

Complete Mitochondrial Genome Sequences of Korean *Phytophthora infestans* Isolates and Comparative Analysis of Mitochondrial Haplotypes

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Potato late blight caused by *Phytophthora infestans* is a destructive disease in Korea. To elucidate the genomic variation of the mitochondrial (mt) genome, we assembled its complete mt genome and compared its sequence among different haplotypes. The mt genome sequences of four Korean *P. infestans* isolates were revealed by Illumina HiSeq. The size of the circular mt genome of the four major genotypes, KR_1_A1, KR_2_A2, SIB-1, and US-11, was 39,872, 39,836, 39,872, and 39,840 bp, respectively. All genotypes contained the same 61 genes in the same order, comprising two RNA-encoding genes, 16 ribosomal genes, 25 transfer RNA, 17 genes encoding electron transport and ATP synthesis, 11 open reading frames of unknown function, and one protein

import-related gene, *tatC*. The coding region comprised 91% of the genome, and GC content was 22.3%. The haplotypes were further analyzed based on sequence polymorphism at two hypervariable regions (HVRi), carrying a 2 kb insertion/deletion sequence, and HVRii, carrying 36 bp variable number tandem repeats (VNTRs). All four genotypes carried the 2 kb insertion/deletion sequence in HVRi, whereas HVRii had two VNTRs in KR_1_A1 and SIB-1 but three VNTRs in US-11 and KR_2_A2. Minimal spanning network and phylogenetic analysis based on 5,814 bp of mtDNA sequences from five loci, KR_1_A1 and SIB-1 were classified as IIa-6 haplotype, and isolates KR_1_A2 and US-11 as haplotypes IIa-5 and IIb-2, respectively. mtDNA sequences of KR_1_A1 and SIB-1 shared 100% sequence identity, and both were 99.9% similar to those of KR_2_A2 and US-11.

Keywords : late blight, minimal spanning network, mitochondrial genome, phylogeny, *Phytophthora infestans*

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The *Phytophthora* genus comprises about 120 pathogens causing late blight disease to major vegetable crops such as pepper, tomato, and potato. Of those, 34 *Phytophthora* species have been identified as major plant pathogens in Korea between 1900 and 2000 (Hyun and Choi, 2014). The

oomycete of *P. infestans* is causing one of the most devastating diseases in potato worldwide, including Korea (Kim et al., 2014). The disease is very infectious and can spread throughout the field within one week, leading to severe yield losses (Fry, 2008). This requires timely application of proper disease control measures at the early incidence stage to minimize crop infestation. The rapid evolution of *P. infestans* led to the prevalence of the virulent A2 mating type. The resistance of new *P. infestans* genotypes to fungicides has rendered difficult disease control challenging (Arora et al., 2014). To develop more effective genetic and chemical disease control methods, the population structure of *P. infestans*, the genetics of both mating types, virulence factors, and fungicide response of each strain have been extensively studied (Cooke and Lees, 2004; Göre et al., 2019; Rekad et al., 2017). Application of molecular techniques such as mitochondrial (mt) DNA sequence haplotyping, restriction fragment length polymorphisms (RFLPs), amplified fragment length polymorphisms, and simple sequence repeat (SSR) accelerated our understanding of the genetics and pathology of *P. infestans* (Drenth et al., 1993, May and Ristaino, 2004, Van der Lee et al., 1997). Carter et al. (1990) used RFLP technique to identify mt haplotypes in relation to mating types. Griffith and Shaw (1998) modified the RFLP method by adopting polymerase chain reaction (PCR) followed by restriction enzyme digestion. By using multiple restriction enzyme combinations, Martin and Tooley (2004) were able to design a PCR-RFLP method applicable to intra- and interspecific mtDNA haplotyping, which they used to calculate genetic relationship within 152 isolates of 31 *Phytophthora* species. This approach has been widely implemented for haplotype identification of regional *P. infestans* strains in South America (Cárdenas et al., 2011), Africa (Shimelash et al., 2016), and Asia (Guo et al., 2010). Mitochondrial haplotypes of *P. infestans* were used to track the origin, migration, and genetic diversity of global *Phytophthora* species (Gómez-Alpizar et al., 2007, Goss et al., 2014; May and Ristaino, 2004; Peters et al., 2014). Later, sequencing of the whole mt genome identified two hypervariable regions (HVRs): HVRi with 1,885 bp no insertion/deletion between *trnY* and *rns* genes, which were used to differentiate type I and II; and HVRii, carrying one, two, or three copies of 36 bp variable number of tandem repeats (VNTRs) between *cox2* and *trnW* genes, which were used to further classify mtDNA as R₁, R₂, and R₃ haplotypes. By this new PCR approach developed by Marin and Tooley (2004), the previously described four mtDNA haplotypes Ia, Ib, IIa, and IIb were reclassified into five types, IR1, IR2, IR3, IIR2, and IIR3 (Yang et al., 2013).

Genetic and pathological evidence supports the intercontinental migration of *P. infestans* further to Asia and displacement of previously dominant haplotypes by new types. Koh et al. (1994) reported the presence of US-1 (mating type A1) and JP-1 (mating type A2) in East Asian countries. The US-1 is found worldwide, including in China, Japan, Korea, and the Philippines, whereas JP-1 is reported only for Korea and Japan. Gotoh et al. (2005) extended their survey to wider Asian regions and discovered 14 new *P. infestans* haplotypes. Two genotypes, JP-1 and KR-1, were identified in Korea, with the latter being specific to Korea. Zhang et al. (2006) conducted extensive analysis to reveal genetic diversity of *P. infestans* populations in South Korea. They collected 367 isolates from major potato production locations between 2002 and 2004 and identified four genotypes, JP-1, US-17, US-14, and KR-1, based on allozyme patterns and mating types; JP-1 was the most frequent type across all locations and years. At least four genotypes, KR_1_A1, KR_2_A2, US-11, and SIB-1, were identified in Korea by 12 SSR markers (Choi et al., 2020).

