodgkin's lymphoma, and leukemia. So far, more than 115 pathogenic gene variants have been reported for *PRF1*, which mainly is the missense and nonsense mutations. Generally, the severity of the disease depends on the residual activity of perforin (Table 1).¹⁷

According to data from European and American HLH registration centers, FHL3 accounts for about 30-50% of FHL, second only to FHL2. However, data from Japan, South Korea, and other countries in Asia show that FHL3 is the most common and can account for up to 89% of FHL in this part of the world. 18 According to the statistics from the Histiocytosis Research Group of the Chinese Academy of Pediatrics in 2016, there were 323 cases of HLH in 12 hospitals in China. Among these cases, FHL accounted for 27.9%, of which 41.7% were FHL3, exceeding FHL2, which accounted for 29.2%. 19 These data indicate that the mutation type in FHL has a geographical location and race difference. FHL3 is an autosomal recessive inherited disease. So far, 112 mutations have been discovered. The UNC13D gene mutation causes the abnormal function of the Munc13-4 protein, leading to severe damage to the killing function of the NK cells. The mutation sites are all over the gene, and nonsense/missense mutations predominate (53.6%).²⁰

Conclusion

In the present case, both the two probands in the family were confirmed with a diagnosis of familial HPS. A pathogenic mutation in the *PRF1* was found in the probands, and there have been related literature reports concerning the heterozygous mutation of c.1349C>T. The pathogenicity of the c.853_855del heterozygous mutation has not been reported in China.

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Table I Mutation of PRFI Gene in Database

Mutation Site Amino Acid Change c.694C>T Arg232Cys²¹ His398fsX32²¹ c.1191insTG Arg410Trp²² c.1228C>T Thr450Met²² c.1349C>T $Met\,I\,Val^{22}$ c.IA>G c.283T>C Trp94Arg²³ Gly220Ser²³ c.658G>A c.662C>T Thr221lle²³ Arg225Trp²³ c.673C>T c.1182insT Gly394FsX²³ $Leu\,I\,7fsX34^{24}$ c.50delT c.207delC Asp70fsX37²⁴ Leu364fsX93²⁴ c.1090-1091delCT Met I Val²⁴ c.3G>A Asp62fsX12²⁴ c.185 195dell1 Val38Met²⁴ c.112G>A Ala9 I Val²⁴ c.272C>T Gly I 49Ser²⁴ c.445G>A Ala262fsX22²⁴ c.786_801del16 Pro201Thr²⁴ c.601C>A Lys285del²⁴ c.853_855delAAG $His 222 G ln^{24} \\$ c.666C>A Arg232His²⁴ c.695G>A Arg240Gly²⁴ c.718C>G Trp374X²⁴ c.1122G>A c.1576delT Leu526fsX87²⁴ L283-L284del²⁵ c.847-852del6 c.657C>A Tyr219X²⁵ Asp430Tyr²⁶ c.1288G>T Glu317Arg²⁶ c.949G>A Ile224Asp²⁷ c.671T>A $Val50Met^{\color{red}27}$ c.148G>A Gly45Arg²⁸ c.133G>A Arg54Ser²⁸ c.160C>T Val38Leu²⁸ c.38G>C Lys284-Lys287del²⁸ c.284 287del12 Thr435Met²⁹ c.1304C>T Phe421Cys²⁹ c.1262T>G Gly305Asp²⁹ c.914G>A $\mathsf{Arg356Trp}^{29}$ c.1066C>T c.755A>G Asn252Ser²⁹ Phe I 93Leu²⁹ c.577T>C c.1229G>C Arg410Pro²⁹ Pro39His²⁹ c.116C>T $Arg361X^{30}$ c.1083delG Cys497X³⁰ c.1491T>A c.1289insG Asp430Gly³⁰ Gln416X³⁰ c.1246C>T Glu253Lys³⁰ c.757G>A c.253G>A Leu85lle³⁰

GIn64X³¹

Table I (Continued).

