



## Dissemination of Carbapenemases (KPC, NDM, OXA-48, IMP, and VIM) Among Carbapenem-Resistant *Enterobacteriaceae* Isolated From Adult and Children Patients in China

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This study aimed to investigate the dissemination and characteristics of bla<sub>KPC</sub>, bla<sub>NDM</sub>, bla<sub>OXA-48-like</sub>. bla<sub>IMP</sub>, and bla<sub>VIM</sub> among the carbapenem-resistant Enterobacteriaceae (CRE) strains isolated from adult and children patients. A total of 935 non-duplicate CRE strains were collected from 36 hospitals in 24 provinces or cities across China from 2016 to 2018. Antimicrobial susceptibility testing was performed by broth microdilution method and carbapenemase genes blaKPC, blaNDM, blaOXA-48-like, blaIMP, and blaVIM were screened by PCR and confirmed by DNA sequencing. Overall, carbapenemases were produced in 97.4% (911/935) of CRE strains, including KPC-2 (51.6%, 482/935), NDM (35.7%, 334/935), and OXA-48-like carbapenemases (7.3%, 68/935). Overall, the most prevalent carbapenemase gene was blaKPC-2 among Klebsiella pneumoniae (64.6%, 457/709) and the CRE strains isolated from adult patients (70.3%, 307/437), and blaNDM among Escherichia coli (96.0%, 143/149) and the CRE strains from children (49.0%, 247/498). The bla<sub>OXA-232</sub>-positive carbapenem-resistant K. pneumoniae (9.3%, 66/709) were all isolated from children. Sixteen strains were positive for bla<sub>IMP</sub> and 9 strains produced multiple carbapenemases. No strain was positive for blavim. Most of the CRE strains (>90%) were resistant to cephalosporins and carbapenems, more than half (>50%) were resistant to aminoglycosides and fluoroquinolones, but the majority (95.8 and 98.4%) were susceptible to polymyxin B and tigecycline. Ceftazidime-avibactam showed excellent in vitro activity against blaKPC-2 and blaOXA-48-like positive strains (100% susceptible). In China, KPC-2, NDM, and OXA-48-like carbapenemases were predominant among CRE clinical isolates. The most prevalent carbapenemase gene was blaKPC-2 among K. pneumoniae isolates from adult patients, and blaNDM among E. coli isolates from children.

Keywords: carbapenem-resistant Enterobacteriaceae, blaKPC-2, blaNDM, blaOXA-48-like, blaIMP

## INTRODUCTION

Enterobacteriaceae are opportunistic pathogens causing severe hospital-acquired infections (Feil, 2016). The spread of carbapenemase-producing Enterobacteriaceae (CPE) has been a global threat to public health. Carbapenems have conventionally been used for treating infections caused by extended-spectrum β-lactamase-producing Escherichia coli and Klebsiella pneumoniae, and are still considered as last resort antibiotics to date (van Duin and Doi, 2016). According to the data from China Antimicrobial Surveillance Network (CHINET, www.chinets.com), the resistance rate of K. pneumoniae to meropenem and imipenem rapidly increased from 2.9 and 3.0% in 2005 to 26.3 and 25% in 2018, respectively. In Europe, carbapenem-resistant K. pneumoniae are most widespread in the Mediterranean and Balkan countries with a prevalence of 60% in Greece and 40% in Italy, respectively (Perez and Villegas, 2015; Feil, 2016). The production of carbapenemases including KPC, NDM, and OXA-48-like is the most common resistance mechanism among carbapenem-resistant Enterobacteriaceae clinical isolates (Nordmann et al., 2012; Goodman et al., 2016). The *bla*<sub>KPC</sub>-positive *Enterobacteriaceae* were widespread in the United States, Latin America, Italy, Greece, the Middle East, and China (Albiger et al., 2015; Feil, 2016; Villegas et al., 2016; Iovleva and Doi, 2017). The bla<sub>NDM</sub>-positive Enterobacteriaceae were widespread in India, Pakistan, Bangladesh, Italy, Poland, Denmark, Latin America, and African countries (Yong et al., 2009; Albiger et al., 2015; van Duin and Doi, 2016). The  $bla_{\rm OXA-48-like}$ -positive strains remained rare in the US, in contrast to the prevalence in Turkey, Spain, France, Belgium, Romania, Middle East, Africa, Asia, and South America as well (Albiger et al., 2015). These infections are usually associated with very poor prognosis and high mortality, especially in neonates or high-risk immunocompromised patients (Falagas et al., 2014; Feil, 2016). In China, the presence of *bla*<sub>KPC</sub> and *bla*<sub>NDM</sub> is responsible for phenotypic resistance in most of the CRE strains (Zhang et al., 2017; Wang et al., 2018). Most researches currently focus on the dissemination of carbapenemases among CRE strains isolated from adult patients, while only a few are available to investigate the distribution of carbapenemases among CRE strains isolated from children. To obtain the comprehensive characteristic of carbapenemases among CRE isolated from both adults and children patients in China, we conducted this study to characterize the dissemination and characteristics of carbapenemases (including KPC, NDM OXA-48, IMP, and VIM) among CRE clinical isolates and the susceptibility to antimicrobial agents.

