

Differentiating allergic and irritant contact dermatitis by high-definition optical coherence tomography: a pilot study

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Received: 13 April 2014/Revised: 24 June 2014/Accepted: 18 August 2014/Published online: 4 September 2014
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Abstract Differentiation of allergic contact dermatitis (ACD) and irritant contact dermatitis (ICD) is important because of different management requirements. Various non-invasive tests have been used in an attempt to improve diagnosis. In irritant dermatitis, thickening of the epidermis has been a constant finding. High-Definition Optical Coherence Tomography (HD-OCT) is a non-invasive real-time three-dimensional imaging technique with cellular resolution for which an adapted algorithmic method for pattern analysis discriminating inflammatory skin diseases has been proposed. The aim of this study was threefold. (1) To evaluate the correlation between HD-OCT features and clinical scores of allergic and irritant patch test reactions. (2) To explore the potential of HD-OCT in optimizing the visual patch test scoring. (3) To assess in vivo the cytological and 3-D micro-architectural differences in skin reaction types between doubtful positive ACD and ICD. Twenty-two volunteers were patch tested using potassium(VI)dichromate, cobalt(II)chloride, nickel(II) sulfate and palladium(II)chloride. Visual patch test scoring and HD-OCT assisted patch test scoring were performed at 48 and 96 h after patch test application according to ECDRG guidelines. Selected HD-OCT features correlated well with clinical severity scores. HD-OCT assessment improved the visual patch test scoring although not significantly. Increased epidermal thickness observed in ICD at first reading was a significant finding useful in differentiating doubtful (+?) ACD from irritant

(IR) ICD reactions. In conclusion, HD-OCT might be a unique tool for in vivo non-invasive real-time three-dimensional epidermal thickness measurements helping to differentiate IR from doubtful (+?) reactions in patch testing. Selected HD-OCT features corresponded well with severity of visual scoring. These features might help to quantify the degree of inflammation in inflammatory skin conditions. HD-OCT might help in optimizing visual patch test scoring in some situations.

Keywords High-definition optical coherence tomography · Patch test scoring · Transition metals · Allergic contact dermatitis · Irritant contact dermatitis

Introduction

Contact dermatitis (CD) is an inflammatory eczematous skin disease mostly caused by chemicals or metal ions that exert toxic effects without inducing a T cell response (contact irritants) or by small reactive chemicals that modify proteins and induce innate and adaptive immune responses (contact allergens) [24]. Irritant and allergic contact dermatitis represent a major health problem. In general, irritant contact dermatitis (ICD) is more frequent than allergic contact dermatitis (ACD) [10]. The prevalence of CD is high as it is suggested that almost 20 % of the general population suffers from CD to at least one chemical, most commonly nickel (Ni) [19]. Differentiation of ACD and ICD is important because of different therapeutic and management options. In most cases contact dermatitis is suspected from anamnestic and clinical findings. Diagnosis is confirmed by patch testing. Recording of patch test reactions follows the guidelines of the International Contact Dermatitis Research Group as shown in

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Table 1 Recording of patch test reactions according to the international Contact Dermatitis Research Group (ICDRG) [4]

+?	Doubtful reaction; faint erythema only
+	Weak positive reaction; erythema, infiltration, possibly papules
++	Strong positive reaction; erythema, infiltration, papules, vesicles
+++	Extreme positive reaction; intense erythema and infiltration and coalescing vesicles
–	Negative reaction
IR	Irritant reaction of different types
NT	Not tested

Table 1 [33]. There is rarely disagreement concerning the reading of obvious irritant and ++ or +++ allergic reactions. By contrast the reading of +? and + reactions and some irritant reactions may cause difficulties. Skin biopsy is of little help in distinguishing the two because histopathologic changes and inflammatory infiltrate are often similar at this stage [21].

Different non-invasive tests have therefore been used in an attempt to improve diagnostic specificity [8, 11, 29, 30]. Among these, reflectance confocal microscopy (RCM) has been shown to be a promising tool for the differentiation of acute allergic and irritant contact dermatitis in vivo [31]. Relevant RCM features for diagnosis of ACD and ICD have been published recently [3]. Conventional optical coherence tomography (OCT) seems to be useful as an objective parameter in grading severity of patch test reactions [11]. Moreover, in irritant contact dermatitis thickening of the epidermis was a constant finding using different techniques and which could be monitored over time [34].

High-definition optical coherence tomography (HD-OCT) is a more recently introduced non-invasive three-dimensional imaging technique with cellular resolution [4–7]. An adapted algorithmic method for pattern analysis in inflammatory skin diseases has been suggested based on inflammation related HD-OCT features [7].

The aim of this pilot study is (1) to evaluate the correlation between selected HD-OCT features with clinical severity of patch test reactions (2) to explore the possible optimization of the visual patch test scoring of reactions to transition metals by HD-OCT and (3) to assess in vivo and real time the cytological and 3-D micro-architectural differences in skin reaction types of doubtful positive ACD and ICD.

Materials and methods

Patients and protocol design

Twenty-two Caucasian subjects with Fitzpatrick skin phototype II or III, aged between 18 and 69 years, were

selected for this study. Inclusion criteria were (1) suspected contact allergy to transition metals (2) patch testing on the back using the European Standard Series including potassium dichromate (VI) $\text{Cr}_2\text{K}_2\text{O}_7$ (in 0.5 % petrolatum), cobalt (II) chloride hexahydrate $\text{CoCl}_2\cdot 6\text{H}_2\text{O}$ (1 % petrolatum) and Nickel(II) sulfate hexahydrate $\text{NiO}_4\text{S}\cdot 6\text{H}_2\text{O}$ (5 % petrolatum) AND extended at least with palladium (II)chloride PdCl_2 (2 % petrolatum) (Chemotechnique Diagnostics, Vellinge, Sweden). (3) Positive irritant or allergic reaction(s) at first and/or second reading to transition metal(s), (4) availability of good quality HD-OCT imaging of reactions at first and second reading, and (5) all subjects provided informed consent for HD-OCT imaging.

The selected transition metals are capable of inducing both ICD and ACD reactions and therefore were considered particularly suitable for this study. Tests were applied using Finn Chambers[®] according to guidelines (SmartPractice, Phoenix, USA). The patch test substances were removed after 48 h and evaluated by clinical assessment and HD-OCT imaging 20 min after removal (first reading). A second reading was performed 48 h after removal of the patch (Table 2) Both readings are essential because of difference in kinetics of allergic and irritant contact reactions in vivo [4, 19]. A control site of normal skin and a site exposed to petrolatum (vehicle) only were also evaluated. Clinical photographs of skin reactions were taken under standardized conditions with a digital camera.

All clinical and HD-OCT assessments have been performed by the same investigator. An obligate irritant agent has not been added because increased epidermal thickness in the case of irritant reactions has already been documented quantitatively by RCM, conventional OCT and high-frequency ultrasound [1, 23, 34] and qualitatively by immunohistochemical findings [22, 23].