However, a few reports exist on complete mt genome sequences of different *P. infestans* isolates (Kubo et al., 2011; Lassiter et al., 2015; Patil et al., 2017) and there are no reports on Korean *P. infestans* isolates. In the present study, we reported whole mt genome sequences of four Korean *P. infestans* isolates and haplotypes, and conducted a comparative analysis using previously reported mt genome sequences to delineate genomic diversity of this species in Korea.

Materials and Methods

Phytophthora infestans isolates and DNA preparation.

Four *P. infestans* isolates, Kpi16-9, Kpi15-14, Kpi15-3, Kpi15-10, representing four different genotypes revealed by 12 SSR markers (Choi et al., 2020) were selected for mtDNA sequence analysis (Table 1). Single isolates were collected after cultivation on V8 fungal culture medium (8.5 g agar, β -sitosterol 0.025 g, 450 ml distilled water, 50 ml V-8 juice, 0.75 g calcium carbonate). The isolates were grown in pea liquid medium (120 g of frozen peas, 1 liter of distilled water) at 18°C with shaking at 160 rpm for 2 weeks. Mycelia were harvested from pea liquid media and washed with distilled water to remove the liquid medium. Mycelia were dried by blotting with sterilized paper towels and freeze-dried in 1.5 ml tubes before storing at -80°C. Total genomic DNA of *P. infestans* were extracted from 50 mg freeze-dried mycelia using a Fungi/Yeast Genomic DNA isolation kit (Norgen Corp., Thorold, ON, Canada)

Table 1. The list of Korean *Phytophthora infestans* isolates for complete mitochondrial genome sequencing

Genotypes ^a	Isolates	Location (latitude, longitude)	Host	Mating type	Response to fungicides	
					Metalaxyl	Dimethomorph
KR_1_A1	Kpi16-9	Pyeongchang (36.7; 128.7)	Potato	A1	Susceptible	Susceptible
US-11	Kpi15-14	Boseong (37.7; 127.1)	Potato	A1	Susceptible	Susceptible
KR_2_A2	Kpi15-3	Miryang (35.4; 128.8)	Potato	A2	Resistances	Susceptible
SIB-1	Kpi15-10	Pyeongchang (35.4; 127.5)	Potato	A1	Susceptible	Susceptible

^aGenotypes were defined with 12 simple sequence repeats markers by Choi et al. (2020).

following manufacturer's instructions.

Mitochondrial genome assembly. The next-generation sequencing (NGS) libraries were constructed using an Illumina Paired-End DNA library kit (Illumina, San Diego, CA, USA) with insert size of ~470 bp. The paired-end (PE) reads were sequenced in sets of approximately 8.0 Gbp using an Illumina HiSeq 4000 platform by Macrogen (Seoul, Korea). The overall *de novo* assembly method was followed by dnaLCW methods (Cho et al., 2015). The low-quality reads (less than Phred score of 20) were filtered using quality_trim script (CLC Assembly Cell package,

version 4.2.1, Qiagen, Aarhus, Denmark). Then, high-quality PE reads were implemented in the *de novo* assembly by *clc_novo_assemble* tool with a distance between the forward start and reverse end reads controlled from 300 to 700 bp, minimum length fraction of read overlap of 0.5, and sequence similarity of 0.8 (CLC Assembly Cell package, version 4.2.1, Qiagen). The mt putative contigs were selected by *nucmer* program in MUMmer package (<http://mummer.sourceforge.net/>) and compared with *P. infestans* (AY898627) mt genome sequence as a reference. The ordering of selected contigs was based on the reference genome. Mapping of the contigs was confirmed by *clc_ref_*

