OPEN

CYP1B1 Gene and Phenotypic Correlation in Patients From Northeastern Brazil With Primary Congenital Glaucoma

Rodrigo E.A. Coêlho, MD,*† Dayse R. Sena, MD, PhD,*‡ Fernando Santa Cruz,§ Bárbara C.F.S. Moura, MD,† Cristal C. Han, BSc,*† Flaviano N. Andrade,§ and Rodrigo P.C. Lira, MD, PhD*

Purpose: To identify variants in the *CYP1B1* gene in northeastern Brazilian patients with primary congenital glaucoma (PCG) and possible genotype-phenotype correlations.

Materials and Methods: This is a cross-sectional observational study of 17 nonrelated patients with PCG, performed at the Altino Ventura Foundation, Recife, Brazil, between December 2017 and February 2018. All patients underwent an examination, including gathering information from their medical records, slit-lamp examination, fundoscopy, tonography, and measuring corneal diameter and thickness.

Results: The mean age at the time of the examination was 27.7 years; 52.9% (n=9) were male, 29.4% (n=5) had history of parental consanguinity. The mean age when the diagnosis was confirmed was 0.53 ± 2.18 years. Horizontal corneal diameter ranged from 12 to 16 mm (mean: 14.05 ± 1.42 mm) and the IOP mean value was 17.31 ± 9.84 mm Hg. Predicted pathogenic variants of the *CYP1B1* gene were identified in 4 patients (23.5%). The differences among all clinical parameters did not reach statistical significance between individuals with and without *CYP1B1* variants (*P*-values > 0.05).

Conclusions: Two variants which had not been previously related to PCG in Brazil (c.182G>A, c.241T>A) were identified. No statistically significant genotype-phenotype correlations were found.

Key Words: primary congenital glaucoma, blindness, genotype, phenotype, *CYP1B1* gene

(J Glaucoma 2019;28:161-164)

P rimary congenital glaucoma (PCG)¹ is a rare and severe genetic disease which is commonly expressed during the first year of life and represents an important cause of childhood blindness worldwide.² The disease is characterized by increased intraocular pressure (IOP) which results in damage

Supported by the MÉC/MCTI/CAPES/CNPQ/FAPS # 735.884; Laboratório de Imunopatologia Keizo Asami (LIKA).

Disclosure: The authors declare no conflict of interest.

Reprints: Fernando Santa Cruz, Av. Prof. Moraes Rego, 1235— Cidade Universitária, Recife, PE 50670-901, Brazil (e-mail: f.santacruzoliveira@gmail.com).

Copyright © 2018 The Author(s). Published by Wolters Kluwer Health, Inc. This is an open-access article distributed under the terms of the Creative Commons Attribution-Non Commercial-No Derivatives License 4.0 (CCBY-NC-ND), where it is permissible to download and share the work provided it is properly cited. The work cannot be changed in any way or used commercially without permission from the journal.

DOI: 10.1097/IJG.000000000001132

to the optic nerve. This increase in the IOP is secondary to an obstruction of the aqueous humor drainage, which is due to a malformation in the trabecular meshwork.^{3–5} Furthermore, edema, corneal opacity, corneal enlargement, Descemet membrane rupture (Haab's Striae), buphthalmos, refractive errors, photophobia, epiphora, and blepharospasm are some other clinical features of PCG.^{6,7}

PCG is the most common type of glaucoma in childhood, with a global incidence rate of $\sim 1:10.000$ births.⁸ This disease exhibits a high prevalence in populations where consanguinity is common,⁹ such as in Saudi Arabia (1:2500)⁶ and among a population in southern India (1:3.300).¹⁰

PCG inheritance is primarily autosomal recessive, with 10% to 40% of its cases being familial.^{11,12} Wiggs et al¹² suggested that the number of individuals with *CYP1B1* deleterious variants might be higher than what was expected among the US population.

