- Bruno MK, Hallett M, Gwinn-Hardy K, et al. Clinical evaluation of idiopathic paroxysmal kinesigenic dyskinesia: new diagnostic criteria. Neurology 2004;63:2280–2287.
- Chen WJ, Lin Y, Xiong ZQ, et al. Exome sequencing identifies truncating mutations in PRRT2 that cause paroxysmal kinesigenic dyskinesia. Nat Genet 2011;43:1252–1255.
- Wang JL, Cao L, Li XH, et al. Identification of PRRT2 as the causative gene of paroxysmal kinesigenic dyskinesias. Brain 2011;134: 3493–3501.
- Méneret A, Grabli D, Depienne C, et al. *PRRT2* mutations: a major cause of paroxysmal kinesigenic dyskinesia in the European population. Neurology 2012;79:170–174.
- Lee YC, Lee MJ, Yu HY, Chen C, Hsu CH, Lin KP. PRRT2 mutations in paroxysmal kinesigenic dyskinesia with infantile convulsions in a Taiwanese cohort. PLoS ONE 2012;7:e38543
- 7. Heron SE, Grinton BE, Kivity S, et al. *PRRT2* mutations cause benign familial infantile epilepsy and infantile convulsions with choreoathetosis syndrome. Am J Hum Genet 2012;90:152–160.
- 8. Lee HY, Huang Y, Bruneau N, et al. Mutations in the gene PRRT2 cause paroxysmal kinesigenic dyskinesia with infantile convulsions. Cell Rep 2012;1:2–12.
- Ebrahimi-Fakhari D, Saffari A, Westenberger A, Klein C. The evolving spectrum of PRRT2-associated paroxysmal diseases. Brain 2015; 138:3476–3495.
- De Gusmao CM, Silveira-Moriyama L. Paroxysmal movement disorders - practical update on diagnosis and management. Expert Rev Neurother 2019;19:807–822.
- 11. Li HF, Chen YL, Zhuang L, et al. TMEM151A variants cause paroxysmal kinesigenic dyskinesia. Cell Discov 2021;7:83
- Li HF, Chen WJ, Ni W, et al. *PRRT2* mutation correlated with phenotype of paroxysmal kinesigenic dyskinesia and drug response. Neurology 2013;80:1534–1535.
- Huang XJ, Wang T, Wang JL, et al. Paroxysmal kinesigenic dyskinesia: clinical and genetic analyses of 110 patients. Neurology 2015; 85:1546–1553.
- Huang XJ, Wang SG, Guo XN, et al. The phenotypic and genetic spectrum of paroxysmal kinesigenic dyskinesia in China. Mov Disord 2020;35:1428–1437.
- 15. Chen YL, Chen DF, Zhao SY, Liu GL, Li HF, Wu ZY. Penetrance estimation of PRRT2 variants in paroxysmal kinesigenic dyskinesia and infantile convulsions. Front Med 2021;15:877–886.
- 16. Richards S, Aziz N, Bale S, et al. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. Genet Med 2015;17: 405–424.
- 17. Erro R, Sheerin UM, Bhatia KP. Paroxysmal dyskinesias revisited: a review of 500 genetically proven cases and a new classification. Mov Disord 2014;29:1108–1116.
- van Vliet R, Breedveld G, de Rijk-van AJ, et al. PRRT2 phenotypes and penetrance of paroxysmal kinesigenic dyskinesia and infantile convulsions. Neurology 2012;79:777–784.
- Li HF, Ni W, Xiong ZQ, Xu J, Wu ZY. PRRT2 c.649dupC mutation derived from de novo in paroxysmal kinesigenic dyskinesia. CNS Neurosci Ther 2013;19:61–65.
- Symonds JD, Zuberi SM, Stewart K, et al. Incidence and phenotypes of childhood-onset genetic epilepsies: a prospective population-based national cohort. Brain 2019;142:2303–2318.
- Mao CY, Shi CH, Song B, et al. Genotype-phenotype correlation in a cohort of paroxysmal kinesigenic dyskinesia cases. J Neurol Sci 2014;340:91–93.

### Supporting Data

Additional Supporting Information may be found in the online version of this article at the publisher's web-site.

# Spontaneous Graft-Induced Dyskinesias Are Independent of 5-HT Neurons and Levodopa Priming in a Model of Parkinson's Disease

Emma L. Lane, PhD,<sup>1</sup> David J. Harrison, PhD Elena Ramos-Varas, MSc,<sup>2</sup> Rachel Hills, BSc, Sophie Turner, BSc,<sup>2</sup> and Mariah J. Lelos, PhD<sup>1\*</sup> (D

<sup>1</sup>School of Pharmacy and Pharmaceutical Sciences, Cardiff University, Cardiff, United Kingdom <sup>2</sup>Brain Repair Group, School of Biosciences, Cardiff University, Cardiff, United Kingdom

**ABSTRACT: Background:** The risk of graftinduced dyskinesias (GIDs) presents a major challenge in progressing cell transplantation as a therapy for Parkinson's disease. Current theories implicate the presence of grafted serotonin neurons, hotspots of dopamine release, neuroinflammation and established levodopa-induced dyskinesia.