#### MATERIALS AND METHODS

#### **Clinical Strains**

From January 2016 to December 2018, a total of 935 nonduplicate sequential CRE strains were collected from 36 hospitals in 24 provinces or cities across China (**Figure 1**), including *K. pneumoniae* (n = 709, 75.8%), *E. coli* (n = 149, 15.9%), *Enterobacter cloacae* (n = 36, 3.9%), *Citrobacter freundii* (n = 14, 1.5%), Serratia marcescens (n = 8, 0.9%), Enterobacter aerogenes (n = 7, 0.7%), Klebsiella oxytoca (n = 7, 0.7%), Morganella morganii (n = 3, 0.3%), Proteus vulgaris (n =1, 0.1%), Providencia rettgeri (n = 1, 0.1%). In this study, 46.7% (437/935) of CRE strains were collected from adult patients and 53.3% (498/935) from children patients. The Enterobacteriaceae strains resistant to at least one of the carbapenem antibiotics (ertapenem, meropenem, doripenem, or imipenem) or producing a carbapenemase (an enzyme that can make them resistant to carbapenem antibiotics) were defined as CRE by Centers for Disease Control and Prevention of USA (https://www.cdc.gov/hai/organisms/cre/technical-info. html#Definition). These CRE strains were isolated from sputum (27.5%), blood (27.1%), urine (17.0%), secreta (6.9%), bile (5.0%), ascites (3.2%), catheter (2.8%), drainage (2.8%), pus (1.4%) and other aseptic body fluid (6.4%). Species identification was confirmed by MALDI-TOF/MS system (bioMérieux, France). E. coli ATCC 25922, E. coli ATCC 35218, and K. pneumoniae ATCC 700603 were tested as the quality control strains for antimicrobial susceptibility testing.

#### Antimicrobial Susceptibility Testing (AST)

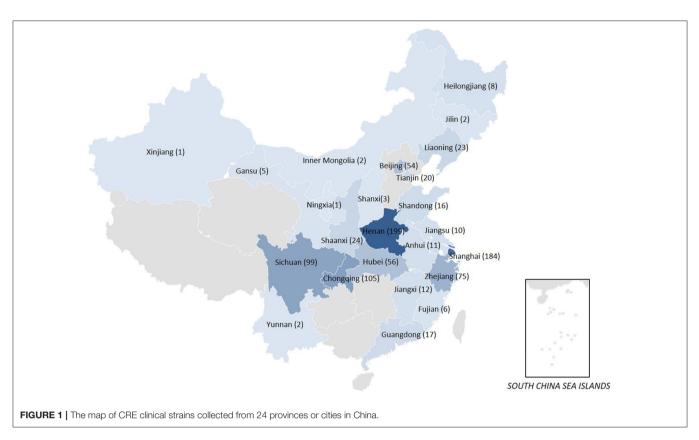
AST was performed by the broth microdilution method recommended by the Clinical and Laboratory Standards Institute. Minimum inhibitory concentrations (MICs) of piperacillin, cefoperazone-sulbactam, piperacillin-tazobactam, cefazolin, cefuroxime, ceftazidime, ceftriaxone, ceftazidime-avibactam, cefepime, cefmetazole, aztreonam, ertapenem, imipenem, meropenem, amikacin, gentamicin, ciprofloxacin, levofloxacin, trimethoprim-sulfamethoxazole, polymyxin B, nitrofurantoin, tigecycline were determined. The MIC breakpoints for *Enterobacteriaceae* (susceptible,  $\leq 2$  mg/L; resistant,  $\geq 8$  mg/L) issued by the Food and Drug Administration were used as the breakpoints for tigecycline.

