

Genome Sequence of the Anterograde-Spread-Defective Herpes Simplex Virus 1 Strain MacIntyre

 Moriah L. Szpara,^{a,c*} Yolanda R. Tafuri,^a Lance Parsons,^b Jacob T. Shreve,^c Esteban A. Engel,^a L. W. Enquist^{a,b}

Department of Molecular Biology, the Princeton Neuroscience Institute^a and Lewis-Sigler Institute for Integrative Genomics,^b Princeton University, Princeton, New Jersey, USA; Department of Biochemistry and Molecular Biology, Pennsylvania State University, University Park, Pennsylvania, USA^c

* Present address: Moriah L. Szpara, Huck Institutes of the Life Sciences, Pennsylvania State University, University Park, Pennsylvania, USA.

We used paired-end Illumina deep sequencing and *de novo* assembly to determine the genome sequence of herpes simplex virus 1 (HSV-1) strain MacIntyre (aka McIntyre). The MacIntyre strain originated from the brain of a patient with lethal HSV encephalitis and has a unique limitation in its neuronal spread, moving solely in the retrograde direction.

Received 29 September 2014 Accepted 8 October 2014 Published 13 November 2014

Citation Szpara ML, Tafuri YR, Parsons L, Shreve JT, Engel EA, Enquist LW. 2014. Genome sequence of the anterograde-spread-defective herpes simplex virus 1 strain MacIntyre. *Genome Announc.* 2(6):e01161-14. doi:10.1128/genomeA.01161-14.

Copyright © 2014 Szpara et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](#).

Address correspondence to Moriah L. Szpara, moriah@psu.edu.

We used Illumina deep sequencing and *de novo* assembly to determine the genome sequence of herpes simplex virus 1 (HSV-1) strain MacIntyre (also spelled MacIntyre B, McIntyre, and McIntyre B). The MacIntyre strain originated from the brain of a patient with lethal HSV encephalitis (1). After isolation, MacIntyre was passaged through multiple cell types and species (1, 2). Dowdle (2, 3) deposited this strain at the ATCC (VR-539), from which we received our stock. Later studies revealed a severe defect in the anterograde spread of HSV-1 MacIntyre in the central nervous systems in rat, mouse, and primate models (4–12). After replication in the nucleus, HSV-1 MacIntyre egresses and infects only neurons that are presynaptic to the infected neuron. This is in contrast to wild-type HSV-1, which egresses and infects neurons both pre- and postsynaptic to the infected one.

We isolated viral nucleocapsid DNA from HSV-1 MacIntyre-infected Vero cells, using published methods (13–15). This was performed according to the manufacturer's protocols (Illumina TruSeq DNA) to produce a bar-coded library of 500-bp fragments and to obtain 100-bp paired-end sequence reads (Illumina HiSeq). A series of quality control filters removed the sequences resulting from Illumina primers and adaptors, contaminating Vero cell DNA, and low-quality terminal bases (16, 17). We then generated eight SSAKE *de novo* assemblies using varied parameters, and we combined these into a draft genome using Celera and GapFiller (18–21). We transferred annotations from the HSV reference genome (strain 17; GenBank accession no. JN555585) to the MacIntyre strain based on sequence homology (22, 23). Three genes, UL46, US7, and US9, contained new stop codon positions due to homopolymer-based frameshifts or single nucleotide variations. Two genes (RL1 and RS1) were undetermined due to gaps in the assembly. As in prior studies (17, 24–29), we found that a majority of HSV MacIntyre proteins have coding variations compared to those of the HSV-1 reference strain 17. We anticipate that the HSV-1 MacIntyre genome contains bystander variations and one or more mutations that directly affect its limited-spread phenotype.