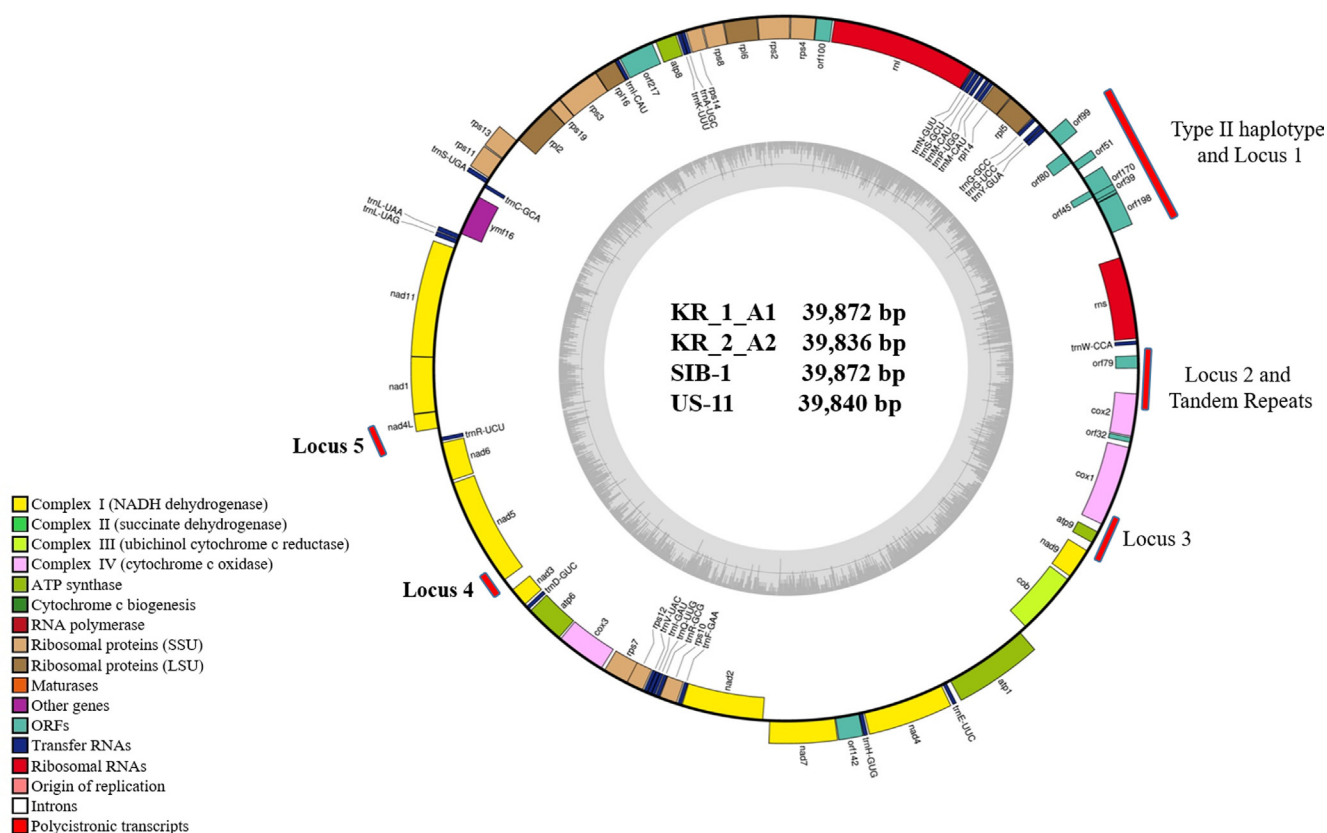


Fig. 1. Mitochondrial genome map of four Korean *Phytophthora infestans* isolates. Five mitochondrial haplotypes in this study were shown in red fill rectangular in outer circle map.

assemble tool in CLC Assembly Cell package (Qiagen). Mapping parameters were the same as those used for *de novo* assembly.

Mitochondrial genome haplotyping. The mtDNA sequences of four Korean *P. infestans* isolates were analyzed to *EcoRI* and *MspI* restriction sites within the target regions (Carter et al., 1990; Griffith and Shaw, 1998) and grouped into Ia, Ib, IIa, or IIb haplotypes based on the sequences of predicted PCR-RFLP haplotypes by *in silico*. In addition to the HVRs carrying the 2 kb insertion at HVRi and 36 bp VNTR at HVRii between *cox* and *orf79* genes (Yang et al., 2013) were also predicted to the haplotypes of mtDNA sequences. Five new haplotype loci developed by Martin et al. (2019) were analyzed (Fig. 1). *In silico* mt genome sequence analysis for haplotyping was conducted by CLC Main Workbench (version 20.0, Qiagen).

Minimal spanning network and phylogenetic analysis.

Phylogenetic analysis of mt genome sequences of 12 *Phytophthora* species (Supplementary Table 1) from Martin et al. (2019) were conducted using maximum likelihood (ML) and maximum parsimony (MP) methods. The best fitting substitution model was K2 + G + I, selected based on the Bayesian information criterion and Akaike information criterion with correction (AICc) in MEGA X (Kumar et al., 2018). Node support was estimated using 1,000 bootstrap replicates in MEGA X (Felsenstein, 1985). The MP analysis was conducted in MEGA X with the subtree-pruning-regrafting algorithm of search level-1. The initial trees were obtained by random addition of sequences (10 replicates).

Results and Discussion

Complete mitochondrial genome sequences.

Mitochondrial genome sequences of the four Korean *P. infestans* isolates were determined by NGS (Table 2). Mapping of the raw data onto the reference mt genome (NC_029397) revealed the average genome coverage of 570, 394, 241, and 389× in KR_1_A1, US-11, KR_2_A2, and SIB-1 *P. infestans* isolates, respectively. All the mt genomes were similar in size and gene content. The mt genome length of KR_1_A1 and SIB-1 (both 39,872 bp) was similar to that of US-11 (39,840 bp) and KR_2_A2 (39,836 bp) (Fig. 1). The KR_1_A1 and SIB-1 mt genome sequences were same in length. Each mt genome contained 61 genes, which included 2 RNA-encoding genes, 16 ribosomal protein genes, 25 transfer RNA and 17 genes encoding electron transport and ATP synthesis (NADH dehydrogenase, cytochrome b, c, ATP synthetase), 11 open reading frames of unknown function, and one import-related protein gene, *tatC* (Table 3). All the genomes were with the same gene order and G + C content of 22.2%. Coding sequences accounted for about 91.2% of the mt genome (Supplementary Table 2). None of the predicted genes contained introns, and ATG and TAA were the start and stop codons for all the genes except *nadh11*, which had TGA as stop codon. Korkmaz et al. (2014) reported that among the three stop codons (TAA, TAG, and TAG), TAA is the most common in bacteria. Similarly, TAA was the most common stop codon in *P. infestans* mtDNA. Seven *orf* genes, *orf99*, *orf80*, *orf51*, *orf171*, *orf45*, *orf39*, and *orf198*, were identified in the 2 kb insertion found in type IIa haplotype previously reported by Avila-Adame et al. (2006).

Comparative analysis among four complete mitochondrial genome sequences of Korean *P. infestans* isolates.