Clinical assessment: (Tables 1, 2)

The allergic patch test reactions were scored by a board-certified specialist of dermatology experienced in reading patch test results, using the visual grading scale recommended by the International Contact Dermatitis Research Group and the North American Contact Dermatitis Group [33]. Irritant reactions were scored according the irritant reaction types proposed by Wahlberg et al. [33].

Cross-sectional (CS) and en-face (EF) imaging by high-definition optical coherence tomography

High-definition optical coherence tomography (Skintell[®] Agfa Healthcare Belgium) uses a combination of parallel time-domain interferometry and adaptive optics resulting in a constant homogenous high resolution of 3 μm in all

Table 2 Visual (V) and HD-OCT assisted (H) patch test scoring

# Subject	First reading after 48 H								Second reading After 96 H							
	Cr		Co		Ni		Pd		Cr		Co		Ni		Pd	
	V	H	V	H	V	H	V	H	V	H	V	H	V	H	V	H
1	-	-	-	-	+?	+?	+?	+?	-	-	-	-	+/?	+/?	+	+
2	+?	+?	IR	IR	-	-	-	-	-	-	IR	IR	-	-	-	-
3	IR	IR	IR	IR	-	-	-	-	IR	IR	IR	IR	-	-	IR	+?
4	++	++	++	++	+	+	-	-	++	++	++	++	+	+	-	-
5	-	-	-	-	IR	IR	-	-	-	-	-	-	+	+	IR	+?
6	+?	<i>IR</i>	-	-	-	-	-	-	IR	IR	-	-	IR	IR	IR	IR
7	IR	IR	IR	IR	-	-	-	-	-	-	IR	IR	-	-	-	-
8	-	-	-	-	+	+	-	-	-	-	-	-	+	+	-	-
9	-	-	+	+	+++	+++	+	+	-	-	+	+	+++	+++	+	+
10	-	-	IR	IR	-	-	-	-	-	-	IR	IR	-	-	-	-
11	IR	IR	IR	IR	+?	+?	+?	<i>IR</i>	IR	IR	IR	IR	+	+	IR	IR
12	IR	IR	IR	IR	IR	IR	-	-	IR	IR	IR	IR	IR	IR	-	-
13	IR	IR	IR	IR	IR	IR	-	-	IR	IR	IR	IR	-	-	IR	IR
14	-	-	IR	IR	-	-	-	-	-	-	-	-	+?	<i>IR</i>	-	-
15	-	-	-	-	+?	+?	-	-	-	-	-	-	+	+	+?	+?
16	IR	IR	-	-	+	+	IR	IR	-	-	-	-	+	+	-	-
17	IR	IR	-	-	-	-	-	-	IR	IR	-	-	IR	+?	-	-
18	IR	IR	-	-	-	-	-	-	IR	IR	-	-	-	-	IR	+?
19	IR	IR	-	-	+/?	+/?	+?	+?	-	-	-	-	++	++	IR	+
20	-	-	IR	IR	+	+	+	+	-	-	+?	<i>IR</i>	++	++	+	+
21	-	-	-	-	+	+	IR	IR	-	-	-	-	+/?	+/?	IR	IR
22	IR	IR	IR	IR	IR	IR	-	-	IR	IR	IR	IR	IR	IR	-	-
Irr/All ratio	10/3	11/2	10/2	10/2	4/10	4/10	2/5	3/4	8/1	8/1	8/3	9/2	4/12	4/12	8/4	4/8
	First reading								Second reading							
	Total number of reactions				46				Total number of reactions				48			
	Irritative reactions visual				26				Irritative reactions visual				28			
	Irritative reactions HD-OCT				28				Irritative reaction HD-OCT				25			
	Allergic reactions visual				20				Allergic reactions visual				20			
	Allergic reactions HD-OCT				18				Allergic reactions HD-OCT				23			
	Misclassification +? → IR ^a				2				Misclassification +? → IR ^a				2			
	Misclassification IR → +? ^b				0				Misclassification IR → +? ^b				5			

^a Italic misclassified lesions: visual (+?) → HD-OCT (IR)

^b Bold misclassified lesions: visual (IR) → HD-OCT (+?)

directions and dimensions, allowing visualization of cells in the epidermis, papillary dermis and superficial reticular dermis in their micro-architectural context at up to 570 μm in depth. A detailed description of the HD-OCT has been published [4–7].

Relevant RCM features [3] for diagnosis of ACD and ICD were studied on the EF HD-OCT images (Table 3). This set of morphological features described for RCM is very similar to those described for HD-OCT [7]. In contrast to RCM, the in-built HD-OCT slice (CS) mode identifies the exact 3-D position of the selected EF image.

High-Definition Optical Coherence Tomography was performed on the patch test, control patch test and unaffected clinically normal looking paralesional skin. The generated images were analyzed with respect to the following criteria: visualization of individual cells in the stratum corneum, stratum granulosum, stratum spinosum and morphology of dermo-epidermal junction, papillary dermis and superficial reticular dermis.

Epidermal thickness (ET) measurements have been performed using the measurement toolbar of the Skintell[®] program. Each voxel of the 3-Dimensional HD-OCT image

Table 3 Relevant RCM features for diagnosis of ACD and ICD respectively

RCM feature	ACD		ICD	
	2d°	4d°	2d°	4d°
Superficial epidermal changes				
Stratum corneum disruption/ detached corneocytes	(-/+)	(-/+)	(+++)	(++)
Parakeratosis	(+/-)	(+/-)	(+++)	(++)
Spongiosis ^a of SG	(++)	(+++)	(++)	(+)
Spongiosis of SS	(++)	(+++)	(++)	(+)
Exocytosis SG	(++)	(+++)	(++)	(+++)
Exocytosis SS	(++)	(+++)	(++)	(+++)
Vesicle formation ^b SG	(++)	(+++)	(+)	(+)
Vesicle formation SS	(++)	(+++)	(+)	(+)
Necrosis	(+)	(+)	(+++)	(+++)
Increased epidermal thickness	(-/+)	(+)	(++)	(+++)
Basal layer brightness	(-/+)	(-/+)	(++)	(+)
Blood vessel dilatation	(+)	(++)	(+)	(+)
Superficial dermal inflammatory infiltrate ^c	(+)	(++)	(+)	(++)

Parakeratotic changes are seen as highly reflective polygonal cells at the level of SC with a central dark or bright nucleus

Intradermal necrosis seen as circumscribed dark spaces with irregular borders with detached KC

d° days after patch test placing

^a Depending on the severity of the clinical reaction: mild, moderate or marked spongiosis

^b Vesicle formation may be microfocal, diffuse or widespread

^c Inflammatory infiltrate either as aggregates or as single cells

is defined by a unique set of x , y , z coordinates. These coordinates permit to measure with great accuracy in vivo and real time the epidermal thickness three-dimensionally and non-invasively. The ET measurement was assessed at the suprapapillary plate in between adnexal epithelium on three different spots on CS image: one with the visual highest ET, one with the visual lowest ET and one spot with intermediate ET. The measurement procedure is demonstrated in Fig. 1. Measurements were carried out in a climate-controlled room, at 24 °C and 40 % relative humidity. On each of the three individual spots ET measurement has been repeated three times. The average of the nine measurements was calculated corresponding to the mean ET. All HD-OCT measurements were performed by the same investigator.