Objective: To elucidate the mechanisms of GIDs.

**Methods:** Neonatally desensitized, dopamine denervated rats received intrastriatal grafts of human embryonic stem cells (hESCs) differentiated into either ventral midbrain dopaminergic progenitor (vmDA) (n = 15) or ventral forebrain cells (n = 14).

**Results:** Of the eight rats with surviving grafts, two vmDA rats developed chronic spontaneous GIDs, which were observed at 30 weeks post-transplantation. GIDs were inhibited by  $D_2$ -like receptor antagonists and not affected by 5-HT1A/1B/5-HT6 agonists/antagonists. Grafts in GID rats showed more microglial activation and lacked serotonin neurons.

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

\*Correspondence to: Dr. M.J. Lelos, Brain Repair Group, School of Biosciences, Cardiff University, Museum Avenue, Cardiff, Wales CF10 3AX, UK; E-mail: lelosmj@cardiff.ac.uk

**Relevant conflicts of interest/financial disclosures:** None to declare. We have not received any funding from any institution, including personal relationships, interests, grants, employment, affiliations, patents, inventions, honoraria, consultancies, royalties, stock options/ownership, or expert testimony for the last 12 months.

**Funding agencies:** M.J.L. was supported by a Parkinson's UK Senior Research Fellowship (F1502); E.L.L. was support by Medical Research Council grant MR/R00630X/1.

Received: 3 June 2021; Revised: 28 September 2021; Accepted: 20 October 2021

Published online 12 November 2021 in Wiley Online Library (wileyonlinelibrary.com). DOI: 10.1002/mds.28856

**Conclusions:** These findings argue against current thinking that rats do not develop spontaneous GID and that serotonin neurons are causative, rather indicating that GID can be induced in rats by hESC-derived dopamine grafts and, critically, can occur independently of both previous levodopa exposure and grafted serotonin neurons. © 2021 The Authors. *Movement Disorders* published by Wiley Periodicals LLC on behalf of International Parkinson and Movement Disorder Society

**Key Words:** Parkinson's disease; dopamine; cell therapy; graft-induced dyskinesias; neuroinflammation; microglia; 5-HTL-dopa

Clinical trials using fetal dopamine (DA) cells as a neuroreparative strategy for Parkinson's disease (PD) have produced remarkable long-term recovery of function in some patients<sup>1</sup> and graft survival for over 20 years.<sup>2</sup> However, the results have been variable and a proportion of patients in three major trials developed persistent abnormal involuntary movements (AIMS), now termed "graft-induced dyskinesias" (GIDs), occurring in the absence of levodopa (L-dopa).<sup>3-5</sup> Pre-existing L-dopa induced dyskinesias (LID) have been identified as a potential risk factor for the development of GIDs, but other potential risk factors include aberrant immune responses,<sup>6,7</sup> incomplete graft innervation,<sup>8</sup> 5-HT neurons in the graft,<sup>9</sup> abnormal ratios of 5-HT/DA receptors,<sup>10-12</sup> or activity of specific 5-HT receptors.<sup>13</sup> Investigations of the mechanisms underlying GIDs have been hampered by rodent models that do not replicate the spontaneous nature of the behaviors. Rather they rely on administration of Ldopa or amphetamine to generate acute, transient GID expression<sup>14,15</sup> in transplanted animals.

With the evolution of human stem cell (hESC)-derived sources of cells for transplantation, it is now possible to generate ventral midbrain DA (vmDA) grafts free of 5-HTergic neurons and to assess if GIDs can occur also in the absence of 5-HT. Intrastriatal grafts of hESC-derived vmDA cells have been widely reported to survive, integrate, release DA, and alleviate functional impairments in rodent models of PD,<sup>16-19</sup> but GIDs have not been explored. To ensure the safety of new cell products for transplantation, it is imperative that we evaluate and understand the potential risk of side effects.

In the process of studying the long-term functional efficacy of hESC-derived vmDA grafts in 6-hydroxydopamine (6-OHDA) lesioned rats, we unexpectedly observed spontaneous, continuous abnormal AIMs in a subset of rats. The rats had been checked weekly for health and welfare purposes, but detailed behavioral analysis commencing at 30 weeks led to the identification of chronic GIDs. We characterized the GIDs pharmacologically, targeting DAergic and 5-HTergic systems. Subsequent immunohistochemical analysis of the brain tissue compared vmDA grafts from GID rats with vmDA grafts from non-GID rats and with control hESC-derived non-DA, forebrain-patterned (vFB) cells.<sup>17</sup>