General characteristics of the complete mt genome sequences of KR_1_A1, KR_2_A2, US-11, and SIB-1 are presented in Table 2. The results showed a high level of

Table 2. Genome size, mitochondrial haplotypes and GenBank accession number of Korean *Phytophthora infestans* isolates

Genotype (isolates)	Genome size (bp)	Average genome coverage (X)	Haplotype		GenBank accession no.
KR_1_A1	39,872	570	IIa-6 ^a	IIR ₃ ^b	MH286884
US-11	39,840	394	IIb-2	IIR ₂	MH286885
KR_2_A2	39,836	241	IIa-5	IIR ₂	MH286886
SIB-1	39,872	389	IIa-6	IIR ₃	MH286887
15/99 ^c	39,870	-	IIa-2	IIR ₃	AY898627
94-52 ^c	39,840	-	IIb-4	IIR ₂	AY898628
80029	37,922	-	Ia-2	IIR ₂	AY894835
West Virginia 4	37,957	-	Ib-22	IIR ₂	NC002387

^{a,b}Haplotypes were identified according to Martin et al. (2019) and Yang et al. (2013), respectively.

^c15/99 and 94-52 isolate collected in United Kingdom and United States was used for references and their mating type were A1.

Table 3. General features of mitochondrial genome in the Korean *Phytophthora infestans* isolates

Category	Function (No. of genes)	Gene
Translation	Transfer RNA tRNA (25)	<i>trnW-CCA, trnY-GUA, trnG-UCC, trnG-GCC, trnM-CAU, trnP-UGG, trnM-CAU, trnS-GCU, trnN-GUU, trnA-UGC, trnK-UUU, trnI-CAU, trnS-UGA, trnC-GCA, trnL-UAA, trnL-UAG, trnR-UCU, trnD-GCU, trnV-UAC, trnI-GAU, trnQ-UUG, trnR-GCG, trnF-GAA, trnH-GUG, trnE-UUC</i>
	Ribosomal protein (18)	<i>rns, rnl, rps4, rps2, rpl6, rps8, rps14, rpl16, rps3, rps19, rpl2, rps13, rps11, rps7, rps12, rps10, rpl5, rpl14</i>
Electron transport and ATP synthesis	Complex V ATP (3)	<i>atp8, atp6, atp1</i>
	Complex I NADH (10)	<i>nad11, nad1, nad4L, nad6, nad5, nad3, nad2, nad7, nad4, nad9</i>
	Complex IV Cytochrome (3)	<i>cox3, cox1, cox2</i>
	Complex III (1)	<i>cob</i>
Protein import	Protein import (1)	<i>ymf16 (tatC)</i>
Hypothetical protein	Unknown (11)	<i>orf79, orf198^a, orf170^a, orf39, orf45, orf51, orf80, orf99, orf100, orf127</i>

^aType IIb has *orf178* and 3 bp longer than *orf170* gene.

similarity in the overall genome composition of the four Korean isolates. There were no differences in total complementary DNA sequences (CDS) (91.2%), G + C content (91.2%), total tRNA bases (1,882), and total tRNA length (75.3) in the four isolates. Moreover, all isolates showed similar CDS bases (36,348) and total repeats (1,738), except for isolate US-11 mt genome sequences (total CDS bases of 36,291 and 1,838 total repeats) (Supplementary Table 2). In the type IIb haplotype, *orf178* gene was 60 bp shorter than *orf198*, which was 597 bp long. *Orf45*

and *orf39* were nested in the *orf170* gene (Table 4). These results indicate that gene content and their order in the mt genome were highly conserved among the four Korean *P. infestans* isolates.

Table 4. The SNPs and InDels of mitochondrial genome sequences among four Korean *Phytophthora infestans* isolate

	SNP	InDel
Genic	24	3 (<i>orf170</i> AAT, <i>orf198</i> 1A, <i>orf79</i> 1A)
Intergenic	6	2 (36 bp VNTR and <i>nad5-nad3</i> 1T)
Total	30	4

The position, transition, transversion were shown in the Supplementary Table 2.

SNP, single nucleotide polymorphism; InDel, insertion and deletion; VNTR, variable number tandem repeat.

Mitochondrial genome haplotyping. The sequence-based methods implemented in the present study identified seven mt haplotypes (Supplementary Data 1). The most polymorphic locus was the 1.1 kb *rns* to *cox2* region (locus 2) with five haplotypes. There were seven single-nucleotide polymorphisms (SNPs; five conserved across type I and II haplotypes), 10 InDels (four 1 bp, one 2 bp, one 3 bp, one 4 bp, two 6 bp, and one 24 bp), and a variable number of the 36 bp repeat (one, two, three, or four copies located between *orf79* and *cox2*) (Fig. 2). The next most polymorphic locus was *nad6-nad4L* (locus 5), the 0.5 kb region present in seven haplotypes, which comprised two, single-base InDels and four SNPs (one conserved between type I and II haplotypes). The remaining three loci *rpl5-rns* (locus 1), *cox1-nad9* (locus3), and *nad3-nad5* (locus 4) were three different haplotypes. The analysis of the five multilocus

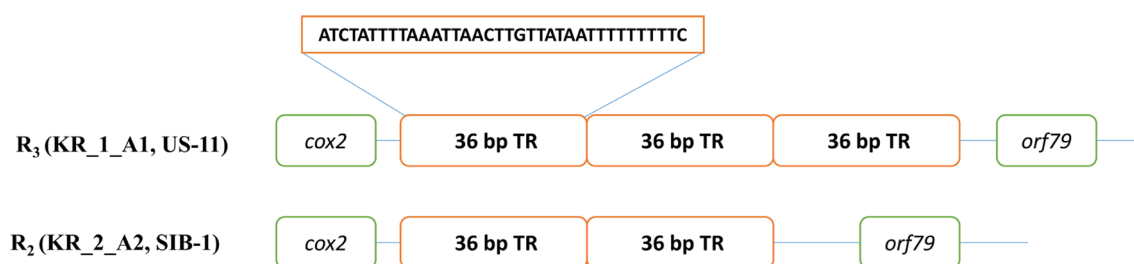
**Fig. 2.** Schematic diagram of mitochondrial genome haplotypes based on the variation number in 36 bp tandem repeats (TR) between *cox2* and *orf79* gene. R₂ and R₃ indicate the two and three TR, respectively.

