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Biomarkers for acute diagnosis and management of stroke in neurointensive care units

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Abstract:

The effectiveness of current management of critically ill stroke patients depends on rapid assessment of the type of stroke, ischemic or hemorrhagic, and on a patient's general clinical status. Thrombolytic therapy with recombinant tissue plasminogen activator (r-tPA) is the only effective treatment for ischemic stroke approved by the Food and Drug Administration (FDA), whereas no treatment has been shown to be effective for hemorrhagic stroke. Furthermore, a narrow therapeutic window and fear of precipitating intracranial hemorrhage by administering r-tPA cause many clinicians to avoid using this treatment. Thus, rapid and objective assessments of stroke type at admission would increase the number of patients with ischemic stroke receiving r-tPA treatment and thereby, improve outcome for many additional stroke patients. Considerable literature suggests that brain-specific protein biomarkers of glial [i.e. S100 calcium-binding protein B (S100B), glial fibrillary acidic protein (GFAP)] and neuronal cells [e.g., ubiquitin C-terminal hydrolase-L1 (UCH-L1), neuron-specific enolase (NSE), all-spectrin breakdown products SBDP120, SBDP145, and SBDP150, myelin basic protein (MBP), neurofilament light chain (NF-L), tau protein, visinin-like protein-1 (VLP 1), NR2 peptide] injury that could be detected in the cerebrospinal fluid (CSF) and peripheral blood might provide valuable and timely diagnostic information for stroke necessary to make prompt management and decisions, especially when the time of stroke onset cannot be determined. This information could include injury severity, prognosis of short-term and long-term outcomes, and discrimination of ischemic or hemorrhagic stroke. This chapter reviews the current status of the development of biomarker-based diagnosis of stroke and its potential application to improve stroke care.

Key words:

Biomarker, blood, cerebrospinal fluid (CSF), blood, clinical trial, intracerebral hemorrhage (ICH), ischemic stroke, serum, transient ischemic attacks (TIAs)

Introduction

In the United States, over 700,000 people are affected by strokes annually, and stroke has remained a major cause of disability for many years.^[1-3] Stroke is currently the fifth leading cause of death in the United States and the third leading cause of death worldwide.[4] Although mortality caused by stroke has significantly declined over recent years, cerebrovascular diseases remain a highly significant global health burden.^[5,6] The World Health Organization (WHO) defines stroke as "rapidly developing clinical signs of focal disturbance of cerebral function lasting more than 24 hours with no apparent cause other than of vascular origin."[7] There are two main types of strokes: ischemic and hemorrhagic. In addition, transient ischemic attacks (TIAs),

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which have traditionally been classified as a separate cerebrovascular disease because their duration (by definition) is less than 24 h, are more appropriately classified as ischemic stroke if brain lesion(s) are evident on magnetic resonance imaging (MRI).^[8]

Ischemic stroke is the most common type, accounting for over 85% of all strokes. The most common causes of ischemic stroke are arterial occlusion from a thrombus or embolus, hypoperfusion from decreased blood pressure, or oxygen deprivation from systemic hypoxia. ^[9] Hemorrhagic stroke is caused by the rupture of an artery or vein in the brain and leaking of blood into the brain tissues. Mass effect from a hematoma may injure neurons from the effects of direct pressure. Elevated blood pressure and cerebral aneurysms are common conditions that can cause a hemorrhagic stroke.

How to cite this article: Glushakova OY, Glushakov AV, Miller ER, Valadka AB, Hayes RL. Biomarkers for acute diagnosis and management of stroke in neurointensive care units. Brain Circ 2016;2:28-47. Hemorrhagic stroke includes intracerebral hemorrhage (ICH) and subarachnoid hemorrhage (SAH). ICH occurs when blood accumulates in the brain parenchyma, whereas SAH refers to bleeding within the space between the brain and the arachnoid. ICH is the most common type of hemorrhagic stroke, comprising 10% of all strokes. SAH represents 1-7% of all strokes and affects about 30,000 people in the United States annually.^{110]} Even though the incidence of hemorrhagic stroke is significantly lower than that of ischemic stroke, the contributions that ICH, SAH, and ischemic stroke make to the total number of years of productive life lost from all strokes are comparable: ICH: 34.2%, SAH: 27.3%, and ischemic stroke: 38.5%.^[11,12]

Diagnosis and Management of the Stroke Patients in Neurointensive Care Unit

Management of a patient who has sustained an acute stroke is focused on stabilizing the patient and restoring cerebral blood flow as soon as possible to prevent further brain damage. Optimal management requires rapid assessment and early intervention to facilitate maximal reperfusion of brain tissue.^[13] Currently, thrombolytic therapy with recombinant tissue plasminogen activator (r-Tpa) remains the only effective treatment option for ischemic stroke patients that is approved by the Food and Drug Administration (FDA).^[14] Administration of r-tPA has a very narrow therapeutic window and requires assessment prior to treatment to rule out a hemorrhagic component of stroke.[15] The use of intravenous r-tPA was approved by the FDA in 1996 based on the National Institute of Neurological Disorders and Stroke r-tPA Stroke Study, in which ischemic stroke patients were treated with r-tPA within 1.5-3 h of symptom onset.^[16] According to the guidelines recently proposed by the American Heart Association/ American Stroke Association based on several clinical trials, intravenous r-tPA is the standard treatment for eligible ischemic stroke patients within 3 h of symptom onset and in selected patients up to 4.5 h after ictus, and selective cerebral intraarterial r-tPA administration has a 6-h time window.^[15] Nevertheless, both intravenous and intraarterial treatments should be applied as soon as feasible to maximize the likelihood of a beneficial outcome.[15]

The workup for a patient with suspected stroke first includes a history (especially the time when neurologic symptoms began), a physical examination [including scoring on the National Institutes of Health Stroke Scale (NIHSS)], and imaging studies (to rule out hemorrhagic components). In addition, several diagnostic studies may be obtained that include prothrombin time and international normalized ratio (PT/INR), glucose, complete blood count, metabolic panel, creatine kinase, electrocardiogram, echocardiogram, lipid panel, carotid Doppler ultrasonography, magnetic resonance angiography (MRA), and computed tomographic angiography (CTA). This assessment should be performed quickly enough to preserve the time-sensitive option of thrombolytic treatment if appropriate. The main goals during the initial assessment of ischemic stroke patients include: 1) exclusion of intracranial hemorrhage, 2) assessment for contraindications to thrombolysis, and 3) characterization of the infarct.^[17]

Neurological assessment plays a central role in both therapeutic decision-making and prognostication. Neurological deterioration is a common complication of acute stroke, occurring in as many as 20-40% of all the cases.^[18,19] However, this rate might be underestimated due to different definitions of neurological deterioration (for example, increase in NIHSS \geq 2 points or \geq 4 points).^[20] There exists clinical evidence that early neurological deterioration may be predictive of a poor short-term prognosis.^[19] Patients who undergo hemorrhagic transformation of an ischemic stroke are at especially high risk of neurological deterioration, with poor outcomes and increased mortality.[21] A retrospective cohort study in ICH patients suggested that relatively easy and effective risk stratification of early neurological deterioration is possible at admission based on the presence of a "spot sign" (suggestive of active hemorrhage) on the initial brain CTA imaging or extensive degree of intraventricular hemorrhage, whereas risk stratification of late neurological deterioration is possible based on specific clinical parameters assessed at admission such as degree of comorbidity (Charlson index), stroke severity, and/ or degree of intraventricular hemorrhage.^[22]

Clinical Implementation of Stroke Biomarkers for Acute Diagnosis in Neurointensive Care Unit

Historically, the use of diagnostic serum biomarkers such as creatinine kinase and troponin for evaluation of possible myocardial ischemia significantly improved diagnostics in the emergency room at admission and thereby expedited timely treatment. Similarly, the use of sensitive and specific central nervous system (CNS)-biomarkers of stroke-associated brain injuries could significantly impact the current treatment of stroke and improve overall patient outcomes. Current diagnostic tests for stroke rely on the neurological assessment, which may have poor interexaminer reliability and on neuroimaging, which might not be promptly available at all locations. Thus, inexpensive blood-based biomarkers could potentially provide a much needed objective assessment tool. Biomarkers could improve stroke care by allowing early diagnosis even by clinical providers without extensive neurological training as well as by facilitating serial monitoring of patients and rapid assessment of the severity of brain injury.

The rapid and objective discrimination of stroke type in acute care environments is likely to increase the number of patients with ischemic stroke receiving thrombolytic therapy, thereby improving overall patient outcome and health care for stroke patients. In addition, using a panel of neuronal injury biomarkers would complement the present neuroimaging modalities for the diagnosis of stroke. These biomarkers would be particularly important in patients with nonlocalizing or transient neurological symptoms, those in whom neuroimaging cannot be obtained, or those who are nondiagnostic. The biomarker assessment could be performed during initial triage, avoiding delays in transporting stroke patients to appropriate care centers and allowing expedited treatment of patients at high risk for early stroke recurrence.

In cases where stroke onset is uncertain, for example, in so-called "wake-up" strokes occurring during sleep, which account for up to 25% of all stroke cases,^[23] the presence of signature biomarkers (those with either acute or delayed

release) or the levels of certain markers that could identify the timing of stroke onset (i.e., recent if acute marker levels are high, or relatively remote if they are low) could help ascertain the time of stroke onset.

Certain biomarker signatures could help the selection of appropriate treatment plans for patients with acute stroke. The information obtained from biomarker measures could be used in conjunction with acute neuroimaging patterns to determine if salvageable tissue is present and potentially to lead to more appropriate therapy.^[24] It is conceivable that early determination of certain biomarkers such as matrix metalloproteinase-9 (MMP-9) or fibronectin could potentially identify patients at risk of secondary complications of stroke, particularly hemorrhagic transformation leading to ICH and edema.^[24] In patients with high levels of such biomarkers, caution may be warranted while low levels of biomarkers may identify patients at lower risk of bleeding who would benefit from more aggressive revascularization measures or thrombolytic treatment. In addition, certain biomarkers of endothelial damage may identify patients at risk for developing malignant edema, for which presently no reliable clinical or imaging predictors exist.^[25]

Monitoring biomarker levels during the first few days of hospitalization may provide further insight into stroke progression and predict or evaluate the possible causes of worsening including infection, fever, metabolic derangement, edema, hemorrhagic transformation in ischemic stroke, or vasospasm in SAH. Serial monitoring of biomarker activity could potentially identify patients with continuing or delayed ischemia who may benefit from more aggressive stroke management.

Although to date no clinically approved biomarker for stroke diagnostics is available, several blood biomarkers associated with different pathophysiological pathways of stroke have been identified as potentially useful in clinical management, possibly contributing additional information to current diagnostics, interventions, risk stratification, and monitoring of efficacy of therapy. Well-designed, large-scale clinical studies addressing relevant clinical questions are needed.^[26]

Selection of Central Nervous System-Specific Protein Biomarkers for Acute Stroke Diagnosis

A wealth of clinical and experimental data indicate that different CNS injuries including ischemic and hemorrhagic strokes and traumatic brain injury (TBI) promote upregulation of specific characteristic proteins and their subsequent release in cerebrospinal fluid (CSF). On the other hand, these CNS injuries may promote upregulation of specific proteolytic enzymes catalyzing the degradation of cell-specific structural and membrane proteins that result in build-up of characteristic proteolysis products. It is widely recognized that the serum levels of many of these proteins or protein fragments may reflect the severity of brain injury, and that their cellular origin might be indicative of the mechanisms or cellular type affected by injury. These specific protein markers accumulate within brain parenchyma and CSF and then leak into the peripheral circulation at levels proportional to those observed in the brain tissue or CSF. Thus, the primary focus of many stroke biomarker studies is to assess brain pathologies and elucidate pathophysiological processes underlying these pathologies to facilitate evidence-based clinical decisions about timely intervention and management in stroke patients. Although a troponin-like diagnostic biomarker for stroke would be of interest, several other biomarker applications are possible and more likely to be of clinical importance.[27] Considerable literature suggests that biomarkers specific for glial and neuronal injuries might provide valuable and timely diagnostic information for stroke such as time of stroke onset when the exact time could not be determined, severity, discrimination of ischemic or hemorrhagic stroke, and short-term and long-term outcomes and prognosis. Because discrimination between ischemic and hemorrhagic strokes is critical to provide clinical decision for appropriate patient management, one of the main focuses of stroke biomarker research is to identify specific biomarkers that reflect ischemic and hemorrhagic insults. Of interest are studies of CNS-specific biomarker dynamics performed in TBI, which share several common features with strokes including brain ischemia, parenchymal brain hemorrhage, and SAH often coexisting at varied degrees; comparisons of the biomarker profiles between these two acute insults would provide valuable information regarding pathophysiological aspects of a specific brain injury phenotype.^[28-30] A number of clinical trials have been performed to establish the utility of several biomarker candidates for stroke diagnosis. The clinical trials with the most promising biomarkers of glial and neuronal brain injuries for diagnosis of acute ischemic stroke and ICH are summarized in Table 1.