Five chromosomal loci are currently associated with the disease: GLC3A (chromosome 2p21), GLC3B (chromosome 1p36.2-p36.1), GLC3C (chromosome 14q24.3), GLC3D (chromosome 14q24.2-q24.3), and GLC3E (chromosome 9p21.2).¹³ Nevertheless, only 3 genes were found to be involved in the development of PCG: *CYP1B1* (cytochrome P450, family 1, subfamily B, polypeptide 1), situated in the GLC3A locus; *LTBP2* (latent protein-binding factor of beta 2 growth), located in the GLC3D; and *TEK* (tyrosine kinase receptor), in the GLC3E. The role that the proteins encoded by these genes play in the disease etiology remains unclear.^{13–15}

It is important to emphasize that > 140 variants in the coding sequence of the *CYP1B1* gene related to the development of PCG have already been reported.^{16,17}

Because of the fact that PCG presents a phenotypic heterogeneity, we believe that ethnically diversified populations, such as the Northeastern Brazilian population, might experience important improvements by obtaining early diagnosis and treatment. This would contribute to lower rates of unnecessary blindness.

MATERIALS AND METHODS

This was a cross-sectional observational study of 17 nonrelated Brazilian patients with PCG diagnosis, whose follow-up was performed at the Altino Ventura Foundation (Recife, Pernambuco, Brazil). This study was realized within the period between December 2017 and February 2018.

The study aims to determine the frequency and variant types of *CYP1B1* in patients from northeastern Brazil with the PCG diagnosis and try to establish a correlation between alterations in *CYP1B1* sequencing and the disease phenotype with regard to the clinical outcomes of PCG.

Received for publication May 18, 2018; accepted November 2, 2018. From the *Hospital das Clínicas, Federal University of Pernambuco (HC—UFPE); †Altino Ventura Foundation (FAV); ‡Laboratory of Immunopathology Keizo Asami (LIKA); and §School of Medicine, Federal University of Pernambuco, Recife, Pernambuco, Brazil.

All patients underwent an ophthalmologic examination through measuring visual acuity, slit-lamp examination, fundoscopy, measuring IOP through a Goldman or Perkins tonometer, corneal pachymetry, and measuring the corneal diameter. Visual acuity was measured using the Snellen chart at 20 feet of distance. The ultrasonic pachymetry was performed using a DGH4000B Pachymeter (DGH Technology Inc., Exton, PA). Ten consecutive measurements were performed in each eye after administering topical ophthalmic anesthetic. The corneal diameters were measured by an ophthalmic compass. Personal information such as sex, family consanguinity, age at disease onset, glaucoma medications and the number of previous ocular surgeries were obtained during the medical interview.

Patients with compatible findings of secondary glaucoma and/or any other ocular or systemic disease that could lead to optic disc abnormalities simulating glaucomatous lesions were excluded from the study. The patients were selected without any sex or age distinctions.

This study followed the tenets of the Declaration of Helsinki and all participants or their legal persons signed the free and informed consent form. This research project was approved by the Ethics Committee for Research involving human beings of the Federal University of Pernambuco's Center of Health Sciences (N. 1.783.121). The authors declare that they have no conflicts of interests, and informed consent was obtained from all individual participants included in the study.

Analysis of CYP1B1 Gene

Genomic DNA was isolated from peripheral blood samples of all patients. The genomic DNA was amplified by polymerase chain reaction (PCR) with GoTag Green Master Mix reagent 2× (Promega, Madison, WI) according to vendor specifications, using 3 pairs of specific primers corresponding to the coding sequence and intronic flanking region of CYP1B1 exons. After confirming the success of the 1% agarose gel amplification, the PCR products of each patient were mixed in a single tube (performing a pool) and quantified by Qubit 2.0 equipment. Each pool of different patients was prepared for second-generation sequencing using a Nextera XT DNA Library Prep Kit (Illumina) that works from the PCR product by tagging and adding adapters, according to the protocol provided by the manufacturer. The second-generation sequencing, also known as next generation sequencing (NGS), was performed in Illumina MiSeq equipment (Illumina, San Diego), and uses the sequencing technology of each amplicon by synthesis in a MiSeq Reagent Kit V2 kit (composed of enzymes, dNTPs, fluorescently labeled ddNTPs and buffers) according to what is recommended by the manufacturer. After the result is acquired by the equipment, the raw data is processed by specific bioinformatics tools generating individual files per patient pool. The use of internal and external quality controls ensures that all steps from sample preparation to release analysis have yielded reliable results. The produced sequences were processed by a bioinformatics team and transformed into analyzable data.