Table 5. Estimates of evolutionary divergence between sequences

	SIB-1	KR_1_A1	KR_2_A2	US-11
SIB-1	-	0.0	5.0	31.0
KR_1_A1	0.0	-	5.0	31.0
KR_2_A2	2.6	2.6	-	30.0
US-11	4.0	4.0	4.3	-

The number of base differences per sequence from between sequences are shown. Standard error estimate(s) are shown below the diagonal and were obtained by a bootstrap procedure (1,000 replicates). The analysis involved 4 nucleotide sequences. All positions containing gaps and missing data were eliminated. There were a total of 39835 positions in the final dataset. Evolutionary analyses were conducted in MEGA X.

haplotypes generated a total of seven *P. infestans* multi-locus mt haplotypes. Haplotype classification as followed by Martin et al. (2019) for example, haplogroup I-2 would be I-4-8-4-1-5, with locus 1 assigned to haplotype 4, locus 2 to haplotype 8 etc., while all the haplotypes belonged to haplotype IIa and IIb. Haplogroup IIa was the most prevalent, with 11 haplotypes among the five isolates (US-13, EU_8_A1, KR_1_A1, KR_2_A2, and SIB-1) compared with IIb, which had five haplotypes within the two isolates (US-11 and US-6). In haplotype IIb alone a US-11 isolate

dominated in the group II haplotype.

Phylogenetic analysis. The estimates of evolutionary divergence between sequences are shown in Table 5. The sequence divergence between Korean and Russian isolates was small compared with the divergence between Korean and US isolates. Both MP and ML analysis resolved Korean isolates (KR_1_A1 and KR_2_A1) more divergent from the US isolate (US-11) than from the Russian isolate (SIB-1); of the two Korean isolates, KR_1_A1 was more closely related to the Russian isolate (SIB-1) (Fig. 3). The haplotype network analysis clearly identified two groups, IIa and IIb haplotypes (Fig. 4), within which Korean isolates and the Russian isolates were clearly divergent from the US isolates. KR_1_A1 and SIB-1 were branched from IIa-6 haplotype, as part of IIa haplotype. The MP analysis placed the Korean isolates with Russian isolates and distant to the US isolate. Within the Korean isolates, KR_1_A1 isolate formed a clade with SIB-1 (Fig. 5).

Migration of *P. infestans* in Korea. The potato was introduced to Korea in 1824 either from Japan in the south or from China in the north, with the latter being the more likely route (Cho et al., 2003). Akino et al. (2014) reported that Japanese populations of *P. infestans* might have been

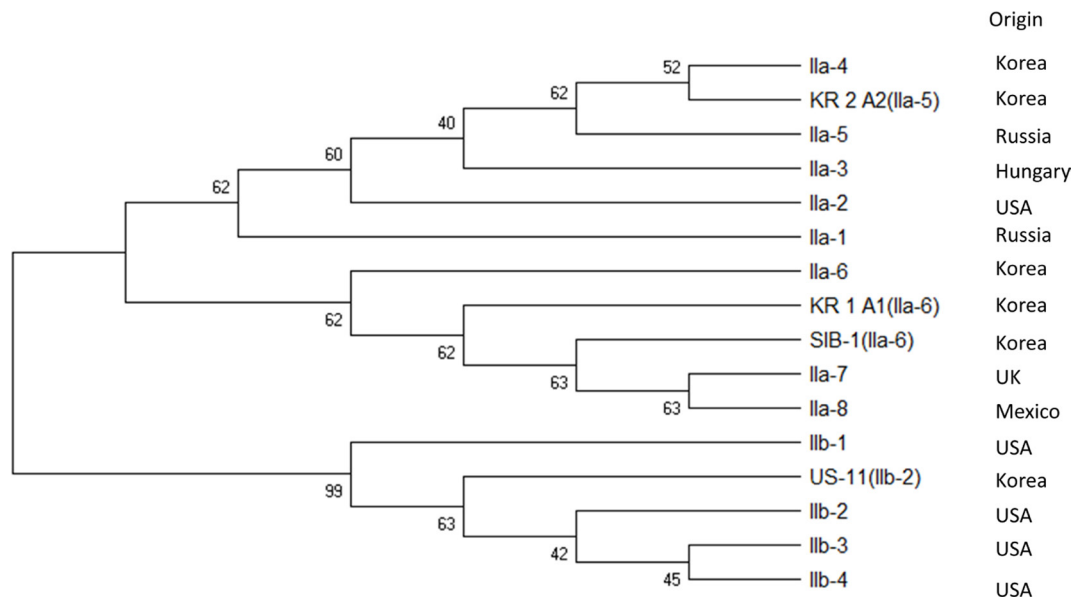


Fig. 3. Evolutionary relationships of 16 *Phytophthora infestans* the evolutionary history was inferred using the UPGMA (unweighted pair-group method with arithmetic mean) method. The optimal tree with the sum of branch length = 0.00253644 is shown. The percentage of replicate trees in which the associated taxa clustered together in the bootstrap test (1,000 replicates) are shown next to the branches. The evolutionary distances were computed using the maximum composite likelihood method and are in the units of the number of base substitutions per site. This analysis involved 16 nucleotide sequences. All ambiguous positions were removed for each sequence pair (pairwise deletion option). There were a total of 5814 positions in the final dataset. Evolutionary analyses were conducted in MEGA X.