Mutation Site	Amino Acid Change
c.548T>G	Val183Gly ³¹
c.835G>A	Cys279Tyr ³¹
c.1286G>A	Gly429Glu ³¹
c.781G>A	Glu261Lys ³²
c.836G>A	Cys279Tyr ³²
c.503G>A	Ser168Asn ³³
c.1177T>C	Cys393Arg ³³
c.83G>A	Arg28His ³⁴
c.632C>T	Ala211Val ³⁴
c.93C>G	Cys31Trp ³⁴
c.145G>A	Asp49Asn ³⁵
c.1442A > C	Gln481Pro ³⁶
c.65delC	Arg22fsX29 ³⁷
c.757G>A	Glu253Lys ³⁸
c.643C>a	Leu215lle ³⁸
c.785C>A	Ala262Asp ³⁸
c.136G>U	Glu46X ³⁸
c.355A>U	Argl 19Trp ³⁸
c.1376C>T	Pro459Leu ³⁸
c.1562A>G	Tyr521Cys ³⁹
c.996_I000delGCCCG	Pro333fsX ³⁹
c.1310C>T	Ala437Val ⁴⁰
c.992C>T	Ser331Leu ⁴⁰
c.1229G>A	Arg410Gln ⁴⁰
c.10C>T	Arg4Cys ⁴⁰
c.655T>A	Tyr219Asn ⁴⁰
c.386G>C	Try129Ser ⁴¹
c.1243G>A	Ala415Thr ⁴¹
c.1471G>A	Asp491Asn ⁴¹
c.833G>A	Ala278Asn ¹⁴
c.658G>C	Gly220Arg ¹⁴
c.1183T>C	Cys395Arg ¹⁴
c.90T>G	Cys31Gly ¹⁴
c.1018G>A	Asp340Asn ¹⁴
c.490C>T	Gln I 64X ¹⁴
c.528_529delinsAA	Cys I 76X ¹⁴
c.1288insG	Asp430Gly fsX28 ¹⁴
c.394G>A	Gly132Arg ⁴²
c.941_948delinsA	L314fsX327 ⁴³
c.1226C>A	Pro409His ⁴³
c.443C>A	Ala I 48Asp ⁴³
c.590T>G	Leu 197Arg ⁴³
c.1314T>A	Tyr438 ⁴³
c.1385C>A	Ser462 ⁴³
c.150delG	Thr51fsX106 ⁴³
c.1337A>C	Gln446Pro ⁴³
c.916G>T	Gly306Cys ⁴³
c.851_862del12	Lys284_Lys287del ⁴³
c.350_356delinsATGC	Vall 17_Arg 1 19 ⁴³
c.785C>T	Ala262Val ⁴³

(Continued) (Continued)

c.190C>T

Table I (Continued).

Mutation Site	Amino Acid Change
c.688G>T	Gly230Cys ⁴³
c.382G>T	Asp128Tyr ⁴³
c.1153C>T	Arg385Cys ⁴³
c.626A>C	Gln209Pro ⁴³
c.886T>C	Tyr296H ⁴³
c.512C>A	Thr171Asn ⁴³
c.895C>A	Arg299Ser ⁴³
c.1081A>T	Arg361Trp ⁴³
c.614A>G	Asn205Ser ⁴³
c.285G>T	Trp95Cys ⁴³
c.140G > T	Gly47Val ⁴⁴
c.IIG>A	Arg4His ⁴⁵
c.403G>A	Vall35Met ⁴⁵
c.434T>C	Val I 45Ala ⁴⁵
c.796G>A	lle266Val ⁴⁵
c.1313A>G	Tyr438Cys ⁴⁶
c.282C>A	Asn94Lys ⁴⁷
c.536ins	[48]
c.1133A>G	Asn378Ser ⁴⁸
c.937A>G	Asn312Asp ⁴⁸
c.1259T>C	lle420Thr ⁴⁸
c.443T>C	Val I 48Ala ⁴⁸
c.422T>G	lle I 48Ser ⁴⁸
c.286G>C	Gln96Arg ⁴⁸
c.626G>A	Arg209GIn ⁴⁸
c.1168C>T	Arg390X ⁴⁹
c.724G>T	Cys242Gly ⁴⁹
c.902C>A	Ser301Tyr ⁴⁹
c.528_529delinsAA	Cys I 76X ⁵⁰
c.762delC	Cys255AlafsX73 ⁵⁰
c.1822del	Val608CysfsX16 ⁵⁰
c.858+1G>A	Splice error ⁵⁰
c.1519G>T	Glu507X ⁵⁰
c.1321A>G	Leu441Val ⁵¹
c.1081A>T	Arg361Trp ⁵²
c.163C>T	Arg55Cys ⁵³

However, it might also cause protein dysfunction, and this might be the cause of HLH in the probands. Meanwhile, detailed etiological diagnosis and genetic counseling has also been provided for the families of the patients.

