

New species and records of *Chapsa* (Graphidaceae) in China

Ming-Zhu Dou^{1*}, Min Li^{1*}, Ze-Feng Jia¹

¹ College of Life Sciences, Liaocheng University, Liaocheng 252059, China

Corresponding author: Ze-Feng Jia (zfa2008@163.com)

Academic editor: Thorsten Lumbsch | Received 5 October 2021 | Accepted 27 November 2021 | Published 10 December 2021

Citation: Dou M-Z, Li M, Jia Z-F (2021) New species and records of *Chapsa* (Graphidaceae) in China MycoKeys 85: 73–85. <https://doi.org/10.3897/mycokeys.85.76040>

Abstract

We studied the genus *Chapsa* in China based on morphological characteristics, chemical traits and molecular phylogenetic analysis. One species new to science (*C. murioelongata* M.Z. Dou & M. Li) and two records new to China were found (*C. wolseleyana* Weerakoon, Lumbsch & Lücking and *C. niveocarpa* Mangold). *Chapsa murioelongata* **sp. nov.** is characterised by its lobed thalline margin, orange discs with white pruina, clear hymenium, and submuriform and long ascospores. *Chapsa wolseleyana* was recombined into *Astrochapsa* based on phenotypic traits. Sequences of this species are for the first time reported here and phylogenetic analyses of three loci (mtSSU, ITS and nuLSU) supported the position of this species within *Chapsa*. A key for the *Chapsa* species known in China is provided.

Keywords

Ascomycota, lichenized fungi, phylogeny, taxonomy

Introduction

The lichen genus *Chapsa* (Graphidaceae) was first established by Massalongo (1860) with *C. indica* as the type species. This genus was ignored for a long time until 2006, when Frisch re-established *Chapsa*, based on the *Chroodiscus*-type apothecia, presence of periphysoids and *Chapsa*-type paraphyses. Frisch (2006) also provided a detailed description and delimitation of the genus *Chapsa*, which was widely recognised by

* These two authors contributed equally to this work.

subsequent researchers (Mangold 2008; Frisch and Kalb 2009; Rivas Plata et al. 2011; Sipman et al. 2012; Xu et al. 2016). The genus *Chapsa* was considered to be monophyletic in the beginning (Frisch 2006) but with further research, it was suspected to be polyphyletic (Mangold 2008; Papong et al. 2010). Subsequently, seven genera, *Astrochapsa* Parnmen, Lücking & Lumbsch, *Crutarndina* Parnmen, Lücking & Lumbsch, *Gintarasia* Kraichak, Lücking & Lumbsch, *Pseudochapsa* Parnmen, Lücking & Lumbsch, *Pseudotopeliopsis* Parnmen, Lücking & Lumbsch, *Myriochapsa* M. Cáceres, Lücking & Lumbsch and *Nitidochapsa* Parnmen, Lücking & Lumbsch were separated from *Chapsa*, based on a combination of molecular evidence, phenotypic and chemical characteristics (Parnmen et al. 2012, 2013; Kraichak et al. 2013).

Although China is rich in lichenised fungal species (Wei 2020), there are few studies and reports on the genus *Chapsa*. More than 60 species of *Chapsa* have been reported in the world, of which only three, *C. indica* A. Massal, *C. mirabilis* A. (Zahlbr.) Lücking and *C. leprocarpa* (Nyl.) Frisch, have so far been found in China (Rivas Plata et al. 2010; Xu et al. 2016; Jia and Lücking 2017; Kalb and Kalb 2017; Wijayawardene et al. 2017; de Lima et al. 2019).

During the study of *Chapsa* A. Massal. in southern China, one species, *C. murioelongata* was found new to science, and two species, *C. niveocarpa* Mangold and *C. wolseleyana* Weerakoon, Lumbsch & Lücking were found new to China. In our study, 26 sequences were newly generated from freshly collected specimens.

Materials and methods

Morphological and chemical analyses

The specimens were collected from southern China and deposited in the Fungarium, College of Life Sciences, Liaocheng University, China (LCUF). Morphological and anatomical characters of thalli and apothecia were examined and photographed under an Olympus SZX16 dissecting microscope and an Olympus BX53 compound microscope. The lichen secondary metabolites were detected and identified by thin-layer chromatography using solvent C (Orange et al. 2010; Jia and Wei 2016).

DNA extraction, PCR sequencing and phylogenetic analysis

Genomic DNA was extracted from ascomata using the Hi-DNA-secure Plant Kit (Tiangen, Beijing, China) according to the manufacturer's protocol. The nuLSU, ITS and mtSSU regions were amplified using the primer pair AL2R/LR6 (Mangold et al. 2008, Vilgalys and Hester 1990), ITS1F/ITS4 (Gardes and Bruns 1993, White et al. 1990) and mrSSU1/mrSSU3R (Zoller et al. 1999), respectively. The PCR amplification progress followed Dou et al. (2018) and the PCR products were sequenced by Biosune Inc. (Shanghai). The newly generated sequences were submitted to GenBank (Table1).

Table 1. Information for the sequences used in this study. Newly generated sequences are shown in bold.

