REVIEW

Dissecting bipolar disorder complexity through epigenomic approach

B Ludwig and Y Dwivedi

In recent years, numerous studies of gene regulation mechanisms have emerged in neuroscience. Epigenetic modifications, described as heritable but reversible changes, include DNA methylation, DNA hydroxymethylation, histone modifications and noncoding RNAs. The pathogenesis of psychiatric disorders, such as bipolar disorder, may be ascribed to a complex gene–environment interaction ($G \times E$) model, linking the genome, environmental factors and epigenetic marks. Both the high complexity and the high heritability of bipolar disorder make it a compelling candidate for neurobiological analyses beyond DNA sequencing. Questions that are being raised in this review are the precise phenotype of the disorder in question, and also the trait versus state debate and how these concepts are being implemented in a variety of study designs.

Molecular Psychiatry (2016) 21, 1490-1498; doi:10.1038/mp.2016.123; published online 2 August 2016

INTRODUCTION

Bipolar disorder (BD) is a chronic, disabling condition that is characterized by recurrent depressive, manic, mixed or hypomanic episodes. The majority of BD patients are either diagnosed with BD I (manic and/or mixed episodes) or BD II (depressed and/or hypomanic episodes). The National Institute of Health conducted an international population-based study and found a 1.4% lifetime prevalence of BD, 2 reflecting the global burden of this chronic disorder.

Today's scientific consensus on the pathogenesis of affective disorders might be best described as genotype-dependent environmental influences on risk for an individual to be affected, although a precise model for the molecular mechanisms behind its interactions has not been established yet. The conventional gene–environment interaction (GxE) model does not specifically include epigenetic modifications, but they might represent the underlying mechanisms of the statistical interaction;³ the importance of epigenetic regulations for complex traits disorders has been acknowledged.⁴

Finding a common definition for epigenetics has been a challenge for the scientific community for some time. In general, the term epigenetics is referred to as long-standing changes in gene expression that are regulated via transcriptional, post-transcriptional, translational and/or post-translational mechanisms (such as DNA methylation, DNA hydroxymethylation, histone modifications and noncoding RNAs for example), which does not entail any change in DNA sequence. The changes beyond DNA sequence can be maintained during the cell cycle (Table 1). A consensus about the question of a transgenerational transmission of epigenetic marks has not been reached yet,⁵ but recent evidence supports this presumption for microRNA⁶ and for DNA methylation.^{7,8} The idea of heritable but reversible changes leads us to the question of how stable these epigenetic changes actually are. It is strongly debated whether these modifications in post-

mortem brain tissue represent a stable disease-associated state or only snapshots of different moments in the course of time. 9 On the one hand, studies suggest that there are subtle differences in the epigenetic landscape of monozygotic twins, taken into account for phenotypical differences such as discordant diagnoses due to non-shared exposures. 10,11 This would set the methylation status further on the stable 'trait' end of the discussion. On the other hand, psychiatric drugs have been shown to influence methylation levels^{12,13} and there is evidence suggesting that different mood episodes are associated with distinct epigenetic alterations 14,15—which ultimately suggests that epigenetic modifications reflect a state rather than a trait. An evolutionary perspective proposes that only specific histone modifications might be stable and conserved between species, depending on factors such as, for example, clustered transcription factor binding sites or high GC content. 16 The majority of publications identify DNA methylation as a long-term and relatively stable epigenetic mark, in contrast to histone modifications that are thought to confer short-term and relatively flexible silencing of gene expression. 17,18

Naturally, there are different approaches to address a complex disorder such as BD; the major part of scientists would approach it as a homogeneous research concept, searching for a trait marker for BD. ^{19–21} Others might divide it into two entities following the DSM-V classification system and compare BD I and BD II. ^{22,23} Scientists searching for a 'state marker' will mainly focus on the diverging emotional states involving manic, mixed and depressive episodes. Last but not least, research facilities with access to both bipolar and schizophrenic patient samples might focus on the psychosis-involving part of the disorder, combining both patient samples to one. ^{24–26}

Numerous studies about gene regulation mechanisms have been emerging in neuroscience over the past few years. In this review, we summarize the current knowledge about epigenetics in BD patients, including DNA methylation, DNA hydroxymethylation

UAB Mood Disorder Program, Department of Psychiatry and Behavioral Neurobiology, University of Alabama at Birmingham, Birmingham, AL, USA. Correspondence: Professor Y Dwivedi, UAB Mood Disorder Program, Department of Psychiatry and Behavioral Neurobiology, University of Alabama at Birmingham, SC711 Sparks Center, 1720 2nd Avenue South, Birmingham, AL 35294, USA.

31

41

22

23

19

97

20

26

21

72

101

100

Findings Gene Reference Human post-mortem brain studies DNA methylation 87.88.90 Promoter hypermethylation Reelin 32 Promoter hypermethylation SLC6A4 97 Promoter hypermethylation in exon 1 to intron 1 HCG9 98 Promoter hypomethylation MB-COMT 100 Higher promoter methylation level in psychotic BD compared with non-psychotic BD DTNBP1 21 Promoter hypomethylation in mixed sample of BD and SZ ST6GALNAC1 Histone modification 58 Increased global histone H3 acetylation 75 Increased H3K4 trimethylation in BD and MDD Peripheral blood DNA methylation 32 Promoter hypermethylation

DNA methylation Promoter hypomethylation Higher promoter methylation level in psychotic BD compared with non-psychotic BD

Abbreviations: BD, bipolar disorder; MDD, major depressive disorder; SZ, schizophrenia.

and histone modifications (Table 1). Questions that are being raised in this review are the precise phenotype of BD and its implications with psychosis and suicide, and also the trait versus state debate and how these concepts are being implemented in a variety of study designs. Furthermore, we briefly describe therapeutic interventions targeting epigenetic mechanisms—from well-established therapeutic drugs to potential agents.

Summary of epigenetic findings in bipolar disorder

Higher promoter methylation in BD II compared with BD I

Promoter hypermethylation in a mixed sample of BD and SZ

Promoter hypomethylation in BD compared with MDD

Promoter hypermethylation from exon 1 to intron 1

EPIGENETIC MECHANISMS AND FINDINGS IN BD

Promoter hypomethylation

Promoter hypomethylation

Histone modification

Saliva

Promoter hypermethylation in BD II

Differential promoter methylation in BD I

Promoter hypomethylation in exon 11

Increased levels of acetylated histone 3

DNA methylation

Table 1.

Methylation at the 5-position of cytosine is a well-studied epigenetic modification. DNA methylation occurs by transfer of a methyl group from S-adenosyl methionine to cytosine residues in the dinucleotide sequence CpG; the majority of the 28 million CpG dinucleotides are methylated. In most of the cases, the level of methylation correlates with the extent of gene inactivation. Early studies focused on CpG islands (CGI) representing DNA regions of a high CpG density, which were shown to be low or unmethylated. Recent work has shown that DNA methylation can also directly silence genes with non-CGI promoters.²⁷ In fact, regions with relatively low CpG density appear to be equally important. For example, in certain disease conditions, differentially methylated regions occur more frequently within CGI shores (< 2kb flanking CGIs) or shelves (< 2kb flanking outwards from a CpG shore) representing relatively low CpG density that flank traditional CGIs compared with within CGIs themselves.^{28–30}

DNA methylation as a trait marker. Twin studies of monozygotic twins regarding discordant phenotypes such as mental health disorder are an excellent design to test genome-wide methods and reduce the noise to a minimum. A study with a sample of monozygotic twins discordant for BD performed methylationsensitive representational difference analysis and found four regions of the genome to have significant alterations in methylation pattern.³¹ Bisulfite sequencing revealed a significantly lower methylation level of peptidylprolyl isomerase E-like (PPIEL) gene in the affected twin. The authors suggest that this gene might be involved in specific neuronal functions, such as dopamine transmission or neuroendocrine systems, as it is highly expressed in the pituitary gland and the substantia nigra.³¹

SICGAA

BDNF exon I

BDNF exon I

BDNF exon I

GPR24, ZNF659

H3K9/K14ac

MB-COMT

DTNBP1

BDNF exon 3 and 5

PPIEL

HCG9

KCNQ3

5HTR1A

Another twin study identified promoter hypermethylation of serotonin transporter gene SLC6A4, which seemed to be associated with BD in a sample of two monozygotic twins.³² This finding led the authors to a case-control study in post-mortem brain samples, which found associations in the serotonergic system, connecting the S/S genotype of HTTLPR to a promoter hypermethylation of *SLC6A4* and finally to the downregulated mRNA on the level of gene expression.³² No additional information about medication or current state of the disorder was given, which certainly constitutes a limitation to the findings.