Abbreviations

FHL, familial hemophagocytic lymphohistiocytosis; pHLH, primary hemophagocytic lymphohistiocytosis; HLH, Hemophagocytic lymphohistiocytosis; HPS, hemophagocytic syndrome; MRI, Magnetic resonance imaging; FHPS, familial haemophagocytosis syndrome.

Ethics Approval and Consent to Participate

This study was conducted with approval from the Ethics Committee of The Affiliated Hospital of Inner Mongolia Medical University. This study was conducted in accordance with the declaration of Helsinki. Informed written consent was obtained from patients and the children's guardians for the publication of the case details and any accompanying images.

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Disclosure

The authors report no conflicts of interest in this work.

References

- Shi LH, Tian L, Huang S, et al. Clinical features of phagocytic syndrome in children and the significance of genetic testing. *Chin J Pract Pediatr*. 2019;34(6):449–452.
- Chen X, Wang F, Zhang Y, et al. Genetic variant spectrum in 265 Chinese patients with hemophagocytic lymphohistiocytosis: molecular analyses of PRF1, UNC13D, STX11, STXBP2, SH2D1A, and XIAP. Clin Genet. 2018;94(2):200–212. doi:10.1111/cge.13363
- Jiang ZF, Shen KL, Shen Y, et al. Chu Fu tang practical pediatrics. 8th ed; 2018. ISBN 978-7-117-19978-0/R-19979.
- Rousselin A, Alavi Z, Le Moigne E, et al. Hemophagocytic syndrome in pregnancy: case report, diagnosis, treatment, and prognosis. *Clin Case Rep.* 2017;5(11):1756–1764. doi:10.1002/ccr3.1172
- Luo ZB, Tang YM. Research progress of primary phagocytic syndrome. Chin J Pediatr. 2014;52(4):267–270.
- Sevim H, Kocaefe YÇ, Onur MA, Uçkan-çetinkaya D, Gürpınar ÖA. Bone marrow derived mesenchymal stem cells ameliorate inflammatory response in an in vitro model of familial hemophagocytic lymphohistiocytosis 2. Stem Cell Res Ther. 2018;9(1):198. doi:10.1186/s13287-018-0941-y
- Mian A, Kumari K, Kaushal S, et al. Fatal familial hemophagocytic lymphohistiocytosis with perforin gene (PRF1) mutation and EBV-associated T-cell lymphoproliferative disorder of the thyroid. *Autops Case Rep.* 2019;9(3):e2019101. doi:10.4322/acr.2019.101
- Ding Q, Guo X, Li Q. [Analysis of PRF1gene variant in a child with late-onset familial hemophagocytic lymphohistiocytosis type 2 and severe central nervous system disease]. *Zhonghua Yi Xue Yi Chuan Xue Za Zhi*. 2019;36(6):592–594. Chinese. doi:10.3760/cma.j. issn.1003-9406.2019.06.015

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9. Viñas-Giménez L, Padilla N, Batlle-Masó L, et al. FHLdb: a comprehensive database on the molecular basis of familial hemophagocytic lymphohistiocytosis. Front Immunol. 2020;31(11):107. doi:10.3389/fimmu.2020.00107