Species	Specimen No.	Locality	ITS	nuLSU	mtSSU
<i>Pseudochapsa phlyctidioides</i>	Lumbsch 20500d	Fiji	–	JX465301	JX421005
<i>Pseudochapsa dilatata</i>	Luecking 32101	Venezuela	–	JX421446	JX420981
<i>Pseudochapsa esslingeri</i>	Caceres s.n.	Brazil	–	–	JX420983
<i>Pseudochapsa esslingeri</i>	Caceres 6006a	Brazil	–	–	JX420984
<i>Pseudochapsa esslingeri</i>	Rivas Plata 107C (F)	Peru	–	–	JX420985
<i>Pseudochapsa esslingeri</i>	Rivas Plata 809a (F)	Peru	–	–	JX420986
<i>Chapsa alborosella</i>	Luecking 31238a	Brazil	–	JX421439	JX420972
<i>Chapsa alborosella</i>	Luecking 25587	Guatemala	–	JX421440	JX420973
<i>Chapsa soredicarpa</i>	Luecking 31200	Brazil	–	JX421462	JX421011
<i>Chapsa soredicarpa</i>	Luecking 31240	Brazil	–	JX421463	JX421012
<i>Chapsa sublilacina</i>	Luecking RLD056	Mexico	–	HQ639624	HQ639600
<i>Chapsa thalotrema</i>	Luecking 32019	Venezuela	–	JX465319	JX421013
<i>Chapsa indica</i>	Parnmen018486(RAMK)	Thailand	–	JX465295	JX465280
<i>Chapsa leprocarpa</i>	GZ19531	China, Guizhou	MW009079	MW007981	MW010276
<i>Chapsa leprocarpa</i>	GZ19537	China, Guizhou	MW009077	MW007984	MW010278
<i>Chapsa leprocarpa</i>	GZ19536	China, Guizhou	MW009080	MW007982	MW010274
<i>Chapsa niveocarpa</i>	HN19508	China, Hainan	MW009076	MW010272	–
<i>Chapsa niveocarpa</i>	Lumbsch_19125k2(F) & Mangold (F)	Australia, Queensland	–	–	EU675274
<i>Chapsa niveocarpa</i>	Lumbsch 19151p & Mangold (F)	Australia, Queensland	–	FJ708487	EU075567
<i>Chapsa patens</i>	FJ19131	China, Fujian	MT995055	MW007979	MW010275
<i>Chapsa patens</i>	FJ19049	China, Fujian	MW007918	MW007980	–
<i>Chapsa woleseleyana</i>	FJ19158	China, Fujian	MW009078	MW010273	MW010277
<i>Chapsa woleseleyana</i>	FJ19148	China, Fujian	MW009106	MW010270	MW010279
<i>Chapsa murioelongata</i>	HN19222	China, Hainan	MW009102	MW010271	–
<i>Chapsa murioelongata</i>	HN19682	China, Hainan	MW009103	MW010269	–
<i>Chapsa pulchra</i>	CHAPUL19129t	Australia	–	KC020261	KC020255
<i>Astrochapsa meridensis</i>	Luecking 17770 (F)	Costa Rica	–	EU075655	EU075610
<i>Astrochapsa mastersonii</i>	Lumbsch 20500f	Fiji	–	–	JX420996
<i>Astrochapsa zablbruckneri</i>	Papong 6516	Thailand	–	JX421467	–
<i>Astrochapsa astroidea</i>	Lumbsch 19166n & Mangold(F)	Australia, Queensland	–	EU075614	EU075566
<i>Astrochapsa astroidea</i>	Lumbsch 19750a	Thailand	–	JX421441	JX420974
<i>Astrochapsa astroidea</i>	Papong 6004	Thailand	–	JX421442	JX420975
<i>Astrochapsa astroidea</i>	Luecking 24006	Thailand	–	JX421443	JX420977
<i>Astrochapsa astroidea</i>	Luecking 24008	Thailand	–	JX421444	JX420978
<i>Astrochapsa astroidea</i>	Luecking 24011	Thailand	–	JX421445	JX465278
<i>Chroodiscus coccineus</i>	Herb. R. Luecking 2000	Costa Rica	–	AF465441	–

Multi-locus (ITS, mtSSU and nuLSU) phylogenetic analysis was performed. The combined analysis included 70 sequences (Table 1) representing 18 in-group taxa and one out-group taxon. As many species as possible of *Chapsa* s. lat. were contained in our data matrix including the taxa that were similar in morphology or sequence to the new species and the two records. We blasted sequences of the three species in GenBank and selected sequence-similar taxa on a pre-determined cut-off.

The alignment was undertaken by applying MAFFT 7 with the option of L-INS-I (Katoh and Standley 2013). The three single-locus alignments were concatenated in PhyloSuite v1.2.2 (Zhang et al. 2020). The concatenated data matrix comprised 3188 nucleotide sites (nuLSU 1405 bp, ITS 647 bp and mtSSU 1136 bp). In order to check the consistency between the three loci, incongruence length difference test (ILD Test) was carried out using PAUP. The P value of ILD Test was 0.65 (>0.5), so the three loci were

rielongata (100%, 1.00), *C. wolseleyana* (99%, 1.00) and *C. niveocarpa* (91%, 1.00). *Chapsa murioelongata* is sister to the clade consisting of *C. wolseleyana* and *C. patens* (Nyl.) Frisch. *Chapsa niveocarpa* HN19508 and *C. niveocarpa* Lumbsch form a well-supported clade and are sisters to *C. leprocarpa*.