Glial-specific biomarkers of acute brain injury *S100 calcium-binding protein B (S100B)*

S100B is a Ca2+-binding protein, which belongs to the S100 EF-hand type calcium-binding protein family and is a commonly used astrocytic marker.^[60] In the CNS, S100B is found primarily in mature astrocytes and NG2 cells.^[61] NG2 cells have an astrocytic appearance but are commonly considered as oligodendrocyte precursor cells based on their NG2 proteoglycan expression, which is characteristic for oligodendrocyte progenitor cells. They do not express glial fibrillary acidic protein (GFAP) or other antigens characteristic of other types of mature glial cell or markers for neurons.[62,63] Under normal physiological conditions, S100B is located predominantly intracellularly and plays important roles in Ca²⁺ homeostasis, astrocytic glutamate uptake, and neurite outgrowth stimulation. However, following brain injuries associated with glutamate excitotoxicity, S100B can be released from astrocytes where it plays complex and differential roles in astrocytic proliferation and differentiation, neuronal survival, and/or apoptotic neuronal and astrocytic cell death.[64] S100B has been identified as a prospective marker to predict early neurological outcomes and potentially discriminate ischemic stroke from TIA and other acute brain injuries including TBL.^[31,32] Thus, S100B levels may have the potential to diagnose and predict human neurological disorders associated with glutamate excitotoxicity and astrogliosis including TBI,^[65,66] traumatic and nontraumatic ICH, and ischemic stroke.[67]

CSF and serum S100 protein and NSE levels have long been considered as markers of glial and neuronal cell damages in the CNS, respectively, in many neurological disorders and brain injuries.^[31,68] Serum S100B levels correlate with neurological

Biomarker (other markers in the study)	Study design	Sample Time point (s)	Patient Groups	Size	Biomarker Levels	Major findings	Clinical implication
Glial-specif	ic biomarkers of a	cute brain in	ijury				
S100B ^[31]	Prospective cohort study	Serial 1-10 d	Ischemic stroke	26	>0.2 µg/L 0.25±0.15 µg/L (day 0) 1.80±3.30 µg/L (day 3) 0.40±0.33 µg/L (day 10)	S100B was elevated in ischemic stroke compared to control S100B correlated with neurological deficits, infarction, and edema at admission but was not significantly correlated with the functional prognosis	A prospective marker for diagnostic applications in ischemic stroke
S100B[32]	Prospective	0-24 h	Ischemic	20	<0.2 µg/∟ 0.08-6.73 pg/	Positive correlation of serum	A prospective marker
3100B ¹⁻¹	cohort study	0-24 11	stroke TIA	18	0.08-8.73 hg/ mL 0.01-0.73 ng/	S100B with neurological deficit in Ischemic stroke	to predict early neurological outcomes
					mL	and TBI patient	and discriminate
			TBI Control	10 28	0.1-3.48 ng/mL 0.01-0.34 ng/ mL	Significant difference in temporal profiles of S100B levels between ischemic stroke and TIA or TBI patients	TIA and other acute brain injuries (i.e., TBI)
S100B (NSE) ^[33]	Prospective cohort study	6-120 h	Ischemic stroke	32		Positive correlation with the neurological deficit and the final infarct volume S100B concentrations at 6 h were associated with the functional outcome S100B above 0.2 µg/L at 48 h indicated a poor functional outcome at 3 months	A prospective marker for diagnostic and prognostic application in ischemic stroke
S100B (MBP, NSE, sTM) ^[34]	Retrospective (samples from NINDS r-tPA trial)	0-24 h	Ischemic stroke (r-tPA) Ischemic stroke (placebo)	359	Baseline: 0.21 ng/mL (0.0- 0.309) 2 h — 0.22 ng/ mL (0.0-0.255) 24 h-0.34 ng/ mL (0.0-0.762)	Higher peak concentrations of S100B was associated with larger CT lesion volumes (r =0.263, P <0.0001) Patients with favorable outcomes had smaller changes in S100B (P <0.05) concentrations in the first 24 h No difference between r-tPA and placebo groups	A prospective marker to predict CT brain lesion and early functional outcome in ischemic stroke Not sensitive for assessment of r-tPA treatment efficacy
S100B (GFAP, NSE, APC- PCI) ^[35]	Prospective multicenter cohort study	24 h	Ischemic stroke ICH	83 14	Median 11.2 μg/L (6.0-40.4) Median 11.7 μg/L (5.9-18.7)	No significant differences in S100B between ischemic stroke and ICH	Not a sensitive mark to discriminate ischemic stroke and ICH in a mixed stroke population
S100B (UCH-L1, GFAP) ^[36]	Randomized controlled trial German Multicenter EPO Stroke Trial	Serial 1-7 d	Ischemic stroke (EPO- treated) Ischemic stroke (placebo)	76 87	~1-1100 pg/mL	AUC for S100B alone corrected for day 1 NIHSS before drug treatment showed a tendency but was not significantly lower in EPO vs placebo groups Composite AUCs of all three markers were different between treatment groups	A prospective marker in biomarker panel (i.e., UCH-L1, GFAP, S100) to assess drug treatment efficacy in ischemic stroke The panel may not provide additional information obtained with UCH-L1 alone

Table	1: (Clinical	trials	of	serum	protein	biomarke	rs of	neuronal	and	glial	injury	for	diagnostic	acute	ischemic
stroke	e an	d ICH														

Table 1: Continued										
Biomarker (other markers in the study)	Study design	Sample Time point (s)	Patient Groups	Size	Biomarker Levels	Major findings	Clinical implication			
S100B (Tau) ^[37]	Prospectively enrolled into the	1-10 d	Ischemic stroke	56	Day 1: 51.0 pg/ mL (30.1-77.2) Day 3: 57.9 pg/ mL (27.5-109.1) Day 5: 52.9 pg/ mL (30.8-92.5) Day 10: 36.0 pg/mL (25.6-59.2)	S100B was detected in all patients on days from 1 to 5 and decreased on day 10. S100B was also detected in the control group (55.2%)	Limited diagnostic value in ischemic stroke			
			Control (crural varices)	38						
S100B (NSE) ^[38]	Prospective cohort study	rospective Serial bhort study 12-48 h	Ischemic stroke ICH	61 79	127,0 nmol/L (57,0-639,5) 183,5 nmol/L	S100B was elevated in stroke vs control S100B correlated with depression symptoms at 60 days	A prospective marker for stroke diagnosis and to predict poststroke depression in ischemic stroke and ICH			
			Control	79	(83,0-3102,5) 84,6 nmol/L (13,6-284,2)					
S100B (RAGE) ^[39]	Prospective cohort study	24 h	Ischemic stroke	776	58.70 pg/mL	S100B levels were significantly increased in ischemic stroke vs ICH (<i>P</i> <0.001)	A prospective marker to discriminate ischemic stroke and ICH			
			ICH	139	107.58 pg/mL		Combination of biomarkers S100B/ RAGE pathway may provide useful information for diagnosis of ischemic stroke vs ICH in the first hours from symptoms onset			
S100B (GFAP,	Prospective single-center	24 h	Ischemic stroke	31	0.069 ng/mL	Serum levels of S100B were significantly higher in stroke	Serum S100B levels is a prospective marker to			
MMP-9, sVCAM-1) ^[40]	pilot study		ICH Vertigo (nonvascular)	12 22	0.047 ng/mL	patients than in nonvascular vertigo patients Detect stroke in vertigo group (sensitivity 94.4%, specificity 31.8%)	distinguish stroke and vertigo of nonvascular causes			
S100B ^[41]	Prospective observational study	48 h	Control Ischemic stroke (66.19%)	15 142	1.12±1.58 ng/ mL	S100B was increased in both ischemic stroke and ICH but not in TIA, compared to the control	A prospective marker to distinguish ischemic stroke and ICH from TIA S100B protein might be a copredictor of outcome in ischemic stroke and ICH			
			TIA (9.15%).		ng/mL ng/mL)					
			Control	40	0.1782±0.1622 ng/mL					
S100B (NSE, HSP70) ^[42]	Prospective cohort study	24 h	ICH	35	0.13±0.03 (day 0) 0.13±0.04 (day 5)	S100B on days 0 and 5 was significantly increased in ICH compared to the control group (<i>P</i> <0.001)	A prospective marker for diagnosis of ICH			
			Control	32	0.08±0.03	Positive correlation with NIHSS and bleeding volume, negative correlation with GCS				

Continued

Tab	le 1	1: (Continued

Biomarker (other markers in the study)	Study design	Sample Time point (s)	Patient Groups	Size	Biomarker Levels	Major findings	Clinical implication
GFAP ^[43]	Prospective cohort study	6 h	Ischemic stroke ICH	93 42	14 ng/L 11 ng/L (0-3096)	GFAP with a cutoff point of 2.9 ng/L detected ICH in acute stroke (sensitivity 0.79, specificity 0.98, positive predictive value 0.94, negative predictive value 0.91; <i>P</i> <0.001)	A prospective marker to discriminate ischemic stroke and ICH
GFAP ^[44]	Prospective cohort study	Serial 1-48 h	Ischemic stroke ICH	45 18		GFAP was not detected in ischemic stroke within 24 h GFAP at 2 h was significantly correlated with ICH volume (r =0.755, P =0.007). Differentiate between ICH and ischemic stroke within 2-6 h (diagnostic accuracy was>0.80)	A prospective marker to discriminate ischemic stroke and ICH
(S100B, NSE, APC- PCI) ^[35]	Prospective multicenter cohort study	24 h	Ischemic stroke	14	Median <30 ng/L (<30-1280) Median 55 ng/L (<30-1850)	Levels of GFAP were significantly higher in ICH patients (<i>P</i> =0.0057) Combination of GFAP and APC-PCI in patients with NIHSS score >3 sensitivity and negative predictive value of 100% for ICH (<i>P</i> =0.0052)	A prospective marker in combination with APC-PCI to discriminate ischemic stroke and ICH prior to neuroimaging in a mixed stroke population
GFAP (UCH-L1, S100B) ^[36]	Randomized controlled trial German Multicenter EPO Stroke Trial	Serial 1-7 d	Ischemic stroke (EPO) Ischemic stroke (placebo)	76 87	~0.1-12 ng/mL	AUC for GFAP alone corrected for day 1 NIHSS before drug treatment showed a tendency but was not significantly lower in EPO vs placebo groups. Composite AUCs of all three markers were different between the treatment groups	A prospective marker in the biomarker panel (i.e., UCH-L1, GFAP, S100) to assess drug treatment efficacy in ischemic stroke The panel may not provide additional information obtained with UCH-L1 alone
GFAP ^[45]	Prospective multicenter cohort study	4.5 h	Ischemic stroke ICH Control (stroke mimic)	163 39 3	Median 1.91 μg/L (0.02-236.27) median 0.08 g/L (0.00-0.97) Median 0.19 μg/L (0.16-0.21)	GFAP cutoff of 0.29 μg/L differentiated ICH from ischemic stroke and stroke mimic (sensitivity 84.2%, specificity 96.3%)	A prospective marker to discriminate ICH from ischemic stroke and stroke mimic
GFAP (S100B, MMP-9, sVCAM-1) ^[40]	Prospective single-center pilot study	24 h	ischemic stroke ICH Vertigo (nonvascular) Control	31 12 22 15	0.069 ng/ml (0.051-0.135) 0.047 ng/mL (0.034-0.06)	No significant differences among groups were found for GFAP levels	Serum GFAP levels is not a sensitive marker to distinguish stroke and vertigo of nonvascular causes
GFAP (NR2 antibodies) ^[46]	Prospective cohort study	12-72 h 1 and 2 w	Ischemic stroke ICH Control	49 23 52		GAFAP and NR2 antibodies when used in combination discriminated ischemic stroke and ICH at 12 h after onset (sensitivity 94%, specificity 91%)	A prospective marker in combination with NR2 antibodies to discriminate ischemic stroke and ICH at 12 h after onset
GFAP ^[47]	Prospective cohort study	2-6 h	Ischemic stroke ICH	65 43	0.6±0.4 ng/mL 1.6±0.8 ng/mL	GFAP in ICH was significantly higher than in ischemic stroke group (<i>P</i> <0.001) GFAP at the cut point of 0.7 ng/mL differentiate ICH from ischemic stroke (sensitivity 86.0%, specificity 76.9%)	A prospective marker to discriminate ischemic stroke and ICH

Table 1: Continued										
Biomarker (other markers in the study)	Study design	Sample Time point (s)	Patient Groups	Size	Biomarker Levels	Major findings	Clinical implication			
GFAP (RPB4) ^[48]	Retrospective cohort study	6 h	Ischemic stroke	36	Median 0.075 ng/mL (0.04-0.68)	Plasma GFAP was significantly higher in ICH vs ischemic stroke (P<0.0001) GEAP and BBP4 showed a	A prospective marker to discriminate ischemic stroke and ICH Combination with BPB4			
				10	ng/mL (0.04-0.04)	specificity 100% for both stroke subtypes	may improve diagnosis within the first hours after stroke			
Neuron-spe	cific biomarkers of	acute brain	i injury							
UCH-L1	Randomized	Serial	Ischemic	76	~0.035-0.36	AUC for UCH-L1 alone corrected	A prospective marker			
(S100B, GFAP) ^[36]	controlled trial German Multicenter EPO	1-7 d	stroke (EPO)		ng/mL	for day 1 NIHSS before drug treatment was significantly lower in EPO vs placebo groups	alone or in biomarker panel (i.e., UCH-L1, GFAP, S100) to assess drug tractment officiant			
	Stroke Trial		Ischemic stroke (placebo)	87		composite AUCs of all three markers were different between treatment groups	in ischemic stroke The panel may not provide additional			
							information obtained with UCH-L1 alone			
NSE (S100B) ^[33]	Prospective cohort study	6-48 h	Ischemic stroke	32		Positive correlation with the neurological deficit and the final infarct volume	A prospective marker for diagnostic and prognostic application in ischemic stroke			
NSE (MBP, S100B, sTM) ^[34]	Retrospective (samples from NINDS r-tPA trial)	0-24 h	ischemic stroke (r-tPA)	359	Baseline: 16,2 ng/mL (2.9-118.7) 2 h-17.3 ng/mL (0.0-172.7) 24 h-15.6 ng/ mL (0.0-189.2)	Higher peak concentrations of MBP was associated with larger CT lesion volumes (<i>r</i> =0.117, <i>P</i> <0.0001) No differences between r-tPA and placebo groups	A prospective marker to predict CT brain lesion and early functional outcome in ischemic stroke Not sensitive for assessment of r-tPA treatment efficacy			
NSE (Tau) ^[49]	Prospective cohort study	3-120 h	Ischemic stroke	66	12.5 μg/L	NSE was associated with the neurovascular status on admission. NSE was significantly correlated with the functional outcome at 3 months (<i>P</i> <0.001)	A prospective marker for diagnostic and prognostic application in ischemic stroke			
NSE (S100B, GFAP.	Prospective multicenter cohort study	24 h	lschemic stroke	83	median 0.11 μg/L (0.02-1.66)	No significant differences in NSE between ischemic stroke and ICH	Not a sensitive mark to discriminate ischemic stroke and ICH in a			
APC-PCI) [35]			ICH	14	Median 0.14 μg/L (0.6-0.96)		mixed stroke population			
NSE ^[50]	Prospective cohort study	72 h	Ischemic stroke	150	>25 ng/mL (>35 ng/mL, <i>n</i> =13).	Positive correlation NSE with degree of disability and neurological worsening	A prospective marker to predict stroke severity and early functional			
			Control	101	≤25 ng/mL	Positive correlation NSE with severity of stroke at the time of admission (r =0.919; P <0.001)	stroke			
						NSE and degree of disability (i.e., mild, moderate, and severe NIHSS score) (χ^2 =94.905, <i>P</i> <0.001)				
NSE (S100B) ^[38]	Prospective cohort study	12-48 h	ischemic stroke	61	11.2 nmol/L (3.3-54.5)	NSE was elevated in stroke vs control	A prospective marker for stroke diagnosis and			
,,	,		ICH	79	19.3 nmol/L (9.7-26.9)	NSE was not associated with stroke severity on admission	to predict functional outcome in ischemic			
			Control (high risk)	79	9.5 nmol/L (2.2-23.0)	NSE was associated with functional neurological outcome at 60 days and to the degree of recovery				