Statistical analysis

Numerical variables such as epidermal thickness measurements were analyzed using the independent two-

sample t test or Welch's t test for unequal sample size and unequal variances. The same parametric procedure was used to evaluate the optimization of the visual patch test scoring by HD-OCT. Pearson's correlation coefficient between clinical scores and selected HD-OCT features was determined.

Results

Visual patch test scoring at first and second reading (Table 2)

The overall irritative/allergic reactions ratio was 26/20 at the first visual reading and 28/20 at the second visual reading. The individual irritative/allergic reactions ratio for each transition metal was 10/3(Cr), 10/2(Co), 4/10(Ni) and 2/5(Pd) at first reading and 8/1(Cr), 8/3(Co), 4/12(Ni) and 8/4(Pd) at second reading.

Normal looking skin assessed by HD-OCT (Fig. 1)

Cross-sectional and EF HD-OCT features of normal skin were similar as those described previously [4]. In the CS mode stratum corneum was defined as a hyporeflexive "band" limited by two thin hyperreflective lines. The upper one corresponded with the entrance signal and the lower one corresponded with an interference signal due to the difference in refraction indices between stratum corneum and stratum granulosum. If stratum corneum was very thin both hyperreflective lines merged to become one thicker line. The epidermis was seen as a more reflective homogeneous band below the stratum corneum. The sub-basal membrane area appeared as a dark zone in between the basal cell layer and the higher reflective papillary dermis. On EF mode the corneocytes and the grainy cells were identified. The typical honeycomb structure of the stratum spinosum cells could be noticed. The basal cells around the dark dermal papillae could be observed. Adnexal structures and blood vessels were differentiated from other dermal microstructures.

HD-OCT features of allergic contact dermatitis (Figs. 2, 3, 4, 5)

Cross-sectional and EF HD-OCT features of ACD were similar as those described previously for acute spongiotic dermatitis [7]. Darker areas relative to the surrounding epithelium of the stratum spinosum corresponding to spongiosis were seen on EF HD-OCT images. In these areas larger intercellular spaces between round to polygonal mildly reflective keratinocytes were observed. On CS HD-OCT images an increase in epidermal thickness but

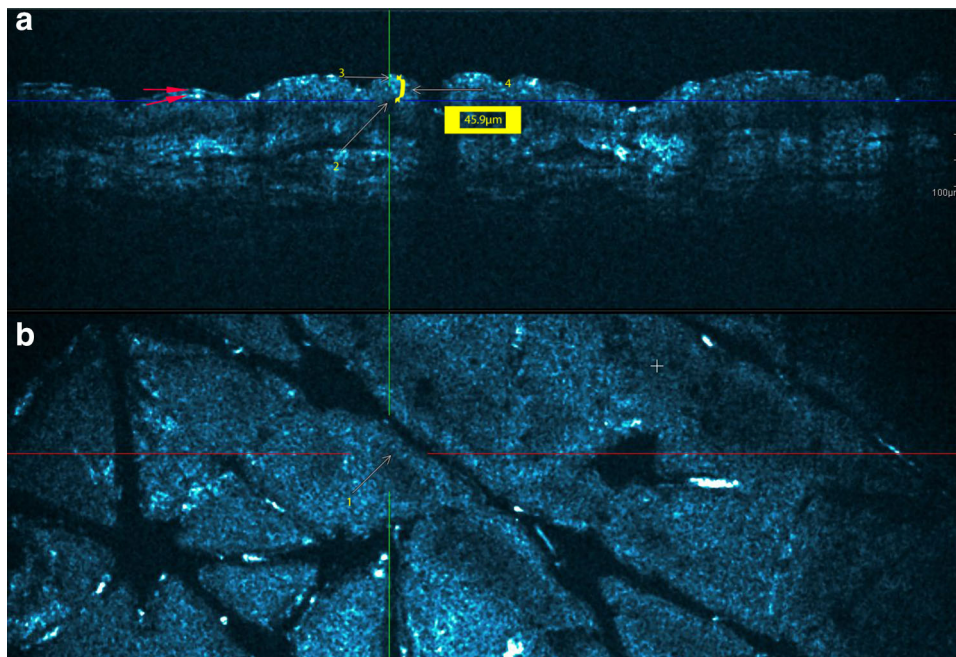


Fig. 1 Epidermal thickness (ET) measurement procedure by High-Definition Optical Coherence Tomography 3-D imaging: **(a)** Cross-sectional (CS) mode and **(b)** En-face (EF) mode. Each voxel of the 3-D HD-OCT imaging is defined by a unique set of x , y , z coordinates. (1) First the basal cell layer at the suprapapillary plate will be searched for in the EF mode at the cross of *green* and *red* line (yellow arrow “1”). (2) The exact position of the basal cell layer in the CS mode can be determined at the cross of *green* and *blue* line (yellow arrow “2”). (3) Stratum corneum is defined as a hyporeflective

“band” limited by two *thin hyperreflective lines* (red arrows). The *upper* one corresponds with the entrance signal and the *lower* one corresponds with an interference signal due to the difference in refraction indices between stratum corneum and stratum granulosum. If stratum corneum is *thin* both *hyperreflective lines* merge to become one *thicker line* (yellow arrow “3”). (4) Epidermal thickness is then measured between this *thicker line* and the *blue line* by using the measurement toolbar of the Skintell(R) program (yellow arrow “4” pointing to the yellow accolade)

decrease in reflectivity could be observed. Smaller but higher reflective cells, single or in small aggregates were observed at the level of the stratum spinosum in both EF and CS modes. Intercellular edema stretching apart keratinocytes sometimes resulted in the formation of intraepidermal vesicles. This vesicle formation secondary to focal areas of spongiosis could be noticed on both EF and CS modes. These vesicles presented as dark round to polylobulated areas. In case of severe allergic reactions macrovesicles were observed. Cells could be observed in- and outside these cavities.

Edema in the dermis appeared to lower dermal reflectivity. In severe forms no dermal reflectivity remained. In moderate forms, dilated signal free cavities in the dermis corresponding to blood vessels were noticed. In these forms the dermal perivascular inflammation was marked with small highly reflective cells.