Taxonomy

New species

Chapsa murioelongata M.Z. Dou & M. Li, sp. nov.

Fungal Names: FN 570754

Figure 2

Etymology. The specific epithet *murioelongata* refers to the elongate, muriform ascospores.

Type. CHINA. Hainan Province: Ledong County, Jianfengling National Forest Park, 18°42'39"N, 108°52'37"E, alt. 760 m, on bark, 09 Dec 2019, Y. H. Ju HN19222 (LCUF: holotype: HN19222; GenBank MW009102 for ITS and MW010271 for LSU).

Description. THALLUS corticolous, crustose, olive-grey, surface dull, smooth to uneven, ecorticate. APOTHECIA erumpent, dispersed or two to four aggregated, rounded, 1–3 mm diam.; THALLINE MARGIN lobed with white felt-like inner surface, lobes

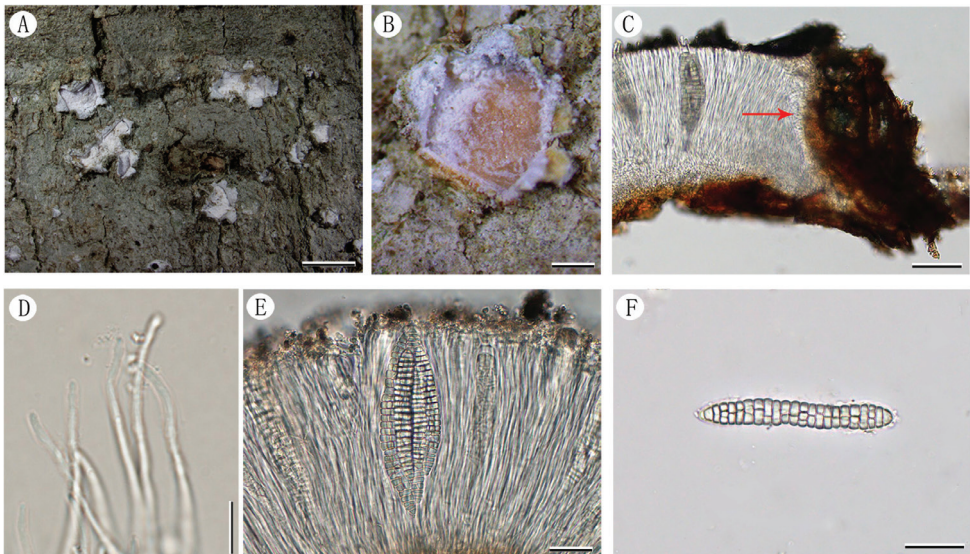


Figure 2. *Chapsa murioelongata* (LCUF HN19222) **A** habit of thallus with apothecia at different developmental stages **B** apothecium (the pruina of the disc partly scraped off) **C** section of apothecium with paraphyses (direction of arrow) **D** paraphyses **E** an ascus containing six ascospores **F** ascospore. Scale bars: 3 mm (**A**); 0.5 mm (**B**); 50 μ m (**C**); 8 μ m (**D**); 30 μ m (**E**); 25 μ m (**F**).

strongly backward curved; DISC flesh-coloured, covered by thick, white pruina. EXCIPIE 80–105 μm wide laterally, dark brown; EPIHYMENIUM 20–40 μm high, with coarse greyish granules; HYMENIUM clear, 110–170 μm high, non-amyloid; HYPOTHECIUM colourless, 10–30 μm high; PARAPHYSES simple, tips unbranched; PERIPHYSOIDES present, 5–30 μm long. ASCI 4–6 (8)-spored, clavate, 100–120 \times 35–50 μm ; ASCOSPORES hyaline, bacillary with rounded to subacute ends, submuriform with 20–25 transverse septa and 0–2 longitudinal septa per segment, 75–105 \times 9.5–16 μm , non-halonate, I-
PYCNIDIA not observed.

Chemistry. Thallus K-, C-, PD-; no compounds detectable by TLC.

Ecology and distribution. On the bark in semi-exposed forest of Hainan Province.

Additional specimens examined. CHINA. Hainan Province: Changjiang County, Bawangling Nature Reserve, Yajia Scenic Area, 10°04'54"N, 109°07'04"E, alt. 810 m, on bark, 08 Dec 2019, Y. H. Ju HN19167 (LCUF); CHINA. Hainan Province: Lingshui County, Diaoluo Mountain, 18°43'35"N, 109°52'02"E, alt. 900 m, on bark, 14 Dec 2019, M. Li HN19682 (LCUF) (GenBank MW009103 for ITS and MW010269 for LSU).