Table 1: Continued

Biomarker (other markers in the study)	Study design	Sample Time point (s)	Patient Groups	Size	Biomarker Levels	Major findings	Clinical implication
NSE ^[51]	Prospective cohort study	24 h	Ischemic stroke	75	15.68-198.42 ng/l	Positive correlation with infarct volume (CT) (<i>r</i> =0.955, <i>P</i> <0.001) Negative correlation with GCS (<i>r</i> =0.806, <i>P</i> <0.001) Positive correlation with functional neurological outcome (mRS) at day 30 (<i>r</i> =0.744, <i>P</i> <0.001)	A prospective marker to predict stroke severity and early functional outcome in ischemic stroke
NSE (IL-10) ^[52]	Prospective cohort study	72 h	lschemic stroke Control	100	17.95±4.54 ng/ mL 7.48±1.51 ng/ mL	Significantly increased in stroke patients compared to control Positive correlation with degree of neurological deficit (NIHSS score) (r =0.8, P ≤0.001)	A prospective marker to predict stroke severity and early functional outcome in ischemic stroke
NSE ^[53]	Prospective cohort study	24 h	Ischemic stroke	83	9.9±1.7 (unchanged) 9.7±3.4 (increased) 10.6±4.5 (decreased) 16.7±6.4 (1 peak) 18.7±6.6 (2 peaks)	NSE levels were stationary [unchanged (26.5% of the patients), increased (10.8%), and decreased (21.7%)] and NSE increase showed 1 peak (20.5%) and 2 peaks (20.5%) NSE increase with 2 peaks correlated with the incidence of atrial fibrillation and hemorrhagic transformation (<i>P</i> =0.02)	A prospective marker for monitoring hemorrhagic transformation and the status of blood-brain barrier disruption in ischemic stroke
NSE (CRP) ^[54]	Prospective cohort study	72 h	Ischemic stroke ICH Control	88 32 50	22.6±7.7 ng/mL	Increased in acute stroke cases compared to the controls (<i>P</i> <0.05) Not statistically different between ischemic stroke and ICH	A prospective marker to predict stroke severity and early functional outcome in ischemic stroke
NSE (S100B, HSP70) ^[42]	Prospective cohort study	24 h	ICH	35 32	Day 0 — 31.66±13.43 Day 5 — 26.56±12.77 21.97±11.13	NSE on day 0 was significantly increased in ICH compared to the control group (<i>P</i> <0.01) No significant correlation with NIHSS score, bleeding volume	A prospective marker for diagnostic of ICH
NSE ^[55]	Prospective study	24 h	Ischemic stroke (r-tPA)	67	15.60 ng/mL (8.480-30.69)	Positive correlation with NIHSS score at 24 h after r-tPA (R =0.342, P =0.005) rather than baseline NIHSS score Serum NSE at 24 h showed an increase patients with atrial fibrillation compared to those without. (18.37±4.83 ng/mL vs 14.64±4.14 ng/mL; P =0.003)	NSE levels at 24 h may help predict long-term outcome of ischemic stroke patients with intravenous rtPA treatment
MBP (NSE, S100B, sTM) ^[34]	Retrospective (samples from NINDS r-tPA trial)	0-24 h	Ischemic stroke (r-tPA) Ischemic stroke (placebo)	178	Baseline — .036 ng/mL (0.0-3.606) 2 h — 0.035 ng/mL (0.0-2.149) 24 h — 0.131 ng/mL (0.0-11.835)	Higher peak concentrations of MBP was associated with larger CT lesion volumes (r =0.209, P<0.0001) Patients with favorable outcomes had smaller changes in MBP (P <0.05) concentrations in the first 24 h No difference between r-tPA and the placebo groups	A prospective marker to predict CT brain lesion and early functional outcome in ischemic stroke Not sensitive for assessment of r-tPA treatment efficacy
MBP (IMA) ^[56]	Prospective study	12 h	lschemic stroke	34	82.4±63.9 pg/ mL	No statistically significant difference between the stroke and control groups Statistically significant	MBP levels do not increase in early period of stroke cases
			Control	34	73±47.5 pg/mL	correlations with and NIHSS score (<i>P</i> =0.002, <i>r</i> =0.43) and IMA (<i>P</i> =0.015, <i>r</i> =0.344) levels	

Biomarker (other markers in the study)	Study design	Sample Time point (s)	Patient Groups	Size	Biomarker Levels	Major findings	Clinical implication
Tau (NSE) ^[49]	Prospective cohort study	3-120 h	Ischemic stroke	66	60 pg/mL	Tau was correlated with severity of neurological deficits and infarct volume (<i>P</i> =0.001) but was not associated with the neurovascular status on admission	A prospective marker for diagnostic and prognostic application in ischemic stroke
Tau (S100B) ^[37]	Prospective study	1-10 d	Ischemic stroke	56	Day 1 — 89.1 pg/mL (77.6- 104.2) day 3 — 87.6 pg/mL (67.1- 140.9) day 5 — 94.2 pg/mL (72.1-197.0) day 10 — 99.6 pg/mL (77.6- 246.3)	Tau protein concentrations measured within the early phase did not correlate with degrees of neurological deficit and disability in the early phase and after 3 months A significant correlation between pau protein level and Barthel Index measured at the same time point was observed only on day 10 (r =0.43; P =0.03	Detection of tau protein in the serum of patients with ischemic stroke but not its concentration can be considered as a bad prognostic factor for the clinical outcome in early and late phases of ischemic stroke
Tau ^[57]	Prospective cohort study	6 h	ICH	176	168.2 pg/mL	Tau levels with cutoff 91.4 pg/mL predicted 3-month poor outcome (sensitivity 83.6%, specificity 75.8%; AUC, 0.826; 95% Cl, 0.762-0.879)	A prospective marker to predict mortality and poor outcomes at 3 months in ICH
VLP-1 ^[58]	Prospective cohort study		Ischemic stroke Control	16 17	1.78 μg/L 0.03 μg/L	VILIP-1 was elevated in ischemic stroke vs control (sensitivity 100%, specificity 100% at 0.093 mcg/L VILIP-1)	A prospective marker for stroke diagnostic in ischemic stroke
NR2 peptide ^[59]	Prospective blinded study	72 h	Ischemic stroke	192	Median 5.44 μg/L (0.1-62.71)	Increased in ischemic stroke Positive correlation with lesion volume (r_s =0.73) At cutoff 1 µg/L, NR2 levels detected ischemic stroke (sensitivity 92%, specificity 96%)	A prospective marker for diagnostic and prognostic application in ischemic stroke

deficit resulting from both stroke and TBI,^[32] and levels have been shown to correlate with the severity of brain injury.[66] S100B has also been reported as a biomarker of blood-brain barrier disruption in various conditions.[69,70] A small study has shown that following aneurysmal SAH, the CSF level of S100B as well as of some other biomarkers mentioned below including UCHL-1, NSE, and spectrin breakdown products (SBDPs) increased up to 100-fold and that a panel of selected neurodegeneration markers might be valuable as surrogate endpoints for assessment in SAH patients.^[71] In addition, changes in S100B levels correlated positively with acute changes in depressive symptoms' severity following coronary artery bypass grafting.^[72] Several studies have demonstrated that serum concentrations of S100B are significantly increased following stroke between days 2 and 4 after symptom onset and peaking after 48 h.^[31,32] There may be a role for serum S100B measured before specific treatment as a copredictor of outcome in patients with acute stroke admitted to a hospital emergency department.^[41,73] Significant correlations between S100B in CSF and volume of infarction have also been demonstrated in early-phase severe TBI.^[74]

Table 1: Continued

Studies performed in TBI patients have shown elevated S100B levels at the acute stage of head trauma. These levels were related to alternations in intracranial pressure and cranial computed tomography (CT) findings, suggesting that S100B may serve as a reliable marker for the evaluation of injury severity^[75,76] and as a potential outcome predictor.^[77] Serum S100B concentrations have been reported to increase after closed head injury in children.^[78] However, temporal profiles of the serum S100B protein levels following stroke and TBI were different, peaking on day 3 or 4 after stroke and on day 1 or 2 after TBI.^[32] S100B serum levels are associated with different types of traumatic intracranial lesions, notably epidural and subdural hematomas, SAH, and brain edema.^[79] The concentration of S100B increases in the plasma of patients with ischemic stroke, and its elevated levels are associated with larger infarct size, poor neurovascular status on admission, and worse outcomes.[33] A recent study including 35 patients with nontraumatic ICH and 32 healthy controls has shown significant increases in S100B levels in the serum of ICH patients at admission and at day 5, and the S100B levels were negatively correlated with Glasgow Coma Scale scores and

positively correlated with NIHSS, bleeding volume, and inhospital mortality.^[42] S100B protein levels were significantly higher in stroke patients compared to the controls, and these levels were significantly higher in the ischemic stroke group as compared to the ICH or TIA groups, with the highest levels in patients who did not survive.^[41] A related study including a total of 80 patients including vertigo patients with ischemic stroke in the posterior circulation (n = 31), ICH (n = 12), and nonvascular cause (n = 22), and matched controls (n = 15) has shown that serum levels of S100B at 24 h of symptom onset were significantly higher in stroke patients than in patients with vertigo from nonvascular causes or controls, whereas the levels of other biomarkers used in the study [i.e., GFAP, soluble vascular cellular adhesion molecule-1 (sVCAM-1), and MMP-9] were not significant in the groups.^[40]

The diagnostic and prognostic values of S100B as single biomarker are limited but it shows great potential as part of a biomarker panel for stroke.^[34,39] S100B levels in combination with levels of other biomarkers such as NSE and GFAP have been reported to predict TBI outcome.[80-84] In ischemic stroke and ICH patients, the levels of S100B at admission were significantly higher than in high-risk controls but not different between the two types of stroke, and the S100B levels correlated with depression symptoms but not with functional recovery at 2 months following stroke onset.[38] Importantly, it has been shown that using a panel comprising S100B and its scavenger receptor, a soluble receptor for advanced glycation end products (sRAGE), allowed differentiation between ischemic stroke and ICH in a sample of 915 stroke patients [area under the curve (AUC) 0.76].^[39] In patients with first-ever acute lacunar stroke,^[85] higher serum levels of S100B and lower levels of sRAGE measured within 7 days of symptom onset were independently associated with the presence and number of cerebral microbleeds assessed by using susceptibility-weighted magnetic resonance imaging (SWI).^[86]

Glial fibrillary acidic protein

Glial fibrillary acidic protein (GFAP) is a brain-specific type III intermediate filament protein found primarily in mature astrocytes and to a lesser extent in some other glial cells.[87] Various types of brain injuries and neurological disorders are associated with gliosis and subsequent GFAP upregulation in the brain. Numerous studies documenting different spatiotemporal profiles of GFAP upregulation in different brain disorders suggest that GFAP could be an attractive marker for screening for the presence of astroglial damage. Several studies have found that changes in GFAP levels have potential clinical utility in acute brain injuries such as TBI and stroke and could provide unique information for determining injury severity and even differential diagnosis of different types of stroke.^[88,89] Recent studies have documented elevated GFAP or its breakdown products (GFAP-BDP) levels in both CSF and serum after mild, moderate, or severe TBI in adult patients, and these elevated GFAP levels correlate with TBI magnitude and outcomes^[66,81,83,90-95] in children as well.^[96-98] These findings are consistent with preclinical models suggesting an association of CSF and serum GFAP levels with TBI outcomes.^[99] Importantly, the aforementioned preclinical study demonstrated significant correlation of GFAP levels in serum and CSF following experimental TBI, suggesting the validity of serum analyses to assess changes of this biomarker in the brain.^[99] Another TBI study has shown that although the levels of GFAP were increased in TBI, the levels GFAP have poor prognostic value.^[100] In addition, clinical studies suggest a possible role for GFAP as a brain-specific marker for malignant gliomas.^[89]

GFAP has long been also suggested as a marker of glial injury in stoke. Even in an early study, GFAP was identified as a biomarker indicative of ICH within 6 h of symptom onset, and a cutoff point of 2.9 ng/L was found to provide a sensitivity of 79% and a specificity of 98% for the differentiation of ICH from ischemic stroke.^[43] It has been suggested that after ICH, GFAP can be found in the serum of patients due to necrosis of brain cells and subsequent blood-brain barrier disruption. A subsequent multicenter study of S100B, NSE, GFAP, and activated protein C-protein C inhibitor complex (APC-PCI) demonstrated a significant ability of GFAP to distinguish ICH from ischemic stroke.^[35] In a recent multicenter cohort study including 205 patients diagnosed with ischemic stroke, ICH, or stroke mimics, plasma GFAP on admission had a sensitivity of 84% and specificity of 96% for differentiating ICH from ischemic stroke and stroke mimics (AUC 0.915, 95% CI 0.847-0.982).[45] GFAP was increased in ischemic stroke and demonstrated its potential application in stroke therapy development studies when used along with other biomarkers.^[36]