A very recently published study assessing a sample of only BD patients draws the attention to the potassium voltage-gated channel gene KCNQ3,²⁰ which has been the focus of genetic linkage studies, 33,34 QTL-mapping 35 and genome-wide association studies³⁶ in BD patients. KCNQ3 has been shown to be involved in the regulation of neuronal excitability by preventing hyperexcitability of neurons, thus increasing their responsiveness.³⁷ The CpG region of exon 11 upstream of the KCNQ3 gene showed significantly lower methylation levels and correspondingly higher mRNA expression in BD patients compared with healthy controls.20

More recently, Perroud *et al.*³⁸ hypothesized that the *5-HT_{3A}R* (5-hydroxytryptamine 3A) methylation status would mediate the effect of childhood trauma on adult psychopathology such as BD, borderline personality disorder and attention deficit/hyperactivity disorder. Among various CpG sites, they were able to associate the methylation status of CpG2 III with the number of previous mood episodes, previous suicide attempts and the polymorphism in single-nucleotide polymorphism rs1062613, regardless of the underlying diagnosis. The authors admit that their results were mainly driven by borderline personality disorder subjects because of the high percentage of childhood maltreatment in this group of patients. The significance of these results for BD patients is certainly limited and not applicable to the question of a trait or state marker debate.

DNA methylation as a state marker. The lack of reliable peripheral blood markers in the psychiatric clinical setting accounts for the continuous attempts to correlate brain tissue findings with expression patterns in the peripheral blood. Peripheral DNA methylation is also an excellent source for studies searching for a trait marker in BD subjects, as peripheral sources can be accessed multiple times. Because of the role played by brain-derived neurotrophic factor (BDNF) in synaptic plasticity and stress response³⁹ and its intraindividual correlation between peripheral and post-mortem brain tissues,⁴⁰ it is a prominent candidate for methylation studies in affective disorder For example, Dell'Osso et al.41 investigated alterations of BDNF exon I promoter methylation levels in blood samples of BD and major depressive disorder (MDD) patients. They compared BD I patients with BD II patients and found higher methylation levels in BD II patients. When MDD patients were compared with healthy subjects, they again found significantly higher levels of BDNF exon I promoter methylation in the MDD patients. Finally, they stratified for the mood state and showed that patients in a depressed state had higher methylation levels compared with patients in a manic/ mixed state. D'Addario's²² group was able to replicate Dell'Osso's results regarding a hypermethylation of the BDNF exon I promoter in BD II patients but not in BD I patients. Additional information regarding the pharmacological treatment was assessed too, demonstrating that the combination of antidepressant agents and mood stabilizers establish higher methylation levels than mood stabilizers only. Carlberg et al.'s 19 findings were also in line with those previously published and suggested that the altered methylation pattern between MDD and BD patients might be associated with the pharmacological treatment, rather than with the diagnosis itself—which again points to the conception of a flexible state marker. BDNF exon IV promoter methylation has been of interest in various psychiatric illnesses including MDD;⁴² however, no data in BD patients are available to date, although preclinical data suggest an involvement of mood stabilizers in BDNF promoter IV methylation. Lithium treatment to hippocampal neurons induced BDNF gene expression, which was accompanied by BDNF exon IV hypomethylation.4

Another innovative approach focused on the global methylation levels and oxidative damage to the DNA of BD patients (measured through 5-methylcytosine and 8-OHdg (8-hydroxy-2'-deoxyguanosine) levels). Compared with the control group, BD patients had higher DNA levels of 8-OHdG.¹⁵ A higher number of previous manic episodes could also predict higher 8-OHdG levels, which was interpreted as a marker for disease progression, although these levels could not be predicted by the number of depressive episodes. No difference in global methylation could be demonstrated.¹⁵ One of the strengths of this study was that all subjects were in the wash-out phase of their medication and the phenotype effect of the disease was not confounded by the effect of the medication. Although the study might not have been designed to find a state marker, higher 8-OHdG levels in patients with previous manic episodes point in this direction.

DNA hydroxymethylation

Hydroxymethylation (5hmc) is another mechanism of epigenetic modification. Similar to cytosine methylation (5mc), hydroxymethylation adds a hydroxymethyl group to the C5 position. Hydroxymethylation is highly enriched at promoters and in intragenic regions but is largely absent from non-gene ⁴ Among various tissues, hydroxymethylation is highly abundant in the brain. 45,46 How hydroxymethylation is associated with cytosine methylation and consequent gene expression is not completely understood yet, but there seems to be a dynamic balance between cytosine methylation and hydroxymethylation.⁴ In this regard, several hypotheses have been proposed. For example, hydroxymethylation is implicated in demethylation⁴⁸ and might be a necessary intermediary for methylation—allowing the promoter sites to be prepared for activation. ⁴⁹ Another model places hydroxymethylation in a correlation with gene expression, depending on the methylation level.⁵⁰ Hydroxymethylation could also be understood as a factor trying to overcompensate for the repressing effect of hypermethylation by increasing gene transcription.⁵¹ The pathophysiological role of hydroxymethylation, especially with regard to neuropsychiatric disorders, has only very recently been introduced, 46,52,53 although its consistency across peripheral tissues has been questioned.⁵⁴ Most of the studies of hydroxymethylation have been carried out in embryonic stem cells^{51,55} or undifferentiated in vitro cells.⁵⁶ Scola et al.⁵⁶ investigated the effect of decreased mitochondrial complex I activity—an established neurobiological finding in BD—on methylation and hydroxymethylation levels in in vitro cortical neuronal rat cells. They found increased methylation and hydroxymethylation (measured by 5hCH), which was successfully prevented by pretreatment with lithium.5

Histone modifications

Post-translational histone modifications are reversible chromatin rearrangements that have an effect on transcription without affecting the DNA sequence. Histones can be acetylated/deactylated,⁵⁷ phosphorylated/dephosphorylated,⁵⁸ methylated/demethylated,⁵⁹ ubiquitinated/deubiquitinated⁶⁰ and sumoylated/desumolyated⁶¹ to regulate gene transcription. The nomenclature of histone modifications indicates which histone tail (H1–4) and which amino acid (R for arginine, K for lysine) is being modified. Additionally, lysines can be monomethylated (me), dimethylated (me2) or trimethylated (me3); the abbreviation Ac is used to refer to the acetyl state. H3K9me3, for example, a mark associated with repressive heterochromatin, stands for a trimethylation of lysine 9 on histone tail 3.⁶²

Histone deacetylation inactivates gene transcription by changing the chromatin structure, and histone deacetylase (HDAC) inhibitors have been discussed as therapeutic targets in the field of cognition and behavior.⁵⁷ At this point, 11 different HDACs have been discovered to interact with chromatin.⁶³ In BD patients, HDAC4 mRNA showed increased expression pattern in a depressed state compared with healthy controls, whereas HDAC6 and HDAC 8 were decreased—suggesting a complex expression pattern overall.⁶⁴ HDACs also seem to interact with other proteins than histones. Histone acetylation of cAMP response elementbinding protein increased its transcription, 65 which is thought to be involved in the pathophysiology of BD.⁶⁶ Epigenetic interactions such as those between histone modifications and methylation state need to be closely examined and revisited.⁶⁷ Another family of deacetylases, sirtuins, also target histone marks: the gene expression of sirtuin 1-7 (ref. 68) has been investigated in mood disorder patients. One study found state-dependent alterations in sirtuin 1, 2 and 6 in peripheral blood cells of BD and MDD patients. ⁶⁹ Duong *et al.* ⁷⁰ aimed to identify mitochondrial complex 1 dysfunction in a BD post-mortem brain sample, but no significant alteration could be determined for Sirt-3.70

A post-mortem brain investigation compared the levels of acetylated histone 3 (H3K9/K14ac) between a mixed patient sample (BD and schizophrenia (SZ)) and controls, targeting psychosis candidate gene promoters; acetylation levels of the mixed patients sample differed significantly to the controls.⁷¹ Another post-mortem study showed increased global histone H3 acetylation in BD subjects compared with age-matched controls.⁵⁸ *In vitro* experiments as well as *in vivo* follow-ups of the same sample showed how HDAC inhibitors increased the levels of H3K9/K14ac.⁷²

H3K4 trimethylation is another highly characterized histone modification, ⁷³ whose mechanism of action is thought to be the opening up of chromatin and thereby facilitating promoter binding and leading to the initiation of transcription. ⁷⁴ H3K4 trimethylation has been studied in synapsin genes (*SYN1*, *SYN2* and *SYN3*) in post-mortem brain samples of BD and MDD patients. This study showed increased H3K4 trimethylation with a distinct synapsin profile for each group. ⁷⁵

The first, above-mentioned acetylation study⁷¹ featured a state marker approach, whereas the second mentioned acetylation study⁵⁸ as well as the trimethylation study⁷⁵ focused on comparing patients with diagnoses—qualifying them for a trait marker approach.