Note. *Chapsa murioelongata* is characterised by its olive-grey thallus; white pruinose discs; distinct periphysoids; clear hymenium; 4–8-spored asci; submuriform ascospores with 20–25 transverse septa and 0–2 longitudinal septa per segment. *Chapsa microspora* Kalb, *C. asteliae* (Kantvilas & Vězda) Mangold, *Astrochapsa elongata* Poengs. & Lumbsch and *C. patens* are morphologically similar to the new species. *Chapsa microspora* can be distinguished from *C. murioelongata* by the smaller apothecia (0.6–1.2 mm diam.), transversely septate and smaller ascospores (7–9 \times 4 μm) (Lumbsch et al. 2011). *Chapsa asteliae* differs in amyloid and shorter ascospores (30–80 μm) (Kantvilas and Vězda 2000; Mangold 2008). *Astrochapsa elongata* differs from *C. murioelongata* in having shorter ascospores (40–65 μm) and less longitudinal septa per segment (0–1) (Poengsungnoen et al. 2019). *Chapsa patens* differs from *C. murioelongata* chiefly in the single-spored asci and broader ascospores (22–35 μm) (Frisch et al. 2006).

Blast searches of nuLSU sequences indicate *Chapsa murioelongata* has close affinities with *C. patens* (98.36% identity), *C. woleleyana* (95.63% identity), *C. leprocarpa* (91.97% identity) and *C. indica* (90.81% identity), so all these species were included in the phylogenetic analyses. *Chapsa murioelongata* was well separated from any other species in the tree and strongly supported as the monophyletic (PP = 1; ML = 100%).

New records

***Chapsa woleleyana* Weerakoon, Lumbsch & Lücking, in Weerakoon, Rivas Plata, Lumbsch & Lücking, Lichenologist 44(3): 377 (2012)**

Figure 3

Astrochapsa woleleyana (Weerakoon, Lumbsch & Lücking) Parmen, Lücking & Lumbsch, in Parmen et al., PLoS ONE 7(12): 10 (2012)

Description. THALLUS crustose, corticolous, grey-brown, surface dull to slightly shiny, uneven, fissured. APOTHECIA erumpent, dispersed, sometimes two or three fused, most-

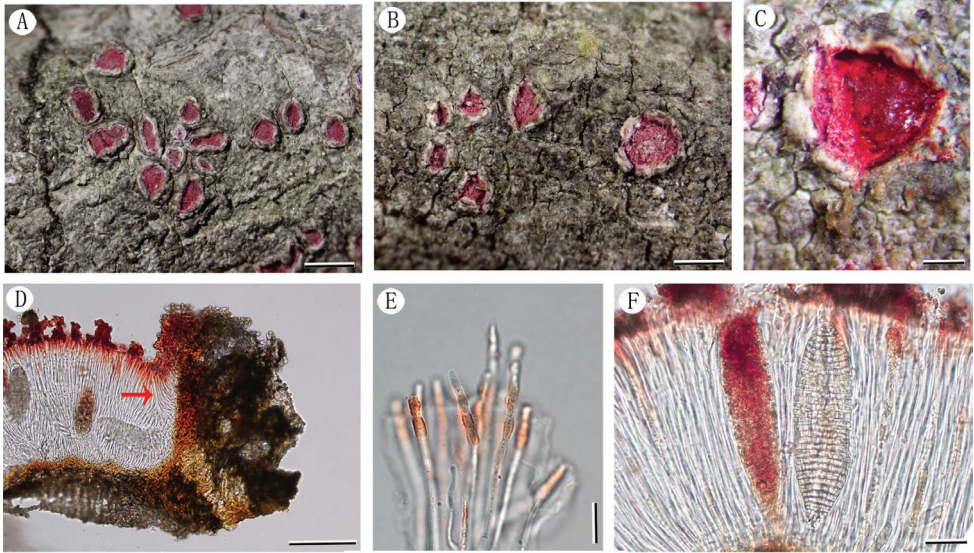


Figure 3. *Chapsa wolseleyana* (LCUF FJ19148-b) **A** habit of thallus with apothecia **B** apothecia at different developmental stages **C** apothecium (part of pruinose scraped off) **D** section of apothecium with periphysoids (direction of arrow) **E** paraphyses **F** young and mature ascospores. Scale bars: 1.5 mm (**A**); 1 mm (**B**); 0.25 mm (**C**); 120 μ m (**D**); 10 μ m (**E**); 25 μ m (**F**).

ly rounded to seldom slightly angular, 0.7–1.2 mm diam.; THALLINE MARGIN raised to lobulate, lobes erected to recurved, inner part brown, covered with rose-red or white pruina; DISC exposed, rose-red, covered with thick, rose-red pruina. EXCIPLE fused, cupular, laterally 180–250 μ m wide, yellowish-brown to brown; EPIHYMENIUM rose-red with granules, 20–50 μ m high, K+ green; HYMENIUM 140–230 μ m high, clear, colourless, non-amyloid; HYPOTHECIUM indistinct; PARAPHYSES septate, tips rose-red and moniliform with oval or rectangular cells; PERIPHYSOIDES present, 50–100 μ m long. ASCI clavate, 1-spored, 110–135 \times 35–50 μ m; ASCOSPORES densely muriform, oblong-ellipsoid, with hemispherical to roundish ends, 105–130 \times 30–45 μ m, first reddish, becoming hyaline to slightly olive-brown at maturity, I-. PYCNIDIA not observed.