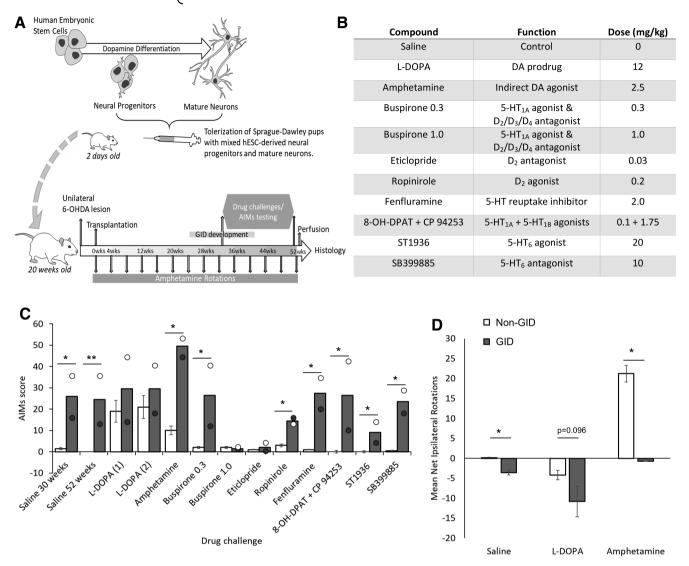
#### Materials and Methods

Experiments were conducted in compliance with the United Kingdom (UK) Animals (Scientific Procedures) Act 1986 under Home Office License No. 30/2498, with the approval of the local Cardiff University Ethics Review Committee.

Sprague–Dawley rat pups (female, n = 29) were neonatally desensitized at 2 days post-birth with mixed hESC-derived neural progenitors and mature neurons (Fig. 1A), as described elsewhere.<sup>20</sup> Rats were housed in groups of 3-4, with a 14-hour:10-hour light:dark cycle. At 20 weeks old, rats received MFB 6-OHDA lesions, as previously described.<sup>18</sup> Rats were sorted into matched groups based on amphetamine-induced rotations (2.5 mg/kg) conducted at 4 weeks post-lesion (mean net rotations/minutes: vmDA =  $15.3 \pm 0.8$ ; vFB =  $14.6 \pm 1.0$ ). Rats received intrastriatal transplants of hESC-derived cells (vmDA or vFB).<sup>17</sup> H9 cells were differentiated for 16 days and made into a suspension of 60,000 cells/µL, as previously described.<sup>17</sup> For both transplanted groups, 4 uL (240,000 total cells/graft) was injected into the neostriatum at the following coordinates: (1) AP: +0.5, ML: -3.0; (2) AP: +1.2, ML: -2.7; (3) DVs: -4/-5.

Rotational behaviors were measured for 90 minutes (Rotorat, Med Associates) after 2.5 mg/kg methamphetamine (Sigma, cat. no. M8750), for 60 minutes after 0.05 mg/kg apomorphine (Sigma, cat. no. Y0001465) and after administration of other pharmacological agents (Fig. 1B). Observations from 30 weeks post-graft revealed spontaneous AIMs. AIMs (as described in Breger et al)<sup>21</sup> were scored every 10 minutes for 60 minutes postadministration of saline or drug. The assessor was blind to both the substance administered and the rat group allocation. During AIMs scoring, the experimenter noted whether normal locomotion and exploratory behaviors were evident in the rotameters, to give confidence that any reduction in GID expression was not the consequence of reduced activity overall. Rotational data were collected in an unbiased manner using automated rotometers.

At 52 weeks post-graft, rats were terminally anaesthetized and transcardially perfused (4% paraformaldehyde [PFA]). Brains were processed for peroxidase-based immunohistochemistry as previously described.<sup>18</sup> CD4<sup>+</sup>, CD8<sup>+</sup>, and 5-HT<sup>+</sup> cells were counted manually within and at the border of the graft. TH<sup>+</sup>, HuNu<sup>+</sup>, Ox42<sup>+</sup>/ CD11b<sup>+</sup>, and GFAP<sup>+</sup> cells were estimated using unbiased stereology. Two-dimensional stereology was performed (Olympus BX50 microscope, Olympus C.A.S.T. imageanalysis software). Ox42 was also measured by optical density (ImageJ, NIH).



**FIG. 1.** (**A**) Schematic representation of the timeline of experimental events. (**B**) Table of pharmacological agents used during abnormal involuntary movement (AIMs) analysis in rats. (**C**) Mean and individual AIMs score for vmDA graft-induced dyskinesias (GID) rats (n = 2). Mean AIMs score for non-GID rats includes small vmDA grafts and vFB grafts, combined into "non-GID" dataset (n = 6), in response to saline or drug challenges. AIMs were stable between 30 and 52 weeks post-graft (both saline datasets, P < 0.05 for non-GID vs. GID rats). Non-GID rats demonstrate AIMs in response to L-dopa, but no change in AIMs is evident in GID rats. For GID rats, 0.3 mg/kg buspirone (5-HT1A agonist/D2-like antagonist), fenfluramine (5-HT reuptake inhibitor), 8-OH-DPAT+CP94253 (5-HT1A/1B agonists), and SB399885 (5-HT6 antagonist) did not alter expression of the AIMs behaviors (all ps < 0.05 for non-GID vs. GID). High dose buspirone and eticlopride both abolish AIMs (ps = n.s. for non-GID vs. GID) and GID rats in response to saline (P < 0.05 for non-GID vs. GID rats), L-dopa (P = 0.096) or amphetamine (P < 0.05). In contrast to non-GID rats, GID rats did not rate in response to saline (P < 0.05 for non-GID vs. GID rats), L-dopa (P = 0.096) or amphetamine (P < 0.05). In contrast to non-GID rats, GID rats did not rotate in response to amphetamine, but did rotate spontaneously and in response to L-dopa. Note: All rats had complete unilateral lesions, as evidenced by their pre-graft rotational response to amphetamine (non-GID rats: mean = 12.3 net turns/minute, SEM = 0.95; GID rats: mean = 14.5 net turns/minute, SEM = 1.27) and the loss of nigral TH<sup>+</sup> neurons (non-GID rats: Error bars = standard errors of the mean. \* $P \le 0.05$ , \*\* $P \le 0.01$ . [Color figure can be viewed at wileyonlinelibrary.com]

