



Genomic Sequence of a New Alphavirus Detected in Comber (Serranus cabrilla)

^(D)Gianpiero Zamperin,^a Adelaide Milani,^a Michele Gastaldelli,^b Rosita Quartesan,^a Andrea Fortin,^a Elisabetta Cappellozza,^c Pierpaolo Patarnello,^d ^(D)Anna Toffan^{a,b}

^aIstituto Zooprofilattico Sperimentale delle Venezie (IZSVe), Department of Comparative Biomedical Sciences, Legnaro, Padua, Italy ^bIstituto Zooprofilattico Sperimentale delle Venezie (IZSVe), Specialistic Aquatic Animal Health Centre, Legnaro, Padua, Italy ^cIstituto Zooprofilattico Sperimentale delle Venezie (IZSVe), Health Awareness and Communication Department, Legnaro, Padua, Italy ^dLocal Veterinary Services (ASL), Lecce, Italy

ABSTRACT The comber alphavirus was isolated from a fish cell line from the brain of an apparently healthy *Serranus cabrilla* specimen collected during wild fish surveillance in southern Italy. The comber alphavirus is a new member of the genus *Alphavirus*, family *Togaviridae*.

Iphavirus is the only genus presently included in the Togaviridae family. Alphaviruses are enveloped single-stranded positive-sense RNA genomes ranging from 9.7 to 11.8 kb (1). This genus comprises a large number of species that are mostly mosquito borne and pathogenic for mammals. In the aquatic environment, the species Salmonid alphavirus can cause severe diseases in Atlantic salmon and rainbow trout (1). Comber (Serranus cabrilla) is a small fish belonging to the family Serranidae, inhabiting the Mediterranean Sea. It can be found along the coast feeding on fish, cephalopods, and crustaceans. From 2010 to 2014, 32 S. cabrilla fish were caught during monitoring activity on the Salento coast (Ionian Sea). Comber brains were collected and subjected to viral isolation on a striped snakehead cell line (SSN-1) (2) at 25°C and a bluegill fry cell line (BF-2) (3) at 20°C according to standard procedures (4, 5). One sample produced clear cytopathic effect on both lines, with round and refractive cells detaching from the bottom of the flask up to complete monolayer destruction after 7 to 10 days. Transmission electron microscope observation revealed that the virus was pleomorphic, enveloped, and 55 to 70 nm in diameter with an icosahedral capsid 40 to 45 nm in diameter, resembling members of the Togaviridae family (Fig. 1). The new virus was called comber alphavirus isolate 12ITT-210.14.

Virions were precipitated from BF-2 cell supernatant with polyethylene glycol and then purified using ultracentrifugation on a 40 to 70% sucrose gradient to obtain a visible viral band, which was diluted in 10 ml of TRIS-HCI-EDTA-NaCl (TEN) buffer, pelleted by ultracentrifugation, and, finally, resuspended in 1 ml of TEN buffer (6). Total RNA was extracted using the total RNA isolation kit NucleoSpin RNA II (Macherey-Nagel) and converted into first- and second-strand cDNA using SuperScript III reverse transcriptase (Invitrogen) and second-strand cDNA synthesis (NEB), respectively. Sequencing libraries were generated using a Nextera DNA XT sample preparation kit (Illumina) and processed with a MiSeq reagent kit v2 (2×250 -bp paired-end [PE] mode; Illumina). Sequencing yielded 1,138,673 251-bp-long paired-end reads, which were quality filtered by (i) removing reads with more than 100 bases with a Q score below 7 and duplicated paired-end reads, (ii) clipping adaptors with Scythe v0.991 (https://github.com/vsbuffalo/scythe), (iii) trimming low-quality 3' ends with Sickle v1.33 (https://github.com/najoshi/sickle), and (iv) discarding reads shorter than 80 bases and unpaired reads. After filtering, we recovered 911,779 high-quality reads, which

Citation Zamperin G, Milani A, Gastaldelli M, Quartesan R, Fortin A, Cappellozza E, Patarnello P, Toffan A. 2020. Genomic sequence of a new *Alphavirus* detected in comber (*Serranus cabrilla*). Microbiol Resour Announc 9:e01294-19. https://doi.org/10.1128/MRA.01294-19.

Editor Jelle Matthijnssens, KU Leuven

Copyright © 2020 Zamperin et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Anna Toffan, atoffan@izsvenezie.it.

Received 14 October 2019 Accepted 6 December 2019 Published 9 January 2020



FIG 1 Togavirus-like particles observed with transmission electron microscopy (Philips EM208S) on BF-2 cell culture infected supernatant at magnifications of $22,000 \times$ (a) and $44,000 \times$ (b).

were taxonomically classified by (i) alignment against the integrated nucleotide (NT) database (26 January 2016 version) using BLAST v2.3.0+ (7) with default parameters and against the integrated nonredundant (NR) database (26 January 2016 version) using DIAMOND v0.8.16 (8) with default parameters, (ii) filtering out alignment hits with an E value larger than 1×10^{-3} , and (iii) feeding the remaining alignment hits to MEGAN v6.5.8 (9). Reads belonging to the *Alphavirus* genus were selected and *de novo* assembled using IDBA-UD v1.1.1 (10) with default parameters and the multi-*k*-mer approach (minimum value, 114; maximum value, 124; increment, 10). Only a single contig with a length comparable to the size of the *Alphavirus* genue was obtained. All reads belonging to the *Alphavirus* genus were subsequently aligned against the longest contig obtained from the *de novo* assembly using Burrows-Wheeler Aligner (BWA) v0.7.12 (11) with standard parameters. The alignment was manually revised with Tablet (12) to avoid the risk of misassembly.