The primer sequences used for *CYP1B1* variants analysis were: CCCAGAGGCTGGGGGTAG, CCTCGGG TCGAGGAAGG, AACTTCTTCACGCGCCAG, AAAC TCAGCATATTCTGTCTCTACTCC, AATGGGAAAG ACAGCATTAGTC, ATGAAGAACCGCTGGGTATG, GCTGGGATTACAAGCCTAAGG, GAAGAAACTCC CACCACAGC, GCGGTAGAGACCCGGAC, GGGAG CCGTTTTAGCCC, AATGCACTTTGCCCTCTCC, GA CAAGCTCCGATGCCC.

Genotype-phenotype Correlation

A comparison was performed between the clinical findings of patients with *CYP1B1* gene variants and patients without an identified variant in order to study the relation between genotype and phenotype. Thus, several clinical parameters were assessed such as age at diagnosis, age at evaluation, IOP at evaluation, horizontal corneal diameter, bilateral disease, corneal thickness, number of surgical interventions, number of antiglaucoma medications, corneal haze, Haab's striae, buphthalmos, cup/disc ratio, sex, blindness, and consanguinity.

Statistical Analysis

A spreadsheet in Microsoft Excel was created in order to analyze the data, which was then transferred into SPSS version 18 software to perform the analysis. Mann-Whitney U test was performed to compare the means of the 2 independent samples, and Fisher exact test was performed to compare categorical variables. A *P*-value <0.05 was used for evidence of statistical significance.

RESULTS

Eight (47.06%) of the 17 patients were female and 9 (52.94%) were male. Among these patients, 15 (88.24%) showed bilateral impairments. The age when the diagnosis was confirmed varied from 0 to 9 (mean 0.53 ± 2.18 y). In the clinical evaluation, horizontal corneal diameter ranged from 12 to 16 mm (mean 14.05 ± 1.42 mm), and the mean IOP value was 17.31 ± 9.84 mm Hg. At least 2 surgical procedures in 18 eyes (52.9%) were required.

Predicted pathogenic variants of the *CYP1B1* gene were identified in 4 patients (23.5%). Most of these were missense variants: c.182G > A, c.241T > A, and c.1310C > T. One nonsense (c.55C > T) and 1 frameshift ($c.1209_{-1210ins}$ TCATGCCACC) were also detected. We also found 1 case of homozygosity (Table 1). Nine single nucleotide variants

Patients	Nucleotide Change	Protein Change	Exon	Zigozity
RC09	c.55C > T	p.Gln19Ter	2nd exon	Heterozygous
	c.241T > A	p.Tyr81Asn	2nd exon	Heterozygous
RC15	c.1310C > T	p.Pro437Leu	3rd exon	Heterozygous
	c.1209_1210insTCAT GCCACC	p.Thr404SerfsTer30	3rd exon	Heterozygous
RC16	c.1310C > T	p.Pro437Leu	3rd exon	Homozygous
RC17	c.1209 1210insTCATGCCACC	p.Thr404SerfsTer30	3rd exon	Heterozygous
	- c.182G > A	p.Gly61Glu	2nd exon	Heterozygous

(SNV) were reported: c.-1-12C > T (intron 1—rs2617266); c.1-14C > T (intron 1—rs4987134); c.1044-141_1044-140insT (intron 2—rs34468862); c.142C > G (R48G-exon 2—rs10012); c.355G > T (A119S—exon 2—rs1056827); c.729G > C (V243V—exon 2—rs9341249); c.1294C > G (L432V—exon 3—rs1056836); c.1347T > C (D449D—exon 3—rs1056837) and c.1358A > G (N453S—exon 3—rs1800440) (dbSNP—www. ncbi.nlm.nih.gov/SNP/).

All patients (100%) in the group with variants developed bilateral PCG, and 11 patients (84.6%) in the group without variants expressed bilateral disease (P = 1.000). However, the differences between individuals with and without CYP1B1 variants did not reach statistical significance for any of the variables (P-values > 0.05 in all comparisons) (Table 2).

DISCUSSION

CYP1B1 variants were identified in 4 patients (23.53%), and only one among these was homozygous. Five different variant types were detected, and all of them had already been described in PCG patients from different populations, $^{18-22}$ although c.182G > A and c.241T > A have never been cited in Brazilian patients.