Pharmacological challenges and rotation data were analyzed using Kruskal-Wallis non-parametric test with Group (non-GID vs. GID) as the factor. Histological data were analyzed by one-way ANOVA with Group (vFB, vmDA non-GID, and vmDA GID) as the factor. Histological and behavioral data were correlated using Spearman's  $\rho$ . All statistical analyses were conducted using IBM SPSS Statistics 25 software.

#### Results

#### Dopaminergic Grafts Can Induce Spontaneous GIDs

To obtain long-term survival of human grafts in a rodent model without the need for daily immunosuppression, we applied a model of neonatal desensitization in rats, which required subcutaneous injection of human cells

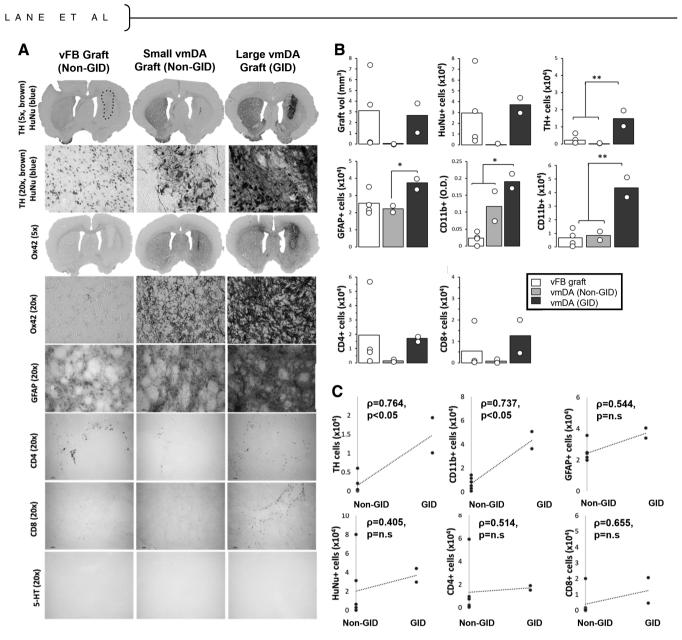


FIG. 2. (A) Immunohistochemical analysis of hESC-derived vmDA and vFB grafts. Dopamine neurons (tyrosine hydroxylase [TH], Millipore, MAB318, 1:2000; brown DAB) and human nuclei (HuNu, Millipore, MAB1281, 1:1000; blue VectorSG) at 5x and 20x magnification in vFB grafts, small vmDA grafts from non-GID rats and large vmDA grafts from GID rats. The HuNu<sup>+</sup> graft core of the vFB graft is delineated with a black dashed line. Activated microglia within the graft core and border (Ox42 [CD11b], Serotec, MCA275G, 1:1000) at 5x and 20x magnification in each graft group. Immunolabeling of reactive astrocytes (GFAP, DAKO, Z0334, 1:1000), CD4+t-lymphocytes (Abcam, Ab33775, 1:100), CD8+t-lymphocytes (Serotec, MC4A48G, 1:500) and 5-HT+ neuron staining (Immunostar, 20,080, 1:10000). 5-HT<sup>+</sup> "control" staining in the raphe nucleus is shown in the left panel, whereas the center and right panels demonstrate no positive staining in the small or large vmDA grafts. (B) Quantification of cells in hESC-derived vmDA and vFB grafts. Graft volume, total HuNu<sup>+</sup> cells, TH<sup>+</sup> cells within the graft, GFAP<sup>+</sup> cells within the striatum, Ox42/CD11b optical density within the graft core/border, Ox42 cells/mm<sup>3</sup> within the graft core/border, CD4<sup>+</sup> cells around the perimeter of the graft and CD8<sup>+</sup> cells around the perimeter of the graft were quantified for vFB grafts, small vmDA grafts (non-GID rats) and large vmDA grafts (GID rats). GID rats had grafts with more TH<sup>+</sup> cells than vFB or small vmDA grafts [F<sub>2.5</sub> = 10.45, P < 0.05; GID vs. non-GID and vFB, ps = 0.01] and more GFAP<sup>+</sup> cells than grafts in non-GID rats [F<sub>2,5</sub> = 4.20, P < 0.05; GID vs. non-GID, P < 0.05]. More activated microglia (Ox42<sup>+</sup>/CD11b<sup>+</sup>) cells were evident in GID rats than in non-GID or vFB grafted rats [F<sub>2.5</sub> = 21.50, P < 0.05; GID vs. non-GID and vFB, ps < 0.05]. No other group effects were significant. (C) Correlations between grafted cells and GID behaviors. Significant correlations were revealed between GID and TH<sup>+</sup> and Ox42<sup>+</sup> cells. No correlations were revealed between GID the total HuNu cell count, GFAP, CD4<sup>+</sup> or CD8<sup>+</sup> cells. For 5× magnification images, all scale bars = 100 µm; For 20× magnification images, all scale bars = 1000 µm. vFB = ventral forebrain control grafts; vmDA = ventral midbrain ventral mesencephalic grafts. Error bars = standard errors of the mean. \* $P \le 0.05$ , \*\* $P \le 0.01$ , \*\*\* $P \le 0.001$ . [Color figure can be viewed at wileyonlinelibrary.com]