SUICIDE IN BD

BD has the highest lifetime risk of suicide within all psychiatric disorders. About 50% of BD patients attempt suicide at least once in their life. Very few articles have been published to date that focus on the epigenetics of suicide in BD patients. Most of the neurobiological studies carried out so far focus on genetic polymorphisms associated with suicide in BD. Among them, associations with serotonin transporter polymorphisms, energy genetic variations in apoptotic regulatory genes such as forkhead box O3a⁷⁹ and polymorphisms of BDNF⁸⁰ were drawn. Gene expression studies showed increased platelet serotonin 2A receptors in suicidal patients with the biggest effect size in suicidal bipolar depression versus normal controls. Dracheva et al. Council altered splicing activity of the 5-hydroxytryptamine 2C receptor in the dorsolateral prefrontal cortex (PFC) of suicidal patients; the effect was independent of their given diagnosis (BD versus SZ).

Post-mortem brain tissue from suicide completers is used for most of current epigenetic studies, not explicitly referring to any diagnosis. For example, Fiori and Turecki⁸³ had a sample combining MDD patients, substance dependent patients and suicide completers. They found a downregulation of the expression of polyamine regulatory gene spermine N1-acetyltransferase 1, which was negatively correlated with CpG methylation in suicide completers. In a similar study with suicide completers, they showed upregulation of another polyamine regulation gene, ornithine decarboxylase antizyme 1, which was associated with increased histone 3 lysine 4 trimethylation in the upstream area. ⁸⁴

Dwivedi *et al.*⁸⁵ found epigenetic alterations of the neurotrophin receptors in the PFC and the hippocampus of suicide subjects: phosphorylation of all tropomyosin receptor kinase receptors was decreased in the hippocampus but not in the PFC; the decreased phosphorylation was noted only for tropomyosin receptor kinases A and B. Increased expression ratios of corresponding mRNA were also observed in the PFC and hippocampus. These changes were interpreted as part of apoptotic programming. Maussion *et al.*⁸⁶ analyzed methylation patterns, and their results point to a hypermethylation of the tropomyosin receptor kinase B-T1 transcript in the 3′-untranslated region in the frontal cortex of suicide completers, again with no specific diagnoses indicated.

PSYCHOSIS IN BD AS A SHARED TRAIT

Many studies investigate both psychotic bipolar patients and schizophrenia patients and refer to the mixed sample of these patients as patients affected by psychotic illnesses.^{24–26} These studies are not explicitly trying to define either a trait or state marker, but one could argue that the mixed samples are pointing towards a common trait for psychotic illness; apart from this, only one of the cited studies qualified for the trait marker search by including patients with current psychotic episodes.

At the turn of the century, the very first studies of epigenetics associated with BD focused on downregulated reelin (*RELN*) and glutamate decarboxylase (*GAD*)67 gene expression in human cortical brain tissue. ^{87–89} *GAD*67 is one of the two decarboxylases that synthesize GABA, whereas *RELN* is an extracellular matrix protein that is preferentially synthesized and secreted by GABAergic interneurons. It was suggested that downregulation of both *RELN* and *GAD*67 genes were associated with hypermethylation of their respective promoter CGIs. Interestingly, hypermethylation of these genes were correlated with increased expression of DNA methyltransferase 1 (DNMT1) in cortical GABAergic interneurons. ^{90–92} Since these studies, *RELN* and *GAD*67 have been investigated extensively showing consistent hypermethylation of the promoter and corresponding mRNA downregulation in the BD and SZ patient population. ^{47,88,93–96}

Given the extent of previous studies, the serotonergic axis is certainly one of the candidates to look for alterations in methylation patterns. Carrard *et al.*²⁶ found a hypermethylation of the serotonin 1A receptor gene promoter in a sample of SZ and BD patients; the downregulation of the corresponding mRNA was already well established.

Choosing a multitissue approach will always provide an additional benefit to the conducted study. Kaminsky *et al.*⁹⁷ analyzed brain tissues, white blood cells and germ cells of 1000 BD patients based on their previous findings in major psychosis patients (mixed sample of BD and SZ patients), which included significant alterations in human leukocyte antigen genes. They found consistent alteration patterns across tissues and were able to establish a logistic regression model based on the covariates age, a single-nucleotide polymorphism (rs1128306) and the DNA methylation pattern at CpGs 5–8 to predict whether a sample was a BD patient or part of the healthy control sample. Gene ontology analyses suggest that the expression of human leukocyte antigen complex group 9 (*HCG9*) is involved with immune system-related functions such as inflammation and regulation of B-cell-based immunological tolerance.⁹⁷

Abdolmaleky *et al.* ^{98–100} conducted various epigenetic studies in BD and SZ subjects. In 2006 they analyzed the methylation of 115 post-mortem frontal cortex samples and showed hypomethylation of catechol-O-methyltransferase (MB-COMT) in both BD and SZ subjects. 98 Nohesara et al. 101 replicated these findings in saliva samples of BD and SZ patients and found that the hyperexpression of *MB-COMT* was even more prominent in drug-free patients.¹⁰¹ Recently, they compared the dystrobrevin binding protein 1 (DTNBP1) promoter methylation in BD patients with and without a psychotic episode in saliva and post-mortem brain tissues. They found that psychotic BD patients had higher methylation rates than BD patients with no current psychotic episode.¹⁰⁰ The *DTNBP1* gene has been the focus of various genome-wide association studies. Both in BD and in SZ patients, genotypic and haplotypic associations between the phenotype and *DTNBP1* have been established. 102–104 It has been suggested that DTNBP1 may have a role in the AMPA receptor complexes, which binds glutamate as an agonist. 105 These findings suggest that genetic variants of the DTNBP1 gene confer to the susceptibility of disorder with psychotic features, whereas the actual risk of having a psychotic episode is bound to the

1494

methylation status of the *DTNBP1* promoter, which interferes with the excitatory glutamate transmission system.

Dempster et al.²¹ studied genome-wide methylation in 22 twin pairs, which were discordant for BD or SZ and found interesting results. According to the diagnostic groups, different results were found: the promoter region of sialyltransferase gene ST6GALNAC1, which has an important role in protein metabolism, showed the most significant difference between the psychosis-associated twins and the healthy twins, and the G-protein-coupled receptor 24 (GPR24) promoter gene, which interacts with the energy metabolism, showed the most significant change in methylation levels between bipolar twins and their healthy twins. Interestingly, there were also CpG sites that ranked highly in both groups (SZ and BD), but displaying opposite changes in methylation pattern. For example, zinc-finger protein 659 (ZNF659) was significantly hypomethylated in BD twins but hypermethylated in SZ twins, whereas in the mixed patients sample it did not even appear in the list of the 100 highest ranked CpG sites as the hyperand hypomethylation leveled each other out. This study supports the claim that changes in methylation levels should at least be investigated in a disorder-specific manner.