- 10. Zhang J, Sun Y, Shi X, et al. Genotype characteristics and immunological indicator evaluation of 311 hemophagocytic lymphohistiocytosis cases in China. Orphanet J Rare Dis. 2020;15(1):112. doi:10.1186/s13023-020-01390-z
- 11. Pérez-Torras S, Mata-Ventosa A, Drögemöller B, et al. Deficiency of perforin and hCNT1, a novel inborn error of pyrimidine metabolism, associated with a rapidly developing lethal phenotype due to multi-organ failure. Biochim Biophys Acta Mol Basis Dis. 2019;1865(6):1182-1191. doi:10.1016/j.bbadis.2019.01.013
- 12. Fager Ferrari M, Leinoe E, Rossing M, et al. Germline heterozygous variants in genes associated with familial hemophagocytic lymphohistiocytosis as a cause of increased bleeding. Platelets. 2018;29 (1):56-64. doi:10.1080/09537104.2017.1293808
- 13. Benson LA, Li H, Henderson LA, et al. Pediatric CNS-isolated hemophagocytic lymphohistiocytosis. Neurol Neuroimmunol Neuroinflamm. 2019;6(3):e560. doi:10.1212/NXI.000000000000560
- 14. Mhatre S, Madkaikar M, Desai M, Ghosh K. Spectrum of perforin gene mutations in familial hemophagocytic lymphohistiocytosis (FHL) patients in India. Blood Cells Mol Dis. 2015;54(3):250-257. doi:10.1016/j.bcmd.2014.11.023
- 15. Sun SW, Guo X, Zhu YP, et al. Clinical phenotype and gene mutation analysis of a familial family with phagocytic syndrome. Chin J Med Gene. 2014;31(05):570–573. doi:10.3760/cma.j.issn.1003-9406.2014.
- 16. Sheth J, Patel A, Shah R, Bhavsar R, Trivedi S, Sheth F. Rare cause of Hemophagocytic Lymphohistiocytosis due to mutation in PRF1 and SH2D1A genes in two children - A case report with a review. BMC Pediatr. 2019;19(1):73. doi:10.1186/s12887-019-1444-4
- 17. Bordbar MR, Modarresi F, Farazi Fard MA, Dastsooz H, Shakib Azad N, Faghihi MA. A case report of novel mutation in PRF1 gene, which causes familial autosomal recessive hemophagocytic lymphohistiocytosis. BMC Med Genet. 2017;18(1):49. doi:10.1186/ s12881-017-0404-9
- 18. Huang JB, Wang J, Jiang L, et al. Late fetal onset of familial phagocytic syndrome - A case report. Chin J Exp Hematol. 2017;25 (6):1825–1828. doi:10.7534/j.issn.1009-2137.2017.06.045
- 19. Xu XJ, Wang HS, Ju XL, et al.; Histiocytosis Study Group of the Chinese Pediatric Society. Clinical presentation and outcome of pediatric patients with hemophagocytic lymphohistiocytosis in China: a retrospective multicenter study. Pediatr Blood Cancer. 2017;64(4):e26264. doi:10.1002/pbc.26264
- 20. Kamoun F, Hsairi M, Grandin V, Ben Ameur S, De Saint Basile G, Hachicha M. Familial haemophagocytosis lymphohisticytosis type 3: a case report. Arch Pediatr. 2017;24(1):33-35. doi:10.1016/j.arcped. 2016.10.021
- 21. Del Giudice E, Savoldi G, Notarangelo LD, et al. Acute inflammatory polyradiculoneuropathy perforin-deficient familial haemophagocytic lymphohistiocytosis. Acta Paediatr. 2003;92(3):398-401. doi:10.1111/j.1651-2227.2003. tb00566 x
- 22. Ishii E, Ueda I, Shirakawa R, et al. Genetic subtypes of familial hemophagocytic lymphohistiocytosis: correlations with clinical features and cytotoxic T lymphocyte/natural killer cell functions. Blood. 2005;105(9):3442-3448. doi:10.1182/blood-2004-08-3296
- 23. Clementi R, Zur Stadt U, Savoldi G. Six novel mutations in the PRF1 gene in children with haemophagocytic lymphohistiocytosis. J Med Genet. 2001;38(9):643-646. doi:10.1136/jmg.38.9.643
- 24. Zur Stadt U, Beutel K, Kolberg S, et al. Mutation spectrum in children with primary hemophagocytic lymphohistiocytosis: molecular and functional analyses of PRF1, UNC13D, STX11, and RAB27A. Hum Mutat. 2006;27(1):62-68. doi:10.1002/ humu.20274