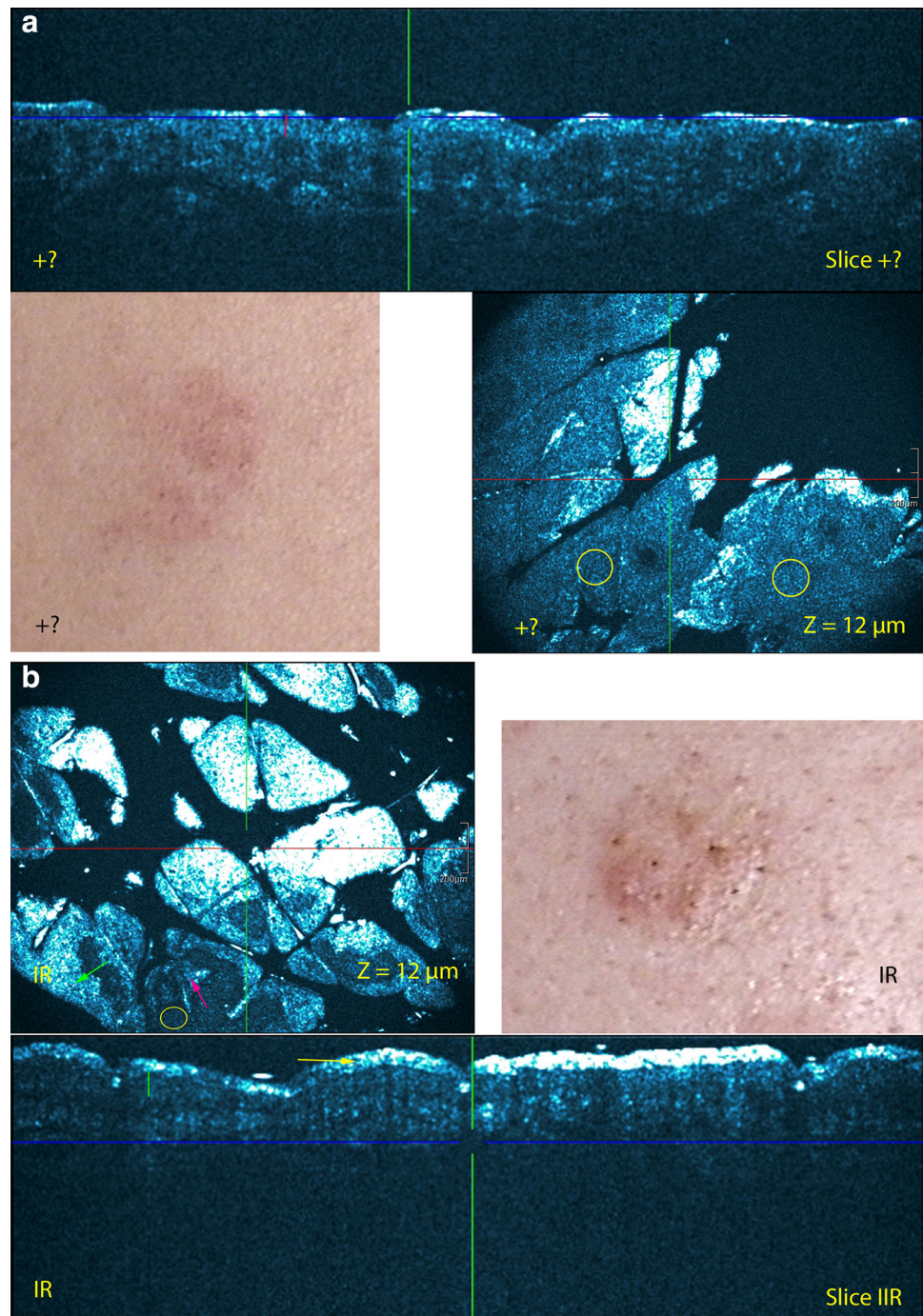
HD-OCT features of irritant contact dermatitis (Figs. 2, 3)

A significant increase in epidermal thickness (ET) was found in ICD by CS HD-OCT measuring the thickness of the suprapapillary ET.

This increase was significant compared to doubtful positive (+?) ACD and normal skin at the first reading ($p < 0.001$) and second reading ($p < 0.001$) (Welch’s t test, unequal sample sizes and unequal variances). The ET of normal skin of the back was $47.38 \mu\text{m}$ [$\pm 1.07 \mu\text{m}$; 95 % confidence interval (CI)]. In ICD at first reading, the ET increased with $12.57 \mu\text{m}$ ($\pm 2.48 \mu\text{m}$; 95 % CI) compared to $2.51 \mu\text{m}$ increase ($\pm 3.71 \mu\text{m}$; 95 % CI) in doubtful positive (+?) ACD. At second reading, the ET increases in ICD measured $9.87 \mu\text{m}$ ($\pm 2.33 \mu\text{m}$; 95 % CI) and in doubtful positive (+?) ACD $2.89 \mu\text{m}$ ($\pm 2.56 \mu\text{m}$; 95 % CI).

Superficial epidermal morphological and cellular changes were also found in ICD compared to ACD. These changes could be observed in combined (3-D) CS and EF HD-OCT mode. The changes consisted of (1) significant parakeratosis whereby parakeratotic cells appeared as bright highly reflective round to oval nucleated structures centrally located within stratum corneum, (2) stratum corneum disruption with disrupted corneocytes presented as single or sheets of multiple discrete mildly bright cellular structures larger than normal keratinocytes without identifiable nucleus, (3) necrotic keratinocytes presented on HD-OCT as polygonal structures brighter and larger than surrounding keratinocytes.

Fig. 2 Visual patch test scoring graded and corresponding HD-OCT images 48 h after patch application: (a) +? for Ni(II) and (b) IR for Cr(VI). On the corresponding CS (slice) HD-OCT imaging a difference in epidermal thickness above the suprapapillary plate is observed between both conditions (*green versus red vertical line*). On the IR CS HD-OCT image a disruption of the stratum corneum is noticed (*yellow arrow*). On the IR EF image a parakeratosis (*green arrow*) is noticed as well as several necrotic keratinocyte (*magenta arrow*). On EF images of both conditions spongiosis is present (*yellow circles*). Z-values = depth in μm of the en-face image