Therapeutic interventions by epigenetic modifications

Therapeutic interventions by epigenetic modifications range from the discovery of the epigenetic mechanism of well-implemented mood stabilizers to substances that are currently being developed or that show promising results in preclinical trials. ¹⁰⁶ Exciting developments are occurring around EpiEffectors, engineered transcription factors such as transcription activator-like effectors or zinc-finger-proteins, which have been designed to bind at specific loci in the genome. These newly introduced chromatin changes have only been tested in animal models so far, but show promising results in the field of neuroscience. ^{107,108} However, there is a debate about the ability to target the appropriate cell type and cause systemic instead of specific alterations. A more thorough understanding of the recent developments of EpiEffectors is provided in a recent review by Kungulovski and Jeltsch. ¹⁰⁹

DNMT inhibitors. DNMTs such as DNMT3A and DNMT3b are involved in the process of *de novo* methylation, whereas DNMT1 adds a methyl group to hemimethylated strands. There is evidence supporting that antidepressants might cause reduced DNMT1 activity by the histone methyltransferase G9a.¹¹⁰ Sales *et al.*¹¹¹ supported this finding by systemic inhibition of DNMT1 in a rodent model, resulting in antidepressant-like effects. However, an *in vitro* study suggested that antidepressant treatment did not change the expression levels of DNMT1.¹¹⁰

HDAC inhibitors. HDAC inhibitors do not only inhibit the removal of acetyl groups as their name suggests. The effects of HDAC inhibitors on DNA demethylation have also been discovered over the past years. 12 De novo methylator DNMT3B 112 or deoxygenase TET1¹¹³ might be involved in the mechanism of DNA methylation by HDAC inhibitors. In particular, the role of valproic acid (2-propylpentanoic acid) as an HDAC inhibitor has been investigated with regard to BD. 114-116 Valproate accelerates the demethylation of previously hypermethylated *RELN* promoter in the frontal cortex of adult mice. 90 Scola *et al.* 56 found methylation and hydroxymethylation levels of frontal cortex cells to be immune to a manipulated increase through retinone, if the cells were pre-treated with lithium. Yasuda et al. 117 showed that not only HDAC inhibitors such as lithium and valproate operate through the activation of BDNF exon IV promoter but butyrate sodium (sodium butanoate) and trichostatin A (7-[4-(dimethylamino)phenyl]-N-hydroxy-4,6-dimethyl-7-oxohepta-2,4-dienamide) also use the same pathway. BDNF transcription may be a key target for the effects of mood stabilizers. 118

HDAC inhibitors such as sirtinol (2-[(2-hydroxynaphthalen-1-ylmethylene)amino]-*N*-(1-phenethyl)benzamide) or MS-275 (pyridin-3-ylmethyl *N*-[[4-[(2-aminophenyl)carbamoyl]phenyl]methyl] carbamate) are also investigated as potential antidepressants in a rodent model.¹¹⁹ To date, no clinical studies have been conducted to evaluate the effect of newly designed HDAC inhibitors in MDD or BD patients.¹²⁰

Histone methyltransferase inhibitors. Histone methyltransferase inhibitors have not yet been associated with the neuropsychiatric field, ¹²¹ but the recent development of small-molecule histone methyltransferase inhibitors as a class of anticancer agents ¹²² makes it likely for them to be considered a future therapeutic target in psychiatric disorders.

Methyl donors. DNA hypomethylation can be corrected by methyl donors. L-Methylfolate has been tested as an adjunctive therapy in many clinical trials, ^{123–127} proving to be safe and effective in MDD patients. Another methyl donor, *S*-adenosyl methionine ((2*S*)-2-amino-4-[[(2*S*,3*S*,4*R*,5*R*)-5-(6-aminopurin-9-yl)-3,4-dihydroxyoxolan-2-yl]methyl-methylsulfonio]butanoate), has been shown to restore normal gene expression in neuroblastoma cells. ¹²⁸ It has also been demonstrated that methionine administration increases the methylation levels of *GAD67* and *RELN* with a consequent downregulation of the corresponding mRNAs. ⁸⁸ Clinical studies of *S*-adenosyl methionine were conducted in the late 1990s, ¹²⁹ but the substance was never introduced to treatment guidelines because of its suggested instability and costs. Recent studies are re-evaluating its feasibility as a therapeutic target in the neuropsychiatric field. ^{130,131}

CONCLUSION AND FUTURE PERSPECTIVES

In this review, we compiled epigenetic studies in BD that were conducted over the past 10 to 15 years, summarizing the results and implications. We also attempted to draw lines between the outcomes and compare different study designs to each other. Methylation, being one of the most studied epigenetic mechanisms, was discussed from various angles: data ranges from exploratory methylation studies in monozygotic twins to the search for methylation patterns as peripheral markers, and the measurement of global methylation levels in bipolar patients. A limitation in methylation research is the fact that most studies depend on patients who receive pharmacological treatment, although these drugs may influence methylation levels. 19,132 The same is true for age-associated methylation changes, although this variable is not as difficult to control for. ^{99,133} Genome-wide approaches in big samples or in monozygotic twin studies should be favored as study designs to broaden the horizons and avoid 'candidate' CpG sites. Another major limitation of previous studies is the consistency of alterations in methylation patterns across multiple tissues. On account of feasibility, most studies use peripheral blood cells as a proxy for expression or methylation levels in the brain. 134 More data are needed to support consistent methylation alterations across different tissues, as previously carried out for *BDNF*, 40 or in a multitissue study by Kaminsky *et al.* 97 Further analyses should also focus on specific cell types. 135

DNA hydroxymethylation has just been discovered with regard to epigenetic modifications. It is likely to have an important role within the network of methylation and transcription, not to mention its abundance in brain tissue compared with other tissues. Further research is needed to specify its role in epigenetic modifications and elucidate its association with affective disorders.

The emergence of microRNA into neuroscience only dates several years back and very few post-mortem brain studies have been conducted with regard to BD.^{136–138} These studies analyzed mixed patient samples, which do not contribute to the

consistency of the results after all. More research is needed to discuss the role and relevance of microRNA in the field of epigenetics.

When discussing the neurobiology of psychiatric disorder, particularly in BD, the state versus trait question is a prominent one. 'State', a stable characteristic, corresponds to BD as a diagnosis in this context, whereas 'trait' refers to a temporary characteristic corresponding to a depressive, manic or mixed episode. Most studies do not differentiate between patients having a manic or depressive episode, thus searching for a trait marker for BD. Interestingly, in methylation studies in particular, a different mental state could be shown as linked to significant alterations in DNA methylation—be it a different state within the same person, before and after medication, or different episodes within a sample of BD patients. Given the limited number of studies conducted regarding hydroxymethylation, histone modifications and small noncoding RNA in BD, this claim cannot be generalized to epigenetic modifications. Thus, more studies are needed with a focus on epigenetic modifications aside from DNA methylation and on state markers instead of diagnoses.

The definition of a bipolar patient sample, as well as the inclusion and exclusion criteria, may influence the outcome of the samples. Especially the grouping of BD with schizophrenia patients is a prevalent scientific practice, although homogeneity of this mixed sample is questioned. The significant difference in methylation patterns between psychotic BD patients and BD patients without any psychotic features¹³⁹ suggest that BD, which of course is not always associated with psychosis, should be investigated as a singular disease entity rather than within a mixed sample. Another study, supporting this paradigm, shows divergent methylation tendencies of BD versus SZ patients within the same CpG sites.²¹ Given that BD patients have the highest risk of attempting suicide in a lifetime among psychiatric disorders, the epigenetics of BD in the context of suicide need to be investigated. Most current studies aim to find distinct epigenetic patterns for suicidality, independent of underlying psychiatric disorders^{84,140}—most likely reflecting the clinical demand for a ubiquitous biomarker.

Maintenance of DNA methylation and histone modifications are critical for the development and proper functioning of the brain. Any aberrations in these modifications can lead to changes in normal brain functioning and in the development of disease phenotype. From the studies mentioned above, it appears that epigenetic modifications in critical genes that are involved in various physiological functions in the brain may have a role in the etiology of BD. Interestingly, some of the epigenetic modifications in genes that occur in BD are also common with SZ, suggesting the involvement of shared trait such as psychosis. Given the complex nature of BD, further studies are needed to dissect the precise role of epigenetic modification in the etiology of this disease phenotype. For example, it will be interesting to know whether there are shared or independent epigenetic changes that may coordinately be having a role in depressed and manic phases of BD. Similarly, the brain regions that are critical in BD need to be studied in a greater detail and in a coherent manner to examine how histone acetylation, methylation and other chromatin modifications exert their effects on gene regulation that may influence the development of BD. Regarding psychiatric disorders from a neurobiological standpoint gives us the rare chance to question rigid diagnoses and to be more precise in terms of phenotypes. A symptom- or state-based approach might help reduce the complexity of neurobiological functioning, especially when it comes to a highly heterogeneous clinical picture such as in BD.

The potential reversibility of epigenetic processes has made them interesting for therapeutic approaches. While HDAC and DNMT inhibitors show promising results in preclinical studies, methyl donors made a comeback to clinical trials, owing to recent scientific contributions to their neurobiological mechanism. A well-known shortcoming of drugs targeting epigenetic modifications is their inability to precisely target a specific cell type. Reservations should be held regarding the global inhibition of HDAC, as it has been shown that the expression pattern of the 11 known HDACs is complex and not necessarily directed in the same way. Far the recent emergence of EpiEffectors surely has the potential of introducing new target-specific psychopharmacological drugs, but a more comprehensive understanding of chromatin biology is needed to make progress in epigenetic editing.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

ACKNOWLEDGMENTS

This research was supported by grants from the National Institute of Mental Health (R01MH082802; R21MH081099; 1R01MH101890; R01MH100616; 1R01MH107183) and American Foundation for Suicide Prevention (SRG-1-042-14) to YD.