# Detection of Carbapenemase and *mcr-1* Genes

All the CRE strains were tested for the presence of the most common carbapenemase genes ( $bla_{\rm KPC}$ ,  $bla_{\rm NDM}$ ,  $bla_{\rm OXA-48-like}$ ,  $bla_{\rm IMP}$ , and  $bla_{\rm VIM}$ ) by polymerase chain reaction (PCR) with specific primers and conditions as described previously (Poirel et al., 2011; Liu et al., 2016). The colistin resistance gene *mcr-1* was also detected by PCR, as previously described (Liu et al., 2016). The positive PCR amplicons were sequenced and compared with the reported sequences from GenBank by Blast (www.ncbi.nlm.nih.gov/blast/).

#### **Statistical Analysis**

Descriptive statistics were used to summarize the epidemiologic characteristics of CRE strains. For categorical variables, the percentage of CRE strains in each category was calculated. All analyses were performed using WHONET (version 5.6) and the IBM SPSS Statistics (version 21).



## RESULTS

## In vitro Antimicrobial Susceptibility

Most of the CRE strains (>90%) were resistant to cephalosporins, piperacillin, cefoperazone-sulbactam, piperacillin-tazobactam, aztreonam, and carbapenems. Overall, 61.4, 50.1, and 45.2% of the strains were susceptible to ceftazidime-avibactam, amikacin, and trimethoprim-sulfamethoxazole, respectively, followed by gentamicin (31.8%), levofloxacin (22.9%), ciprofloxacin (19%), and nitrofurantoin (18.8%). Polymyxin B and tigecycline showed excellent antibacterial activity against CRE strains (95.8 and 98.4% susceptible, respectively) (Table 1). Ceftazidimeavibactam had potent activity against both KPC-2-producing and OXA-48-like producing Enterobacteriaceae (100% susceptible) and inhibited all of blaKPC-2-positive and blaOXA-48-likepositive strains at 8 mg/L. However, all NDM-producing Enterobacteriaceae were resistant to ceftazidime-avibactam  $(MIC_{90} > 32 \text{ mg/L})$ . The MICs of ceftazidime-avibactam were higher than 32 mg/L against IMP- and multi-carbapenemase producing Enterobacteriaceae (KPC and NDM co-producers, NDM and OXA-48 co-producer). Most of the *bla*<sub>NDM</sub>-positive strains were susceptible to amikacin (86.2% susceptible) (Table 1).

#### Prevalence of *bla*<sub>KPC</sub>, *bla*<sub>NDM</sub>, *bla*<sub>OXA-48</sub>, *bla*<sub>IMP</sub>, and *bla*<sub>VIM</sub> Carbapenemase and *mcr-1* Genes

Carbapenemase gene was positive in 97.4% (911/935) of the CRE strains, including  $bla_{\rm KPC-2}$  in 51.6% (482/935),  $bla_{\rm NDM}$  in 35.7% (334/935),  $bla_{\rm OXA-48-like}$  in 7.3% (68/935),  $bla_{\rm IMP}$  in

1.7% (16/935),  $bla_{\text{KPC}}$  and  $bla_{\text{NDM}}$  in 1.0% (9/935),  $bla_{\text{NDM-24}}$ and  $bla_{\text{OXA-48}}$  in 0.1% (1/935),  $bla_{\text{NDM-1}}$  and  $bla_{\text{IMP-4}}$  in 0.1% (1/935) of the strains (**Table 2**). KPC-2 was the most prevalent carbapenemase among *K. pneumoniae* (64.5%, 457/709) and *S. marcescens* (100%, 8/8) strains. NDM-5 was the predominant type carbapenemase among *E. coli* (74.5%, 111/149), *E. cloacae* (66.7%, 24/36) and *C. freundii* (64.3%, 9/14). Among all OXA-48like producing *K. pneumoniae*, PCR and DNA sequencing results showed the presence of  $bla_{\text{OXA-232}}$  (97.1%, 66/68) and  $bla_{\text{OXA-48}}$ (2.9%, 2/68) (**Table 2**).