at P0-5.<sup>20,22</sup> In adulthood, the desensitized rats received unilateral lesions and subsequently hESC-derived vmDA or vFB intrastriatal transplants. Only a subset of the transplanted rats (4/15) had surviving vmDA grafts and, of these, two displayed spontaneous contralateral

rotations, a trend for L-dopa-induced rotations and reduced amphetamine-induced rotations (Fig. 1D). These two rats also developed spontaneous AIMs at 30 weeks post-graft (Fig. 1C and Supporting Data). These were compared to rats with smaller surviving vmDA grafts (n = 2) and rats with non-DA, vFB grafts (n = 4/14) (neither the smaller vmDA nor the vFB graft groups displayed spontaneous AIMs or reduced amphetamine-induced rotations).

To elucidate the neurobiological basis of the GIDs, pharmacological challenges were undertaken using receptor agonists and antagonists. AIMs were observed in both the home cage and rotometers (Supplementary Video S1), and scored blind twice post-saline administration (at 30 and 52 weeks post-graft) and twice post-L-dopa administration (Fig. 1C). These sequential observations suggest that GIDs were chronic, stable, and minimally affected by stress or repeated exposure to rotometers. For GID rats, 0.3 mg/kg buspirone (5-HT<sub>1A</sub> agonist/D<sub>2</sub>-like antagonist), fenfluramine (5-HT reuptake inhibitor), 8-OH-DPAT+CP94253 (5-HT<sub>1A/1B</sub> agonists) and SB399885 (5-HT<sub>6</sub> antagonist) did not alter expression of the AIMs behaviors. High dose 1 mg/kg buspirone<sup>23</sup> and eticlopride (D<sub>2</sub>/D<sub>3</sub> antagonist) largely eliminated GID expression without reducing normal motor activity.

#### Larger vmDA Grafts Induce Neuroinflammation and GIDs

Larger vmDA grafts were identified in GID-expressing rats and small vmDA grafts in non-GID rats. The vFB grafts were a similar volume to large vmDA grafts, containing similar numbers of human cells, but with an absence of mature DA neurons (Fig. 2A,B). Similar numbers of CD4<sup>+</sup> and CD8<sup>+</sup> t-lymphocytes were present in large vFB and vmDA grafts, but more reactive astrocytes were observed in GID vmDA rats (Fig. 2A,B). No 5-HT<sup>+</sup> cells were observed in any of the grafts (Fig. 2A), despite positive control staining evident within the raphe nuclei.

Large vmDA grafts were associated with high levels of microglial activation in both hemispheres, whereas small vmDA grafts induced modest microglial activation in both hemispheres. In contrast, large vFB grafts induced almost no microglial activation (Fig. 2A,B). Neither graft volume, total HuNu<sup>+</sup> cells, GFAP, CD4<sup>+</sup>, nor CD8<sup>+</sup> t-lymphocyte infiltration correlated with the development of spontaneous AIMs (Fig. 2C). High DA neuron content (tyrosine hydroxylase [TH]) and markedly increased microglial activation (Ox42) correlated significantly with GIDs (Fig. 2C).

### Discussion

This is the first report of measurable spontaneous GIDs occurring following transplantation of hESCderived vmDA neurons into an animal model. These behaviors occurred without prior exposure to L-dopa or LID "priming" and with grafts devoid of 5-HT neurons. Given that they were abolished by the  $D_2$  receptor antagonists eticlopride and by buspirone, which acts as a  $D_2$  receptor antagonist at high doses, this suggests that DA itself, and particularly  $D_2$ -like family receptors, play a role in mediating GIDs.

These long-term (1 year post-graft) pilot data are important insofar as they suggest that GID emergence is possible in clinical trials of hESC-derived vmDA grafts. This is despite the evidence that hESC-derived cell preparations for clinical use will contain no/limited 5-HT neurons and patient selection will prioritize patients without severe LIDs. Important parallels can be drawn with recently published details on five patients who developed persistent GIDs in a double-blind United States (US) fetal cell transplant study.<sup>5</sup> Both of these datasets challenge current assumptions in the field, which suggest that GIDs expression may be dependent on 5-HT expression.<sup>24-27</sup> Dopamine is directly implicated in both studies: (1) the presentation of GIDs resembles some elements of classic LIDs; (2) anti-dopaminergic drugs reduce GIDs; (3) GIDs correlate with measures of dopamine (fluorodopa in clinical trials/TH<sup>+</sup> immunohistochemistry here), and (4) GIDs occur in association with functional improvements (Unified Parkinson Disease Rating Scale [UPDRS] in clinical studies, amphetamine rotations here). Although in the present study, grafts contained ~14,000 TH+ cells (whereas rats have ~10-12,000 TH<sup>+</sup> cells/hemisphere),<sup>28,29</sup> in patients GIDs have been induced from grafts of  $\sim$ 37,000 TH cells<sup>5</sup> (human brain harbors ~170,000 substantia nigra pars compacta [SNpc] DA neurons)<sup>30</sup> and without evidence of excess dopamine in PET scans.