REFERENCES

- 1 American Psychiatric Association. *Diagnostic and Statistical Manual of Mental Disorders*, 5th edn. American Psychiatric Publishing: Arlington, VA, USA, 2013.
- 2 Merikangas KR, Jin R, He JP, Kessler RC, Lee S, Sampson NA et al. Prevalence and correlates of bipolar spectrum disorder in the world mental health survey initiative. Arch Gen Psychiatry 2011; 68: 241–251.
- 3 Heim C, Binder EB. Current research trends in early life stress and depression: review of human studies on sensitive periods, gene-environment interactions, and epigenetics. *Exp Neurol* 2012; **233**: 102–111.
- 4 Gelernter J. Genetics of complex traits in psychiatry. *Biol Psychiatry* 2015; **77**: 36–42.
- 5 Bohacek J, Gapp K, Saab BJ, Mansuy IM. Transgenerational epigenetic effects on brain functions. *Biol Psychiatry* 2013; 73: 313–320.
- 6 Gapp K, Jawaid A, Sarkies P, Bohacek J, Pelczar P, Prados J et al. Implication of sperm RNAs in transgenerational inheritance of the effects of early trauma in mice. Nat Neurosci 2014; 17: 667–669.
- 7 Anway MD, Cupp AS, Uzumcu M, Skinner MK. Epigenetic transgenerational actions of endocrine disruptors and male fertility. Science 2005; 308: 1466–1469.
- 8 Dias BG, Ressler KJ. Parental olfactory experience influences behavior and neural structure in subsequent generations. *Nat Neurosci* 2014; 17: 89–96.
- 9 Houston I, Peter CJ, Mitchell A, Straubhaar J, Rogaev E, Akbarian S. Epigenetics in the human brain. *Neuropsychopharmacology* 2013; **38**: 183–197.
- 10 Haque FN, Gottesman II, Wong AH. Not really identical: epigenetic differences in monozygotic twins and implications for twin studies in psychiatry. Am J Med Genet C 2009; 151C: 136–141.
- 11 Czyz W, Morahan JM, Ebers GC, Ramagopalan SV. Genetic, environmental and stochastic factors in monozygotic twin discordance with a focus on epigenetic differences. BMC Med 2012; 10: 93.
- 12 Guidotti A, Grayson DR. DNA methylation and demethylation as targets for antipsychotic therapy. *Dialog Clin Neurosci* 2014; 16: 419–429.
- 13 Houtepen LC, van Bergen AH, Vinkers CH, Boks MP. DNA methylation signatures of mood stabilizers and antipsychotics in bipolar disorder. *Epigenomics* 2016; 8: 197–208
- 14 Higuchi F, Uchida S, Yamagata H, Otsuki K, Hobara T, Abe N et al. State-dependent changes in the expression of DNA methyltransferases in mood disorder patients. J Psychiatr Res 2011; 45: 1295–1300.
- 15 Soeiro-de-Souza MG, Andreazza AC, Carvalho AF, Machado-Vieira R, Young LT, Moreno RA. Number of manic episodes is associated with elevated DNA oxidation in bipolar I disorder. *Int J Neuropsychopharmacol* 2013; 16: 1505–1512.
- 16 Woo YH, Li WH. Evolutionary conservation of histone modifications in mammals. Mol Biol Evol 2012: 29: 1757–1767.
- 17 Reik W. Stability and flexibility of epigenetic gene regulation in mammalian development. *Nature* 2007: **447**: 425–432.
- 18 Blomen VA, Boonstra J. Stable transmission of reversible modifications: maintenance of epigenetic information through the cell cycle. *Cell Mol Life Sci* 2011; 68: 27–44.
- 19 Carlberg L, Scheibelreiter J, Hassler MR, Schloegelhofer M, Schmoeger M, Ludwig B et al. Brain-derived neurotrophic factor (BDNF)-epigenetic regulation in unipolar and bipolar affective disorder. J Affect Disord 2014; 168: 399–406.

- 20 Kaminsky Z, Jones I, Verma R, Saleh L, Trivedi H, Guintivano J *et al.* DNA methylation and expression of KCNQ3 in bipolar disorder. *Bipolar Disord* 2015; **17**: 150–159
- 21 Dempster EL, Pidsley R, Schalkwyk LC, Owens S, Georgiades A, Kane F et al. Disease-associated epigenetic changes in monozygotic twins discordant for schizophrenia and bipolar disorder. Hum Mol Genet 2011; 20: 4786–4796.
- 22 D'Addario C, Dell'Osso B, Palazzo MC, Benatti B, Lietti L, Cattaneo E et al. Selective DNA methylation of BDNF promoter in bipolar disorder: differences among patients with BDI and BDII. Neuropsychopharmacology 2012; 37: 1647–1655
- 23 Strauss JS, Khare T, De Luca V, Jeremian R, Kennedy JL, Vincent JB *et al.* Quantitative leukocyte BDNF promoter methylation analysis in bipolar disorder. *Int J Bipolar Disord* 2013: **1**: 28.
- 24 Chen C, Zhang C, Cheng L, Reilly JL, Bishop JR, Sweeney JA et al. Correlation between DNA methylation and gene expression in the brains of patients with bipolar disorder and schizophrenia. Bipolar Disord 2014; 16: 790–799.
- 25 Ruzicka WB, Subburaju S, Benes FM. Circuit- and diagnosis-specific DNA methylation changes at gamma-aminobutyric acid-related genes in postmortem human hippocampus in schizophrenia and bipolar disorder. *JAMA Psychiatry* 2015; 72: 541–551.
- 26 Carrard A, Salzmann A, Malafosse A, Karege F. Increased DNA methylation status of the serotonin receptor 5HTR1A gene promoter in schizophrenia and bipolar disorder. J Affect Disord 2011; 132: 450–453.
- 27 Han H, Cortez CC, Yang X, Nichols PW, Jones PA, Liang G. DNA methylation directly silences genes with non-CpG island promoters and establishes a nucleosome occupied promoter. Hum Mol Genet 2011: 20: 4299–4310.
- 28 Doi A, Park IH, Wen B, Murakami P, Aryee MJ, Irizarry R et al. Differential methylation of tissue- and cancer-specific CpG island shores distinguishes human induced pluripotent stem cells, embryonic stem cells and fibroblasts. Nat Genet 2009; 41: 1350–1353.
- 29 Irizarry RA, Ladd-Acosta C, Wen B, Wu Z, Montano C, Onyango P et al. The human colon cancer methylome shows similar hypo- and hypermethylation at conserved tissue-specific CpG island shores. Nat Genet 2009: 41: 178–186.
- 30 Edgar R, Tan PP, Portales-Casamar E, Pavlidis P. Meta-analysis of human methylomes reveals stably methylated sequences surrounding CpG islands associated with high gene expression. *Epigenet Chromatin* 2014; **7**: 28.
- 31 Kuratomi G, Iwamoto K, Bundo M, Kusumi I, Kato N, Iwata N et al. Aberrant DNA methylation associated with bipolar disorder identified from discordant monozygotic twins. Mol Psychiatry 2008; 13: 429–441.
- 32 Sugawara H, Iwamoto K, Bundo M, Ueda J, Miyauchi T, Komori A *et al.* Hypermethylation of serotonin transporter gene in bipolar disorder detected by epigenome analysis of discordant monozygotic twins. *Transl Psychiatry* 2011; 1: e24.
- 33 Avramopoulos D, Willour VL, Zandi PP, Huo Y, MacKinnon DF, Potash JB *et al.* Linkage of bipolar affective disorder on chromosome 8q24: follow-up and parametric analysis. *Mol Psychiatry* 2004; **9**: 191–196.
- 34 Zhang P, Xiang N, Chen Y, Sliwerska E, McInnis MG, Burmeister M et al. Family-based association analysis to finemap bipolar linkage peak on chromosome 8q24 using 2,500 genotyped SNPs and 15,000 imputed SNPs. Bipolar Disord 2010; 12: 786–792.
- 35 de Mooij-van Malsen AJ, van Lith HA, Oppelaar H, Hendriks J, de Wit M, Kostrzewa E et al. Interspecies trait genetics reveals association of Adcy8 with mouse avoidance behavior and a human mood disorder. Biol Psychiatry 2009; 66: 1123–1130.
- 36 Judy JT, Seifuddin F, Pirooznia M, Mahon PB, Bipolar Genome Study C, Jancic D et al. Converging evidence for epistasis between ANK3 and potassium channel gene KCNQ2 in bipolar disorder. Front Genet 2013; 4: 87.
- 37 Fidzinski P, Korotkova T, Heidenreich M, Maier N, Schuetze S, Kobler O *et al.* KCNQ5 K(+) channels control hippocampal synaptic inhibition and fast network oscillations. *Nat Commun* 2015; **6**: 6254.
- 38 Perroud N, Zewdie S, Stenz L, Adouan W, Bavamian S, Prada P et al. Methylation of serotonin receptor 3a in Adhd, borderline personality, and bipolar disorders: link with severity of the disorders and childhood maltreatment. *Depress Anxiety* 2016: **33**: 45–55.
- 39 Dwivedi Y. Brain-derived neurotrophic factor: role in depression and suicide. Neuropsychiatr Dis Treat 2009; 5: 433–449.
- 40 Stenz L, Zewdie S, Laforge-Escarra T, Prados J, La Harpe R, Dayer A et al. BDNF promoter I methylation correlates between post-mortem human peripheral and brain tissues. Neurosci Res 2015; 91: 1–7.
- 41 Dell'Osso B, D'Addario C, Carlotta Palazzo M, Benatti B, Camuri G, Galimberti D et al. Epigenetic modulation of BDNF gene: differences in DNA methylation between unipolar and bipolar patients. J Affect Disord 2014; 166: 330–333.
- 42 Januar V, Ancelin ML, Ritchie K, Saffery R, Ryan J. BDNF promoter methylation and genetic variation in late-life depression. *Transl Psychiatry* 2015; 5: e619.