Of the CRE strains isolated from adult patients, 70.3% (307/437) were KPC-2-producers; 20.6% (90/437) were NDM-producers (including 12.1% of NDM-1-producers, 8.2% of NDM-5-producers, and 0.2% of NDM-3-producer); and 0.5% (2/437) were OXA-48-producers (**Table 3, Figure 2**) (P < 0.01). However, of the CRE strains isolated from children, 49.0% (244/498) were NDM-producers (including 32.9% of NDM-5-producers, 15.9% of NDM-1-producers and 0.2% of NDM-3-producer); 35.1% (175/498) were KPC-2-producers and 13.3% (66/498) were OXA-232-producers (**Table 3, Figure 2**) (P < 0.01). The  $bla_{OXA-232}$ -positive *K. pneumoniae* were only isolated from children patients while  $bla_{OXA-48}$ -positive *K. pneumoniae* were isolated from adults. One polymyxin B resistant *E. coli* was positive for *mcr-1* with co-producing  $bla_{NDM-5}$ .

## DISCUSSION

Previous studies have proved that the presence of carbapenemase genes, including  $bla_{\text{KPC-2}}$  and  $bla_{\text{NDM}}$ , was the major mechanism of carbapenem resistance among CRE strains in China, which

#### TABLE 1 | Antimicrobial susceptibility testing results of clinical CRE strains (MICs, mg/L).

Antimicrobial agent	All CRE ( <i>n</i> = 935)				KPC-producers ( $n = 482$ )			NDM-producers ( $n = 334$ )			OXA-48-like producers ( $n = 68$ )						
	MIC range	MIC <sub>50</sub>	MIC <sub>90</sub>	%R	%S	MIC <sub>50</sub>	MIC <sub>90</sub>	%R	%S	MIC <sub>50</sub>	MIC <sub>90</sub>	%R	%S	MIC <sub>50</sub>	MIC <sub>90</sub>	% <b>R</b>	%S
Piperacillin	4->256	>256	>256	98.9	0.9	>256	>256	99.4	0.2	>256	>256	99.7	0.3	>256	>256	100	0
Cefoperazone-sulbactam	1->128	>128	>128	98.3	1.2	>128	>128	98.1	1.2	>128	>128	99.4	0	>128	>128	100	0
Piperacillin-tazobactam	2->256	>256	>256	97.2	1.5	>256	>256	98.8	0.6	>256	>256	99.4	0	>256	>256	100	0
Cefazolin	32->32	>32	>32	100	0	>32	>32	100	0	>32	>32	100	0	>32	>32	100	0
Cefuroxime	2->64	>64	>64	99.9	0.1	>64	>64	100	0	>64	>64	100	0	>64	>64	100	0
Ceftazidime	0.5->32	>32	>32	98.6	0.7	>32	>32	98.1	0.8	>32	>32	99.7	0	>32	>32	100	0
Ceftriaxone	0.12-64	>32	>32	99.4	0.6	>32	>32	99.4	0.6	>32	>32	99.7	0.3	>32	>32	100	0
Ceftazidime-avibactam	0.25->32	2	>32	38.6	61.4	2	4	0	100	>32	>32	100	0	0.5	4	0	100
Cefepime	0.25->32	>32	>32	98.1	0.9	>32	>32	97.9	1	>32	>32	99.4	0	>32	>32	100	0
Cefmetazole	1->64	>64	>64	92.7	4.5	>64	>64	92.3	5.6	>64	>64	97.6	1.2	64	>64	73.5	13.2
Aztreonam	0.25->128	>128	>128	93.2	4.2	>128	>128	99	0.8	>128	>128	85.3	7.8	>128	>128	100	0
Ertapenem	0.25->32	>32	>32	98.9	1	>32	>32	99	1	>32	>32	99.7	0.3	>32	>32	100	0
Imipenem	0.12->16	>16	>16	96.1	2.1	>16	>16	99.2	0.6	16	>16	99.4	0.3	>16	>16	73.5	17.6
Meropenem	0.12->16	>16	>16	97	1.9	>16	>16	98.1	1.5	>16	>16	99.7	0.3	>16	>16	85.3	4.4
Amikacin	1->128	16	>128	49.6	50.1	>128	>128	69.7	29.9	1	>128	13.8	86.2	>128	>128	100	0
Gentamicin	1->128	128	>128	67.9	31.8	>128	>128	83.8	16	1	128	40.4	59.3	>128	>128	100	0
Ciprofloxacin	0.06->8	>8	>8	78.4	19	>8	>8	95.6	3.7	8	>8	53.6	41.3	>8	>8	100	0
Levofloxacin	0.06->16	>16	>16	76.3	22.9	>16	>16	94.6	4.8	4	>16	49.4	49.7	>16	>16	100	0
Trimethoprim- Sulfamethoxazole	0.25->32	32	>32	54.8	45.2	1	>32	47.9	52.1	>32	>32	54.5	45.5	>32	>32	100	0
Polymyxin B	0.125->16	0.25	1	4	95.8	0.25	1	4.4	95.4	0.25	1	3.6	96.1	0.5	0.5	1.5	98.5
Nitrofurantoin	4->128	>128	>128	64.1	18.8	>128	>128	92.9	4.6	64	>128	22.8	41.9	128	>128	64.7	8.8
Tigecycline	0.12-8	0.5	2	0.3	98.4	0.5	2	0.4	97.7	0.5	1	0.3	99.1	1	2	0	100