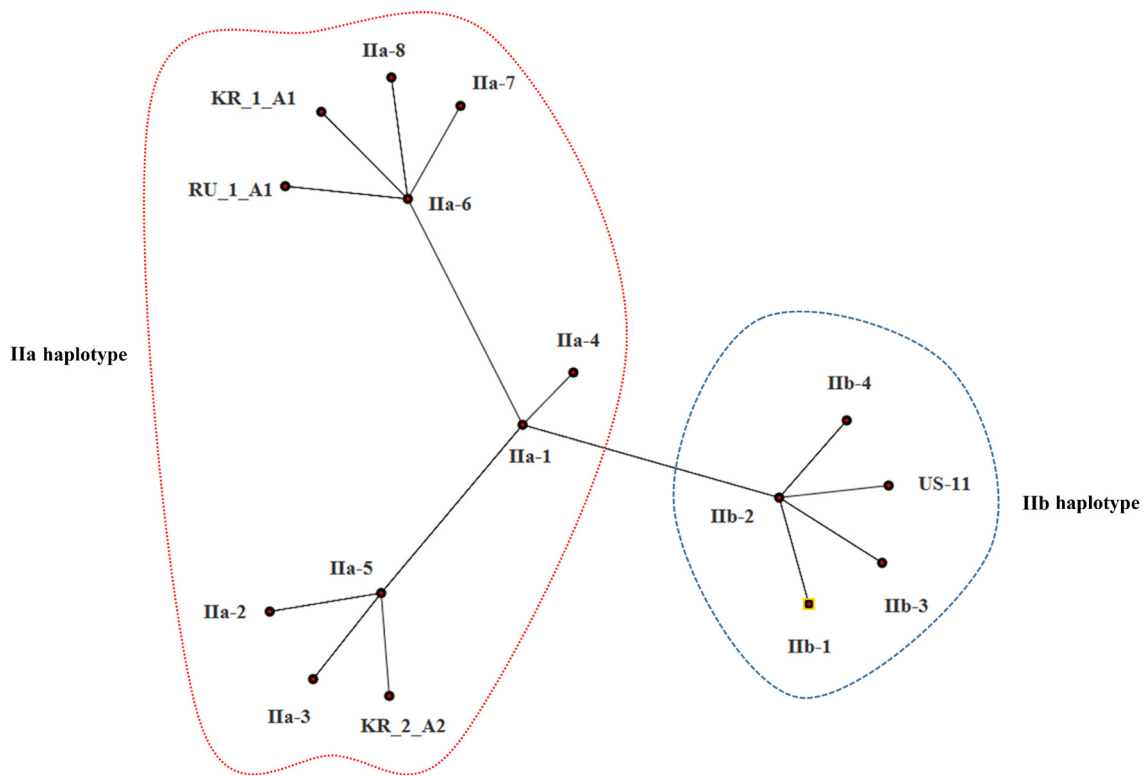


Fig. 4. Minimal spanning network (MSN) tree of mitochondrial haplotypes in four Korean *Phytophthora infestans* isolates with 12 mt haplotypes. MSN was analyzed SplitsTree ver. 4.0 with uncorrected *P* with gaps and a neighbor joining network.

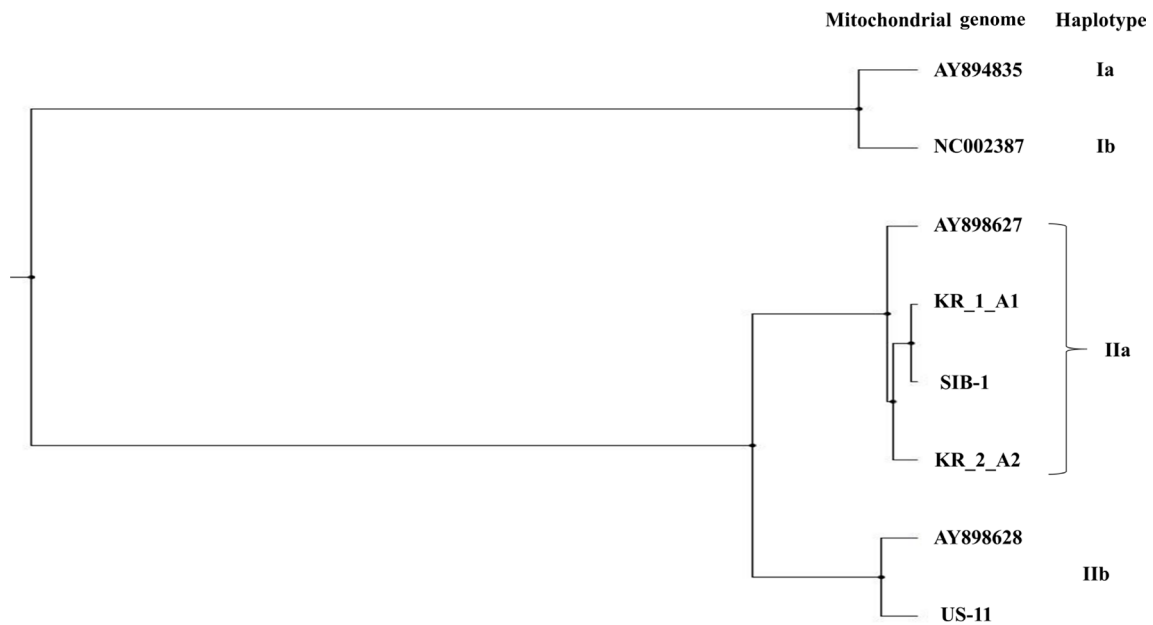


Fig. 5. Maximum parsimony analysis of four Korean *Phytophthora infestans* isolates with whole mitochondrial genome sequences. Four mt genome sequences (AY894835, AY898627, AY898628, and NC002387) were retrieved from GenBank (<http://ncbi.nlm.nih.gov>). Eight mt genomes were haplotyped as Ia, Ib, IIa and IIb by Griffith and Shaw (1998) method.