A frequent comparator for HSV-1 is the distantly related swine alphaherpesvirus pseudorabies virus (PRV) (30, 31). HSV-1 MacIntyre resembles the PRV vaccine strain Bartha in terms of its defect in anterograde spread, extensive passage history, and attenuated virulence (32, 33). We recently sequenced the full genome of PRV strain Bartha, allowing us to explore how these two viruses converged on the same phenotype of defective spread in neurons (16). The anterograde spread defect of PRV-Bartha results from loss of three proteins, gE (US6), gI (US7), and US9. The loss of US9 alone strongly affects sorting into neuronal axons (34–37). Our sequence data reveal that HSV-1 MacIntyre contains a single nucleotide polymorphism in US9, which creates a premature stop codon (C172T or R58Stop). We have confirmed this by PCR and Western blot analysis (data not shown). Curiously, an identical US9 mutation was previously described in two additional unrelated HSV-1 strains (38, 39). Further characterization of this and other differences in HSV-1 MacIntyre is under way with an aim of illuminating the mechanisms of neuronal sorting and egress for HSV-1.

Nucleotide sequence accession number. The HSV-1 MacIntyre strain genome sequence has been deposited at GenBank under the accession no. [KM222720](#).

ACKNOWLEDGMENTS

This work was supported by the Virus Pathogens Resource (ViPR), a Bioinformatics Resource Center (BRC) funded by the NIAID. Additional support was provided by a Center Grant (NIH/NIGMS P50 GM071508) and NIH grant P40 RR 018604 (to L.W.E. and M.L.S.). E.A.E. is a Pew Latin American Fellow in the Biomedical Sciences (grant 2010-000225-002), supported by the Pew Charitable Trusts.

REFERENCES

1. Sosa-Martinez J, Lennette EH. 1955. Studies on a complement fixation test for herpes simplex. *J. Bacteriol.* 70:205–215.
2. Dowdle WR, Nahmias, Harwell RW, Pauls FP. 1967. Association of antigenic type of *Herpesvirus hominis* with site of viral recovery. *J. Immunol.* 99:974–980.