CRE, carbapenem-resistant Enterobacteriaceae; MIC<sub>50/90</sub>, 50%/90% minimum inhibitory concentration; %R, % of isolates resistant; %S, % of isolates susceptible.

Species	Strains tested, N	bla <sub>КРС-2</sub> , n (%)	<i>bla</i> <sub>NDM</sub> , n (%)	bla <sub>OXA-48-like</sub> , n (%)	bla <sub>IMP</sub> , n (%)	Two genes, <i>n</i> (%)	Any gene, n (%
K. pneumoniae 70	709	457 (64.5)	<i>bla</i> <sub>NDM-1</sub> , 64 (9.0)	<i>bla</i> <sub>OXA-48</sub> , 2 (0.3)	<i>bla</i> IMP-4, 6 (0.8)	<i>bla</i> <sub>KPC-2</sub> + <i>bla</i> <sub>NDM-1</sub> , 6 (0.8)	693 (97.7)
			<i>bla<sub>NDM-5</sub></i> , 85 (12.0)	<i>bla</i> <sub>OXA-232</sub> , 66 (9.3)	<i>bla</i> <sub>IMP-69</sub> , 3 (0.4)	<i>bla</i> <sub>KPC-2</sub> + <i>bla</i> <sub>NDM-5</sub> , 1 (0.1)	
			<i>bla</i> <sub>NDM-3</sub> , 1 (0.1)			<i>bla</i> <sub>NDM-1</sub> + <i>bla</i> <sub>IMP-4</sub> , 1 (0.1)	
						<i>bla</i> <sub>NDM-24</sub> + <i>bla</i> <sub>OXA-48</sub> , 1 (0.1)	
E. coli 149	149	4 (2.7)	<i>bla<sub>NDM-1</sub></i> , 31 (20.8)				147 (98.7)
			<i>bla<sub>NDM-5</sub></i> , 111 (74.5)				
			<i>bla</i> <sub>NDM-3</sub> , 1 (0.7)				
E. cloacae 36	36	3 (8.3)	<i>bla</i> <sub>NDM-1</sub> , 24 (66.7)		<i>bla</i> <sub>IMP-4</sub> , 4 (11.1)	<i>bla</i> <sub>KPC-2</sub> + <i>bla</i> <sub>NDM-1</sub> , 1 (2.8)	36 (100)
			<i>bla</i> <sub>NDM-5</sub> , 3 (8.3)		<i>bla</i> IMP-6, 1 (2.8)		
C. freundii	14	3 (21.4)	<i>bla</i> <sub>NDM-1</sub> , 9 (64.3)				12 (85.7)
S. marcescens	8	8 (100)					8 (100)
E. aerogenes	7	1 (14.3)	<i>bla</i> <sub>NDM-1</sub> , 1 (14.3)				3 (42.9)
			<i>bla<sub>NDM-5</sub></i> , 1 (14.3)				
K. oxytoca	7	3 (42.9)	<i>bla</i> <sub>NDM-1</sub> , 2 (28.6)		<i>bla</i> <sub>IMP-4</sub> , 1 (14.3)	<i>bla</i> <sub>KPC-2</sub> + <i>bla</i> <sub>NDM-1</sub> , 1 (14.3)	7 (100)
M. morganii	3	2 (66.7)	<i>bla</i> <sub>NDM-1</sub> , 1 (33.3)				3 (100)
P. vulgaris	1	1 (100)					1 (100)
P. rettgeri	1				<i>bla</i> <sub>IMP-4</sub> , 1 (100)		1 (100)
Total	935	482 (51.6)	334 (35.7)	68 (7.3)	16 (1.7)	11 (1.2)	911 (97.4)