There has remained an unanswered question about the temporal pattern of GID emergence clinically. One supposition is that it relates to the withdrawal of immunosuppression, potentially triggering inflammatory drivers,<sup>6</sup> which would be consistent with data demonstrating that DA can mediate immune responses.<sup>31</sup> In the current study, the large microglial inflammatory response was specific to brains with vmDA-containing grafts although a direct causal relationship between neuroinflammation and GIDs could not be determined. Alternatively, GID emergence could relate to the time that it takes for DAergic neurons to begin significant maturation and outgrowth as a similar timescale occurred in a trial, which did not immunosuppress graft recipients.<sup>5</sup>

The use of neonatal desensitization makes this the first behavioral study of extended (12 months) human vmDA graft survival times in immunologically intact animals (in the absence of immunosuppression). Graft survival was low in this study (n = 4 grafts/group), but variability within this model has been reported previously.<sup>32,33</sup> Moreover, all published literature using this model has reported graft survival for a maximum of 6 months postgraft.<sup>20,22</sup> The data presented here are, to our knowledge, the first report of human-to-rat graft survival up to 12 months post-graft. Even if protection conferred by neonatal desensitization may reduce with time, this model has nevertheless been highly effective in revealing a previously unobserved phenomenon not readily investigated in standard immune-compromised/suppressed animals. The severity of spontaneous GIDs in these rats were mild-to-moderate in magnitude when compared to LID established in standard protocols in rodents<sup>21</sup> and did not appear to worsen over the observed time. Importantly, if LIDs are not the sole driver for GIDs, then the risks to late-stage patients may not be significantly elevated, whereas their benefits, such as reductions in LID, could be substantially more than early-stage patients. Therefore, this therapy may be open to a broader population of people with PD.

Importantly for future clinical trials, although stemcell based therapies can be designed to be completely devoid of 5-HT neurons and, therefore, may be considered safer from a GID perspective,<sup>24</sup> our data suggest that this may not completely eliminate the risk for GIDs. The mechanism of action of spontaneous GIDs needs further investigation in larger studies, but if they can be mediated through excess dopamine and/or inflammatory responses, it may be possible to mitigate these through design of the therapeutic delivery.

#### Conclusion

This is the first study to demonstrate that stem cellbased therapies can induce spontaneous GIDs in a rodent model of PD, simultaneous with functional recovery. The data demonstrate that GID onset can occur independently of L-dopa exposure and 5-HT neurons in the graft. Instead, the data presented here suggest involvement of D<sub>2</sub>-like family receptors and suggest that GIDs may be associated with graft inflammation.

**Acknowledgments:** We thank Prof. Malin Parmar, Dr. Agnete Kirkeby and Dr. Sara Nolbrant for the generous supply of the hESC-derived dopamine cells for tolerization and transplantation. We also thank Dr. Thomas Bjorklund for the kind gift of ST1936 drug.

#### **Data Availability Statement**

Data available on request from the authors

#### References

- Barker RA, Barrett J, Mason SL, Björklund A. Fetal dopaminergic transplantation trials and the future of neural grafting in Parkinson's disease. Lancet Neurol 2013;12:84–91.
- Li W, Englund E, Widner H, et al. Extensive graft-derived dopaminergic innervation is maintained 24 years after transplantation in the degenerating parkinsonian brain. Proc Natl Acad Sci 2016;113: 6544–6549.
- Hagell P, Piccini P, Björklund A, et al. Dyskinesias following neural transplantation in Parkinson's disease. Nat Neurosci 2002;5: 627–628.
- Olanow CW, Gracies J-M, Goetz CG, et al. Clinical pattern and risk factors for dyskinesias following fetal nigral transplantation in Parkinson's disease: a double blind video-based analysis. Mov Disord 2009;24:336–343.
- Greene PE, Fahn S, Eidelberg D, Bjugstad KB, Breeze RE, Freed CR. Persistent dyskinesias in patients with fetal tissue transplantation for Parkinson disease. npj Parkinson's Dis 2021;7:38