Chemistry. No substances detected by TLC but apothecial disc with pigment producing K+ yellow-green efflux, suggesting presence of isohypocrelline.

Ecology and distribution. Growing on bark exposed to wind and high light intensity in montane forests. Worldwide distribution: Sri Lanka (Weerakoon et al. 2012) and newly reported for China.

Selected specimens examined. CHINA. Fujian Province: Quanzhou City, Jiuxian Mountain, Reflecting Pool, 25°42'57"N, 118°07'14"E, alt. 1540 m, on bark, 5 Jul 2019, F.Y. Liu FJ19148-b (LCUF) (GenBank MW009106 for ITS, MW010270 for LSU and MW010279 for SSU); CHINA. Fujian Province: Quanzhou City, Jiuxian Mountain, Natural Observation Path, 25°42'44"N, 118°07'17"E, alt. 1460 m, on bark, 25 Jul 2019, F.Y. Liu FJ19158 (LCUF) (GenBank MW009078 for ITS, MW010273 for LSU and MW010277 for SSU). CHINA. Fujian Province: Quanzhou City, Jiuxian Mountain,

Reflecting Pool, 25°42'57"N, 118°07'14"E, alt. 1540 m, on bark, 25 Jul 2019, F.Y. Liu FJ19127-2, same locality, FJ19128-2, FJ19141-2 (LCUF).

Note. *Chapsa walseleyana* is characterised by its grey-brown, uneven thallus, apothecia with raised to lobed thalline margin, rose-red discs with similar coloured pruina, rose-red epihymenium and paraphyses tips, distinct periphysoids, 1-spored asci, muriform ascospores, red when young and hyaline to olive-brown when old. Only a few species of *Chapsa* have pigmented discs and among them *C. rubropulveracea* Hale ex Mangold, Lücking & Lumbsch is morphologically most similar to *C. walseleyana*, but its thallus is farinose and its ascospores are 8 per ascus, smaller (15–20 × 5–6 µm) and transversely septate (Lumbsch et al. 2011).

Chapsa walseleyana was transferred to *Astrochapsa*, based on a phenotype-based analysis (not molecular phylogeny) (Parnmen et al. 2012). However, our phylogenetic analysis shows that this species belongs in *Chapsa*, rather than *Astrochapsa*. *Chapsa walseleyana* was associated phylogenetically with a strongly-supported clade (100/1) with *C. patens*, but with sufficient distance to be considered a distinct species. In addition, the latter differs from *C. walseleyana* in having larger pale brown apothecia (up to 2 mm diam.) with white pruina, unpigmented epihymenium and unpigmented paraphyses adspersed with fine greyish to brownish granules, hyaline ascospores (Frisch et al. 2006; Joshi et al. 2012; Joshi et al. 2018).

***Chapsa niveocarpa* Mangold in Mangold, Elix & Lumbsch, Flora of Australia, 57:654 (2009)**

Figure 4

Description. THALLUS corticolous, crustose, pale grayish-green surface dull and fluctuating along the bark. APOTHECIA erumpent, solitary to fused, angular rounded to slightly elongate, 0.5–1.8 × 0.5–1.2 mm; THALLINE MARGIN split and recurved, inside with thick white pruina; DISC exposed, yellowish-brown, covered by white pruina. EXCIPLE laterally 12–75 µm wide, dark brown; EPIHYMENIUM 10–20 µm high; HYMENIUM 120–200 µm high, grey-brown, interspersed by granules, non-amyloid; HYPOTHECIUM indistinct; PARAPHYSES unbranched; tips distinctly thickened; PERIPHYSOIDES present, but obscured by granular inclusions. ASCI 1-spored, clavate, 120–140 × 27–36 µm; ASCOSPORES densely muriform, with thick halo at both ends, oblong, hyaline, 115–135 × 25–34 µm, I-. PYCNIDIA not observed.

Chemistry. Thallus K-, C-, PD-; no compounds detectable by TLC.

Ecology and distribution. Growing on tree bark in tropical rainforests in altitudes ranging from 500 to 1100 m. Australia, Queensland (Mangold 2008); newly reported for China.

Selected specimens examined. CHINA. Hainan Province: Wuzhishan City, Wuzhishan Nature Reserve, 18°54'13"N, 109°41'04"E, alt. 870 m, on bark, 12 Dec 2019,