TABLE 2 | Prevalence of different carbapenemase genes among 935 CRE strains.

CRE, carbapenem-resistant Enterobacteriaceae.

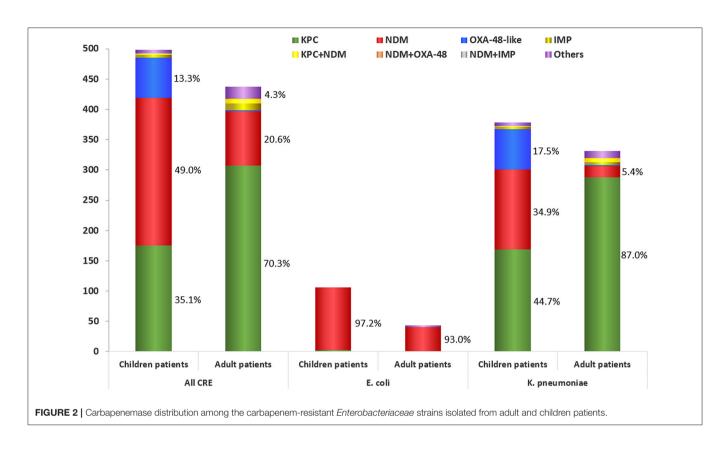
TABLE 3 | Distribution of different carbapenemase genes in 935 CRE strains isolated from adults and children patients.

Carbapenemase genes	All CRE	, n (%)	E. coli,	n (%)	K. pneumoniae, n (%)		
	From children	From adults	From children	From adults	From children	From adults	
bla <sub>KPC-2</sub>	175 (35.1)	307 (70.3)	3 (2.8)	1 (2.3)	169 (44.7)	288 (87.0)	
bla <sub>NDM-1</sub>	79 (15.9)	53 (12.1)	21 (19.8)	10 (23.3)	50 (13.2)	14 (4.2)	
bla <sub>NDM-5</sub>	164 (32.9)	36 (8.2)	82 (77.4)	29 (67.4)	81 (21.4)	4 (1.2)	
bla <sub>NDM-3</sub>	1 (0.2)	1 (0.2)		1 (2.3)	1 (0.3)		
bla <sub>OXA-48</sub>		2 (0.5)				2 (0.6)	
bla <sub>OXA-232</sub>	66 (13.3)				66 (17.5)		
bla <sub>IMP-4</sub>	3 (0.6)	9 (2.1)			2 (0.5)	4 (1.2)	
bla <sub>IMP-6</sub>		1 (0.2)					
bla <sub>IMP-69</sub>	2 (0.4)	1 (0.2)			2 (0.5)	1 (0.3)	
bla <sub>KPC-2</sub> +bla <sub>NDM-1</sub>	2 (0.4)	6 (1.4)			1 (0.3)	5 (1.5)	
bla <sub>KPC-2</sub> +bla <sub>NDM-5</sub>		1 (0.2)				1 (0.3)	
bla <sub>NDM-1</sub> +bla <sub>IMP-4</sub>	1 (0.2)				1 (0.3)		
bla <sub>NDM-24</sub> + bla <sub>OXA-48</sub>		1 (0.2)				1 (0.3)	
Others	5 (1.0)	19 (4.3)		2 (4.7)	5 (1.3)	11 (3.3)	
Total	498	437	106	43	378	331	