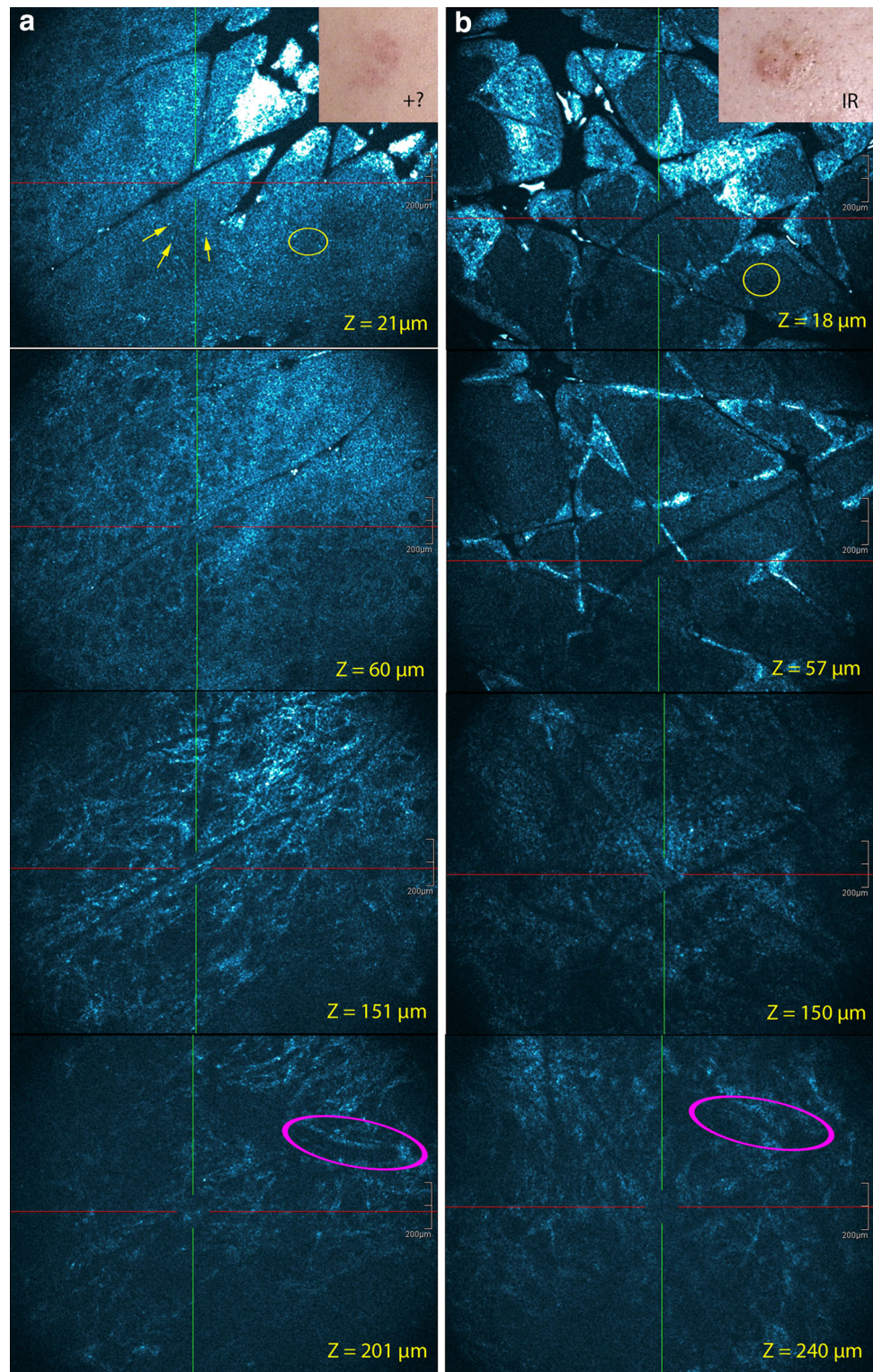


HD-OCT features: correlation with clinical/visual severity. Grading of inflammation by HD-OCT

Two groups of features describing inflammation could be determined. The first group consisted of features linked to water accumulation: spongiosis, intraepidermal vesicle formation and dermal edema. The second group consisted of features related to cellular infiltrate: exocytosis and

superficial dermal inflammatory infiltrate associated with blood vessel dilatation. The degree of epidermal changes related to water accumulation correlated well with clinical severity scores. The degree of dermal edema also correlated with clinical severity scores. The more severe the dermal edema, the less the blood vessel dilatation and superficial dermal inflammatory infiltrate could be perceived Figs. 4, 5.

Fig. 3 Visual patch test scoring graded and corresponding EF HD-OCT images 48 h after patch application: (a) +? for Ni(II) and (b) IR for Cr(VI). Pronounced spongiosis of stratum granulosum and stratum spinosum is observed in both reaction types (yellow circles). No vesicle formation is noticed. Exocytosis in stratum granulosum and stratum spinosum is more pronounced in the +?-reaction (yellow arrows). Blood vessel dilatation and superficial dermal inflammatory infiltrate is similar in both reaction types (magenta circles). Dermal edema is almost absent because dermal structures are visible at higher Z-values. Z-values = depth in μm of the en-face image

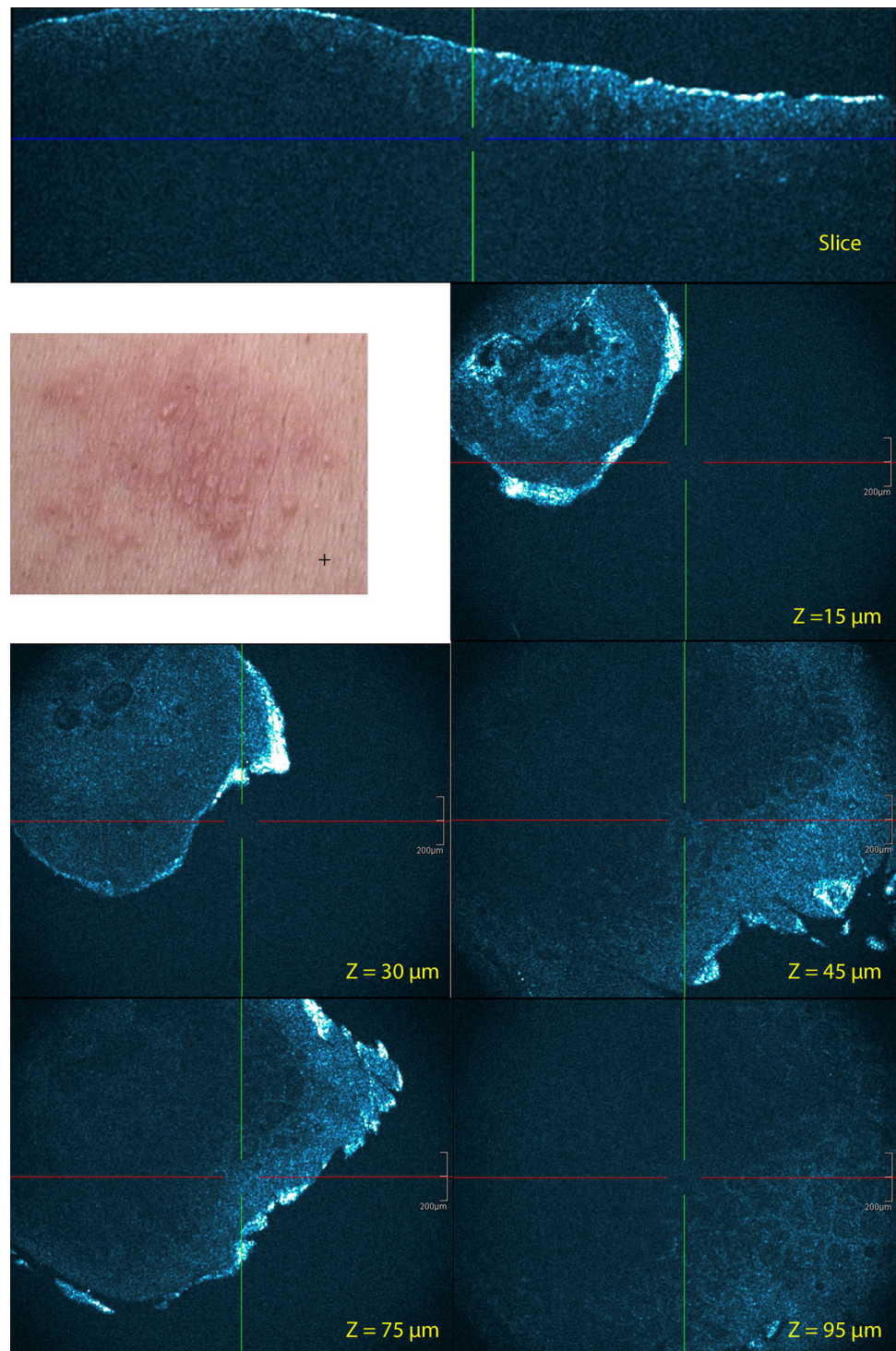


Optimization of the visual patch test scoring by HD-OCT

Table 2 displays the HD-OCT assisted patch test scoring at the first and second reading. The difference was not