introduced to Japan from other countries such as Korea or China. SIB-1 is widely spread in northern China and north-west Russia (Siberia region), where it is a predominant strain (Elansky et al., 2001; Guo et al., 2010). This suggests that SIB-1 was introduced to Korea from the north, China or Russia, through contaminated potato seeds and then settled and spread throughout Korea. Considering the 100% match of mt genome sequences between SIB-1 and KR_1_A1, the Korean specific genotype likely originated from SIB-1. KR_2_A2, also found in Korea, might have been introduced from other countries or by changes in mating type that were triggered by exposure to fungicides (Goodwin and Drenth, 1997).

Conflicts of Interest

No potential conflict of interest relevant to this article was reported.

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Electronic Supplementary Material

Supplementary materials are available at The Plant Pathology Journal website (<http://www.ppjonline.org/>).

References

- Akino, S., Takemoto, D. and Hosaka, K. 2014. *Phytophthora infestans*: a review of past and current studies on potato late blight. *J. Gen. Plant Pathol.* 80:24-37.
- Arora, R. K., Sharma, S. and Singh, B. P. 2014. Late blight disease of potato and its management. *Potato J.* 41:16-40.
- Avila-Adame, C., Gómez-Alpizar, L., Zismann, V., Jones, K. M., Buell, C. R. and Ristaino, J. B. 2006. Mitochondrial genome sequences and molecular evolution of the Irish potato famine pathogen. *Phytophthora infestans. Curr. Genet.* 49:39-46.
- Cárdenas, M., Grajales, A., Sierra, R., Rojas, A., González-Almarino, A., Vargas, A., Marín, M., Fermín, G., Lagos, L. E., Grünwald, N. J., Bernal, A., Salazar, C. and Restrepo, S. 2011. Genetic diversity of *Phytophthora infestans* in the Northern Andean region. *BMC Genet.* 12:23.
- Carter, D. A., Archer, S. A., Buck, K. W., Shaw, D. S. and Shattock, R. C. 1990. Restriction fragment length polymorphisms of mitochondrial DNA of *Phytophthora infestans*. *Mycol. Res.* 94:1123-1128.
- Cho, H. M., Park, Y. E., Cho, J. H. and Kim, S. Y. 2003. Historical review of land race potatoes in Korea. *J. Korean Soc. Hortic. Sci.* 44:838-845 (in Korean).
- Cho, K.-S., Yun, B.-K., Yoon, Y.-H., Hong, S.-Y., Mekapogu, M., Kim, K.-H. and Yang, T.-J. 2015. Complete chloroplast genome sequence of tartary buckwheat (*Fagopyrum tataricum*) and comparative analysis with common buckwheat (*F. esculentum*). *PLoS ONE* 10:e0125332.
- Choi, J.-G., Hong, S.-Y., Kessel, G. J. T., Cooke, D. E. L., Vossen, J. H., Cho, J.-H., Im, J.-S., Park, Y.-E. and Cho, K.-S. 2020. Genotypic and phenotypic characterization of *Phytophthora infestans* in South Korea during 2009-2016 reveals clonal reproduction and absence of EU_13_A2 genotype. *Plant Pathol.* 69:932-943.
- Cooke, D. E. L. and Lees, A. K. 2004. Markers, old and new, for examining *Phytophthora infestans* diversity. *Plant Pathol.* 53:692-704.
- Drenth, A., Turkensteen, L. J. and Govers, F. 1993. The occurrence of the A2 mating type of *Phytophthora infestans* in the Netherlands: significance and consequences. *Neth. J. Plant Pathol.* 99:57-67.
- Elansky, S., Smirnov, A., Dyakov, Y., Dolgova, A., Filippov, A., Kozlovsky, B., Kozlovskaya, I., Russo, P., Smart, C. and Fry, W. 2001. Genotypic analysis of Russian isolates of *Phytophthora infestans* from the Moscow region, Siberia and Far East. *J. Phytopathol.* 149:605-611.
- Felsenstein, J. 1985. Confidence limits on phylogenies: an approach using the bootstrap. *Evolution* 39:783-791.
- Fry, W. 2008. *Phytophthora infestans*: the plant (and R gene) destroyer. *Mol. Plant Pathol.* 9:385-402.
- Gómez-Alpizar, L., Carbone, I. and Ristaino, J. B. 2007. An Andean origin of *Phytophthora infestans* inferred from mitochondrial and nuclear gene genealogies. *Proc. Natl. Acad. Sci. U. S. A.* 104:3306-3311.
- Goodwin, S. B. and Drenth, A. 1997. Origin of the A2 mating type of *Phytophthora infestans* outside Mexico. *Phytopathology* 87:992-999.
- Göre, M. E., Altın, N., Yaman, T., Myers, K., Çağlı, A., Cooke, D. E. L., Pırlak, U., Alkan, M., Kabakçı, H., Zencirci, N., Fry, W. E. and Özer, G. 2019. Severe outbreaks of *Phytophthora infestans* on potato in Turkey caused by recent changes in the pathogen population structure. *Phytoparasitica* 47:693-709.
- Goss, E. M., Tabima, J. F., Cooke, D. E. L., Restrepo, S., Fry, W. E., Forbes, G. A., Fieland, V. J., Cardenas, M. and Grünwald, N. J. 2014. The Irish potato famine pathogen *Phytophthora infestans* originated in central Mexico rather than the Andes. *Proc. Natl. Acad. Sci. U. S. A.* 111:8791-8796.
- Gotoh, K., Akino, S., Maeda, A., Kondo, N., Naito, S., Kato, M. and Ogoshi, A. 2005. Characterization of some Asian isolates of *Phytophthora infestans*. *Plant Pathol.* 54:733-739.
- Griffith, G. W. and Shaw, D. S. 1998. Polymorphisms in *Phytophthora infestans*: four mitochondrial haplotypes are detected after PCR amplification of DNA from pure cultures or from host lesions. *Appl. Environ. Microbiol.* 64:4007-4014.