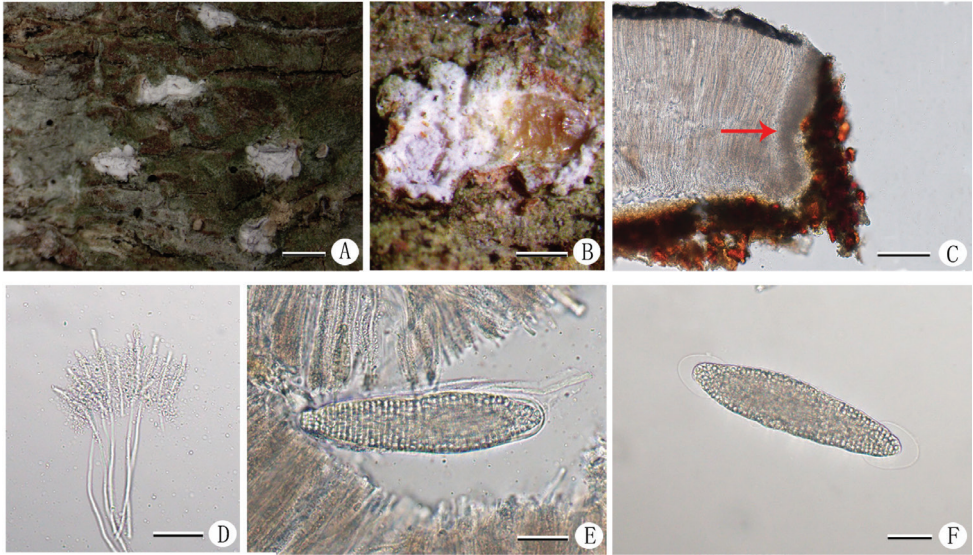


Figure 4. *Chapsa niveocarpa* (LCUF HN19508) **A** habit of thallus with apothecia **B** apothecium (part of pruinose scraped off) **C** section of apothecium with periphysoids (direction of arrow) **D** paraphyses with hyaline granules **E** ascus **F** ascospore with halo. Scale bars: 1 mm (**A**); 0.5 mm (**B**); 50 μ m (**C**); 25 μ m (**D**); 30 μ m (**E**); 25 μ m (**F**).

M. Li HN19508 (LCUF) (GenBank MW009076 for ITS and MW010272 for LSU); CHINA. Hainan Province: Wuzhishan City, Wuzhishan Nature Reserve, 18°53'13"N, 109°41'04"E, alt. 1020 m, on bark, 12 Dec 2019, M. Li HN19530 (LCUF); CHINA. Hainan Province: Wuzhishan City, Wuzhishan Nature Reserve, 18°54'13"N, 109°41'04"E, alt. 870 m, on bark, 12 Dec 2019, M. Li HN19499 (LCUF); CHINA. Hainan Province: Lingshui County, Diaoluo Mountain, 18°43'35"N, 109°52'02"E, alt. 900 m, on bark, 14 Dec 2019, M. Li HN19687 (LCUF); CHINA. Hainan Province: Lingshui County, Diaoluo Mountain, 18°43'35"N, 109°52'02"E, alt. 900 m, on bark, 14 Dec 2019, M. Li HN19679 (LCUF).

Note. *Chapsa niveocarpa* is characterised by its crustose, pale greyish-green thallus; rounded to elongate apothecia, yellowish-brown discs with white pruinose, obscured periphysoids, interspersed hymenium, 1-spored (rare 2-spored) ascus and muriform and hyaline ascospores with halo. *Chapsa niveocarpa* is morphologically similar and phylogenetically related to *C. leprocarpa*, and both species occur on bark in tropical forests (Frisch 2006; Mangold 2008; Parmen et al. 2012). *Chapsa leprocarpa* differs from *C. niveocarpa* in having a lower hymenium (100–130 μ m) and smaller ascospores (up to 111 μ m long) (Frisch 2006). The specimen (HN19508) we collected in China is allocated phylogenetically to a strongly-supported (1/91) clade with *C. niveocarpa*. The collections cited above are the first reports for China.

Key to *Chapsa* in China

- 1 Disc with red pruina; ascospores 1/ascus, muriform, 105–135 × 30–50 µm.
..... *C. wolseleyana* **2**
- Disc with white pruina **2**
- 2 Ascospores transversely septate; ascospores 4–8/ascus, 50–110 × 6–12 µm....
..... *C. indica* **3**
- Ascospores (sub)muriform **3**
- 3 Hamathecium inspersed; ascospores 1/ascus, 80–190 × 20–50 µm
..... **4**
- Hamathecium clear **5**
- 4 Ascospores 1/ascus, 80–190 × 20–50 µm *C. niveocarpa* **5**
- Ascospores 8/ascus, 40–50 × 11–15 µm *C. mirabilis* **5**
- 5 Asci 4–6 (8)-spored; ascospores oblong to cylindrical with rounded to subacute ends, submuriform with 20–25 transverse septa and 0–2 longitudinal septa per segment, 75–105 × 9.5–16 µm *C. murioelongata* **5**
- Asci 4-spored; ascospores oblong to slightly ellipsoid, with roundish ends, 60–130 × 20–40 µm *C. leprocarpa* **5**

Acknowledgements

This study was supported by the National Natural Science Foundation of China (31750001), Doctoral Initiation Fund of Liaocheng University (318051813) and Research Fund of Liaocheng University (318012011).