The prevalence of *CYP1B1* variants that were found in the present study (23.53%) is comparable to what was observed in other populations such as in Mexico (10%), Japan (20%), and Portugal (28.57%).^{23–25} In contrast, there is a discrepancy between these numbers and those obtained in populations with high consanguinity rates such as in Morocco (40%), Iran (70%), Kuwait (70.6%), and Saudi Arabia.^{6,26–28} Different results were found in previous Brazilian studies such as those performed by De Melo et al²⁹ and Della Paolera et al³⁰ with 44% and 30%, respectively. Nevertheless, these studies did not include patients from northeastern Brazil in their samples, which is the object of the present study.

The 6 SNVs identified in exons in this study do not alter the disease phenotype and were previously described in affected individuals and healthy controls. Likewise, 3 SNVs were found in introns; 2 of them (rs4987134 and rs34468862) had no citation found in the literature, but the other one (rs2617266) was already described.³¹

Almost all *CYP1B1* gene variants identified in this study were heterozygous, showing that there is an allelic heterogeneity in patients with PCG from Northeastern Brazil, which is in accordance with the low consanguinity frequency in this region.

Martin et al³² speculated that individuals with compound heterozygous *CYP1B1* gene variants may exhibit a less severe form of the disease than those with homozygous alterations. On the other hand, in a cohort study, Lim et al³³ observed that both compound heterozygous and homozygous cases had indiscriminate clinical courses of the disease. This divergence in results is not well explained and could possibly be a consequence of regional differences, consanguine marriage percentage, variety of surgical techniques, immediate access to health services, postoperative follow-up time, population composition, studied samples or other factors not yet established.

Regarding the genotype-phenotype correlation, a great variability was shown in relation to the *CYP1B1* gene and the PCG. Weisschuh et al³⁴ compared a group of PCG patients with *CYP1B1* variants, and they did not observe significant differences between them regarding the age of disease onset, the severity of the condition or the response to the treatment. Campos-Mollo et al²¹ did not show significant differences between the presence or not of variants in relation to the ocular involvement, age at diagnosis, sex or number of surgical interventions. However, Abu-Amero et al²⁸ observed that individuals with pathogenic variants of *CYP1B1* had higher rates of postoperative visits and a greater necessity of antiglaucomatous drugs than individuals without pathogenic variants.

The present study did not find significant differences in the phenotype among the PCG population, with and without *CYP1B1* variants, although the limited costs of the research and consequently the limited number

TABLE 2. Clinical Parameters and Demographic Profile of Patients From Northeastern of Brazil With PCG and Variants of the CYP1	31
Gene and Patients Without Identified CYP71B1 Gene Variants	

Clinical Parameters	Patients With Variants $(n = 4)$	Patients Without Variants (n = 13)	Р
Sex [n (%)]			
Female	3 (75.0)	5 (38.5)	0.294†
Male	1 (25.0)	8 (61.5)	
Consanguinity [n (%)]	2 (50.0)	3 (23.1)	0.538†
Age at diagnosis (y)	2.3 ± 4.5	0.00 ± 0.00	*
Age at evaluation (y)	25.2 ± 15.6	28.5 ± 10.5	0.071*
No. antiglaucoma medications required	3.5 ± 1.0	2.5 ± 1.3	0.111*
IOP at evaluation (mm Hg) (RE/LE)	$15.0 \pm 2.4/12.5 \pm 3.5$	$19.3 \pm 9.7/16.6 \pm 12.6$	0.569/0.746*
Blindness in at least one eye	75.0	91.7	0.450†
Horizontal corneal diameter (mm) (RE/LE)	$13.6 \pm 1.3/13.5 \pm 2.1$	$14.0 \pm 1.4/14.3 \pm 1.6$	0.605/0.478*
Corneal thickness (µm) (RE/LE)	$524.3 \pm 20.6/523.0 \pm 17.0$	$598.4 \pm 111.0/557.6 \pm 117.9$	0.117/ 0.814*
No. surgical interventions (RE/LE) [n (%)]			
0-1	1 (25.0)/3 (75.0)	6 (46.2)/6 (46.2)	0.603/0.576†
≥ 2	3 (75.0)/1 (25.0)	7 (53.8)/7 (53.8)	
Corneal haze (RE/LE) [n (%)]	1 (25.0)/0 (0,0)	5 (38.5)/4 (30.8)	1.000/0.519†
Haab' strie (RE/LE) [n (%)]	0 (0.0)/0 (0.0)	3 (23.1)/6 (46.2)	0.541/0.237†
Buphthalmos (RE/LE) [n (%)]	1 (25.0)/0 (0.0)	4 (30.8)/4 (30.8)	1.000/0.519†
Cup/disc ratio (RE/LE)	$0.5 \pm 0.4/0. \pm 0.4$	$0.8 \pm 0.2/0.9 \pm 0.3$	0.404/0.073*

Values are mean \pm SD

*Mann-Whitney Test. †Fisher Exact Test.