significant between visual scoring and HD-OCT evaluation at the first reading. HD-OCT assisted scoring at the second reading, however suggested that five irritant reactions could be re-classified as allergic reactions and two doubtful reactions as irritant reactions. At second reading, reactions

Fig. 4 Visual patch test scoring and corresponding cross-sectional and EF (at different Z-values) HD-OCT images of Ni (II) allergic reaction graded (+) 48 h after patch application. Spongiosis and dermal edema is more severe compared to +? reactions. Intraepidermal visicle formation is observed. Due to dermal edema the visualization of dilated blood vessels and superficial dermal inflammatory infiltrate is lesser. Z-values = depth in μm of the en-face image

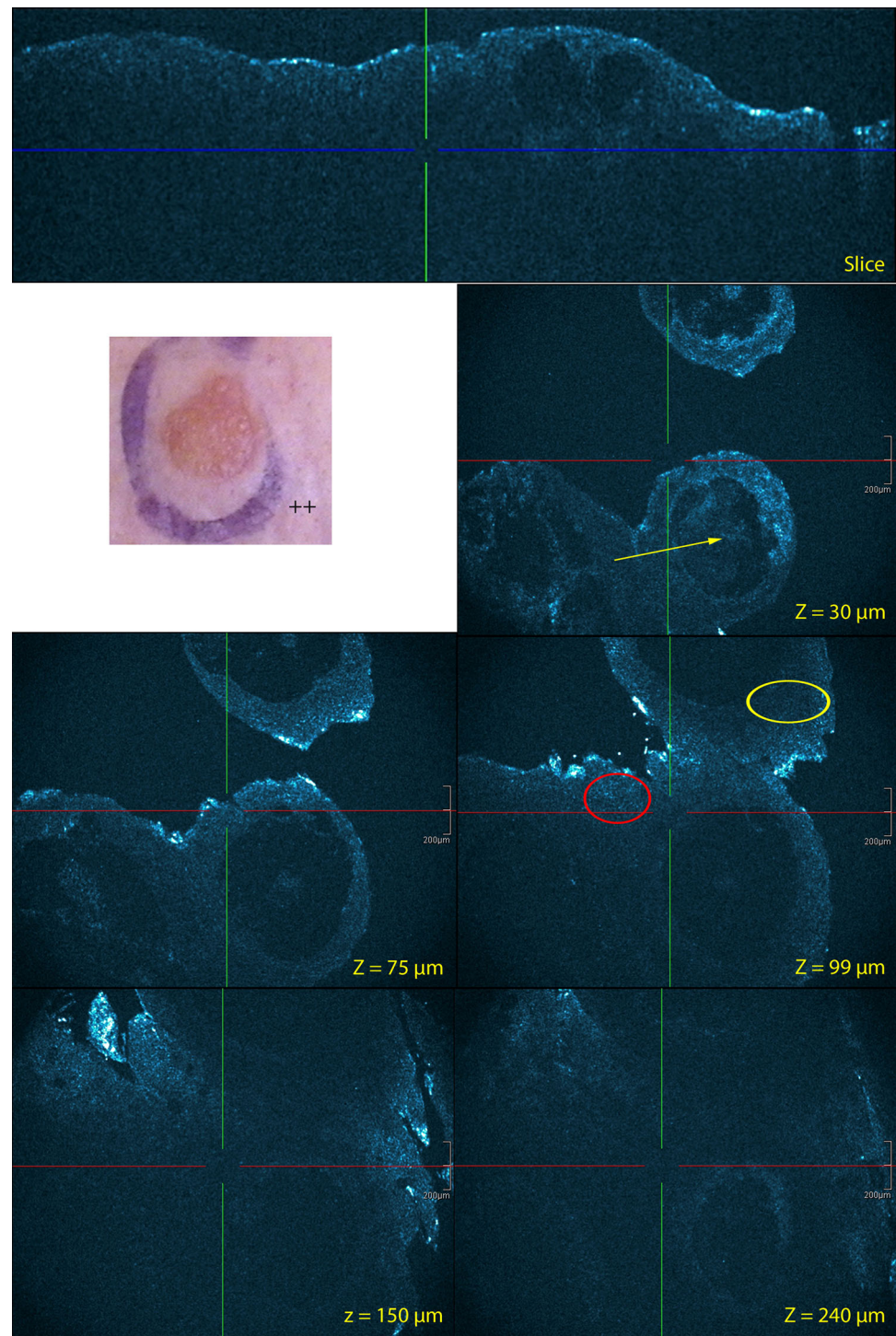


to Pd were clinically most often misclassified as IR instead of doubtful (+?) reactions. Although HD-OCT improved patch test scoring, the difference between visual scoring and HD-OCT assisted scoring was not significant.

Discussion

Differentiating ACD and ICD remains a challenge for clinicians and researchers [26]. A perfect golden standard

Fig. 5 Visual patch test scoring and corresponding CS and EF (at different Z-values) HD-OCT images of Cr(VI) allergic reaction graded (++) 48 h after patch application. The more severe the reaction the more intense the spongiosis (*yellow circle*), vesicle formation (*yellow arrow*) and exocytosis (*red circle*). The more severe the dermal edema the lesser the blood vessel dilatation and superficial dermal inflammatory infiltrate could be perceived. Z-values = depth in μm of the en-face image



permitting differentiation between ACD and ICD does not exist.

Both conditions can be clinically and histologically indistinguishable despite their different pathogenesis [17, 28, 35]. However, two hallmarks of ICD are perturbation of the skin barrier and the epidermal regenerative

hyperproliferation [9, 25]. The predominant ultrastructural change in the epidermis of acute ACD is spongiosis [20].

This paper suggests that the increased epidermal thickness (ET) observed in ICD at first reading is a significant finding useful in differentiating doubtful (+?) ACD from (IR) ICD reactions, following exposure to transition metals.

The increased ET is probably the result of acute regenerative hyperproliferation and disturbed differentiation expressed by parakeratosis. [9, 28] Normal values however, have to be established for reference when direct comparisons are not made in an experimental set-up. The increased ET in the case of irritant reactions has already been documented quantitatively by RCM, conventional OCT and high-frequency ultrasound [1, 23, 34] and qualitatively by immunohistochemical findings [22, 23]. Conventional OCT has been used for the quantification and monitoring of the epidermal and dermal changes in irritant contact dermatitis. Repeated measurements were performed in healthy volunteers after experimental induction of irritant contact dermatitis. A thickening of the epidermis was a constant finding and could be monitored over time [34].