3. Pauls FP, Dowdle WR. 1967. A serologic study of *Herpesvirus hominis* strains by microneutralization tests. *J. Immunol.* 98:941–947.
4. Cook ML, Stevens JG. 1973. Pathogenesis of herpetic neuritis and ganglionitis in mice: evidence for intra-axonal transport of infection. *Infect. Immun.* 7:272–288.
5. Pepose JS, Foos RY, Stevens JG. 1986. Herpes simplex virus immediate-early protein ICP4 in murine models of latency. *Graefes Arch. Clin. Exp. Ophthalmol.* 224:341–345. <http://dx.doi.org/10.1007/BF02150027>.
6. McFarland DJ, Hotchin J. 1987. Contrasting patterns of virus spread and neuropathology following microinjection of herpes simplex virus into the hippocampus or cerebellum of mice. *J. Neurol. Sci.* 79:255–265. [http://dx.doi.org/10.1016/0022-510X\(87\)90233-4](http://dx.doi.org/10.1016/0022-510X(87)90233-4).
7. McLean JH, Shipley MT, Bernstein DI. 1989. Golgi-like, transneuronal retrograde labelling with CNS injections of herpes simplex virus type 1. *Brain. Res. Bull.* 22:867–881. [http://dx.doi.org/10.1016/0361-9230\(89\)90032-4](http://dx.doi.org/10.1016/0361-9230(89)90032-4).
8. Krinke GJ, Dietrich FM. 1990. Transneuronal spread of intraperitoneally administered herpes simplex virus type 1 from the abdomen via the vagus nerve to the brains of mice. *J. Comp. Pathol.* 103:301–306. [http://dx.doi.org/10.1016/S0021-9975\(08\)80050-3](http://dx.doi.org/10.1016/S0021-9975(08)80050-3).
9. Garner JA, LaVail JH. 1999. Differential anterograde transport of HSV type 1 viral strains in the murine optic pathway. *J. Neurovirol.* 5:140–150. <http://dx.doi.org/10.3109/13550289909021996>.
10. LaVail JH, Topp KS, Giblin PA, Garner JA. 1997. Factors that contribute to the transneuronal spread of herpes simplex virus. *J. Neurosci. Res.* 49:485–496. [http://dx.doi.org/10.1002/\(SICI\)1097-4547\(19970815\)49:4<485::AID-JNR9>3.3.CO;2-N](http://dx.doi.org/10.1002/(SICI)1097-4547(19970815)49:4<485::AID-JNR9>3.3.CO;2-N).
11. Zemanick MC, Strick PL, Dix RD. 1991. Direction of transneuronal transport of herpes simplex virus 1 in the primate motor system is strain-dependent. *Proc. Natl. Acad. Sci. U. S. A.* 88:8048–8051. <http://dx.doi.org/10.1073/pnas.88.18.8048>.
12. Hoover JE, Strick PL. 1993. Multiple output channels in the basal ganglia. *Science* 259:819–821. <http://dx.doi.org/10.1126/science.7679223>.
13. Szpara ML, Tafuri YR, Enquist LW. 2011. Preparation of viral DNA from nucleocapsids. *J. Vis. J. Vis. Exp.* (54):3151. <http://dx.doi.org/10.3791/3151>.
14. Smith GA, Enquist LW. 1999. Construction and transposon mutagenesis in *Escherichia coli* of a full-length infectious clone of pseudorabies virus, an alphaherpesvirus. *J. Virol.* 73:6405–6414.
15. Enquist LW, Madden MJ, Schiop-Stanley P, Vande Woude GF. 1979. Cloning of herpes simplex type 1 DNA fragments in a bacteriophage lambda vector. *Science* 203:541–544. <http://dx.doi.org/10.1126/science.216076>.
16. Szpara ML, Tafuri YR, Parsons L, Shamim SR, Verstrepen KJ, Legendre M, Enquist LW. 2011. A wide extent of inter-strain diversity in virulent and vaccine strains of alphaherpesviruses. *PLoS Pathog.* 7:e1002282. <http://dx.doi.org/10.1371/journal.ppat.1002282>.
17. Szpara ML, Parsons L, Enquist LW. 2010. Sequence variability in clinical and laboratory isolates of herpes simplex virus 1 reveals new mutations. *J. Virol.* 84:5303–5313. <http://dx.doi.org/10.1128/JVI.00312-10>.
18. Myers EW, Sutton GG, Delcher AL, Dew IM, Fasulo DP, Flanigan MJ, Kravitz SA, Mobarry CM, Reinert KH, Remington KA, Anson EL, Bolanos RA, Chou HH, Jordan CM, Halpern AL, Lonardi S, Beasley EM, Brandon RC, Chen L, Dunn PJ, Lai Z, Liang Y, Nusskern DR, Zhan M, Zhang Q, Zheng X, Rubin GM, Adams MD, Venter JC. 2000. A whole-genome assembly of *Drosophila*. *Science* 287:2196–2204. <http://dx.doi.org/10.1126/science.287.5461.2196>.
19. Boetzer M, Pirovano W. 2012. Toward almost closed genomes with GapFiller. *Genome Biol.* 13:R56. <http://dx.doi.org/10.1186/gb-2012-13-6-r56>.
20. Warren RL, Sutton GG, Jones SJM, Holt Ra. 2007. Assembling millions of short DNA sequences using SSAKE. *Bioinformatics* 23:500–501. <http://dx.doi.org/10.1093/bioinformatics/btl629>.