- 43 Dwivedi T, Zhang H. Lithium-induced neuroprotection is associated with epigenetic modification of specific BDNF gene promoter and altered expression of apoptotic-regulatory proteins. Front Neurosci 2014; 8: 457.
- 44 Jin SG, Wu X, Li AX, Pfeifer GP. Genomic mapping of 5-hydroxymethylcytosine in the human brain. *Nucleic Acids Res* 2011; **39**: 5015–5024.
- 45 Li W, Liu M. Distribution of 5-hydroxymethylcytosine in different human tissues. J Nucleic Acids 2011: **2011**: 870726.
- 46 Kriaucionis S, Heintz N. The nuclear DNA base 5-hydroxymethylcytosine is present in Purkinje neurons and the brain. Science 2009; 324: 929–930.
- 47 Grayson DR, Guidotti A. The dynamics of DNA methylation in schizophrenia and related psychiatric disorders. *Neuropsychopharmacology* 2013; **38**: 138–166.
- 48 Bogdanovic O, Smits AH, de la Calle Mustienes E, Tena JJ, Ford E, Williams R et al. Active DNA demethylation at enhancers during the vertebrate phylotypic period. Nat Genet 2016: 48: 417–426.
- 49 Pastor WA, Pape UJ, Huang Y, Henderson HR, Lister R, Ko M et al. Genome-wide mapping of 5-hydroxymethylcytosine in embryonic stem cells. Nature 2011; 473: 394–397.
- 50 Massart R, Suderman M, Provencal N, Yi C, Bennett AJ, Suomi S et al. Hydroxymethylation and DNA methylation profiles in the prefrontal cortex of the nonhuman primate rhesus macaque and the impact of maternal deprivation on hydroxymethylation. *Neuroscience* 2014; 268: 139–148.
- 51 Wu H, D'Alessio AC, Ito S, Xia K, Wang Z, Cui K *et al.* Dual functions of Tet1 in transcriptional regulation in mouse embryonic stem cells. *Nature* 2011; **473**: 389–393.
- 52 Kato T, Iwamoto K. Comprehensive DNA methylation and hydroxymethylation analysis in the human brain and its implication in mental disorders. *Neuro-pharmacology* 2014; 80: 133–139.
- 53 Lister R, Mukamel EA. Turning over DNA methylation in the mind. Front Neurosci 2015: 9: 252.
- 54 Godderis L, Schouteden C, Tabish A, Poels K, Hoet P, Baccarelli AA et al. Global methylation and hydroxymethylation in DNA from blood and saliva in healthy volunteers. Biomed Res Int 2015; 2015: 845041.
- 55 Tahiliani M, Koh KP, Shen Y, Pastor WA, Bandukwala H, Brudno Y et al. Conversion of 5-methylcytosine to 5-hydroxymethylcytosine in mammalian DNA by MLL partner TET1. Science 2009: 324: 930–935.
- 56 Scola G, Kim HK, Young LT, Salvador M, Andreazza AC. Lithium reduces the effects of rotenone-induced complex I dysfunction on DNA methylation and hydroxymethylation in rat cortical primary neurons. *Psychopharmacology (Berl)* 2014; 231: 4189–4198.
- 57 Machado-Vieira R, Frey BN, Andreazza AC, Quevedo J. Translational research in bipolar disorders. *Neural Plast* 2015; **2015**: 576978.
- 58 Rao JS, Keleshian VL, Klein S, Rapoport SI. Epigenetic modifications in frontal cortex from Alzheimer's disease and bipolar disorder patients. *Transl Psychiatry* 2012: **2**: e132.
- 59 Lachner M, Jenuwein T. The many faces of histone lysine methylation. Curr Opin Cell Biol 2002: 14: 286–298.
- 60 Cao J, Yan Q. Histone ubiquitination and deubiquitination in transcription, DNA damage response, and cancer. Front Oncol 2012; 2: 26.
- 61 Seeler JS, Dejean A. Nuclear and unclear functions of SUMO. Nat Rev Mol Cell Biol 2003; 4: 690–699.
- 62 Bannister AJ, Kouzarides T. Regulation of chromatin by histone modifications. Cell Res 2011; 21: 381–395.
- 63 de Ruijter AJ, van Gennip AH, Caron HN, Kemp S, van Kuilenburg AB. Histone deacetylases (HDACs): characterization of the classical HDAC family. *Biochem J* 2003; 370(Part 3): 737–749.
- 64 Hobara T, Uchida S, Otsuki K, Matsubara T, Funato H, Matsuo K *et al.* Altered gene expression of histone deacetylases in mood disorder patients. *J Psychiatr Res* 2010; **44**: 263–270.
- 65 Lu Q, Hutchins AE, Doyle CM, Lundblad JR, Kwok RP. Acetylation of cAMPresponsive element-binding protein (CREB) by CREB-binding protein enhances CREB-dependent transcription. J Biol Chem 2003; 278: 15727–15734.
- 66 Gaspar L, van de Werken M, Johansson AS, Moriggi E, Owe-Larsson B, Kocks JW et al. Human cellular differences in cAMP—CREB signaling correlate with light-dependent melatonin suppression and bipolar disorder. Eur J Neurosci 2014; 40: 2206–2215.
- 67 Dong E, Guidotti A, Grayson DR, Costa E. Histone hyperacetylation induces demethylation of reelin and 67-kDa glutamic acid decarboxylase promoters. *Proc Natl Acad Sci USA* 2007; **104**: 4676–4681.
- 68 Bosch-Presegue L, Vaquero A. Sirtuin-dependent epigenetic regulation in the maintenance of genome integrity. FEBS J 2015; 282: 1745–1767.
- 69 Abe N, Uchida S, Otsuki K, Hobara T, Yamagata H, Higuchi F et al. Altered sirtuin deacetylase gene expression in patients with a mood disorder. J Psychiatr Res 2011; 45: 1106–1112.
- 70 Duong A, Che Y, Ceylan D, Pinguelo A, Andreazza AC, Trevor Young L *et al.*Regulators of mitochondrial complex I activity: a review of literature and