The final genome obtained resulted in a consensus sequence 11,604 nucleotides long showing a sequence coverage of 30,221-fold and a 56.4% G+C content. A comparison performed using BLAST (online version, blastn algorithm) with full *Alpha-virus* genomes available in GenBank (July 2019) revealed 59% query coverage and 67.2% identity with *Salmonid alphavirus* subtype 3 (GenBank accession number KC122924).

Open reading frames (ORFs) were determined using the NCBI tool ORFfinder (https://www.ncbi.nlm.nih.gov/orffinder/). It detected two distinct ORFs, an ORF in the 5' two-thirds of the genome and a second one in the 3' one-third of the genome, in agreement with the genomic content organization of other known alphaviruses.

Herein, we report the first description and the almost full-genome sequence of a new member of the genus *Alphavirus*, isolated from a wild Mediterranean fish.

Data availability. MiSeq raw data were submitted to the NCBI Sequence Read Archive (SRA) under the accession number SRR9718464. The comber *Alphavirus* genome sequence has been deposited in GenBank under the accession number MN207265.

ACKNOWLEDGMENTS

The comber samples were collected during wild fish surveillance activities funded by project IZSVE RC 07/10, and the nearly complete genome sequence was funded by project IZSVE RC 08/16, both from the Italian Ministry of Health.

REFERENCES

- Chen R, Mukhopadhyay S, Merits A, Bolling B, Nasar F, Coffey LL, Powers A, Weaver SC, ICTV Report Consortium. 2018. ICTV virus taxonomy profile: *Togaviridae*. J Gen Virol 99:761–762. https://doi.org/10.1099/jgv .0.001072.
- Frerichs GN, Rodger HD, Peric Z. 1996. Cell culture isolation of piscine neuropathy nodavirus from juvenile seabass, *Dicentrarchus labrax*. J Gen Virol 77:2067–2071. https://doi.org/10.1099/0022-1317-77-9-2067.
- Wolf K, Quimby MC. 1962. Established eurythermic line of fish cells in vitro. Science 135:1065–1066. https://doi.org/10.1126/science.135.3508 .1065.
- 4. OIE (World Organization for Animal Health). 2017. Chapter 2.3.12. Viral encephalopathy and retinopathy. Manual of diagnostic tests for aquatic animals. World Organisation for Animal Health, Paris, France.
- Pascoli F, Bilò F, Nonnis Marzano F, Borghesan F, Mancin M, Manfrin A, Toffan A. 2015. Susceptibility of genotyped marble trout *Salmo marmoratus* (Cuvier, 1829) strains to experimental challenge with European viral hemorrhagic septicemia virus (VHSV) and infectious hematopoietic necrosis virus (IHNV). Aquaculture 435:152–156. https://doi.org/10.1016/ j.aquaculture.2014.09.038.
- Panzarin V, Toffan A, Abbadi M, Buratin A, Mancin M, Braaen S, Olsen CM, Bargelloni L, Rimstad E, Cattoli G. 2016. Molecular basis for antigenic diversity of genus *Betanodavirus*. PLoS One 11:e0158814. https://doi.org/ 10.1371/journal.pone.0158814.

A Microbiolog

- Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. J Mol Biol 215:403–410. https://doi.org/10.1016/ S0022-2836(05)80360-2.
- Buchfink B, Xie C, Huson DH. 2015. Fast and sensitive protein alignment using DIAMOND. Nat Methods 12:59–60. https://doi.org/10.1038/nmeth .3176.
- Huson DH, Beier S, Flade I, Górska A, El-Hadidi M, Mitra S, Ruscheweyh HJ, Tappu R. 2016. MEGAN community edition: interactive exploration and analysis of large-scale microbiome sequencing data. PLoS Comput Biol 12:e1004957-12. https://doi.org/10.1371/journal.pcbi.1004957.
- Peng Y, Leung HCM, Yiu SM, Chin F. 2012. IDBA-UD: a *de novo* assembler for single-cell and metagenomic sequencing data with highly uneven depth. Bioinformatics 28:1420–1428. https://doi.org/10.1093/bioinformatics/ bts174.
- Li H, Durbin R. 2010. Fast and accurate long-read alignment with Burrows-Wheeler transform. Bioinformatics 26:589–595. https://doi.org/ 10.1093/bioinformatics/btp698.
- Milne I, Stephen G, Bayer M, Cock PJA, Pritchard L, Cardle L, Shaw PD, Marshall D. 2013. Using Tablet for visual exploration of secondgeneration sequencing data. Brief Bioinform 14:193–202. https://doi.org/ 10.1093/bib/bbs012.