- Piccini P, Pavese N, Hagell P, et al. Factors affecting the clinical outcome after neural transplantation in Parkinson's disease. Brain 2005;128:2977–2986.
- Soderstrom KE, Meredith G, Freeman TB, et al. The synaptic impact of the host immune response in a parkinsonian allograft rat model: influence on graft-derived aberrant behaviors. Neurobiol Dis 2008; 32:229–242.
- Soderstrom KE, O'Malley JA, Levine ND, Sortwell CE, Collier TJ, Steece-Collier K. Impact of dendritic spine preservation in medium spiny neurons on dopamine graft efficacy and the expression of dyskinesias in parkinsonian rats. Eur J Neurosci 2010;31:478–490.
- Politis M, Wu K, Loane C, et al. Serotonergic neurons mediate dyskinesia side effects in Parkinson's patients with neural transplants. Sci Transl Med 2010;2:38ra46
- Pagano G, Niccolini F, Politis M. The serotonergic system in Parkinson's patients with dyskinesia: evidence from imaging studies. J Neural Transm 2018;125:1217–1223.
- Shin E, Garcia J, Winkler C, Björklund A, Carta M. Serotonergic and dopaminergic mechanisms in graft-induced dyskinesia in a rat model of Parkinson's disease. Neurobiol Dis 2012;47:393–406.
- Politis M, Oertel WH, Wu K, et al. Graft-induced dyskinesias in Parkinson's disease: high striatal serotonin/dopamine transporter ratio. Mov Disord 2011;26:1997–2003.
- Aldrin-Kirk P, Heuer A, Wang G, Mattsson B, Lundblad M, Parmar M, Björklund T. DREADD modulation of transplanted DA neurons reveals a novel parkinsonian dyskinesia mechanism mediated by the serotonin 5-HT6 receptor. Neuron 2016;90:955–968.
- Lane EL, Vercammen L, Cenci MA, Brundin P. Priming for L-DOPA-induced abnormal involuntary movements increases the severity of amphetamine-induced dyskinesia in grafted rats. Exp Neurol 2009;219:355–358.
- Steece-Collier K, Collier TJ, Danielson PD, Kurlan R, Yurek DM, Sladek JR Jr. Embryonic mesencephalic grafts increase levodopainduced forelimb hyperkinesia in Parkinsonian rats. Mov Disord 2003;18:1442–1454.
- Kriks S, Shim J-W, Piao J, et al. Dopamine neurons derived from human ES cells efficiently engraft in animal models of Parkinson's disease. Nature 2011;480:547–551.
- Kirkeby A, Grealish S, Wolf DA, et al. Generation of regionally specified neural progenitors and functional neurons from human embryonic stem cells under defined conditions. Cell Rep 2012;1:703–714.
- Lelos MJ, Morgan RJ, Kelly CM, Torres EM, Rosser AE, Dunnett SB. Amelioration of non-motor dysfunctions after transplantation of human dopamine neurons in a model of Parkinson's disease. Exp Neurol 2016;278:54–61.
- Grealish S, Heuer A, Cardoso T, et al. Monosynaptic tracing using modified rabies virus reveals early and extensive circuit integration of human embryonic stem cell-derived neurons. Stem Cell Rep 2015;4:975–983.
- Heuer A, Kirkeby A, Pfisterer U, Jönsson ME, Parmar M. hESCderived neural progenitors prevent xenograft rejection through neonatal desensitisation. Exp Neurol 2016;282:78–85.
- Breger LS, Dunnett SB, Lane EL. Comparison of rating scales used to evaluate l-DOPA-induced dyskinesia in the 6-OHDA lesioned rat. Neurobiol Dis 2013;50:142–150.
- Kelly CM, Precious SV, Scherf C, et al. Neonatal desensitization allows long-term survival of neural xenotransplants without immunosuppression. Nat Methods 2009;6:271–273.
- 23. Loane C, Politis M. Buspirone: what is it all about? Brain Res 2012; 1461:111–118.
- 24. Henchcliffe C, Parmar M. Repairing the brain: cell replacement using stem cell-based technologies. J Parkinsons Dis 2018;8: S131–S137.
- 25. Barker RA, TRANSEURO Consortium. Designing stem-cell-based dopamine cell replacement trials for Parkinson's disease. Nat Med 2019;25:1045–1053.
- Doi D, Samata B, Katsukawa M, et al. Isolation of human induced pluripotent stem cell-derived dopaminergic progenitors by cell sorting for successful transplantation. Stem Cell Rep 2014;2:337–350.

- 27. Fan Y, Winanto, Ng SY. Replacing what's lost: a new era of stem cell therapy for Parkinson's disease. Transl Neurodegener 2020;9:2
- Nair-Roberts RG, Chatelain-Badie SD, Benson E, White-Cooper H, Bolam JP, Ungless MA. Stereological estimates of dopaminergic, GABAergic and glutamatergic neurons in the ventral tegmental area, substantia nigra and retrorubral field in the rat. Neuroscience 2008; 152:1024–1031.
- 29. Healy-Stoffel M, Omar Ahmad S, Stanford JA, Levant B. Differential effects of intrastriatal 6-hydroxydopamine on cell number and morphology in midbrain dopaminergic subregions of the rat. Brain Res 2014;1574:113–119.
- Chu Y, Kompoliti K, Cochran EJ, Mufson EJ, Kordower JH. Agerelated decreases in Nurr1 immunoreactivity in the human substantia nigra. J Comp Neurol 2002;450:203–214.
- Matt SM, Gaskill PJ. Where is dopamine and how do immune cells see it?: dopamine-mediated immune cell function in health and disease. J Neuroimmune Pharmacol 2019;15:114–164. https://doi.org/ 10.1007/s11481-019-09851-4
- 32. Janowski M, Jablonska A, Kozlowska H, et al. Neonatal desensitization does not universally prevent xenograft rejection. Nat Methods 2012;9:856–858.
- Roberton VH, Rosser AE, Kelly CM. Neonatal desensitization for the study of regenerative medicine. Regen Med 2015;10:265–274.