- 25. Marcenaro S, Gallo F, Martini S, et al. Analysis of natural killer-cell function in familial hemophagocytic lymphohistiocytosis (FHL): defective CD107a surface expression heralds Munc13-4 defect and discriminates between genetic subtypes of the disease. Blood. 2006;108(7):2316-2323. doi:10.1182/blood-2006-04-015693
- 26. Bryceson YT, Rudd E, Zheng C, et al. Defective cytotoxic lymphocyte degranulation in syntaxin-11 deficient familial hemophagocytic lymphohistiocytosis 4 (FHL4) patients. Blood.(6):1906–1915. doi:10.1182/blood-2007-02-074468
- 27. Göransdotter Ericson K, Fadeel B, Nilsson-Ardnor S, et al. Spectrum of perforin gene mutations in familial hemophagocytic lymphohistiocytosis. Am J Hum Genet. 2001;68(3):590-597. doi:10.1086/318796
- 28. Horne A, Ramme KG, Rudd E, et al. Characterization of PRF1, STX11 and UNC13D genotype-phenotype correlations in familial hemophagocytic lymphohistiocytosis. Br J Haematol. 2008;143 (1):75-83. doi:10.1111/j.1365-2141.2008.07315.x
- 29. Chia J, Yeo KP, Whisstock JC, Dunstone MA, Trapani JA, Voskoboinik I. Temperature sensitivity of human perforin mutants unmasks subtotal loss of cytotoxicity, delayed FHL, and a predisposition to cancer. Proc Natl Acad Sci U S A. 2009;106 (24):9809–9814. doi:10.1073/pnas.0903815106
- 30. Nagai K, Yamamoto K, Fujiwara H, et al. Subtypes of familial hemophagocytic lymphohistiocytosis in Japan based on genetic and functional analyses of cytotoxic T lymphocytes. PLoS One. 2010;5
- 31. Suga N, Takada H, Nomura A, et al. Perforin defects of primary haemophagocytic lymphohistiocytosis in Japan. Br J Haematol. 2002;116(2):346-349. doi:10.1046/j.1365-2141.2002.03266.x
- 32. Ueda I, Morimoto A, Inaba T, et al. Characteristic perforin gene mutations of haemophagocytic lymphohistiocytosis patients in Japan. Br J Haematol. 2003;121(3):503-510. doi:10.1046/j.1365-2141.2003.04298.x
- 33. Wang JS, Wang Z, Wu L, Chen X, Feng C. Etiology analysis of 38 cases of hemophagocytic syndrome. Chin j Exp Hematol. 2010;18 (05):1316-1320.
- 34. Zhizhuo H, Junmei X, Yuelin S, et al. Screening the PRF1, UNC13D, STX11, SH2D1A, XIAP, and ITK gene mutations in Chinese chilwith Epstein-Barr virus-associated hemophagocytic lymphohistiocytosis. Pediatr Blood Cancer. 2012;58(3):410-414. doi:10.1002/pbc.23216
- 35. Chia J, Thia K, Brennan AJ, et al. Fatal immune dysregulation due to a gain of glycosylation mutation in lymphocyte perforin. Blood. 2012;119(7):1713-1716. doi:10.1182/blood-2011-08-374355
- 36. Sumegi J, Nestheide SV, Barnes MG, et al. Gene-expression signatures differ between different clinical forms of familial hemophagocytic lymphohistiocytosis. *Blood*. 2013;121(7):e14–24. doi:10.1182/ blood-2012-05-425769
- 37. Seo JY, Song JS, Lee KO, et al.; Korea Histiocytosis Working Party. Founder effects in two predominant intronic mutations of UNC13D, c.118-308C>T and c.754-1G>C underlie the unusual predominance of type 3 familial hemophagocytic lymphohistiocytosis (FHL3) in Korea. Ann Hematol. 2013;92(3):357-364. doi:10.1007/s00277-012-1628-6
- 38. Dias C, McDonald A, Sincan M, et al. Recurrent subacute post-viral onset of ataxia associated with a PRF1 mutation. Eur J Hum Genet. 2013;21(11):1232-1239. doi:10.1038/ejhg.2013.20
- 39. Wada T, Sakakibara Y, Nishimura R, et al. Down-regulation of CD5 expression on activated CD8+ T cells in familial hemophagocytic lymphohistiocytosis with perforin gene mutations. Hum Immunol. 2013;74(12):1579-1585. doi:10.1016/j.humimm.2013. 09 001
- 40. Zhang K, Chandrakasan S, Chapman H, et al. Synergistic defects of different molecules in the cytotoxic pathway lead to clinical familial hemophagocytic lymphohistiocytosis. Blood. 2014;124(8):1331-13 34. doi:10.1182/blood-2014-05-573105