Furthermore, this paper suggests that HD-OCT might be an useful and unique tool for non-invasive in vivo real-time three-dimensional ET measurements. In vivo measurements of ET have been systematically studied in healthy skin using RCM [27] and conventional OCT [12, 13, 18, 32]. Epidermal thickness measurement by RCM is based on at least one vertical (z -axis) stack of 21 EF images collected at 5 μm intervals from the skin surface to a depth of at least 100 μm . The ET corresponds to the distance from the first living cells of the granular layer to the basal cells lying on the top of the dermal papillae. Although some epidermal properties vary by site, the most important limitation is the degree of within-site variation accounting for between 50 and 74 % of the total variation in ET observed. This variation was not due to measurement errors but corresponds to topographical irregularity. This fine-scale variation limits the use of RCM for quantitative studies of epidermis [27].

Conventional optical coherence tomography is an interesting tool for ET measurements. Epidermal thickness is determined in the OCT image (B-scan) on the computer screen using the integrated measure tool (ruler). ET is manually measured in the OCT image from the skin surface reflection (entrance signal) to the first well-demarcated change of reflectance intensity as expressed in a more signal-poor zone [13]. Local positioning of the dermo-epidermal junction between observers was often different. Therefore, a tendency to overestimate the thickness of the epidermis was noticed [12].

High-definition optical coherence tomography offers some practical advantages in ET measurement by providing three micrometer lateral and axial resolution data in the 3D micro-anatomical context [4–7]. EF HD-OCT imaging permits to define with precision both the position of the junction between stratum corneum and granular layer and the position of the basal cell layer in contact with the dermo-epidermal junction. Once both positions have been

determined ET measurement can be performed on CS HD-OCT imaging with higher precision than conventional OCT because of the higher axial resolution of the former (3 μm compared to the 7.5 μm axial resolution of OCT). In a recent paper, HD-OCT measurement of ET in relatively thin lesions appears to create valid results compared with routine histopathology indicating that this method could be well suited to skin monitoring purposes and therapy outcome assessments [14].

A previous study suggested that HD-OCT features of inflammatory skin conditions with epidermal alteration correlate well with dermatopathologic descriptors as defined in RCM [7]. The morphological CS and EF features of acute spongiotic skin conditions imaged by HD-OCT in this study were very similar to those demonstrated for conventional OCT and RCM, respectively [11, 16].

Reflectance confocal microscopy has been used in non-invasive in vivo studies to demonstrate intraepidermal and superficial dermal cellular changes and micro-architectural alterations in ACD and ICD. These features were later confirmed by correlating them with histopathology [16]. Furthermore, the kinetics of ACD and ICD over time, as well as the individual ethnic susceptibility to ICD have also been examined by using RCM [1, 2]. The sensitivity and specificity of a series of 12 RCM criteria to differentiate ACD and ICD in reference to patch testing have been determined [3].

This study demonstrates that the degree of changes in some epidermal HD-OCT features of acute spongiotic skin conditions, such as spongiosis, vesicle formation and exocytosis, correlates well with clinical severity scores confirming the findings of previous histopathological studies and RCM studies [1, 3, 36]. Furthermore, the degree of changes in dermal HD-OCT features correlates also with clinical severity scores. Dermal edema correlates with clinical severity scores which was clearly observed in the CS HD-OCT images. The more severe the dermal edema, the less the blood vessel dilatation and superficial dermal inflammatory infiltrate could be perceived. This inflammatory infiltrate in HD-OCT is very similar in ICD and (+?) or (+) ACD reactions which is in agreement with the literature [21].

The overall irritative/allergic reactions ratio was not significantly different between visual scoring and HD-OCT evaluation. At second reading, reactions to Pd were most often misclassified as IR instead of doubtful (+?) reactions. The morphology of patch test reactions can vary with different metals. Some allergens such as Pd report to show a delay in the development of positive tests [15]. This could probably explain why Pd “behaves” as Cr/Co at first reading with high irritative/allergic reactions ratio and as Ni at second reading with low irritative/allergic reactions

ratio. The allergic reactions to Pd are significantly more present at second reading in this study. At first reading we probably observe indirectly the stimulation of innate immunity by Pd [24]. As the strength of the innate inflammatory response is much higher for Ni than for Pd, the threshold for activation of the adaptive immune system is eventually much higher for Pd [15]. As a result, it takes more time before an allergic reaction to Pd develops.

We found an optimization of the visual patch test scoring by HD-OCT assessment although not significant. A possible explanation could be the experience in visual patch test scoring. Dermatologists not so familiar with patch test reading could be assisted by HD-OCT to increase their diagnostic accuracy. The qualitative and quantitative differences found between the reactions may imply that earlier readings of patch tests may be possible if supplemented by HD-OCT. HD-OCT assisted patch test scoring could be very useful when it is difficult to assess erythema; e.g. when dealing with patients with Fitzpatrick phototype IV or higher [2]. Additional studies are essential to validate these suggestions.

Some important drawbacks need to be mentioned. Firstly an obligate irritant agent has not been included into the test substances because the increased ET in the case of irritant reactions has already been documented previously by other authors [1, 22, 23, 34]. In addition, the use of transition metals was thought to reflect a real-world situation better. Secondly the visual patch test scoring as well as the HD-OCT assisted patch test scoring was performed only by one observer. However, as mentioned in the method section on three different spots in between adnexal epithelium and at the suprapapillary plate, ET measurement has been repeated three times. The average of the nine measurements was calculated corresponding to the mean ET.

In conclusion, HD-OCT may be considered as a unique tool for in vivo, non-invasive and real time three-dimensional epidermal thickness measurements. Epidermal thickness measurements by HD-OCT help to differentiate IR from doubtful (+?) reactions in patch testing and may be performed repeatably in real time. Selected HD-OCT features corresponded well with severity of visual scoring. These features might help to quantify the degree of inflammation in inflammatory skin conditions. HD-OCT might help in optimizing visual patch test scoring in some situations.

Conflict of interest None declared.