## Supporting Data

Additional Supporting Information may be found in the online version of this article at the publisher's web-site.

# Dysprosody in Isolated REM Sleep Behavior Disorder with Impaired Olfaction but Intact Nigrostriatal Pathway

Jan Rusz, PhD,<sup>1,2\*</sup> <sup>(1)</sup> Annette Janzen, MD,<sup>3</sup> <sup>(1)</sup> Tereza Tykalová, PhD,<sup>1</sup> Michal Novotný, PhD,<sup>1</sup> David Zogala, PhD,<sup>4</sup> Lars Timmermann, MD,<sup>3</sup> Evžen Růžička, DSc,<sup>2</sup> <sup>(2)</sup> Karel Šonka, DSc,<sup>2</sup> Petr Dušek, MD,<sup>2</sup> <sup>(1)</sup> and Wolfgang Oertel, MD<sup>3</sup>

\*Correspondence to: Dr. Jan Rusz, Department of Circuit Theory, Faculty of Electrical Engineering, Czech Technical University in Prague, Technická 2, 160 00, Prague 6, Czech Republic; E-mail: rusz. mz@gmail.com;ruszjan@fel.cvut.cz

Jan Rusz and Annette Janzen contributed equally to this work.

Petr Dušek and Wolfgang Oertel jointly supervised this work.

Relevant conflicts of interest/financial disclosures: L.T. received payments as a consultant for Boston Scientific between September 2018 and September 2021 and received honoraria as a speaker on symposia sponsored by UCB, Desitin, Boston Scientific, AbbVie, Novartis, GlaxoSmithKline, and DIAPLAN. The institution of L.T., not L.T. personally, received funding from Boston Scientific, the German Research Foundation, the German Ministry of Education and Research, and the Deutsche Parkinson Vereinigung. Neither Lars Timmermann nor any member of his family holds stocks, stock options, patents, or financial interests in any of the aforementioned companies or their competitors. W.O. was <sup>1</sup>Department of Circuit Theory, Faculty of Electrical Engineering, Czech Technical University in Prague, Prague, Czech Republic <sup>2</sup>Department of Neurology and Centre of Clinical Neuroscience, First Faculty of Medicine, Charles University and General University Hospital, Prague, Czech Republic <sup>3</sup>Department of Neurology, Philipps University Marburg, Marburg, Germany <sup>4</sup>Institute of Nuclear Medicine, First Faculty of Medicine, Charles University and General University Hospital, Prague, Czechia

**ABSTRACT: Background:** Impairments of olfactory and speech function are likely early prodromal symptoms of  $\alpha$ -synucleinopathy.

**Objective:** The aim of this study is to assess whether dysprosody is present in isolated rapid eye movement sleep behavior disorder (iRBD) with hyposmia/anosmia and a normal nigrostriatal system.

**Methods:** Pitch variability during speech was investigated in 17 iRBD subjects with normal olfactory function (iRBD-NOF), 30 iRBD subjects with abnormal olfactory function (iRBD-AOF), and 50 healthy controls. iRBD subjects were evaluated using the University of Pennsylvania Smell Identification Test and [123I]-28-carbomethoxy-38-(4-iodophenyl)-*N*-(3-fluoropropyl)-nortropane dopamine transporter single-photon emission computed tomography (DAT-SPECT). All iRBD subjects completed the 24-month follow-up with DAT-SPECT, speech, and olfactory testing.

**Results:** At baseline, only iRBD-AOF showed monopitch when compared to iRBD-NOF (P = 0.04) and controls (P = 0.03), with no difference between iRBD-NOF and controls (P = 1). At follow-up, dysprosody progressed only in iRBD-AOF with abnormal DAT-SPECT (P = 0.03). **Conclusion:** Prosody is impaired in hyposmic but not in normosmic iRBD subjects before the nigrostriatal dopaminergic transmission is affected (Braak stage 2). © 2021 International Parkinson and Movement Disorder Society

Key Words: dysarthria; speech disorder; olfactory; dopamine transporter imaging; prodromal synucleinopathy biomarker

supported by scientific grants from the Charitable Hertie Foundation, the German Research Foundation, The Michael J. Fox Foundation, Stichting ParkinsonFonds the Netherlands, and ParkinsonFonds Deutschland and received payments as a consultant for Adamas and Modag and honoraria as a speaker on symposia sponsored by AbbVie, Bial, Desitin, and Roche. The other authors have nothing to disclose.

**Funding agencies:** This study was supported by the Czech Ministry of Health (grant nos. NU20-08-00445 and MH CZ-DRO-VFN64165). W.O. is Hertie-Senior-Research Professor, supported by the Charitable Hertie Foundation, Frankfurt/Main, Germany. W.O. and A.J. were supported by the ParkinsonFonds Deutschland.

Received: 16 September 2021; Revised: 1 November 2021; Accepted: 8 November 2021

Published online 27 November 2021 in Wiley Online Library (wileyonlinelibrary.com). DOI: 10.1002/mds.28873