Neuron-specific biomarkers of acute brain injury

Ubiquitin C-terminal hydrolase L1

Ubiquitin C-terminal hydrolase-L1 (UCH-L1), also known as neuronal-specific protein gene product 9.5, is an abundant protein localized in the cell body of neurons in CNS. Thus, UCH-L1 been proposed as a biomarker specific to neuronal injury following ischemic insult in a preclinical model of ischemic stroke induced by middle cerebral artery occlusion (MCAO).^[101] A recent study demonstrated elevation of CSF and serum UCH-L1 levels in a preclinical model of ischemic stroke induced by middle cerebral artery occlusion, whereas no change was observed in a rat ICH model induced by collagenase injection into the striatum.^[102] A related study in a preclinical piglet model also suggested the utility of serum UCH-L1 to predict neuronal apoptosis induced by deep hypothermic circulatory arrest, which is routinely used in cardiac surgery.^[103] Several studies have demonstrated the utility of CSF and serum levels of UCHL-1 as diagnostic and prognostic biomarkers of severe and mild TBIs.[66,100,104-106, 108] Another TBI study has shown that although serum UCH-L1 levels were elevated in military personnel with low-level occupational blast exposure, the correlation of UCH-L1 levels with blast magnitude was weak and inconsistent, and no correlation has been found between UCH-L1 levels and deficits in behavioral measures.^[109] Interestingly, a recent TBI study suggests that serum UCH-L1 concentrations might be able to discriminate TBI patients with abnormal and normal CT findings.^[94]

In addition, a significant correlation between serum and CSF levels of UCH-L1 and outcome has been demonstrated in a preclinical TBI model, and the increases in both serum and CSF levels reflected brain pathology.^[99] CSF levels of UCH-L1 and other neurodegenerative biomarkers such as phosphorylated neurofilament H (pNF-H) and S100B are elevated in patients with aneurysmal SAH, and the levels of these biomarkers are correlated with neurological outcomes and mortality.^[71,110] A study performed using serum samples from patients enrolled in the trial of erythropoietin for stroke^[111] has shown that the levels of UCH-L1, S100B, and GFAP were increased in ischemic stroke samples, and the decrease in the UCH-L1 levels was associated with beneficial outcomes of erythropoietin treatment.^[36]

A study performed in 125 patients with dizziness, hypertension, type 2 diabetes mellitus, or dyslipidemia found that UCH-L1 levels in serum were significantly different between the controls and patients with white matter lesions (i.e., patients without white matter lesions; n = 46, P < 0.05), and these increased UCH-L1 levels were correlated with white matter lesion severity, suggesting that UCH-L1 could serve as a biomarker of neuronal damage associated with white matter lesions.^[112] Subgroup analysis of patients with different types of lesions has shown that serum UCH-L1 levels were significantly higher in the patients with subcortical white matter lesions but not in the patients with paraventricular lesions compared to the controls.[112] Interestingly, in the same study an attempt was made also to correlate UCH-L1 levels in the urine of patients with white matter lesions with that of controls but the urine UCH-L1 levels were similar between these groups (P > 0.05).^[112]

Neuron-specific enolase

Neuron-specific enolase (NSE) is a dimeric isoform of the glycolytic enzyme enolase found mainly in neurons. Although NSE is relatively specific for neuronal cells, it is also found in neuroendocrine carcinomas. Several early studies documented elevated NSE concentrations in CSF and blood samples following different brain disorders including stroke [65,113] and related preclinical models.[114-118] Increased serum concentrations of NSE have been reported in TBI patients, and the NSE levels were correlated with neurological outcomes, suggesting that NSE could be a diagnostic and prognostic biomarker of TBI.^[65] An earlier study in acute ischemic stroke found that peak levels of NSE in plasma samples were observed on the second day after symptom onset, and some correlation was found between the levels of NSE and infarct volumes (r = 0.37, P < 0.05) but not with clinical outcome (r = 0.18, P > 0.05).^[119] In contrast, a further study demonstrated that serum NSE and its peak levels correlated with baseline NIHSS scores but not with infarct volume.^[34] Elevated NSE levels in serum within the first 24 h after symptom onset have been reported only in a small fraction of stroke patients, [34,49,68,120-122] probably due to the wide range of NSE levels in the normal population.^[123] In addition, no clear relationship has been found between serum and CSF levels of NSE in subjects without neurological disease.^[123] Further studies have shown that serum NSE levels at 72 h after stroke significantly correlate with worse neurological outcomes, and these levels have high predictive value for determining stroke severity.[50-52,54] It has been shown that serum levels of NSE in the first few days after ischemic stroke can serve as a useful marker to predict stroke severity and early functional outcome.^[51] A study performed in 100 ischemic stroke patients and controls without stroke demonstrated that serum concentrations of NSE were significantly increased (17.95 \pm 4.54 pg/mL vs 7.48 \pm 1.51 pg/mL, $P \le 0.05$) and concentrations of interleukin-10 (IL-10) significantly decreased (11.79 ± 2.77 pg/mL vs $15.72 \pm 2.69 \text{ pg/mL}, P \leq 0.05$) in the blood samples from stroke patients taken within the first 72 h after stroke onset

as compared to NSE and IL-10 levels in control patients.^[52] This study also demonstrated a significant correlation of NSE concentrations with NIHSS scores, with concentrations of both NSE and IL-10 having a high predictive value for early neurobehavioral outcomes.^[52] The maximum serum NSE and C-reactive protein levels within 72 h have been shown to be significantly correlated with the severity of neurological disability, and both markers have high predictive value for early neurobehavioral outcome after acute ischemic stroke.^[54] Furthermore, a recent study of 83 ischemic stroke patients found that the temporal profile of NSE levels varied widely among different patients: no changes in NSE levels in 26.5% of the patients, increase in 10.8% of the patients, decrease in 21.7% of the patients, and transient changes with a single peak in 20.5% of the patients or two peaks in 20.5% of the patients. The presence of an NSE peak was significantly correlated with the incidence of atrial fibrillation and hemorrhagic transformation.[53] A study including 44 ischemic stroke patients and 17 ICH patients found that in serum samples from patients with ischemic stroke and ICH, the median concentrations of NSE were significantly higher than in samples collected from a high-risk group (n = 79), and the median NSE concentration in ICH patients was significantly higher compared to the ischemic stroke group.^[38] However, the NSE levels were not associated with stroke severity at admission though the NSE levels were predictive of functional recovery at 60 days following stroke onset.[38] A recent prospective study has shown that serum NSE concentrations in ischemic stroke patients (n = 67) assessed within the first 4.5 h of symptom onset were correlated with neurological outcomes assessed using NIHSS at 24 h after r-tPA therapy (R = 0.342, P = 0.005), and the patients with favorable neurological outcomes after 90 days (n = 32) had lower serum NSE levels and NIHSS scores.^[55]

In addition, a small trial provided evidence that a panel of biomarkers including NSE may have predictive value for infarct volume, incidence of vasospasm, and overall outcome in patients with SAH.^[71] In patients with nontraumatic ICH, the serum levels of NSE were significantly higher at admission but not at day 5 as compared to the controls though no significant correlation has been found between NSE levels and Glasgow Coma Scale scores, NIHSS, bleeding volume, or in-hospital mortality.^[42]

α II-spectrin breakdown products SBDP120, SBDP145, and SBDP150

In CNS, α II-spectrin is a major structural component of the neuronal cytoskeleton and is especially prevalent in axons. It plays a critical role in neuronal integrity. After injury, α IIspectrin can become a major substrate for both calpain and caspase-3 proteases, which play an important role in oncotic necrosis and apoptotic cell death, respectively. Although calpain activation is primarily associated with oncotic cell death, it can also contribute in some situations to apoptotic cell death.^[124]

Different SBDPs produced by calpain and caspase-3 specific proteolysis have long been studied as biomarkers of axonal damage following brain injuries. Moreover, there exists substantial experimental evidence showing that α II-spectrin is specifically processed to signature cleavage products that

vary with the particular protease that is involved, including 150 kDa (SBDP150) and 145 kDa (SBDP145) fragments produced by calpain proteolysis, and a major cleavage product of 120 kDa (SBDP120) produced by caspase-3.^[125] Studies performed in TBI patients have shown that the temporal profiles of the levels of the various SBDPs in CSF correlate with the acute diagnosis of severe TBI.^[126,127]

These temporal profiles suggest that neuronal death (before 72 h) is mostly due to necrosis, and secondary neuronal death (after 72 h) is primarily due to apoptosis. Furthermore, the severity of brain injury correlates with increased levels of SBDPs.^[126-129]

Several SBDPs are detectable in CSF after in patients with aneurysmal SAH.^[128] Quantitative analyses of SBDPs in brains from rats that underwent middle cerebral artery occlusion (MCAO) revealed that SBDP150 and SBDP145 were strongly increased 6 h after experimental ischemia. SBDP145 level was elevated after 6 h after ischemic stroke detected by Western blot and enzyme-linked immunosorbent assay (ELISA).^[102] Other preclinical studies in animal stroke models demonstrated accumulation of calpain and caspase-3 proteolytic fragments of brain-derived αII-spectrin in CSF.^[130,131]

The major advantage of α II-spectrin over other related biomarkers is its ability to reveal both calpain and caspase-3 activation within the acute period after CNS injury. SBDP120 is a sensitive marker of caspase-3 activation exclusively associated with apoptotic cell death following severe TBI.^[126] SBDP150 and SBDP145 provide a highly sensitive measure of calpain activation primarily associated with oncotic cell death, and to lesser extent the changes in the levels of these SBDPs might be also be indicative of apoptotic cell death.^[124] A significant increase in SBDP150 and SBDP120 levels was observed following SAH.^[128] This variability emphasizes the heterogeneous nature of CNS pathology after stroke and points to important implications for individualized treatment of braininjured patients tailored to specific neurochemical cascades in the injured brain.

Importantly, a recent small study performed in patients with aneurysmal SAH showed that the changes in the CSF levels of SBDPs used in a biomarker panel significantly correlated with brain infarction, cerebral vasospasm, and poor outcomes.^[71]

Myelin basic protein and myelin basic protein autoantibody

Myelin basic protein (MBP) is a structural hydrophilic protein that plays a critical role in organization of the structure of myelin sheaths of oligodendrocytes and Schwann cells in the CNS. MBP has long been considered as a marker of active demyelination.^[132] It has been shown that the MBP levels in TBI patients with severe intracerebral damage were significantly increased at the time of admission and remained elevated for up to 2 weeks following injury, and higher serum MBP levels were associated with poor outcomes.^[133] Recent studies suggested a prospective prognostic value of serum MBP levels in adult and pediatric TBIs.^[65] Studies from the 1980s have shown increased levels of MBP in the CSF of patients with ischemic stroke and ICH.^[134,135] MBP was found to be elevated in CSF samples from patients with ICH collected at a mean of 1 day after stroke (range: days 0-3). MBP levels increased in samples from both ischemic stroke and ICH patients collected at a mean of day 5 (range: days 4-8) as compared to day 1, whereas no change in MBP levels was found in patients with TIAs in whom the symptoms resolved within 24 h.^[135] In this early study, the MBP levels correlated with brain lesions, and higher values were predictive of poor short-term prognosis.^[135] These findings were confirmed in a study with 28 ischemic stroke patients, which showed that MBP concentrations in serial CSF specimens were significantly increased at 7 days but not at 48 h or at 18-21 days after stroke onset.[136] Another study found elevated MPB concentrations in 39% (n = 28) of the samples collected from ischemic stroke patients at admission.^[137] In a study using retrospective serum samples from subjects enrolled in the original NINDS r-tPA Stroke Study,^[16] peak MBP concentrations at 24 h after stroke onset were associated with higher NIHSS baseline scores and larger CT lesion volumes.[34] However, in this study, no significant changes in MBP or other biomarkers (e.g., tau protein, NSE, S100B) were found between the treatment groups despite significantly improved outcomes in the r-tPA-treated group.^[34] The results of a recent study of ischemic stroke patients included within 5-10 days of symptom onset demonstrated significantly higher levels of CSF MBP than in the normal population, and the levels of MBP and other markets used in this study (e.g., tau protein, GFAP) correlated with clinical stroke severity.^[138] However, in acute ischemic stroke patients, the serum levels of MBP within 12 h of symptom onset were not significantly different compared to the MBP levels in the samples from the healthy control group.^[56] Thus, the delay in increase of MBP levels might be a factor limiting its diagnostic application for acute stroke management.

Interestingly, a recent study including subjects with ischemic stroke and controls (n = 112 and n = 40, respectively) demonstrated that although there was a global decrease in MBP autoantibody titers at acute time points after stroke, patients with white matter lesions demonstrated an increased titer of MBP autoantibodies at 1 month after stroke and also demonstrated worse neurological outcomes, suggesting possible pathological consequences involving these autoantibodies.^[139] These findings were consistent with a recent preclinical study showing that rats developing Th1 or Th17 responses to MBP had a worse outcome after ischemic stroke.^[140] On the other hand, measuring MBP autoantibodies titers in stroke patients could be an alternative strategy for MBP-based diagnosis of white matter injury.

Neurofilament light chain

Neurofilament light chain (NF-L) is a light chain subunit of a structural triplet protein that forms the core of the neurofilament, which is an essential structural element of axons and to lesser extent of the cell body and dendrites. NF-L binds to several structural proteins including spectrin and microtubule-associated protein-2 (MAP-2).^[141] Thus, the appearance of NLF in CSF has been postulated to be specific for axonal injury. Elevated NF-L levels in CSF have been reported in different chronic neurodegenerative disorders and following acute brain injuries including ischemic stroke SAH and TBI.^[142,143] A recent study demonstrated that patients with ischemic stroke had significantly higher CSF levels of NF-L and that it correlated with the severity of white matter lesions.^[138] A study of patients with cervical artery dissection compared to patients with local symptoms only (n = 8), TIAs, (n = 10) or ischemic stroke (n = 31) found a significant increase in serum NF-L levels in patients with stroke as compared to patients with local symptoms or TIAs, and the NF-L level correlated with clinical stroke severity and poor outcome at 3 months.^[144] In addition, the CSF level of NF-L correlates with outcome after aneurysmal SAH.^[107,145] Analysis of serial ventricular CSF samples collected from 35 aneurysmal SAH patients for up to 15 days demonstrated elevated NF-L concentrations in all patients, and the NF-L levels were higher in the patients with early cerebral ischemia defined by CT scans within the first 3 days.^[107] However the dynamics of NF-L release in neurodegenerative disorders and following CNS injuries are still poorly understood, and further studies are warranted to establish the validity of NF-L as a diagnostic biomarker for acute stroke.