References

- Dou MZ, Wu XH, Li M, Zhao X, Jia ZF (2018) *Gyalecta caudiospora* sp. nov. from China. Mycotaxon 133(4): 721–727. <https://doi.org/10.5248/133.721>
- Frisch A (2006) Contribution towards a new systematic of the lichen family Thelotremataceae. I. The lichen family Thelotremataceae in Africa. Bibliotheca Lichenologica 92: 1–370.
- Frisch A, Kalb K (2009) *Chapsa* species (Thelotremataceae) from Brazil. Bibliotheca Lichenologica 99: 133–142.
- Gardes M, Bruns TD (1993) ITS primers with enhanced specificity for Basidiomycetes – application to the identification of mycorrhiza and rusts. Molecular Ecology 2: 113–118. <https://doi.org/10.1111/j.1365-294X.1993.tb00005.x>
- Jia ZF, Lücking R (2017) Resolving the genus *Phaeographina* Müll. Arg. in China. Mycokeys 21(1): 13–32.
- Jia ZF, Wei JC (2016) Flora Lichenum Sinicorum, vol. 13. – Ostropales (I) – Graphidaceae 1. Science Press, Beijing, 21–22.
- Joshi S, Upreti DK, Nayaka S (2012) The lichen genus *Chapsa* (Graphidaceae) in India. Mycotaxon 120: 23–24, 27, 31, 32. <https://doi.org/10.5248/120.23>

- Joshi S, Upreti DK, Divakar PK, Lumbsch HT, Lücking R (2018) A re-evaluation of thelotremoid Graphidaceae (lichenized Ascomycota: Ostropales) in India. *The Lichenologist* 50(6): 634–639. <https://doi.org/10.1017/S0024282918000439>
- Kalb J, Kalb K (2017) New lichen species from Thailand, new combinations and new additions to the Thai lichen biota. *Phytotaxa* 332 (2): 141–156. <https://doi.org/10.11646/phytotaxa.332.2.2>
- Kantvilas G, Vězda A (2000) Studies on the lichen family Thelotremataceae in Tasmania. The genus *Chroodiscus* and its relatives. *Lichenologist* 32(4): 325–357. <https://doi.org/10.1006/lich.2000.0274>
- Katoh K, Standley DM (2013) MAFFT multiple sequence alignment software version 7: improvements in performance and usability. *Molecular Biology and Evolution* 30: 772–780. <https://doi.org/10.1093/molbev/mst010>
- Kraichak E, Parnmen S, Lücking R, Lumbsch HT (2013) *Gintarasia* and *Xalocoa*, two new genera to accommodate temperate to subtropical species in the predominantly tropical Graphidaceae (Ostropales, Ascomycota). *Australian Systematic Botany* 26: 466–474. <https://doi.org/10.1071/SB13038>
- Lanfear R, Frandsen PB, Wright AM, Senfeld T, Calcott B (2017) PartitionFinder 2: new methods for selecting partitioned models of evolution for molecular and morphological phylogenetic analyses. *Molecular Biology and Evolution* 34(3): 772–773. <https://doi.org/10.1093/molbev/msw260>
- Lima EL, Maia LC, Martins MCB, da Silva NL, Lücking R, da Silva Cáceres ME (2019) Five new species of Graphidaceae from the Brazilian Northeast, with notes on *Diorygma alagoense*. *The Bryologist* 122(3): 414–422. <https://doi.org/10.1639/0007-2745-122.3.414>
- Lumbsch HT, Ahti T, Altermann S, De Paz GA, Aptroot A, Arup U et al. (2011) One hundred new species of lichenized fungi: a signature of undiscovered global diversity. *Phytotaxa* 18: 36–40. <https://doi.org/10.11646/phytotaxa.18.1.1>
- Mangold A (2008) Taxonomic studies on members of Thelotrematoid Ostropales (Lichenized Ascomycota) in Australia. PhD Thesis, University of Duisburg, Essen.
- Massalongo AB (1860) Esame comparativo di alcune genere di licheni. *Atti dell'Istituto Veneto Scienze* 5: 247–276.
- Orange A, James PW, White FJ (2010) *Microchemical methods for the identification of lichens*. British Lichen Society, London, 101 pp.
- Papong K, Boonpragob K, Mangold A, Divakar PK, Lumbsch HT (2010) Thelotremoid lichen species recently described from Thailand: a re-evaluation. *The Lichenologist* 42(2): 131–137. <https://doi.org/10.1017/S0024282909990405>
- Parnmen S, Lücking R, Lumbsch HT (2012) Phylogenetic classification at generic level in the absence of distinct phylogenetic patterns of phenotypical variation: a case study in Graphidaceae (Ascomycota). *PLoS ONE* 7(12): e51392. <https://doi.org/10.1371/journal.pone.0051392>
- Parnmen S, Cáceres MES, Lücking R, Lumbsch HT (2013) *Myriochapsa* and *Nitidochapsa*, two new genera in Graphidaceae (Ascomycota: Ostropales) for chroodiscoid species in the *Ocellularia* clade. *The Bryologist* 116(2): 127–133. <https://doi.org/10.1639/0007-2745-116.2.127>
- Poengsungnoen V, Buaruang K, Vongshewarat K, Sangvichien K, Boonpragob K, Mongkolsuk P, Lumbsch HT (2019) Three new crustose lichens from Thailand. *The Bryologist* 122(3): 451–456. <https://doi.org/10.1639/0007-2745-122.3.451>