- Mhatre S, Madkaikar M, Jijina F, Ghosh K. Unusual clinical presentations of familial hemophagocytic lymphohistiocytosis type-2. *J Pediatr Hematol Oncol*. 2014;36(8):e524–7. doi:10.1097/MPH.00 00000000000102
- Hussein AA, Hamadah T, Qandeel M, et al. Hematopoietic stem cell transplantation of an adolescent with neurological manifestations of homozygous missense PRF1 mutation. *Pediatr Blood Cancer*. 2014;61(12):2313–2315. doi:10.1002/pbc.25166
- Abdalgani M, Filipovich AH, Choo S, et al. Accuracy of flow cytometric perforin screening for detecting patients with FHL due to PRF1 mutations. *Blood*. 2015;126(15):1858–1860. doi:10.1182/blood-2015-06-648659
- 44. Romero CA, Sánchez IP, Gutierrez-Hincapié S, et al. A novel pathogenic variant in PRF1 associated with hemophagocytic lymphohistiocytosis. *J Clin Immunol*. 2015;35(5):501–511. doi:10. 1007/s10875-015-0169-x
- Willenbring RC, Ikeda Y, Pease LR, Johnson AJ. Human perforin gene variation is geographically distributed. *Mol Genet Genomic Med.* 2018;6(1):44–55. doi:10.1002/mgg3.344
- 46. Willenbring RC, Johnson AJ. Finding a balance between protection and pathology: the dual role of perforin in human disease. *Int J Mol Sci.* 2017;18(8):1608. doi:10.3390/ijms18081608
- Liu C, Li M, Wu X, Yao X, Zhao L. Type 2 familial hemophagocytic lymphohistiocytosis in half brothers: a case report. *Medicine*. 2018;97 (30):e11577. doi:10.1097/MD.000000000011577

- 48. Almalky MA, Saleh SHA, Baz EG, Fakhr AE. Clinico-laboratory profile and perforin gene mutations of pediatric hemophagocytic lymphohistiocytosis cases: a five-year single center study. *Pan Afr Med J.* 2020;27(36):354.
- Sato H, Kawasaki N, Kawasaki M, et al. Three consecutive cases of familial hemophagocytic lymphohistiocytosis, including a case due to maternal uniparental disomy. *J Pediatr Hematol Oncol*. 2020;42(8): e819–e821. doi:10.1097/MPH.000000000001681
- Shabrish S, Kelkar M, Yadav RM, et al. The spectrum of clinical, immunological, and molecular findings in familial hemophagocytic lymphohistiocytosis: experience from India. *Front Immunol*. 2021;5 (12):612583. doi:10.3389/fimmu.2021.612583
- 51. Godby RC, Kraemer RR, May J, et al. Co-occurrence of familial hemophagocytic lymphohistiocytosis Type 2 and chronic active Epstein-Barr virus in adulthood. *Am J Med Sci.* 2021;361 (3):388–393. doi:10.1016/j.amjms.2020.10.004
- Ahmari AA, Alsmadi O, Sheereen A, et al. Genetic and clinical characteristics of pediatric patients with familial hemophagocytic lymphohistiocytosis. *Blood Res.* 2021;56(2):86–101. doi:10.5045/ br.2021.2020308
- 53. Liu XY, Nie YB, Chen XJ, Gao XH, Zhai LJ, Min FL. Adult onset type 2 familial hemophagocytic lymphohistiocytosis with PRF1 c.65delC/c.163C>T compound heterozygous mutations: a case report. World J Clin Cases. 2021;9(10):2289–2295. doi:10.12998/wjcc.v9.i10.2289

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