Tau protein

Tau is a highly soluble structural protein that belongs to the neuron-specific type II microtubule-associated protein family. In the CNS, tau protein is predominantly expressed in neurons and to a lesser extent in astrocytes and oligodendrocytes. Tau plays a critical role in the structural stabilization of microtubules by interaction with tubulin and formation of the neuronal cytoskeleton. Under pathological conditions, tau forms insoluble aggregates (pathologies collectively known as tauopathies), which are believed to play role in the pathophysiology of many neurodegenerative disorders.^[146] Tau aggregates were first described in Alzheimer's disease, and it was suggested that these aggregates resulted from abnormal modification of tau protein mainly by its hyperphosphorylation.[147,148] Levels of both phosphorylated and total tau proteins in the CSF have long been considered as promising markers to discriminate neurodegenerative disorders from healthy aging.[149,150] Further studies have shown that tau protein also increases in CSF following acute brain injuries including stroke^[37,151,152] and TBI.^[153,154] Increased levels of tau protein in serum have been demonstrated in patients following stroke, and mild and severe TBI.[155-157] Interestingly, these studies have also shown that CSF and serum concentrations of tau protein in TBI patients measured initially after injury are correlated with short-term and longterm outcomes.[154,157]

After ischemic stroke, tau protein levels in CSF showed a marked transient increase 2-3 days after symptom onset, peak at 1 week, and return to normal levels after 3-5 months.^[151,152] A recent study demonstrated that CSF levels of total tau protein in samples collected within 5-10 days of stroke onset as well as levels of other biomarkers including NF-L, MBP, and GFAP were significantly higher in patients with ischemic stroke as compared to controls and that tau protein levels, along with the levels of MBP and GFAP, significantly correlated with clinical stroke severity.^[138]

Similarly, serum concentrations of tau proteins in blood samples of patients with ischemic stroke collected at extended time points have shown correlation with stroke outcomes, whereas tau protein concentrations in the majority of the samples collected within 24 h of symptom onset were within normal range.^[49] Serum tau protein was detected in 47.8% of ischemic stroke patients, and these patients developed more severe neurological deficits, suggesting that the presence of tau protein in the serum itself, rather that its exact concentration, at early time points after stroke has a predictive value as a prognostic indicator associated with infarction volume, neurological deficits, and functional status measured in the early and late phases of ischemic stroke.^[37] In ICH, a study performed with 176 patients demonstrated that serum tau levels measured at admission had a prognostic value for mortality and poor neurological outcomes at 3 months after symptom onset.^[57]

Visinin-like protein-1

Visinin-like protein-1 (VLP-1) is a member of the family of neuronal intracellular calcium sensor visinin-like proteins (also known as VILIPs) that are abundantly expressed in the brain and play a role in Ca²⁺-dependent cell signaling involved in the modulation of cyclic adenosine monophosphate (cAMP) production.[158] Initially, VLP-1 was cloned using cDNA from a rat brain library, which was further demonstrated to have 100% homology with human VLP-1.[159,160] Although VLP-1 is preferentially expressed primarily in CNS neurons, it is also found in pancreatic islets.^[161] Initial studies have shown timedependent increase of VLP-1 levels in a preclinical model of ischemic stroke and in human plasma samples retrospectively collected from stroke patients.^[162] In human plasma samples, VLP-1 was not detected in ~92% of unaffected controls (36 from 39 samples tested), whereas VLP-1 was detected in up to 100% of the samples from patients with confirmed stroke after 48 h after stroke onset but VLP-1 was detected only in 44% of the plasma samples from these patients taken within the first 24 h after symptom onset.^[162] However, another pilot study performed using serum samples taken after 3 h of stroke onset demonstrated that the VLP-1 level of $0.093 \,\mu g/m$ significantly (P < 0.01) differentiated healthy individuals (n = 17) from stroke patients (n = 16) with 100% sensitivity and 100% specificity.^[58]

In addition, recent studies have shown that serum and CSF levels of VLP-1 might be diagnostic of Alzheimer's disease^[163,164] and that serum and CSF VLP-1 levels might be diagnostic biomarkers of acute encephalopathy with biphasic seizures and late reduced diffusion (AESD).^[163,164]

NR2 peptide and NR2 autoantibody

The N-methyl-D-aspartate (NMDA) receptor is a subtype of ionotropic glutamate receptor that plays a critical role in learning and memory. The NMDA receptor is a tetramer comprising two glycine-binding NR1 and two glutamate-binding NR2 subunits.^[165] NMDA receptors are expressed predominantly in CNS neurons though the expression of different NR1 and NR2 subunit receptors has been demonstrated in cerebral epithelial cells,^[166] oligodendrocytes,^[167] and in neurons of enteric nervous system.^[168,169] Alterations in the expression of NMDA receptor subunits have been demonstrated in different neurological disorders. Following cerebral ischemia, the expression of NR2 subunit is upregulated,[170] whereas the expression of the NR1 subunit is downregulated.[171] In addition, there exists evidence that cerebral ischemia and excitotoxicity induce calpain-mediated cleavage of both NR2A and NR2B subunits.[172,173]

A study including 105 stroke patients [i.e., TIAs (n = 56), acute ischemic stroke (n = 31), ICH (n = 18)] and 255 controls has shown that the levels of the autoantibodies to NR2A/2B NMDA receptor subunits in plasma are increased in patients with TIAs and ischemic stroke as compared to controls, and that the levels of NR2A/2B autoantibodies measured within 72 h differentiated ischemic stroke from ICH.[174] Another study from the same group using serum samples collected from patients with suspected TIA and ischemic stroke within 72 h of stroke onset demonstrated that levels of NR2 autoantibodies were elevated in females with acute strokes and in both males and females with multiple recent strokes as compared to patients without stroke or to healthy controls, and the levels of NR2 autoantibodies in stroke patients correlated in a gender-dependent manner with several risk factors for stroke including hypertension, diabetes mellitus, and atrial fibrillation, suggesting that levels of NR2 autoantibodies might be indicative of a history of multiple strokes and be a predictive factor for stroke.[175]

Levels of NR2 peptide (probably a product of calpain-mediated proteolysis of the NR2A/2B subunits^[176]) have been shown to be elevated in plasma samples collected at admission from patients with ischemic stroke.^[59] This study enrolled 292 subjects with ischemic stroke, TIAs, or vascular stroke factors (ICH patients were excluded from the study) as well as healthy controls. NR2 peptides may have a role as a diagnostic biomarker for acute stroke.

Use of Blood Glial Fibrillary Acidic Protein Levels for Early Discrimination of Ischemic Stroke and Intracerebral Hemorrhage

Rapid differentiation between ICH and ischemic stroke using biomarker testing would facilitate prehospital, cause-specific management of stroke patients. It has been postulated that the more sudden disruption of astrocytes and blood-brain barrier after ICH may be responsible for the rapid appearance of GFAP in serum, in contrast to more delayed astrocytic damage and subsequent release of GFAP in ischemic stroke.^[43,89] The predictive value of GFAP for discriminating between ischemic stroke and ICH could be increased when used with an additional biomarker or biomarker panel. An exploratory multicenter study with 97 stroke patients enrolled within 24 h of symptom onset (83 ischemic stroke and 14 ICH) showed that blood levels of GFAP and (APC-PCI) might be useful to rule out ICH in a mixed stroke population prior to neuroimaging, whereas S100B and NSE had no such predictive value.[35]

Elevated serum and CSF levels of GFAP are hallmarks of brain injuries and are associated with overall outcome and prognosis. GFAP may be useful as a surrogate marker for the management of hemorrhagic stroke.^[43] Previous studies have suggested that the serum level of GFAP appeared to be a more sensitive marker of brain injury in small lesions and minor strokes as compared to S100B^[27] but its application as a diagnostic tool might be limited by its delayed rise in concentration following mild injuries.^[43,177] However, a recent study provided strong evidence that the serum levels of GFAP can differentiate ICH and ischemic stroke. Meta-analyses of recent clinical trials suggested that serum GFAP is a sensitive and specific test for differentiating ICH and ischemic stroke in patients within 1-6 h of stroke onset.^[178,179] GFAP level is increased after both ischemic stroke and ICH but its concentrations are significantly higher in early ICH patients.^[44]

A multicenter cohort study including 205 patients diagnosed with either ischemic stroke, ICH, or stroke mimics suggested that plasma GFAP analysis performed within 4.5 h of symptom onset can differentiate ICH and ischemic stroke with a sensitivity of 84% and specificity of 96%.[45] Similar results were obtained in a study of Chinese patients using blood samples collected within 2-6 h after the onset of symptoms. A blood GFAP level at the cut point of 0.7 ng/mL yielded an AUC of 0.901 (95% CI 0.828-0.950) with high sensitivity (86.0%) and specificity (76.9%) to differentiate ICH from ischemic stroke.^[47] In addition, in the ICH group, a significant correlation was observed between GFAP levels and neurological deficits assessed by NIHSS, and between GFAP levels and hemorrhage volume, and the highest GFAP levels were predictive of poor short-term functional outcomes.[47] Furthermore, a recent study suggested that GFAP used in combination with NR2 can differentiate between ischemic stroke and ICH at a time point extended to 12 h after the ictus with a sensitivity of 94% and specificity of 91%.[46] Another study showed that plasma concentrations of retinol binding protein 4 and GFAP at concentrations >61 µg/mL and <0.07 ng/mL, respectively, demonstrated 100% specificity, and these biomarkers might be independent predictors to discriminate stroke subtype, improving discrimination by 29% (P < 0.0001).^[48]

Conclusion

Recent clinical trials have identified several prospective brainspecific biomarkers for which their appearance in the peripheral circulation reflects their postinjury dynamics in the brain tissue and CSF. Although many of these biomarkers demonstrate independent diagnostic and prognostic values in ischemic stroke and ICH, the heterogeneity of stroke phenotypes suggests that useful clinical information can be obtained only from a panel of selected biomarkers. Some promising biomarkers are still at an early stage of development, and further clinical trials are warranted to determine their utility in clinical practice. Of interest, especially in the neurointensive care unit (NICU) applications, are early biomarkers capable of estimating the time of stroke onset or discriminating ischemic from hemorrhagic stroke to facilitate clinical decision-making such as whether to administer thrombolytic therapy. Future trials will provide answers to this and many other questions in this rapidly developing area.

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References

1. Rosamond W, Flegal K, Furie K, Go A, Greenlund K, Haase N, *et al.*; American Heart Association Statistics Committee and Stroke

Statistics Subcommittee. Heart disease and stroke statistics — 2008 update: A report from the American Heart Association Statistics Committee and Stroke Statistics Subcommittee. Circulation 2008;117:e25-146.

- 2. Kochanek KD, Murphy SL, Xu J, Arias E. Mortality in the United States, 2013. NCHS Data Brief 2014:1-8.
- 3. Mozaffarian D, Benjamin EJ, Go AS, Arnett DK, Blaha MJ, Cushman M, *et al.*; American Heart Association Statistics Committee and Stroke Statistics Subcommittee. Heart disease and stroke statistics — 2015 update: A report from the American Heart Association. Circulation 2015;131:e29-322.
- GBD 2013 Mortality and Causes of Death Collaborators. Global, regional, and national age-sex specific all-cause and cause-specific mortality for 240 causes of death, 1990-2013: A systematic analysis for the Global Burden of Disease Study 2013. Lancet 2015;385: 117-71.
- Feigin VL, Forouzanfar MH, Krishnamurthi R, Mensah GA, Connor M, Bennett DA, *et al.*; Global Burden of Diseases, Injuries, and Risk Factors Study 2010 (GBD 2010) and the GBD Stroke Experts Group. Global and regional burden of stroke during 1990-2010: Findings from the Global Burden of Disease Study 2010. Lancet 2014;383:245-54.
- Johnston SC, Mendis S, Mathers CD. Global variation in stroke burden and mortality: Estimates from monitoring, surveillance, and modelling. Lancet Neurol 2009;8:345-54.
- The World Health Organization MONICA Project (monitoring trends and determinants in cardiovascular disease): A major international collaboration. WHO MONICA Project Principal Investigators. J Clin Epidemiol 1988;41:105-14.
- 8. Easton JD, Saver JL, Albers GW, Alberts MJ, Chaturvedi S, Feldmann E, et al.; American Heart Association; American Stroke Association Stroke Council; Council on Cardiovascular Surgery and Anesthesia; Council on Cardiovascular Radiology and Intervention; Council on Cardiovascular Nursing; Interdisciplinary Council on Peripheral Vascular Disease. Definition and evaluation of TIA: A scientific statement for healthcare professionals from the American Heart Association/American Stroke Association Stroke Council; Council on Cardiovascular Surgery and Anesthesia; Council on Cardiovascular Surgery and Anesthesia; Council on Cardiovascular Radiology and Intervention; Council on Cardiovascular Radiology and Intervention; Council on Cardiovascular Nursing; and the Interdisciplinary Council on Peripheral Vascular Disease. The American Academy of Neurology affirms the value of this statement as an educational tool for neurologists. Stroke 2009;40:2276-93.
- Deb P, Sharma S, Hassan KM. Pathophysiologic mechanisms of acute ischemic stroke: An overview with emphasis on therapeutic significance beyond thrombolysis. Pathophysiology 2010;17: 197-218.
- Feigin VL, Lawes CM, Bennett DA, Anderson CS. Stroke epidemiology: A review of population-based studies of incidence, prevalence, and case-fatality in the late 20th century. Lancet Neurol 2003;2:43-53.
- Johnston SC, Selvin S, Gress DR. The burden, trends, and demographics of mortality from subarachnoid hemorrhage. Neurology 1998;50:1413-8.
- Feigin VL, Rinkel GJ, Lawes CM, Algra A, Bennett DA, van Gijn J, et al. Risk factors for subarachnoid hemorrhage: An updated systematic review of epidemiological studies. Stroke 2005;36: 2773-80.
- 13. Benavente O, Hart RG. Stroke: Part II. Management of acute ischemic stroke. Am Fam Physician 1999;59:2828-34 concl.
- 14. Emberson J, Lees KR, Lyden P, Blackwell L, Albers G, Bluhmki E, *et al.*; Stroke Thrombolysis Trialists' Collaborative Group. Effect of treatment delay, age, and stroke severity on the effects of intravenous thrombolysis with alteplase for acute ischaemic stroke: A meta-analysis of individual patient data from randomised trials. Lancet 2014;384:1929-35.
- 15. Jauch EC, Saver JL, Adams HP, Jr., Bruno A, Connors JJ, Demaerschalk BM, *et al.*; American Heart Association Stroke

Council; Council on Cardiovascular Nursing; Council on Peripheral Vascular Disease; Council on Clinical Cardiology. Guidelines for the early management of patients with acute ischemic stroke: A guideline for healthcare professionals from the American Heart Association/American Stroke Association. Stroke 2013;44:870-947.