- Rivas Plata R, Lücking R, Lumbsch HT (2011) A new classification for the family Graphidaceae (Ascomycota: Lecanoromycetes: Ostropales). *Fungal Diversity* 52(1): 107–121. <https://doi.org/10.1007/s13225-011-0135-8>
- Rivas Plata R, Lücking R, Sipman HJM, Mangold A, Klab K, Lumbsch HT (2010) A worldwide key to the thelotremoid Graphidaceae, excluding the Ocellularia-Myriotrema-Stegobolus clade. *The Lichenologist* 42(2): e139185.
- Ronquist F, Teslenko M, van der Mark P, Ayres DL, Darling A, Höhna S, Larget B, Liu L, Suchard MA, Huelsenbeck JP (2012) MrBayes 3.2: efficient Bayesian phylogenetic inference and model choice across a large model space. *Systematic Biology* 61: 539–542. <https://doi.org/10.1093/sysbio/sys029>
- Sipman HJM, Lücking R, Aptroot A, et al. (2012) A first assessment of the Ticolichen biodiversity inventory in Costa Rica and adjacent areas: the thelotremoid Graphidaceae (Ascomycota: Ostropales). *Phytotaxa* 55: 1–214. <https://doi.org/10.11646/phytotaxa.55.1.1>
- Stamatakis A (2014) RAxML version 8: A tool for phylogenetic analysis and post-analysis of large phylogenies. *Bioinformatics* 30(9): 1312–1313. <https://doi.org/10.1093/bioinformatics/btu033>
- Vilgalys R, Hester M (1990) Rapid genetic identification and mapping of enzymatically amplified ribosomal DNA from several *cryptococcus* species. *Journal of Bacteriology* 172(8): 4238–4246. <https://doi.org/10.1128/JB.172.8.4238-4246.1990>
- Weerakoon G, Rivas Plata, Lumbsch HT, Lücking R (2012) Three new species of *Chapsa* (lichenized Ascomycota: Ostropales: Graphidaceae) from tropical Asia. *The Lichenologist* 44(3): 373–379. <https://doi.org/10.1017/S0024282911000892>
- Wei JC (2020) The enumeration of lichenized fungi in China. China Forestry Publishing House, Beijing, 606 pp.
- White T, Bruns T, Lee S, Taylor J (1990) Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: Innis MA, Gelfand DH, Sninsky JJ, White TJ (Eds) *PCR Protocols: a guide to methods and applications*. Academic Press, New York, 315–322. <https://doi.org/10.1016/B978-0-12-372180-8.50042-1>
- Wijayawardene NN, Hyde KD, Rajeshkumar KC, Hawksworth DL, Madrid H, Kirk PM, Braun U, Singh RV, Crous PW, Kukwa M, Lücking R, Kurtzman CP, Yurkov A, Haelewaters D, Aptroot A, Lumbsch HT, Timdal E, Ertz D, Etayo J, Phillips AJL, Groenewald JZ, Papizadeh M, Selbmann L, Dayarathne MC, Weerakoon G, Jones EBG, Suetrong S, Tian Q, Castañeda-Ruiz RF, Bahkali AH, Pang KL, Tanaka K, Dai DQ, Sakayaroj J, Hujislová M, Lombard L, Shenoy BD, Suija A, Maharachchikumbura SSN, Thambugala KM, Wanasinghe DN, Sharma BO, Gaikwad S, Pandit P, Zucconi L, Onofri S, Egidí E, Raja HA, Kodsueb R, Cáceres MES, Pérez-Ortega S, Fiuza PO, Monteiro JS, Vasilyeva LN, Shivas RG, Prieto M, Wedin M, Olariaga I, Lateef AA, Agrawal Y, Fazeli SAS, Amoozegar MA, Zhao GZ, Pfliegler WP, Sharma G, Oset M, Abdel-Wahab MA, Takamatsu S, Bensch K, de Silva NI, Kesel AD, Karunarathna A, Boonmee S, Pfister DH, Lu YZ, Luo ZL, Boonyuen N, Daranagama DA, Senanayake IC, Jayasiri SC, Samarakoon MC, Zeng XY, Doilom M, Quijada L, Rampadarath S, Heredia G, Dissanayake AJ, Jayawardana RS, Perera RH, Tang LZ, Phukhamsakda C, Hernández-Restrepo M, Ma X, Tibpromma S, Gusmao LFP,

- Weerahewa D, Karunarathna SC (2017) Notes for genera: Ascomycota. Fungal Diversity 86: e96. <https://doi.org/10.1007/s13225-017-0386-0>
- Xu LL, Wu QH, Wang QD, Jia ZF (2016) *Chapsa* (Graphidaceae, Ostropales), A lichen genus new to China. Journal of Tropical and Subtropical Botany 24(5): 495–498.
- Zhang D, Gao FL, Jakovli I, Zou H, Wang GT (2020). Phylsuite: an integrated and scalable desktop platform for streamlined molecular sequence data management and evolutionary phylogenetics studies. Molecular Ecology Resources 20(1): 348–355. <https://doi.org/10.1111/1755-0998.13096>
- Zoller S, Scheidegger C, Sperisen C (1999) PCR primers for the amplification of mitochondrial small subunit ribosomal DNA of lichen-forming ascomycetes. Lichenologist 31: 511–516. <https://doi.org/10.1006/lich.1999.0220>