- Guo, L., Zhu, X.-Q., Hu, C.-H. and Ristaino, J. B. 2010. Genetic structure of *Phytophthora infestans* populations in China indicates multiple migration events. *Phytopathology* 100:997-1006.
- Hyun, I.-H. and Choi, W. 2014. *Phytophthora* species, new threats to the plant health in Korea. *Plant Pathol. J.* 30:331-342.
- Kim, J.-S., Lee, Y.-G., Kwon, M., Kim, J.-I., Jee, S. and Park K.-H. 2014. Mating types of *Phytophthora infestans* isolates and their responses to metalaxyl and dimethomorph in Korea. *Res. Plant Dis.* 20:25-30.
- Koh, Y. J., Goodwin, S. B., Dyer, A. T., Cohen, B. A., Ogoshi, A., Sato, N. and Fry, W. E. 1994. Migrations and displacements of *Phytophthora-infestans* populations in East Asian countries. *Phytopathology* 84:922-927.
- Korkmaz, G., Holm, M., Wiens, T. and Sanyal, S. 2014. Comprehensive analysis of stop codon usage in bacteria and its correlation with release factor abundance. *J. Biol. Chem.* 289:30334-30342.
- Kubo, T., Kitazaki, K., Matsunaga, M., Kagami, H. and Mikami, T. 2011. Male sterility-inducing mitochondrial genomes: how do they differ? *Crit. Rev. Plant Sci.* 30:378-400.
- Kumar, S., Stecher, G., Li, M., Knyaz, C. and Tamura, K. 2018. MEGA X: molecular evolutionary genetics analysis across computing platforms. *Mol. Biol. Evol.* 35:1547-1549.
- Lassiter, E. S., Russ, C., Nusbaum, C., Zeng, Q., Saville, A., Olarte, R., Carbone, I., Hu, C.-H., Seguin-Orlando, A., Samaniego, J. A., Thorne, J. L. and Ristaino, J. B. 2015. Mitochondrial genome sequences reveal evolutionary relationships of the *Phytophthora* Ic clade species. *Curr. Genet.* 61:567-577.
- Martin, F. N. and Tooley, P. W. 2004. Identification of phytophthora isolates to species level using restriction fragment length polymorphism analysis of a polymerase chain reaction-amplified region of mitochondrial DNA. *Phytopathology* 94:983-991.
- Martin, F. N., Zhang, Y., Cooke, D. E. L., Coffey, M. D., Grünwald, N. J. and Fry, W. E. 2019. Insights into evolving global populations of *Phytophthora infestans* via new complementary mtDNA haplotype markers and nuclear SSRs. *PLoS ONE* 14:e0208606.
- May, K. J. and Ristaino, J. B. 2004. Identity of the mtDNA haplotype(s) of *Phytophthora infestans* in historical specimens from the Irish potato famine. *Mycol. Res.* 108:471-479.
- Patil, V. U., Vanishree, G., Pattanayak, D., Sharma, S., Bhardwaj, V., Singh, B. P. and Chakrabarti, S. K. 2017. Complete mitogenome mapping of potato late blight pathogen, *Phytophthora infestans* A₂ mating type. *Mitochondrial DNA Part B Resour.* 2:90-91.
- Peters, R. D., Al-Mughrabi, K. I., Kalischuk, M. L., Dobinson, K. F., Conn, K. L., Alkher, H., Islam, M. R., Daayf, F., Lynn, J., Bizimungu, B., De Koeyer, D., Lévesque, C. A. and Kawchuk, L. M. 2014. Characterization of *Phytophthora infestans* population diversity in Canada reveals increased migration and genotype recombination. *Can. J. Plant Pathol.* 36:73-82.
- Rekad, F. Z., Cooke, D. E. L., Puglisi, I., Randall, E., Guenaoui, Y., Bouznad, Z., Evoli, M., Pane, A., Schena, L., di San Lio, G. M. and Cacciola, S. O. 2017. Characterization of *Phytophthora infestans* populations in northwestern Algeria during 2008-2014. *Fungal Biol.* 121:467-477.
- Shimelash, D., Hussien, T., Fininsa, C., Forbes, G. and Yuen, J. 2016. Mitochondrial DNA assessment of *Phytophthora infestans* isolates from potato and tomato in Ethiopia reveals unexpected diversity. *Curr. Genet.* 62:657-667.
- Van der Lee, T., De Witte, I., Drenth, A., Alfonso, C. and Govers, F. 1997. AFLP linkage map of the oomycete *Phytophthora infestans*. *Fungal Genet. Biol.* 21:278-91.
- Yang, Z.-H., Qi, M.-X., Qin, Y.-X., Zhu, J.-H., Gui, X.-M., Tao, B., Xu, X.-H. and Zhang, F.-G. 2013. Mitochondrial DNA polymorphisms in *Phytophthora infestans*: new haplotypes are identified and re-defined by PCR. *J. Microbiol. Methods* 95:117-121.
- Zhang, X.-Z., Kim, H.-Y. and Kim, B.-S. 2006. Analysis of genetic diversity of *Phytophthora infestans* in Korea by using molecular markers. *J. Microbiol. Biotechnol.* 16:423-430.