- Tissue plasminogen activator for acute ischemic stroke. The National Institute of Neurological Disorders and Stroke rt-PA Stroke Study Group. N Engl J Med 1995;333:1581-7.
- 17. Albertson M, Sharma J. Stroke: Current concepts. S D Med 2014;67:455, 457-61, 463-5.
- Davalos A, Toni D, Iweins F, Lesaffre E, Bastianello S, Castillo J. Neurological deterioration in acute ischemic stroke: Potential predictors and associated factors in the European cooperative acute stroke study (ECASS) I. Stroke 1999;30:2631-6.
- Kwan J, Hand P. Early neurological deterioration in acute stroke: Clinical characteristics and impact on outcome. QJM 2006;99: 625-33.
- Siegler JE, Martin-Schild S. Early Neurological Deterioration (END) after stroke: The END depends on the definition. Int J Stroke 2011;6:211-2.
- 21. Fiorelli M, Bastianello S, von Kummer R, del Zoppo GJ, Larrue V, Lesaffre E, *et al*. Hemorrhagic transformation within 36 hours of a cerebral infarct: Relationships with early clinical deterioration and 3-month outcome in the European Cooperative Acute Stroke Study I (ECASS I) cohort. Stroke 1999;30:2280-4.
- Ovesen C, Christensen AF, Havsteen I, Krarup Hansen C, Rosenbaum S, Kurt E, *et al.* Prediction and prognostication of neurological deterioration in patients with acute ICH: A hospitalbased cohort study. BMJ Open 2015;5:e008563.
- Rimmele DL, Thomalla G. Wake-up stroke: Clinical characteristics, imaging findings, and treatment option - an update. Front Neurol 2014;5:35.
- 24. Montaner J. Stroke biomarkers: Can they help us to guide stroke thrombolysis? Drug News Perspect 2006;19:523-32.
- 25. Castellanos M, Serena J. Applicability of biomarkers in ischemic stroke. Cerebrovasc Dis 2007;24(Suppl 1):7-15.
- Senn R, Elkind MS, Montaner J, Christ-Crain M, Katan M. Potential role of blood biomarkers in the management of nontraumatic intracerebral hemorrhage. Cerebrovasc Dis 2014;38:395-409.
- 27. Maas MB, Furie KL. Molecular biomarkers in stroke diagnosis and prognosis. Biomark Med 2009;3:363-83.
- 28. Prakash R, Carmichael ST. Blood-brain barrier breakdown and neovascularization processes after stroke and traumatic brain injury. Curr Opin Neurol 2015;28:556-64.
- Belavić M, Jančić E, Mišković P, Brozović-Krijan A, Bakota B, Žunić J. Secondary stroke in patients with polytrauma and traumatic brain injury treated in an Intensive Care Unit, Karlovac General Hospital, Croatia. Injury 2015;46(Suppl 6):S31-5.
- 30. Shi H, Hu X, Leak RK, Shi Y, An C, Suenaga J, *et al.* Demyelination as a rational therapeutic target for ischemic or traumatic brain injury. Exp Neurol 2015;272:17-25.
- Buttner T, Weyers S, Postert T, Sprengelmeyer R, Kuhn W. S-100 protein: Serum marker of focal brain damage after ischemic territorial MCA infarction. Stroke 1997;28:1961-5.
- 32. Elting JW, de Jager AE, Teelken AW, Schaaf MJ, Maurits NM, van der Naalt J, *et al.* Comparison of serum S-100 protein levels following stroke and traumatic brain injury. J Neurol Sci 2000;181:104-10.
- 33. Wunderlich MT, Wallesch CW, Goertler M. Release of neurobiochemical markers of brain damage is related to the neurovascular status on admission and the site of arterial occlusion in acute ischemic stroke. J Neurol Sci 2004;227:49-53.
- 34. Jauch EC, Lindsell C, Broderick J, Fagan SC, Tilley BC, Levine SR; NINDS rt-PA Stroke Study Group. Association of serial biochemical markers with acute ischemic stroke: The National Institute of Neurological Disorders and Stroke recombinant

tissue plasminogen activator Stroke Study. Stroke 2006;37: 2508-13.

- Unden J, Strandberg K, Malm J, Campbell E, Rosengren L, Stenflo J, *et al.* Explorative investigation of biomarkers of brain damage and coagulation system activation in clinical stroke differentiation. J Neurol 2009;256:72-7.
- Ehrenreich H, Kästner A, Weissenborn K, Streeter J, Sperling S, Wang KK, et al. Circulating damage marker profiles support a neuroprotective effect of erythropoietin in ischemic stroke patients. Mol Med 2011;17:1306-10.
- Bielewicz J, Kurzepa J, Czekajska-Chehab E, Stelmasiak Z, Bartosik-Psujek H. Does serum Tau protein predict the outcome of patients with ischemic stroke? J Mol Neurosci 2011;43:241-5.
- González-García S, González-Quevedo A, Fernández-Concepción O, Peña-Sánchez M, Menéndez-Saínz C, Hernández-Díaz Z, et al. Short-term prognostic value of serum neuron specific enolase and S100B in acute stroke patients. Clin Biochem 2012;45:1302-7.
- Montaner J, Mendioroz M, Delgado P, García-Berrocoso T, Giralt D, Merino C, *et al.* Differentiating ischemic from hemorrhagic stroke using plasma biomarkers: The S100B/RAGE pathway. J Proteomics 2012;75:4758-65.
- Purrucker JC, Herrmann O, Lutsch JK, Zorn M, Schwaninger M, Bruckner T, *et al.* Serum protein S100 is a diagnostic biomarker for distinguishing posterior circulation stroke from vertigo of nonvascular causes. Eur Neurol 2014;72:278-84.
- 41. Kumar H, Lakhotia M, Pahadiya H, Singh J. To study the correlation of serum S-100 protein level with the severity of stroke and its prognostic implication. J Neurosci Rural Pract 2015;6: 326-30.
- 42. Alatas ÖD, Gürger M, Ateşçelik M, Yildiz M, Demir CF, Kalayci M, et al. Neuron-specific enolase, S100 calciumbinding protein B, and heat shock protein 70 levels in patients with intracranial hemorrhage. Medicine (Baltimore) 2015;94:e2007.
- Foerch C, Curdt I, Yan B, Dvorak F, Hermans M, Berkefeld J, et al. Serum glial fibrillary acidic protein as a biomarker for intracerebral haemorrhage in patients with acute stroke. J Neurol Neurosurg Psychiatry 2006;77:181-4.
- 44. Dvorak F, Haberer I, Sitzer M, Foerch C. Characterisation of the diagnostic window of serum glial fibrillary acidic protein for the differentiation of intracerebral haemorrhage and ischaemic stroke. Cerebrovasc Dis 2009;27:37-41.
- 45. Foerch C, Niessner M, Back T, Bauerle M, De Marchis GM, Ferbert A, *et al.* Diagnostic accuracy of plasma glial fibrillary acidic protein for differentiating intracerebral hemorrhage and cerebral ischemia in patients with symptoms of acute stroke. Clin Chem 2012;58:237-45.
- 46. Stanca DM, Mărginean IC, Sori ău O, Drago C, Mărginean M, Mure anu DF, et al. GFAP and antibodies against NMDA receptor subunit NR2 as biomarkers for acute cerebrovascular diseases. J Cell Mol Med 2015;19:2253-61.
- 47. Xiong L, Yang Y, Zhang M, Xu W. The use of serum glial fibrillary acidic protein test as a promising tool for intracerebral hemorrhage diagnosis in Chinese patients and prediction of the short-term functional outcomes. Neurol Sci 2015;36:2081-7.
- Llombart V, García-Berrocoso T, Bustamante A, Giralt D, Rodriguez-Luna D, Muchada M, et al. Plasmatic RBP4 and GFAP as biomarkers to differentiate ischemic stroke and intracerebral hemorrhage. J Neurochem 2015. [Epub ahead of print].
- 49. Wunderlich MT, Lins H, Skalej M, Wallesch CW, Goertler M. Neuron-specific enolase and tau protein as neurobiochemical markers of neuronal damage are related to early clinical course and long-term outcome in acute ischemic stroke. Clin Neurol Neurosurg 2006;108:558-63.
- 50. Bharosay A, Bharosay VV, Varma M, Saxena K, Sodani A, Saxena R. Correlation of Brain Biomarker Neuron Specific Enolase

(NSE) with degree of disability and neurological worsening in cerebrovascular stroke. Indian J Clin Biochem 2012;27:186-90.

- Zaheer S, Beg M, Rizvi I, Islam N, Ullah E, Akhtar N. Correlation between serum neuron specific enolase and functional neurological outcome in patients of acute ischemic stroke. Annals of Indian Academy of Neurology 2013;16:504-8.
- Singh HV, Pandey A, Shrivastava AK, Raizada A, Singh SK, Singh N. Prognostic value of neuron specific enolase and IL-10 in ischemic stroke and its correlation with degree of neurological deficit. Clin Chim Acta 2013;419:136-8.
- 53. Kim BJ, Kim YJ, Ahn SH, Kim NY, Kang DW, Kim JS, *et al.* The second elevation of neuron-specific enolase peak after ischemic stroke is associated with hemorrhagic transformation. J Stroke Cerebrovasc Dis 2014;23:2437-43.
- 54. Pandey A, Shrivastava AK, Saxena K. Neuron specific enolase and c-reactive protein levels in stroke and its subtypes: Correlation with degree of disability. Neurochem Res 2014;39:1426-32.
- Lu K, Xu X, Cui S, Wang F, Zhang B, Zhao Y. Serum neuron specific enolase level as a predictor of prognosis in acute ischemic stroke patients after intravenous thrombolysis. J Neurol Sci 2015;359:202-6.
- Can S, Akdur O, Yildirim A, Adam G, Cakir DU, Karaman HI. Myelin basic protein and ischemia modified albumin levels in acute ischemic stroke cases. Pak J Med Sci 2015;31:1110-4.
- 57. Hu HT, Xiao F, Yan YQ, Wen SQ, Zhang L. The prognostic value of serum tau in patients with intracerebral hemorrhage. Clin Biochem 2012;45:1320-4.
- 58. Stejskal D, Sporova L, Svestak M, Karpisek M. Determination of serum visinin like protein-1 and its potential for the diagnosis of brain injury due to the stroke: A pilot study. Biomed Pap Med Fac Univ Palacky Olomouc Czech Repub 2011;155:263-8.
- Dambinova SA, Bettermann K, Glynn T, Tews M, Olson D, Weissman JD, et al. Diagnostic potential of the NMDA receptor peptide assay for acute ischemic stroke. PloS One 2012;7:e42362.
- Baudier J, Glasser N, Gerard D. Ions binding to S100 proteins. I. Calcium- and zinc-binding properties of bovine brain S100 alpha alpha, S100a (alpha beta), and S100b (beta beta) protein: Zn2+ regulates Ca2+ binding on S100b protein. J Biol Chem 1986;261:8192-203.
- 61. Deloulme JC, Raponi E, Gentil BJ, Bertacchi N, Marks A, Labourdette G, *et al*. Nuclear expression of S100B in oligodendrocyte progenitor cells correlates with differentiation toward the oligodendroglial lineage and modulates oligodendrocytes maturation. Mol Cell Neurosci 2004;27:453-65.
- 62. Nishiyama A, Chang A, Trapp BD. NG2+ glial cells: A novel glial cell population in the adult brain. J Neuropathol Exp Neurol 1999;58:1113-24.
- Levine JM, Card JP. Light and electron microscopic localization of a cell surface antigen (NG2) in the rat cerebellum: Association with smooth protoplasmic astrocytes. J Neurosci 1987;7: 2711-20.
- 64. Donato R. S100: A multigenic family of calcium-modulated proteins of the EF-hand type with intracellular and extracellular functional roles. Int J Biochem Cell Biol 2001;33:637-68.
- Berger RP, Beers SR, Richichi R, Wiesman D, Adelson PD. Serum biomarker concentrations and outcome after pediatric traumatic brain injury. J Neurotrauma 2007;24:1793-801.
- 66. Lee JY, Lee CY, Kim HR, Lee CH, Kim HW, Kim JH. A role of serum-based neuronal and glial markers as potential predictors for distinguishing severity and related outcomes in traumatic brain injury. J Korean Neurosurg Soc 2015;58:93-100.
- Yardan T, Erenler AK, Baydin A, Aydin K, Cokluk C. Usefulness of S100B protein in neurological disorders. J Pak Med Assoc 2011;61:276-81.
- Persson L, Hårdemark HG, Gustafsson J, Rundström G, Mendel-Hartvig I, Esscher T, *et al.* S-100 protein and neuron-

specific enolase in cerebrospinal fluid and serum: Markers of cell damage in human central nervous system. Stroke 1987;18:911-8.