21. Cunningham C, Gatherer D, Hilfrich B, Baluchova K, Derrick J, Thomson M, Griffiths PD, Wilkinson GWG, Schulz TF, Dargan DJ, Davison AJ. 2009. Sequences of complete human cytomegalovirus genomes from infected cell cultures and clinical specimens. *J. Gen. Virol.* 91:605–615. <http://dx.doi.org/10.1099/vir.0.015891-0>.
22. McGeoch DJ, Dalrymple MA, Davison AJ, Dolan A, Frame MC, McNab D, Perry LJ, Scott JE, Taylor P. 1988. The complete DNA sequence of the long unique region in the genome of herpes simplex virus type 1. *J. Gen. Virol.* 69:1531–1574. <http://dx.doi.org/10.1099/0022-1317-69-7-1531>.
23. Mcgeoch DJ, Dolan A, Donald S, Street C. 1986. Complete DNA sequence of the short repeat region in the genome of herpes simplex virus type 1. *Nucleic Acids Res.* 14:1727–1746. <http://dx.doi.org/10.1093/nar/14.4.1727>.
24. Kolb AW, Adams M, Cabot EL, Craven M, Brandt CR. 2011. Multiplex sequencing of seven ocular herpes simplex virus type-1 genomes: phylogeny, sequence variability, and SNP distribution. *Invest. Ophthalmol. Vis. Sci.* 52:9061–9073. <http://dx.doi.org/10.1167/iov.11-7812>.
25. Watson G, Xu W, Reed A, Babra B, Putman T, Wick E, Wechsler SL, Rohrman GF, Jin L. 2012. Sequence and comparative analysis of the genome of HSV-1 strain McKrae. *Virology* 433:528–537. <http://dx.doi.org/10.1016/j.virol.2012.08.043>.
26. Macdonald SJ, Mostafa HH, Morrison La, Davido DJ. 2012. Genome sequence of herpes simplex virus 1 strain KOS. *J. Virol.* 86:6371–6372. <http://dx.doi.org/10.1128/JVI.00646-12>.
27. Macdonald SJ, Mostafa HH, Morrison La, Davido DJ. 2012. Genome sequence of herpes simplex virus 1 strain McKrae. *J. Virol.* 86:9540–9541. <http://dx.doi.org/10.1128/JVI.01469-12>.
28. Ushijima Y, Luo C, Goshima F, Yamauchi Y, Kimura H, Nishiyama Y. 2007. Determination and analysis of the DNA sequence of highly attenuated herpes simplex virus type 1 mutant HF10, a potential oncolytic virus. *Microbes Infect.* 9:142–149. <http://dx.doi.org/10.1016/j.micinf.2006.10.019>.
29. Szpara ML, Gatherer D, Ochoa A, Greenbaum B, Dolan A, Bowden RJ, Enquist LW, Legendre M, Davison AJ. 2014. Evolution and diversity in human herpes simplex virus genomes. *J. Virol.* 88:1209–1227. <http://dx.doi.org/10.1128/JVI.01987-13>.
30. Ugolini G, Wouterlood FG. 2010. Advances in viral transneuronal tracing. *J. Neurosci. Methods* 194:1–19. <http://dx.doi.org/10.1016/j.jneumeth.2009.12.001>.
31. Curanovic D, Enquist LW. 2009. Directional transneuronal spread of alpha-herpesvirus infection. *Future Virol.* 4:591–603. <http://dx.doi.org/10.2217/fvl.09.62>.
32. Bartha A. 1961. Experimental reduction of virulence of Aujeszky's disease virus. *Magy. Allatorv. Lapja* 16:42–45.
33. Lomniczi B, Blankenship ML, Ben-Porat T. 1984. Deletions in the genomes of pseudorabies virus vaccine strains and existence of four isomers of the genomes. *J. Virol.* 49:970–979.
34. Brideau AD, Card JP, Enquist LW. 2000. Role of pseudorabies virus Us9, a type II membrane protein, in infection of tissue culture cells and the rat nervous system. *J. Virol.* 74:834–845. <http://dx.doi.org/10.1128/JVI.74.2.834-845.2000>.
35. Brideau AD, Eldridge MG, Enquist LW. 2000. Directional transneuronal infection by pseudorabies virus is dependent on an acidic internalization motif in the Us9 cytoplasmic tail. *J. Virol.* 74:4549–4561. <http://dx.doi.org/10.1128/JVI.74.10.4549-4561.2000>.
36. Lyman MG, Feierbach B, Curanovic D, Bisher M, Enquist LW. 2007. Pseudorabies virus Us9 directs axonal sorting of viral capsids. *J. Virol.* 81:11363–11371. <http://dx.doi.org/10.1128/JVI.01281-07>.
37. Lomniczi B, Watanabe S, Ben-Porat T, Kaplan AS. 1987. Genome location and identification of functions defective in the Bartha vaccine strain of pseudorabies virus. *J. Virol.* 61:796–801.
38. Haugo AC, Szpara ML, Parsons L, Enquist LW, Roller RJ. 2011. Herpes simplex virus 1 pUL34 plays a critical role in cell-to-cell spread of virus in addition to its role in virus replication. *J. Virol.* 85:7203–7215. <http://dx.doi.org/10.1128/JVI.00262-11>.
39. Negatsch A, Mettenleiter TC, Fuchs W. 2011. Herpes simplex virus type 1 strain KOS carries a defective US9 and a mutated US8A gene. *J. Gen. Virol.* 92:167–172. <http://dx.doi.org/10.1099/vir.0.026484-0>.