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References

1. Astner S, Gonzalez E, Cheung AC, Rius-Diaz F, Mihm MC, Doukas A, Gonzalez S (2005) The kinetics of allergic and irritant contact dermatitis in vivo—a non-invasive evaluation. *J Invest Dermatol* 124:351–359
2. Astner S, Burnett N, Cheung AC, Rius-Diaz F, Doukas AG, Gonzalez S, Gonzalez E (2006) The impact of skin color on the susceptibility to irritant contact dermatitis, a non-invasive evaluation. *J Am Acad Dermatol* 54:458–465
3. Astner S, Ulrich M (2012) Spongiotic dermatitis. In: Hofmann-Wellenhof R, Pellacani G, Malvehy J, Soyer HP (eds) *Reflectance confocal microscopy for skin diseases*, 1st edn. Springer-Verlag, Berlin Heidelberg, pp 381–389
4. Boone M, Jemec GB, Del Marmol V (2012) High-definition optical coherence tomography enables visualization of individual cells in healthy skin, comparison to reflectance confocal microscopy. *Exp Dermatol* 21:740–744
5. Boone M, Norrenberg S, Jemec GB, Del Marmol V (2012) Imaging of basal cell carcinoma by high-definition optical coherence tomography. *Histomorphologic correlation. A pilot study. Br J Dermatol* 167:856–864
6. Boone M, Norrenberg S, Jemec GB, Del Marmol V (2013) Imaging actinic keratosis by high-definition optical coherence tomography. *Histomorphologic correlation. A pilot study. Exp Dermatol* 22:93–97
7. Boone M, Norrenberg S, Jemec GB, Del Marmol V (2013) High-definition optical coherence tomography, adapted algorithmic method for pattern analysis of inflammatory skin diseases. A pilot study. *Arch Derm Res* 305:283–297
8. Ermolli M, Menné C, Pozzi G, Serra MA, Clerici LA (2001) Nickel, cobalt and chromium-induced cytotoxicity and intracellular accumulation in human haca1 keratinocytes. *Toxicology* 159:23–31
9. Fartasch M (1997) Ultrastructure of the epidermal barrier after irritation. *Microsc Res Tech* 37:193–199
10. Frosch JP, John SW (2006) Clinical aspects of irritant contact dermatitis. In: Frosch JP, Menné T, Lepoittevin JP (eds) *contact dermatitis*, 4th edn. Springer-Verlag, Berlin Heidelberg, pp 255–294
11. Gambichler T, Moussa G, Sand M, Sand D, Orlikov A, Altmeyer P, Hoffmann K (2005) Correlation between clinical scoring of allergic patch test reactions and optical coherence tomography. *J Biomed Opt* 10(6):064030
12. Gambichler T, Boms S, Stücker M, Kreuter A, Moussa G, Sand M, Altmeyer P, Hoffmann K (2006) Epidermal thickness assessed by optical coherence tomography and routine histology: preliminary results of method comparison. *J Eur Acad Dermatol Venereol* 20(7):791–795
13. Gambichler T, Matip R, Moussa G, Altmeyer P, Hoffmann K (2006) In vivo data of epidermal thickness evaluated by optical coherence tomography: effects of age, gender, skin type and anatomic site. *J Dermatol Sci* 44:145–152
14. Gambichler T, Valavanis K, Plura I, Georgas D, Kampilafkos, Stücker M (2014) In vivo determination of epidermal thickness using high-definition optical coherence tomography. *Br J Dermatol* 170:737–739
15. Gawkrödger DJ, Lewis FM, Shah M (2000) Contact sensitivity to nickel and other metals in jewelry reactions. *J Am Acad Dermatol* 43:31–36
16. Gonzalez S, Gonzalez E, White WM, Rajadhyaksha M, Anderson RR (1999) Allergic contact dermatitis, correlation of in vivo confocal imaging to routine histology. *J Am Acad Dermatol* 40:708–713

17. Judge MR, Griffiths HA, Basketter DA, White IR, Rycroft RJ, McFadden JP (1996) Variation in response of human skin to irritant challenge. *Contact Dermat* 34:115–117
18. Josse G, George J, Black D (2011) Automatic measurement of epidermal thickness from optical coherence tomography images using a new algorithm. *Skin Res Technol* 17(3):314–319
19. Kornik R, Zug KA (2008) *Nickel Dermat* 19:3–8
20. Komura J, Ofuji S (1980) Ultrastructural studies of allergic contact dermatitis in man. *Arch Dermatol Res* 267:275–282
21. Lachapelle JM, Marot L (2006) Histopathological and immunohistopathological features of irritant and allergic contact dermatitis. In: Frosch JP, Menné T, Lepoittevin JP (eds) *Contact dermatitis*, 4th edn. Springer-Verlag, Berlin Heidelberg, pp 108–116
22. Le TKM, Schalkwijk J, van de Kerkhof PC, van Haelst U, van der Valk PG (1998) A histological and immunohistochemical study on chronic irritant contact dermatitis. *Am J Contact Dermat* 9:23–28
23. Le TKM, van der Valk PG, Schalkwijk J, van de Kerkhof PC (1995) Changes in epidermal proliferation and differentiation in allergic and irritant contact dermatitis reactions. *Br J Dermatol* 133:236–240
24. Martin SF (2012) Contact dermatitis, from pathomechanisms to immunotoxicology. *Exp Dermatol* 21:382–389
25. Medenica M, Rostenberg A Jr (1971) A comparative light and electron microscopic study of primary irritant contact dermatitis and allergic contact dermatitis. *J Invest Dermatol* 56:259–271
26. Nosbaum A, Vocanson M, Rozieres A, Hennino A, Nicolas JF (2009) Allergic and irritant contact dermatitis. *Eur J Dermatol* 19:325–332
27. Robertson K, Rees JL (2010) Variation in epidermal morphology in human skin at different body sites as measured by reflectance confocal microscopy. *Act Derm Venereol* 90:368–373
28. Roediger B, Weninger W (2011) How nickel turns on innate immune cells. *Immunol Cell Biol* 89:1–2
29. Sakanashi NE, Matsumura M, Kikuchi K, Ikeda M, Miura H (2010) A comparative study of allergic contact dermatitis by patch test versus reflectance confocal laser microscopy, with nickel and cobalt. *Eur J Dermatol* 20:705–711
30. Serup J, Staberg B (1987) Ultrasound for assessment of allergic and irritant patch test reactions. *Contact Dermat* 17(2):80–84
31. Swindells K, Burnett N, Rius-Diaz F, Gonzalez E, Mihm MC, Gonzalez S (2004) Reflectance confocal microscopy may differentiate acute allergic and irritant contact dermatitis in vivo. *J Am Acad Dermatol* 50:220–228
32. Tsugita T, Nishijima T, Kitahara T, Takema Y (2013) Positional differences and aging changes in Japanese woman epidermal thickness and corneus thickness determined by OCT (Optical coherence tomography). *Skin Res Technol* 19:242–250
33. Wahlberg JE, Lindberg M (2006) Patch testing. In: Frosch JP, Menné T, Lepoittevin JP (eds) *Contact dermatitis*, 4th edn. Springer-Verlag, Berlin Heidelberg, pp 366–390
34. Welzel J, Bruhns M, Wolff HH (2003) Optical coherence tomography in contact dermatitis and psoriasis. *Arch Dermatol Res* 295:50–55
35. Willis CM, Young E, Brandon DR, Wilkinson JD (1986) Immunopathological findings in human allergic and irritant contact dermatitis. *Br J Dermatol* 115:305–316
36. Willis CM, Stephens CJ, Wilkinson JD (1989) Epidermal damage induced by irritants in man, a light and electron microscopic study. *J Invest Dermatol* 93:695–699