- evaluation in postmortem prefrontal cortex from patients with bipolar disorder. *Psychiatry Res* 2016: **236**: 148–157.
- 71 Tang B, Dean B, Thomas EA. Disease- and age-related changes in histone acetylation at gene promoters in psychiatric disorders. *Transl Psychiatry* 2011; 1: e64
- 72 Gavin DP, Kartan S, Chase K, Grayson DR, Sharma RP. Reduced baseline acetylated histone 3 levels, and a blunted response to HDAC inhibition in lymphocyte cultures from schizophrenia subjects. *Schizophr Res* 2008; **103**: 330–332.
- 73 Ruthenburg AJ, Allis CD, Wysocka J. Methylation of lysine 4 on histone H3: intricacy of writing and reading a single epigenetic mark. Mol Cell 2007; 25: 15–30.
- 74 Kouzarides T. Chromatin modifications and their function. *Cell* 2007; **128**: 693–705
- 75 Cruceanu C, Alda M, Nagy C, Freemantle E, Rouleau GA, Turecki G. H3K4 trimethylation in synapsin genes leads to different expression patterns in bipolar disorder and major depression. *Int J Neuropsychopharmacol* 2013; 16: 289–299.
- 76 Abreu LN, Lafer B, Baca-Garcia E, Oquendo MA. Suicidal ideation and suicide attempts in bipolar disorder type I: an update for the clinician. *Rev Bras Psiquiatr* 2009: 31: 271–280.
- 77 Jamison KR. Suicide and bipolar disorder. *J Clin Psychiatry* 2000; **61**(Suppl 9): 47–51
- 78 Neves FS, Malloy-Diniz LF, Romano-Silva MA, Aguiar GC, de Matos LO, Correa H. Is the serotonin transporter polymorphism (5-HTTLPR) a potential marker for suicidal behavior in bipolar disorder patients? J Affect Disord 2010; 125: 98–102.
- 79 Magno LA, Santana CV, Sacramento EK, Rezende VB, Cardoso MV, Mauricio-da-Silva L et al. Genetic variations in FOXO3A are associated with Bipolar Disorder without confering vulnerability for suicidal behavior. J Affect Disord 2011; 133: 633–637.
- 80 Neves FS, Malloy-Diniz L, Romano-Silva MA, Campos SB, Miranda DM, De Marco L et al. The role of BDNF genetic polymorphisms in bipolar disorder with psychiatric comorbidities. *J Affect Disord* 2011; **131**: 307–311.
- 81 Pandey GN. Biological basis of suicide and suicidal behavior. *Bipolar Disord* 2013; 15: 524–541.
- 82 Dracheva S, Chin B, Haroutunian V. Altered serotonin 2C receptor RNA splicing in suicide: association with editing. *Neuroreport* 2008; 19: 379–382.
- 83 Fiori LM, Turecki G. Epigenetic regulation of spermidine/spermine N1-acetyltransferase (SAT1) in suicide. J Psychiatr Res 2011; **45**: 1229–1235.
- 84 Fiori LM, Gross JA, Turecki G. Effects of histone modifications on increased expression of polyamine biosynthetic genes in suicide. Int J Neuropsychopharmacol 2012; 15: 1161–1166.
- 85 Dwivedi Y, Rizavi HS, Zhang H, Mondal AC, Roberts RC, Conley RR *et al.* Neurotrophin receptor activation and expression in human postmortem brain: effect of suicide. *Biol Psychiatry* 2009; **65**: 319–328.
- 86 Maussion G, Yang J, Suderman M, Diallo A, Nagy C, Arnovitz M et al. Functional DNA methylation in a transcript specific 3'UTR region of TrkB associates with suicide. Epigenetics 2014; 9: 1061–1070.
- 87 Chen Y, Sharma RP, Costa RH, Costa E, Grayson DR. On the epigenetic regulation of the human reelin promoter. *Nucleic Acids Res* 2002; **30**: 2930–2939.
- 88 Guidotti A, Auta J, Davis JM, Di-Giorgi-Gerevini V, Dwivedi Y, Grayson DR *et al.* Decrease in reelin and glutamic acid decarboxylase67 (GAD67) expression in schizophrenia and bipolar disorder: a postmortem brain study. *Arch Gen Psychiatry* 2000; **57**: 1061–1069.
- 89 Smalheiser NR, Costa E, Guidotti A, Impagnatiello F, Auta J, Lacor P et al. Expression of reelin in adult mammalian blood, liver, pituitary pars intermedia, and adrenal chromaffin cells. Proc Natl Acad Sci USA 2000; 97: 1281–1286.
- 90 Dong E, Ruzicka WB, Grayson DR, Guidotti A. DNA-methyltransferase1 (DNMT1) binding to CpG rich GABAergic and BDNF promoters is increased in the brain of schizophrenia and bipolar disorder patients. Schizophr Res 2015; 167: 35–41.
- 91 Veldic M, Caruncho HJ, Liu WS, Davis J, Satta R, Grayson DR *et al.* DNA-methyltransferase 1 mRNA is selectively overexpressed in telencephalic GABAergic interneurons of schizophrenia brains. *Proc Natl Acad Sci USA* 2004; **101**: 348–353.
- 92 Veldic M, Kadriu B, Maloku E, Agis-Balboa RC, Guidotti A, Davis JM *et al.* Epigenetic mechanisms expressed in basal ganglia GABAergic neurons differentiate schizophrenia from bipolar disorder. *Schizophr Res* 2007; **91**: 51–61.
- 93 Guidotti A, Auta J, Chen Y, Davis JM, Dong E, Gavin DP et al. Epigenetic GABAergic targets in schizophrenia and bipolar disorder. *Neuropharmacology* 2011; 60: 1007–1016.
- 94 Tamura Y, Kunugi H, Ohashi J, Hohjoh H. Epigenetic aberration of the human REELIN gene in psychiatric disorders. *Mol Psychiatry* 2007; **12**: 519, 593–600.
- 95 Abdolmaleky HM, Cheng KH, Russo A, Smith CL, Faraone SV, Wilcox M *et al.* Hypermethylation of the reelin (RELN) promoter in the brain of schizophrenic patients: a preliminary report. *Am J Med Genet B* 2005; **134B**: 60–66.

- 96 Akbarian S, Huang HS. Molecular and cellular mechanisms of altered GAD1/ GAD67 expression in schizophrenia and related disorders. *Brain Res Rev* 2006; 52: 293–304
- 97 Kaminsky Z, Tochigi M, Jia P, Pal M, Mill J, Kwan A *et al.* A multi-tissue analysis identifies HLA complex group 9 gene methylation differences in bipolar disorder. *Mol Psychiatry* 2012; **17**: 728–740.
- 98 Abdolmaleky HM, Cheng KH, Faraone SV, Wilcox M, Glatt SJ, Gao F et al. Hypomethylation of MB-COMT promoter is a major risk factor for schizophrenia and bipolar disorder. Hum Mol Genet 2006: 15: 3132–3145.
- 99 Abdolmaleky HM, Yaqubi S, Papageorgis P, Lambert AW, Ozturk S, Sivaraman V et al. Epigenetic dysregulation of HTR2A in the brain of patients with schizophrenia and bipolar disorder. Schizophr Res 2011; 129: 183–190.
- 100 Abdolmaleky HM, Pajouhanfar S, Faghankhani M, Joghataei MT, Mostafavi A, Thiagalingam S. Antipsychotic drugs attenuate aberrant DNA methylation of DTNBP1 (dysbindin) promoter in saliva and post-mortem brain of patients with schizophrenia and psychotic bipolar disorder. Am J Med Genet B 2015; 168: 687–696.
- 101 Nohesara S, Ghadirivasfi M, Mostafavi S, Eskandari MR, Ahmadkhaniha H, Thiagalingam S et al. DNA hypomethylation of MB-COMT promoter in the DNA derived from saliva in schizophrenia and bipolar disorder. J Psychiatr Res 2011; 45: 1432–1438.
- 102 Gaysina D, Cohen-Woods S, Chow PC, Martucci L, Schosser A, Ball HA et al. Association of the dystrobrevin binding protein 1 gene (DTNBP1) in a bipolar case-control study (BACCS). Am J Med Genet B 2009; 150B: 836–844.
- 103 Guo AY, Sun J, Riley BP, Thiselton DL, Kendler KS, Zhao Z. The dystrobrevinbinding protein 1 gene: features and networks. *Mol Psychiatry* 2009; 14: 18–29.
- 104 Tan GK, Tee SF, Tang PY. Genetic association of single nucleotide polymorphisms in dystrobrevin binding protein 1 gene with schizophrenia in a Malaysian population. *Genet Mol Biol* 2015; **38**: 138–146.
- 105 Orozco IJ, Koppensteiner P, Ninan I, Arancio O. The schizophrenia susceptibility gene DTNBP1 modulates AMPAR synaptic transmission and plasticity in the hippocampus of juvenile DBA/2J mice. Mol Cell Neurosci 2014; 58: 76–84.
- 106 Peedicayil J. Epigenetic approaches for bipolar disorder drug discovery. *Expert Opin Drug Discov* 2014; **9**: 917–930.
- 107 Heller EA, Cates HM, Pena CJ, Sun H, Shao N, Feng J et al. Locus-specific epigenetic remodeling controls addiction- and depression-related behaviors. Nat Neurosci 2014; 17: 1720–1727.
- 108 Laganiere J, Kells AP, Lai JT, Guschin D, Paschon DE, Meng X et al. An engineered zinc finger protein activator of the endogenous glial cell line-derived neurotrophic factor gene provides functional neuroprotection in a rat model of Parkinson's disease. J Neurosci 2010; 30: 16469–16474.
- 109 Kungulovski G, Jeltsch A. Epigenome editing: state of the art, concepts, and perspectives. *Trends Genet* 2016; **32**: 101–113.
- 110 Zimmermann N, Zschocke J, Perisic T, Yu S, Holsboer F, Rein T. Antidepressants inhibit DNA methyltransferase 1 through reducing G9a levels. *Biochem J* 2012; 448: 93–102.
- 111 Sales AJ, Biojone C, Terceti MS, Guimaraes FS, Gomes MV, Joca SR. Antidepressant-like effect induced by systemic and intra-hippocampal administration of DNA methylation inhibitors. Br J Pharmacol 2011; 164: 1711–1721.
- 112 Xiong Y, Dowdy SC, Podratz KC, Jin F, Attewell JR, Eberhardt NL et al. Histone deacetylase inhibitors decrease DNA methyltransferase-3B messenger RNA stability and down-regulate de novo DNA methyltransferase activity in human endometrial cells. Cancer Res 2005; 65: 2684–2689.
- 113 Lu HG, Zhan W, Yan L, Qin RY, Yan YP, Yang ZJ et al. TET1 partially mediates HDAC inhibitor-induced suppression of breast cancer invasion. Mol Med Rep 2014; 10: 2595–2600.
- 114 Arent CO, Valvassori SS, Fries GR, Stertz L, Ferreira CL, Lopes-Borges J et al. Neuroanatomical profile of antimaniac effects of histone deacetylases inhibitors. Mol Neurobiol 2011; 43: 207–214.
- 115 Kao CY, Hsu YC, Liu JW, Lee DC, Chung YF, Chiu IM. The mood stabilizer valproate activates human FGF1 gene promoter through inhibiting HDAC and GSK-3 activities. *J Neurochem* 2013; **126**: 4–18.
- 116 Kanai H, Sawa A, Chen RW, Leeds P, Chuang DM. Valproic acid inhibits histone deacetylase activity and suppresses excitotoxicity-induced GAPDH nuclear accumulation and apoptotic death in neurons. *Pharmacogenomics J* 2004; 4: 336–344.
- 117 Yasuda S, Liang MH, Marinova Z, Yahyavi A, Chuang DM. The mood stabilizers lithium and valproate selectively activate the promoter IV of brain-derived neurotrophic factor in neurons. *Mol Psychiatry* 2009; **14**: 51–59.
- 118 Grande I, Fries GR, Kunz M, Kapczinski F. The role of BDNF as a mediator of neuroplasticity in bipolar disorder. *Psychiatry Investig* 2010; 7: 243–250.
- 119 Covington HE 3rd, Maze I, LaPlant QC, Vialou VF, Ohnishi YN, Berton O *et al.*Antidepressant actions of histone deacetylase inhibitors. *J Neurosci* 2009; **29**: 11451–11460.