- 69. Koh SX, Lee JK. S100B as a marker for brain damage and blood-brain barrier disruption following exercise. Sports Med 2014;44:369-85.
- Li X, Wilder-Smith CH, Kan ME, Lu J, Cao Y, Wong RK. Combattraining stress in soldiers increases S100B, a marker of increased blood-brain-barrier permeability, and induces immune activation. Neuro Endocrinol Lett 2014;35:58-63.
- Siman R, Giovannone N, Toraskar N, Frangos S, Stein SC, Levine JM, *et al.* Evidence that a panel of neurodegeneration biomarkers predicts vasospasm, infarction, and outcome in aneurysmal subarachnoid hemorrhage. PloS One 2011; 6:e28938.
- 72. Pearlman DM, Brown JR, MacKenzie TA, Hernandez F Jr, Najjar S. Blood levels of S-100 calcium-binding protein B, high-sensitivity C-reactive protein, and interleukin-6 for changes in depressive symptom severity after coronary artery bypass grafting: Prospective cohort nested within a randomized, controlled trial. PloS One 2014;9:e11110.
- Abraha HD, Butterworth RJ, Bath PM, Wassif WS, Garthwaite J, Sherwood RA. Serum S-100 protein, relationship to clinical outcome in acute stroke. Ann Clin Biochem 1997;34:546-50.
- Hayakata T, Shiozaki T, Tasaki O, Ikegawa H, Inoue Y, Toshiyuki F, et al. Changes in CSF S100B and cytokine concentrations in early-phase severe traumatic brain injury. Shock 2004;22:102-7.
- Raabe A, Kopetsch O, Woszczyk A, Lang J, Gerlach R, Zimmermann M, *et al.* Serum S-100B protein as a molecular marker in severe traumatic brain injury. Restor Neurol Neurosci 2003;21:159-69.
- Nylen K, Ost M, Csajbok LZ, Nilsson I, Hall C, Blennow K, *et al.* Serum levels of S100B, S100A1B and S100BB are all related to outcome after severe traumatic brain injury. Acta Neurochir (Wien) 2008;150:221-7.
- Thelin EP, Johannesson L, Nelson D, Bellander BM. S100B is an important outcome predictor in traumatic brain injury. J Neurotrauma 2013;30:519-28.
- Berger RP, Pierce MC, Wisniewski SR, Adelson PD, Kochanek PM. Serum S100B concentrations are increased after closed head injury in children: A preliminary study. J Neurotrauma 2002;19:1405-9.
- Wolf H, Frantal S, Pajenda G, Leitgeb J, Sarahrudi K, Hajdu S. Analysis of S100 calcium binding protein B serum levels in different types of traumatic intracranial lesions. J Neurotrauma 2015;32:23-7.
- Bohmer AE, Oses JP, Schmidt AP, Peron CS, Krebs CL, Oppitz PP, et al. Neuron-specific enolase, S100B, and glial fibrillary acidic protein levels as outcome predictors in patients with severe traumatic brain injury. Neurosurgery 2011;68:1624-31.
- Papa L, Silvestri S, Brophy GM, Giordano P, Falk JL, Braga CF, et al. GFAP out-performs S100 in detecting traumatic intracranial lesions on computed tomography in trauma patients with mild traumatic brain injury and those with extracranial lesions. J Neurotrauma 2014;31:1815-22.
- Papa L, Robinson G, Oli M, Pineda J, Demery J, Brophy G, *et al.* Use of biomarkers for diagnosis and management of traumatic brain injury patients. Expert Opin Med Diagn 2008;2:937-45.
- Vos PE, Jacobs B, Andriessen TM, Lamers KJ, Borm GF, Beems T, et al. GFAP and S100B are biomarkers of traumatic brain injury: An observational cohort study. Neurology 2010;75: 1786-93.
- Goyal A, Failla MD, Niyonkuru C, Amin K, Fabio A, Berger RP, et al. S100b as a prognostic biomarker in outcome prediction for patients with severe traumatic brain injury. J Neurotrauma 2013;30:946-57.
- Bamford J, Sandercock P, Jones L, Warlow C. The natural history of lacunar infarction: The Oxfordshire Community Stroke Project. Stroke 1987;18:545-51.

- Xiao L, Sun W, Lan W, Xiong Y, Duan Z, Zhang Z, et al. Correlation between cerebral microbleeds and S100B/RAGE in acute lacunar stroke patients. J Neurol Sci 2014;340:208-12.
- Eng LF. Glial fibrillary acidic protein (GFAP): The major protein of glial intermediate filaments in differentiated astrocytes. J Neuroimmunol 1985;8:203-14.
- Schiff L, Hadker N, Weiser S, Rausch C. A literature review of the feasibility of glial fibrillary acidic protein as a biomarker for stroke and traumatic brain injury. Mol Diagn Ther 2012;16:79-92.
- 89. Foerch C, Pfeilschifter W, Zeiner P, Brunkhorst R. Glial fibrillary acidic protein in patients with symptoms of acute stroke: Diagnostic marker of cerebral hemorrhage. Nervenarzt 2014;85:982-9.
- 90. Czeiter E, Mondello S, Kovacs N, Sandor J, Gabrielli A, Schmid K, *et al.* Brain injury biomarkers may improve the predictive power of the IMPACT outcome calculator. J Neurotrauma 2012;29:1770-8.
- 91. Okonkwo DO, Yue JK, Puccio AM, Panczykowski DM, Inoue T, McMahon PJ, et al.; Transforming Research and Clinical Knowledge in Traumatic Brain Injury (TRACK-TBI) Investigators. GFAP-BDP as an acute diagnostic marker in traumatic brain injury: Results from the prospective transforming research and clinical knowledge in traumatic brain injury study. J Neurotrauma 2013;30:1490-7.
- 92. Honda M, Tsuruta R, Kaneko T, Kasaoka S, Yagi T, Todani M, *et al.* Serum glial fibrillary acidic protein is a highly specific biomarker for traumatic brain injury in humans compared with S-100B and neuron-specific enolase. J Trauma 2010;69:104-9.
- 93. Papa L, Lewis LM, Falk JL, Zhang Z, Silvestri S, Giordano P, et al. Elevated levels of serum glial fibrillary acidic protein breakdown products in mild and moderate traumatic brain injury are associated with intracranial lesions and neurosurgical intervention. Ann Emerg Med 2011;59:471-83.
- 94. Welch RD, Ayaz SI, Lewis LM, Unden J, Chen JY, Mika VH, et al. Ability of serum glial fibrillary acidic protein, Ubiquitin C-Terminal Hydrolase-L1, and S100B to differentiate normal and abnormal head computed tomography findings in patients with suspected mild or moderate traumatic brain injury. J Neurotrauma 2016;33:203-14.
- 95. McMahon PJ, Panczykowski DM, Yue JK, Puccio AM, Inoue T, Sorani MD, et al.; TRACK-TBI Investigators. Measurement of the glial fibrillary acidic protein and its breakdown products GFAP-BDP biomarker for the detection of traumatic brain injury compared to computed tomography and magnetic resonance imaging. J Neurotrauma 2015;32:527-33.
- Mannix R, Eisenberg M, Berry M, Meehan WP 3rd, Hayes RL. Serum biomarkers predict acute symptom burden in children after concussion: A preliminary study. J Neurotrauma 2014;31:1072-5.
- 97. Fraser DD, Close TE, Rose KL, Ward R, Mehl M, Farrell C, *et al.*; Canadian Critical Care Translational Biology Group. Severe traumatic brain injury in children elevates glial fibrillary acidic protein in cerebrospinal fluid and serum. Pediatr Crit Care Med 2011;12:319-24.
- Hayes RL, Mondello S, Wang K. Glial fibrillary acidic protein: A promising biomarker in pediatric brain injury. Pediatr Crit Care Med 2011;12:603-4.
- Huang XJ, Glushakova O, Mondello S, Van K, Hayes RL, Lyeth BG. Acute temporal profiles of serum levels of UCH-L1 and GFAP and relationships to neuronal and astroglial pathology following traumatic brain injury in rats. J Neurotrauma 2015;32:1179-89.
- 100. Takala RS, Posti JP, Runtti H, Newcombe VF, Outtrim J, Katila AJ, *et al.* GFAP and UCH-L1 as outcome predictors in traumatic brain injury. World Neurosurg 2015. [Epub ahead of print].
- 101. Liu MC, Akinyi L, Scharf D, Mo J, Larner SF, Muller U, *et al.* Ubiquitin C-terminal hydrolase-L1 as a biomarker for ischemic and traumatic brain injury in rats. Eur J Neurosci 2010;31: 722-32.

- 102. Ren C, Zoltewicz S, Guingab-Cagmat J, Anagli J, Gao M, Hafeez A, et al. Different expression of ubiquitin C-terminal hydrolase-L1 and αII-spectrin in ischemic and hemorrhagic stroke: Potential biomarkers in diagnosis. Brain Res 2013;1540:84-91.
- 103. Zhang YP, Zhu YB, Duan DD, Fan XM, He Y, Su JW, et al. Serum UCH-L1 as a novel biomarker to predict neuronal apoptosis following deep hypothermic circulatory arrest. Int J Med Sci 2015;12:576-82.
- 104. Papa L, Akinyi L, Liu MC, Pineda JA, Tepas JJ 3rd, Oli MW, et al. Ubiquitin C-terminal hydrolase is a novel biomarker in humans for severe traumatic brain injury. Crit Care Med 2010;38:138-44.
- 105. Diaz-Arrastia R, Wang KK, Papa L, Sorani MD, Yue JK, Puccio AM, et al.; TRACK-TBI Investigators. Acute biomarkers of traumatic brain injury: Relationship between plasma levels of ubiquitin C-terminal hydrolase-L1 and glial fibrillary acidic protein. J Neurotrauma 2014;31:19-25.
- 106. Berger RP, Hayes RL, Richichi R, Beers SR, Wang KK. Serum concentrations of ubiquitin C-terminal hydrolase-L1 and αII-spectrin breakdown product 145 kDa correlate with outcome after pediatric TBI. J Neurotrauma 2012;29:162-7.
- 107. Zanier ER, Refai D, Zipfel GJ, Zoerle T, Longhi L, Esparza TJ, et al. Neurofilament light chain levels in ventricular cerebrospinal fluid after acute aneurysmal subarachnoid haemorrhage. J Neurol Neurosurg Psychiatry 2011;82:157-9.
- 108. Brophy GM, Mondello S, Papa L, Robicsek SA, Gabrielli A, Tepas J 3rd, *et al.* Biokinetic analysis of ubiquitin C-terminal hydrolase-L1 (UCH-L1) in severe traumatic brain injury patient biofluids. J Neurotrauma 2011;28:861-70.
- 109. Carr W, Yarnell AM, Ong R, Walilko T, Kamimori GH, da Silva U, et al. Ubiquitin carboxy-terminal hydrolase-l1 as a serum neurotrauma biomarker for exposure to occupational low-level blast. Front Neurol 2015;6:49.
- 110. Lewis SB, Wolper R, Chi YY, Miralia L, Wang Y, Yang C, et al. Identification and preliminary characterization of ubiquitin C terminal hydrolase 1 (UCHL1) as a biomarker of neuronal loss in aneurysmal subarachnoid hemorrhage. J Neurosci Res 2010;88:1475-84.
- 111. Ehrenreich H, Weissenborn K, Prange H, Schneider D, Weimar C, Wartenberg K, *et al.*; EPO Stroke Trial Group. Recombinant human erythropoietin in the treatment of acute ischemic stroke. Stroke 2009;40:e647-56.
- 112. Li Y, Sun Y, Li J, Wang Z, Lin Y, Tang L, *et al*. Changes of ubiquitin C-terminal hydrolase-L1 levels in serum and urine of patients with white matter lesions. J Neurol Sci 2015;357:215-21.
- 113. Cunningham RT, Young IS, Winder J, O'Kane MJ, McKinstry S, Johnston CF, *et al*. Serum neurone specific enolase (NSE) levels as an indicator of neuronal damage in patients with cerebral infarction. Eur J Clin Invest 1991;21:497-500.
- 114. Steinberg R, Gueniau C, Scarna H, Keller A, Worcel M, Pujol JF. Experimental brain ischemia: Neuron-specific enolase level in cerebrospinal fluid as an index of neuronal damage. J Neurochem 1984;43:19-24.
- 115. Hårdemark HG, Persson L, Bolander HG, Hillered L, Olsson Y, Påhlman S. Neuron-specific enolase is a marker of cerebral ischemia and infarct size in rat cerebrospinal fluid. Stroke 1988;19:1140-4.
- 116. Hårdemark HG, Ericsson N, Kotwica Z, Rundström G, Mendel-Hartvig I, Olsson Y, *et al.* S-100 protein and neuron-specific enolase in CSF after experimental traumatic or focal ischemic brain damage. J Neurosurg 1989;71:727-31.
- 117. Hatfield RH, McKernan RM. CSF neuron-specific enolase as a quantitative marker of neuronal damage in a rat stroke model. Brain Res 1992;577:249-52.
- 118. Barone FC, Clark RK, Price WJ, White RF, Feuerstein GZ, Storer BL, *et al.* Neuron-specific enolase increases in cerebral and systemic circulation following focal ischemia. Brain Res 1993;623:77-82.