CRE, carbapenem-resistant Enterobacteriaceae.

were the most prevalent in *K. pneumoniae* and *E. coli*, respectively (Zhang et al., 2017; Wang et al., 2018). However, the researches on CRE strains isolated from children patients are limited in China. This study provided a comprehensive and updated carbapenemase profile of 935 CRE strains isolated from both adult and children patients. We found that  $bla_{\rm KPC-2}$  (51.6%) and  $bla_{\rm NDM}$  (35.7%) were the most common carbapenemase genes among CRE strains, while the emergence of  $bla_{\rm OXA-232}$ ,  $bla_{\rm IMP}$ , and other multi-carbapenemase genes have been increasing in recent years. KPC-2 was the most frequently detected

carbapenemase gene in *K. pneumoniae*, while NDM was the most prevalent one in *E. coli*. This pattern in China is significantly different from that in Europe. In Europe, the prevalence of OXA-48-like producing *Enterobacteriaceae* was 38% (333/927), next to KPC- (42%, 393/927), but higher than NDM-producing *Enterobacteriaceae* (12%, 113/927) (Grundmann et al., 2017). The distribution of carbapenemase-producers also varied with bacterial species. In *K. pneumoniae*, KPC-producers were the most prevalent, followed by OXA-48-like (37%, 310/850) and NDM-producers (11%, 93/850). In *E. coli*, OXA-48-like



producers were the most prevalent (56%, 43/77), followed by NDM- (26%, 20/77) and KPC-producers (18%, 14/77). *K. pneumoniae* and *E. coli* were the two main species in China with a ratio of 5:1 (4:1 in children, 8:1 in adults) in this study, which differed from the prevalence trends (ratio of 11:1) in EuSCAPE (Grundmann et al., 2017).

Notably, KPC-2-producers were widespread in adult patients, followed by NDM-producers, while NDM-producers were prevalent in children patients, followed by KPC-2- and OXA-48-like producers. These findings described the different patterns of carbapenemases among CRE strains from adults and children. In contrast to the previous finding that NDM-1 was the most common carbapenemase among children patients, we have found that NDM-5-producers (32.9%) were most frequently detected CRE strains from children (Tian et al., 2018; Yin et al., 2018; Zhang et al., 2018). The outbreak of NDM-5-producing ST48 *K. pneumoniae* was first reported in Shanghai (Tian et al., 2018). We speculated that outbreak of NDM-5-producers accounted for the spread of NDM-5 among children patients in this study (Tian et al., 2018; Li et al., 2020). Further study is needed to track the type of plasmids harboring these carbapenemase genes.

Unlike the previous report that few OXA-48-like producing *Enterobacteriaceae* (0.1%, 2/1801) were detected in China from 2012 to 2016 (Wang et al., 2018), we found 7.3% (68/935) OXA-48-like producing *K. pneumoniae* between 2016 and 2018. Since the first OXA-232-producing *K. pneumoniae* isolated from neonate in 2017, the outbreaks of OXA-232-producing *Enterobacteriaceae* have been successively reported in children

patients (Yin et al., 2017; Tian et al., 2018). Subsequently, 10 strains of OXA-232-producing *K. pneumoniae* were isolated from elderly patients in the intensive care unit in 2019 and the  $bla_{OXA-232}$  was located in a 6.1-kb ColKP3-type non-conjugative plasmid, which was highly similar to the pkNICU5 first reported (similarity about 99%) in 2017 (Yin et al., 2017; Shu et al., 2019). We speculated that the presence of  $bla_{OXA-232}$  on a mobile element and its spread among different strains were responsible for the recent dissemination of OXA-232-producing *Enterobacteriaceae*, which would make it possible to become the "third epidemic" carbapenemase after KPC-2 and NDM in China (Yin et al., 2017; Tian et al., 2018).