1498

- 120 Fuchikami M, Yamamoto S, Morinobu S, Okada S, Yamawaki Y, Yamawaki S. The potential use of histone deacetylase inhibitors in the treatment of depression. Prog Neuropsychopharmacol Biol Psychiatry 2016; 64: 320–324.
- 121 Hu J, Chen S, Kong X, Zhu K, Cheng S, Zheng M et al. Interaction between DNA/histone methyltransferases and their inhibitors. Curr Med Chem 2015; 22: 360–372.
- 122 McCabe MT, Ott HM, Ganji G, Korenchuk S, Thompson C, Van Aller GS et al. EZH2 inhibition as a therapeutic strategy for lymphoma with EZH2-activating mutations. Nature 2012; 492: 108–112.
- 123 Fava M, Shelton RC, Zajecka JM. Evidence for the use of I-methylfolate combined with antidepressants in MDD. J Clin Psychiatry 2011; 72: e25.
- 124 Wade RL, Kindermann SL, Hou Q, Thase ME. Comparative assessment of adherence measures and resource use in SSRI/SNRI-treated patients with depression using second-generation antipsychotics or L-methylfolate as adjunctive therapy. J Manag Care Pharm 2014; 20: 76–85.
- 125 Papakostas GI, Shelton RC, Zajecka JM, Bottiglieri T, Roffman J, Cassiello C et al. Effect of adjunctive ∟-methylfolate 15 mg among inadequate responders to SSRIs in depressed patients who were stratified by biomarker levels and genotype: results from a randomized clinical trial. J Clin Psychiatry 2014; 75: 855–863.
- 126 Papakostas Gl, Shelton RC, Zajecka JM, Etemad B, Rickels K, Clain A *et al.* L-Methylfolate as adjunctive therapy for SSRI-resistant major depression: results of two randomized, double-blind, parallel-sequential trials. *Am J Psychiatry* 2012; **169**: 1267–1274.
- 127 Shelton RC, Pencina MJ, Barrentine LW, Ruiz JA, Fava M, Zajecka JM *et al.*Association of obesity and inflammatory marker levels on treatment outcome: results from a double-blind, randomized study of adjunctive L-methylfolate calcium in patients with MDD who are inadequate responders to SSRIs. *J Clin Psychiatry* 2015; **76**: 1635–1641.
- 128 Fuso A, Seminara L, Cavallaro RA, D'Anselmi F, Scarpa S. S-adenosylmethionine/ homocysteine cycle alterations modify DNA methylation status with consequent deregulation of PS1 and BACE and beta-amyloid production. *Mol Cell Neurosci* 2005; 28: 195–204.
- 129 Carney MW, Toone BK, Reynolds EH. S-adenosylmethionine and affective disorder. Am J Med 1987; 83: 104–106.
- 130 Young SN, Shalchi M. The effect of methionine and S-adenosylmethionine on S-adenosylmethionine levels in the rat brain. J Psychiatry Neurosci 2005; 30: 44–48.
- 131 Mischoulon D, Price LH, Carpenter LL, Tyrka AR, Papakostas Gl, Baer L et al. A double-blind, randomized, placebo-controlled clinical trial of

- S-adenosyl-ı-methionine (SAMe) versus escitalopram in major depressive disorder. J Clin Psychiatry 2014; **75**: 370–376.
- 132 Melka MG, Laufer BI, McDonald P, Castellani CA, Rajakumar N, O'Reilly R et al. The effects of olanzapine on genome-wide DNA methylation in the hippocampus and cerebellum. Clin Epigenet 2014; 6: 1.
- 133 Abdolmaleky HM, Smith CL, Zhou JR, Thiagalingam S. Epigenetic alterations of the dopaminergic system in major psychiatric disorders. *Methods Mol Biol* 2008; 448: 187–212.
- 134 Plume JM, Beach SR, Brody GH, Philibert RA. A cross-platform genome-wide comparison of the relationship of promoter DNA methylation to gene expression. Front Genet 2012; 3: 12.
- 135 Klengel T, Binder EB. Gene–environment interactions in major depressive disorder. Can J Psychiatry 2013; 58: 76–83.
- 136 Smalheiser NR, Lugli G, Zhang H, Rizavi H, Cook EH, Dwivedi Y. Expression of microRNAs and other small RNAs in prefrontal cortex in schizophrenia, bipolar disorder and depressed subjects. PLoS One 2014; 9: e86469.
- 137 Kim AH, Reimers M, Maher B, Williamson V, McMichael O, McClay JL et al. MicroRNA expression profiling in the prefrontal cortex of individuals affected with schizophrenia and bipolar disorders. Schizophr Res 2010; 124: 183–191.
- 138 Moreau MP, Bruse SE, David-Rus R, Buyske S, Brzustowicz LM. Altered microRNA expression profiles in postmortem brain samples from individuals with schizophrenia and bipolar disorder. Biol Psychiatry 2011; 69: 188–193.
- 139 Abdolmaleky HM, Zhou JR, Thiagalingam S. An update on the epigenetics of psychotic diseases and autism. *Epigenomics* 2015; **7**: 427–449.
- 140 Poulter MO, Du L, Weaver IC, Palkovits M, Faludi G, Merali Z et al. GABAA receptor promoter hypermethylation in suicide brain: implications for the involvement of epigenetic processes. Biol Psychiatry 2008; 64: 645–652.

This work is licensed under a Creative Commons Attribution-NonCommercial-NoDerivs 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit http://creativecommons.org/licenses/by-nc-nd/4.0/

© The Author(s) 2016