IOP indicates intraocular pressure; PCG, primary congenital glaucoma.

Copyright © 2018 The Author(s). Published by Wolters Kluwer Health, Inc.

of participants may have contributed to this conclusion. Other studies have reported similar results.^{6,29}

The major limitation of this study was the small sample size due to the high costs of genetic testing. Nevertheless, it showed important results. Two variants that had not been previously related to PCG in Brazil (c.182G>A, c.241T>A) were identified.

In this study, there were no statistically significant differences between the clinical findings of the cases with and without variants. In addition, knowledge about PCG genetics is still far from complete and remains to be a challenging subject for further research. Therefore, early recognition of PCG signs and symptoms, and identification of families bearing pathogenic variants might have a significant impact on the prediction of disease severity and may help predict surgical outcomes. More efforts are needed to provide effectiveness, timely screening, and appropriate allocation of resources to enable health professionals to reduce the rates of avoidable blindness in Brazil and worldwide.

REFERENCES

- Amberger JSS, Bocchini CAA, Schiettecatte F, et al. OMIM. org: Online Mendelian Inheritance in Man (OMIM[®]), an Online catalog of human genes and genetic disorders. *Nucleic Acids Res.* 2015;43:D789–D798.
- Lewis CJ, Hedberg-Buenz A, DeLuca AP, et al. Primary congenital and developmental glaucomas. *Hum Mol Genet.* 2017; R1:R28–R36.
- Sarfarazi M, Stoilov I. Molecular genetics of primary congenital glaucoma. *Eye*. 2000;14:422–428.
- Bakunowicz-Lazarczyk A, Sulkowska M, Sulkowski S, et al. Ultrastructural changes in the trabecular meshwork of congenital glaucoma. J Submicrosc Cytol Pathol. 2001;33:17–22.
- deLuise VP, Anderson DR. Primary infantile glaucoma (congenital glaucoma). Surv Ophthalmol. 1983;28:1–19.
- Bejjani BA, Stockton DW, Lewis RA, et al. Multiple CYP1B1 mutations and incomplete penetrance in an inbred population segregating primary congenital glaucoma suggest frequent de novo events and a dominant modifier locus. *Hum Mol Genet*. 2000;9:367–374.
- Geyer O, Wolf A, Levinger E, et al. Genotype/phenotype correlation in primary congenital glaucoma patients from different ethnic groups of the israeli population. *Am J Ophthalmol.* 2011;151:263–271.
- Sarfarazi M, Stoilov I, Schenkman JB. Genetics and biochemistry of primary congenital glaucoma. *Ophthalmol Clin North Am.* 2003;16:543–554.
- Sena D, Finzi S, Rodgers K, et al. Founder mutations of *CYP1B1* gene in patients with congenital glaucoma from the United States and Brazil. *J Med Genet*. 2004;41:e6.
- Dandona L, Williams JD, Williams BC, et al. Population-based assessment of childhood blindness in southern India. Arch Ophthalmol. 1998;116:545–546.
- Ho C, Walton D. Primary congenital glaucoma: 2004 update. J Pediatr Ophthalmol Strabismus. 2004;41:271–288.
- Wiggs JL, Langgurth AM, Allen KF. Carrier frequency of CYP1B1 mutations in the United States (an American Ophthalmological Society thesis). *Trans Am Ophthalmol Soc.* 2014;112:94–102.
- 13. Chen Y, Jiang D, Yu L, et al. CYP1B1 and MYOC mutations in 116 Chinese patients with primary congenital glaucoma. *Arch Ophthalmol.* 2008;126:1443–1447.
- 14. Stoilov I, Akarsu AN, Alozie I, et al. Sequence analysis and homology modeling suggest that primary congenital glaucoma on 2p21 results from mutations disrupting either the hinge region or the conserved core structures of cytochrome P4501B1. *Am J Hum Genet*. 1998;62:573–584.