- Missler U, Wiesmann M, Friedrich C, Kaps M. S-100 protein and neuron-specific enolase concentrations in blood as indicators of infarction volume and prognosis in acute ischemic stroke. Stroke 1997;28:1956-60.
- Stevens H, Jakobs C, de Jager AE, Cunningham RT, Korf J. Neuronespecific enolase and N-acetyl-aspartate as potential peripheral markers of ischaemic stroke. Eur J Clin Invest 1999;29:6-11.
- 121. Anand N, Stead LG. Neuron-specific enolase as a marker for acute ischemic stroke: A systematic review. Cerebrovasc Dis 2005;20:213-9.
- 122. Fassbender K, Schmidt R, Schreiner A, Fatar M, Mühlhauser F, Daffertshofer M, *et al.* Leakage of brain-originated proteins in peripheral blood: Temporal profile and diagnostic value in early ischemic stroke. J Neurol Sci 1997;148:101-5.
- 123. Casmiro M, Maitan S, De Pasquale F, Cova V, Scarpa E, Vignatelli L; NSE Study Group. Cerebrospinal fluid and serum neuron-specific enolase concentrations in a normal population. Eur J Neurol 2005;12:369-74.
- 124. Dash PK, Zhao J, Hergenroeder G, Moore AN. Biomarkers for the diagnosis, prognosis, and evaluation of treatment efficacy for traumatic brain injury. Neurotherapeutics 2010;7:100-14.
- 125. Zhang Z, Larner SF, Liu MC, Zheng W, Hayes RL, Wang KK. Multiple alphaII-spectrin breakdown products distinguish calpain and caspase dominated necrotic and apoptotic cell death pathways. Apoptosis 2009;14:1289-98.
- 126. Brophy GM, Pineda JA, Papa L, Lewis SB, Valadka AB, Hannay HJ, et al. alphaII-Spectrin breakdown product cerebrospinal fluid exposure metrics suggest differences in cellular injury mechanisms after severe traumatic brain injury. J Neurotrauma 2009;26:471-9.
- 127. Pineda JA, Lewis SB, Valadka AB, Papa L, Hannay HJ, Heaton SC, et al. Clinical significance of alphaII-spectrin breakdown products in cerebrospinal fluid after severe traumatic brain injury. J Neurotrauma 2007;24:354-66.
- 128. Lewis SB, Velat GJ, Miralia L, Papa L, Aikman JM, Wolper RA, *et al.* Alpha-II spectrin breakdown products in aneurysmal subarachnoid hemorrhage: A novel biomarker of proteolytic injury. J Neurosurg 2007;107:792-6.
- 129. Mondello S, Robicsek SA, Gabrielli A, Brophy GM, Papa L, Tepas J, *et al.* αII-spectrin breakdown products (SBDPs): Diagnosis and outcome in severe traumatic brain injury patients. J Neurotrauma 2010;27:1203-13.
- 130. Pike BR, Flint J, Dave JR, Lu XC, Wang KK, Tortella FC, *et al.* Accumulation of calpain and caspase-3 proteolytic fragments of brain-derived alphaII-spectrin in cerebral spinal fluid after middle cerebral artery occlusion in rats. J Cereb Blood Flow Metab 2004;24:98-106.
- Zhang C, Siman R, Xu YA, Mills AM, Frederick JR, Neumar RW. Comparison of calpain and caspase activities in the adult rat brain after transient forebrain ischemia. Neurobiol Dis 2002;10:289-05.
- Cohen SR, Herndon RM, McKhann GM. Myelin basic protein in cerebrospinal fluid as an indicator of active demyelination. Trans Am Neurol Assoc 1976;101:45-7.
- 133. Thomas DG, Palfreyman JW, Ratcliffe JG. Serum-myelin-basicprotein assay in diagnosis and prognosis of patients with head injury. Lancet 1978;1:113-5.
- 134. Matias-Guiu J, Martinez-Vazquez J, Ruibal A, Colomer R, Boada M, Codina A. Myelin basic protein and creatine kinase BB isoenzyme as CSF markers of intracranial tumors and stroke. Acta Neurol Scand 1986;73:461-5.
- 135. Strand T, Alling C, Karlsson B, Karlsson I, Winblad B. Brain and plasma proteins in spinal fluid as markers for brain damage and severity of stroke. Stroke 1984;15:138-44.
- 136. Aurell A, Rosengren LE, Karlsson B, Olsson JE, Zbornikova V, Haglid KG. Determination of S-100 and glial fibrillary acidic protein concentrations in cerebrospinal fluid after brain infarction. Stroke 1991;22:1254-8.

- 137. Hill MD, Jackowski G, Bayer N, Lawrence M, Jaeschke R. Biochemical markers in acute ischemic stroke. CMAJ 2000;162: 1139-40.
- 138. Hjalmarsson C, Bjerke M, Andersson B, Blennow K, Zetterberg H, Aberg ND, et al. Neuronal and glia-related biomarkers in cerebrospinal fluid of patients with acute ischemic stroke. J Cent Nerv Syst Dis 2014;6:51-8.
- Shibata D, Cain K, Tanzi P, Zierath D, Becker K. Myelin basic protein autoantibodies, white matter disease and stroke outcome. J Neuroimmunol 2012;252:106-12.
- 140. Zierath D, Kunze A, Fecteau L, Becker K. Promiscuity of autoimmune responses to MBP after stroke. J Neuroimmunol 2015;285:101-5.
- 141. Frappier T, Stetzkowski-Marden F, Pradel LA. Interaction domains of neurofilament light chain and brain spectrin. Biochem J 1991;275:521-7.
- 142. Van Geel WJ, Rosengren LE, Verbeek MM. An enzyme immunoassay to quantify neurofilament light chain in cerebrospinal fluid. J Immunol Methods 2005;296:179-85.
- 143. Rosengren LE, Karlsson JE, Karlsson JO, Persson LI, Wikkelsø C. Patients with amyotrophic lateral sclerosis and other neurodegenerative diseases have increased levels of neurofilament protein in CSF. J Neurochem 1996;67:2013-8.
- 144. Traenka C, Disanto G, Seiffge DJ, Gensicke H, Hert L, Grond-Ginsbach C, *et al*. Serum neurofilament light chain levels are associated with clinical characteristics and outcome in patients with cervical artery dissection. Cerebrovasc Dis 2015;40:222-7.
- 145. Nylen K, Csajbok LZ, Ost M, Rashid A, Karlsson JE, Blennow K, et al. CSF -neurofilament correlates with outcome after aneurysmal subarachnoid hemorrhage. Neurosci Lett 2006;404:132-6.
- Avila J, Lucas JJ, Perez M, Hernandez F. Role of tau protein in both physiological and pathological conditions. Physiol Rev 2004;84:361-84.
- 147. Grundke-Iqbal I, Iqbal K, Tung YC, Quinlan M, Wisniewski HM, Binder LI. Abnormal phosphorylation of the microtubuleassociated protein tau (tau) in Alzheimer cytoskeletal pathology. Proc Natl Acad Sci U S A 1986;83:4913-7.
- 148. Grundke-Iqbal I, Iqbal K, Quinlan M, Tung YC, Zaidi MS, Wisniewski HM. Microtubule-associated protein tau. A component of Alzheimer paired helical filaments. J Biol Chem 1986;261:6084-9.
- 149. Andreasen N, Minthon L, Clarberg A, Davidsson P, Gottfries J, Vanmechelen E, et al. Sensitivity, specificity, and stability of CSF-tau in AD in a community-based patient sample. Neurology 1999;53:1488-94.
- 150. Hampel H, Blennow K, Shaw LM, Hoessler YC, Zetterberg H, Trojanowski JQ. Total and phosphorylated tau protein as biological markers of Alzheimer's disease. Exp Gerontol 2010;45:30-40.
- 151. Hesse C, Rosengren L, Vanmechelen E, Vanderstichele H, Jensen C, Davidsson P, *et al*. Cerebrospinal fluid markers for Alzheimer's disease evaluated after acute ischemic stroke. J Alzheimers Dis 2000;2:199-206.
- 152. Hesse C, Rosengren L, Andreasen N, Davidsson P, Vanderstichele H, Vanmechelen E, *et al.* Transient increase in total tau but not phospho-tau in human cerebrospinal fluid after acute stroke. Neurosci Lett 2001;297:187-90.
- 153. Franz G, Beer R, Kampfl A, Engelhardt K, Schmutzhard E, Ulmer H, *et al.* Amyloid beta 1-42 and tau in cerebrospinal fluid after severe traumatic brain injury. Neurology 2003;60: 1457-61.
- 154. Ost M, Nylén K, Csajbok L, Ohrfelt AO, Tullberg M, Wikkelsö C, et al. Initial CSF total tau correlates with 1-year outcome in patients with traumatic brain injury. Neurology 2006;67:1600-4.
- 155. Bulut M, Koksal O, Dogan S, Bolca N, Ozguc H, Korfali E, *et al.* Tau protein as a serum marker of brain damage in mild traumatic brain injury: Preliminary results. Adv Ther 2006;23:12-22.

- 156. Kavalci C, Pekdemir M, Durukan P, Ilhan N, Yildiz M, Serhatlioglu S, *et al*. The value of serum tau protein for the diagnosis of intracranial injury in minor head trauma. Am J Emerg Med 2007;25:391-5.
- 157. Liliang PC, Liang CL, Weng HC, Lu K, Wang KW, Chen HJ, *et al.* Tau proteins in serum predict outcome after severe traumatic brain injury. J Surg Res 2010;160:302-7.
- 158. Braunewell KH, Gundelfinger ED. Intracellular neuronal calcium sensor proteins: A family of EF-hand calcium-binding proteins in search of a function. Cell Tissue Res 1999;295:1-12.
- 159. Kuno T, Kajimoto Y, Hashimoto T, Mukai H, Shirai Y, Saheki S, et al. cDNA cloning of a neural visinin-like Ca(2+)-binding protein. Biochem Biophys Res Commun 1992;184:1219-25.
- 160. Polymeropoulos MH, Ide S, Soares MB, Lennon GG. Sequence characterization and genetic mapping of the human VSNL1 gene, a homologue of the rat visinin-like peptide RNVP1. Genomics 1995;29:273-5.
- 161. Dai FF, Zhang Y, Kang Y, Wang Q, Gaisano HY, Braunewell KH, et al. The neuronal Ca2+ sensor protein visinin-like protein-1 is expressed in pancreatic islets and regulates insulin secretion. J Biol Chem 2006;281:21942-53.
- 162. Laterza OF, Modur VR, Crimmins DL, Olander JV, Landt Y, Lee JM, *et al*. Identification of novel brain biomarkers. Clin Chem 2006;52:1713-21.
- 163. Tarawneh R, D'Angelo G, Macy E, Xiong C, Carter D, Cairns NJ, et al. Visinin-like protein-1: Diagnostic and prognostic biomarker in Alzheimer disease. Ann Neurol 2011;70:274-85.
- Tarawneh R, Lee JM, Ladenson JH, Morris JC, Holtzman DM. CSF VILIP-1 predicts rates of cognitive decline in early Alzheimer disease. Neurology 2012;78:709-19.
- 165. Furukawa H, Singh SK, Mancusso R, Gouaux E. Subunit arrangement and function in NMDA receptors. Nature 2005;438:185-92.
- 166. Sharp CD, Fowler M, Jackson TH 4th, Houghton J, Warren A, Nanda A, et al. Human neuroepithelial cells express NMDA receptors. BMC Neurosci 2003;4:28.
- 167. Karadottir R, Cavelier P, Bergersen LH, Attwell D. NMDA receptors are expressed in oligodendrocytes and activated in ischaemia. Nature 2005;438:1162-6.
- 168. Del Valle-Pinero AY, Suckow SK, Zhou Q, Perez FM, Verne GN, Caudle RM. Expression of the N-methyl-D-aspartate receptor NR1 splice variants and NR2 subunit subtypes in the rat colon. Neuroscience 2007;147:164-73.
- Burns GA, Stephens KE, Benson JA. Expression of mRNA for the N-methyl-D-aspartate (NMDAR1) receptor by the enteric neurons of the rat. Neurosci Lett 1994;170:87-90.
- Gappoeva MU, Izykenova GA, Granstrem OK, Dambinova SA. Expression of NMDA neuroreceptors in experimental ischemia. Biochemistry (Mosc) 2003;68:696-702.
- 171. Gascon S, Deogracias R, Sobrado M, Roda JM, Renart J, Rodríguez-Peña A, *et al.* Transcription of the NR1 subunit of the N-methyl-D-aspartate receptor is down-regulated by excitotoxic stimulation and cerebral ischemia. J Biol Chem 2005;280:35018-27.
- 172. Gascon S, Sobrado M, Roda JM, Rodríguez-Peña A, Díaz-Guerra M. Excitotoxicity and focal cerebral ischemia induce truncation of the NR2A and NR2B subunits of the NMDA receptor and cleavage of the scaffolding protein PSD-95. Mol Psychiatry 2008;13:99-114.
- 173. Dong YN, Waxman EA, Lynch DR. Interactions of postsynaptic density-95 and the NMDA receptor 2 subunit control calpainmediated cleavage of the NMDA receptor. J Neurosci 2004;24:11035-45.
- 174. Dambinova SA, Khounteev GA, Izykenova GA, Zavolokov IG, Ilyukhina AY, Skoromets AA. Blood test detecting autoantibodies to N-methyl-D-aspartate neuroreceptors for evaluation of patients with transient ischemic attack and stroke. Clin Chem 2003;49: 1752-62.

- 175. Weissman JD, Khunteev GA, Heath R, Dambinova SA. NR2 antibodies: Risk assessment of transient ischemic attack (TIA)/stroke in patients with history of isolated and multiple cerebrovascular events. J Neurol Sci 2011;300:97-102.
- 176. Guttmann RP, Sokol S, Baker DL, Simpkins KL, Dong Y, Lynch DR. Proteolysis of the N-methyl-d-aspartate receptor by calpain *in situ*. J Pharmacol Exp Ther 2002;302:1023-30.
- 177. Herrmann M, Vos P, Wunderlich MT, de Bruijn CH, Lamers KJ. Release of glial tissue-specific proteins after acute stroke: A

comparative analysis of serum concentrations of protein S-100B and glial fibrillary acidic protein. Stroke 2000;31:2670-7.

- 178. Zhang H, Kang T, Li L, Zhang J. Electroacupuncture reduces hemiplegia following acute middle cerebral artery infarction with alteration of serum NSE, S-100B and endothelin. Curr Neurovasc Res 2013;10:216-21.
- 179. Sun Y, Qin Q, Shang YJ, Fang CP, Zhang WW, Gu ML, et al. The accuracy of glial fibrillary acidic protein in acute stroke differential diagnosis: A meta-analysis. Scand J Clin Lab Invest 2013;73:601-6.