- López-Garrido MP, Medina-Trillo C, Morales-Fernandez L, et al. Null CYP1B1 genotypes in primary congenital and nondominant juvenile glaucoma. *Ophthalmology*. 2013;120:716–723.
- Stenson PD, Mort M, Ball EV, et al. The Human Gene Mutation Database: towards a comprehensive repository of inherited mutation data for medical research, genetic diagnosis and nextgeneration sequencing studies. *Hum Genet*. 2017;136:665–677.
- Cooper DN, Ball EV, Stenson PD, et al. The Human Gene Mutation Database (HGMD®). Available at: www.hgmd.cf. ac.uk/ac/index.php. Accessed September 9, 2017.
- Suh W, Kee C. A clinical and molecular genetics study of primary congenital glaucoma in South Korea. *Br J Ophthalmol.* 2012;96:1372–1377.
- Chavarria-Soley G, Michels-Rautenstrauss K, Pasutto F, et al. Primary congenital glaucoma and Rieger's anomaly: extended haplotypes reveal founder effects for eight distinct CYP1B1 mutations. *Mol Vis.* 2006;12:523–531.
- Reddy ABM, Kaur K, Mandal AK, et al. Mutation spectrum of the CYP1B1 gene in Indian primary congenital glaucoma patients. *Mol Vis.* 2004;10:696–702.
- Campos-Mollo E, Blanco-Marchite C, Garcia-Feijoo J, et al. CYP1B1 mutations in Spanish patients with primary congenital glaucoma: phenotypic and functional variability. *Mol Vis.* 2009;15: 417–431.
- Hollander DA, Sarfarazi M, Stoilov I, et al. Genotype and phenotype correlations in congenital glaucoma. *Trans Am Ophthalmol Soc.* 2006;104:183–195.
- Ohtake Y, Tanino T, Suzuki Y, et al. Phenotype of cytochrome P4501B1 gene (CYP1B1) mutations in Japanese patients with primary congenital glaucoma. *Br J Ophthalmol.* 2003;87: 302–304.
- Cardoso MS, Anjos R, Vieira L, et al. CYP1B1 gene analysis and phenotypic correlation in Portuguese children with primary congenital glaucoma. *Eur J Ophthalmol.* 2015;25:474–477.
- Zenteno JC, Hernandez-Merino E, Mejia-Lopez H, et al. Contribution of CYP1B1 mutations and founder effect to primary congenital glaucoma in Mexico. J Glaucoma. 2008;17:189–192.
- Belmouden A, Melki R, Hamdani M, et al. A novel frameshift founder mutation in the cytochrome P450 1B1 (CYP1B1) gene is associated with primary congenital glaucoma in Morocco. *Clin Genet*. 2002;62:334–339.
- Chitsazian F, Tusi BK, Elahi E, et al. CYP1B1 mutation profile of Iranian primary congenital glaucoma patients and associated haplotypes. J Mol Diagn. 2007;9:382–393.
- Abu-Amero KK, Osman EA, Mousa A, et al. Screening of CYP1B1 and LTBP2 genes in Saudi families with primary congenital glaucoma: genotype-phenotype correlation. *Mol Vis.* 2011;17:2911–2919.
- De Melo MB, Mandal AK, Tavares IM, et al. Genotypephenotype correlations in *CYP1B1*-associated primary congenital glaucoma patients representing two large cohorts from India and Brazil. Hejtmancik JF, ed. *PLoS ONE*. 2015;10:e0127147.
- Della Paolera M, de Vasconcellos JPC, Umbelino CC, et al. CYP1B1 gene analysis in primary congenital glaucoma Brazilian patients: novel mutations and association with poor prognosis. J Glaucoma. 2010;19:176–182.
- Tanwar M, Dada T, Sihota R, et al. Mutation spectrum of CYP1B1 in North Indian congenital glaucoma patients. *Mol Vis.* 2009;15:1200–1209.
- 32. Martin SN, Sutherland J, Levin AV, et al. Molecular characterisation of congenital glaucoma in a consanguineous Canadian community: a step towards preventing glaucoma related blindness. J Med Genet. 2000;37:422–427.
- Lim SH, Yanovitch TL, Freedman SF, et al. CYP1B1, MYOC, and LTBP2 mutations in primary congenital glaucoma patients in the United States. *Am J Ophthalmol.* 2012;155:508–517.
- Weisschuh N, Wolf C, Wissinger B, et al. A clinical and molecular genetic study of German patients with primary congenital glaucoma. *Am J Ophthalmol.* 2009;147:744–753.