All of the CRE strains were highly resistant to cephalosporins, carbapenems, aminoglycosides, and fluoroquinolones but susceptible to polymyxin B and tigecycline. Ceftazidimeavibactam, launched last year in China, showed excellent in vitro antibacterial activity against both KPC-2- and OXA-48like producers, but not active against metallo-β-lactamases producers. Most (86.2%) of NDM-producers were susceptible to amikacin. In addition, we found a blaNDM-5 and mcr-1 co-harboring E. coli resistant to polymyxin B. These findings limited the utility of ceftazidime-avibactam and polymyxin B and prompted the development of novel or combinational therapies to combat CRE strains. For example, aztreonam plus meropenem-vaborbactam and aztreonam plus ceftazidimeavibactam showed good antibacterial activity against NDMand non-OXA-48-like producing Enterobacteriaceae (Biagi et al., 2019). The combination of colistin and amikacin showed

consistently bactericidal against NDM-5-bearing *mcr-1*-positive *E. coli*, which might be an alternative therapeutic option for the treatment of lethal infections (Zhou et al., 2017).

## CONCLUSIONS

In conclusion, KPC-2, NDM, and OXA-48-like enzymes were the most prevalent carbapenemases among CRE clinical isolates in China. The most prevalent carbapenemase gene was  $bla_{\rm KPC-2}$  among *K. pneumoniae* isolated from adult patients, and  $bla_{\rm NDM}$  among *E. coli* isolates from both children and adult patients. The  $bla_{\rm OXA-232}$  was only detected among *K. pneumoniae* isolates from children.

## DATA AVAILABILITY STATEMENT

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation, to any qualified researcher.

## **ETHICS STATEMENT**

The study protocol was approved by the Institutional Review Board of Huashan Hospital, Fudan University (Number: 2018-408).

## **AUTHOR CONTRIBUTIONS**

FH and RZ designed the study. RH, QS, SW, and MP performed the experimental work. RH and DY collected the data. FH analyzed the data. All authors read and approved the final manuscript. All authors contributed to the article and approved the submitted version.

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## REFERENCES

- Albiger, B., Glasner, C., Struelens, M. J., Grundmann, H., and Monnet, D. L. (2015). Carbapenemase-producing *Enterobacteriaceae* in Europe: assessment by national experts from 38 countries, May 2015. *Euro. Surveill.* 20. doi: 10.2807/1560-7917.ES.2015.20.45.30062
- Biagi, M., Wu, T., Lee, M., Patel, S., Butler, D., and Wenzler, E. (2019). Searching for the optimal treatment for metallo- and serine-beta-lactamase producing *Enterobacteriaceae*: Aztreonam in combination with ceftazidime-avibactam

or meropenem-vaborbactam. *Antimicrob. Agents Chemother*. 63:e01426–19. doi: 10.1128/AAC.01426-19

- Falagas, M. E., Tansarli, G. S., Karageorgopoulos, D. E., and Vardakas, K. Z. (2014). Deaths attributable to carbapenem-resistant *Enterobacteriaceae* infections. *Emerg. Infect. Dis.* 20, 1170–1175. doi: 10.3201/eid2007. 121004
- Feil, E. J. (2016). Enterobacteriaceae: joining the dots with pan-European epidemiology. Lancet Infect Dis. 17, 118–119. doi: 10.1016/S1473-3099(16)30333-4

- Goodman, K. E., Simner, P. J., Tamma, P. D., and Milstone, A. M. (2016). Infection control implications of heterogeneous resistance mechanisms in carbapenemresistant *Enterobacteriaceae* (CRE). *Expert. Rev. Anti. Infect. Ther.* 14, 95–108. doi: 10.1586/14787210.2016.1106940
- Grundmann, H., Glasner, C., Albiger, B., Aanensen, D. M., Tomlinson, C. T., Andrasevic, A. T., et al. (2017). Occurrence of carbapenemaseproducing *Klebsiella pneumoniae* and *Escherichia coli* in the European survey of carbapenemase-producing *Enterobacteriaceae* (EuSCAPE): a prospective, multinational study. *Lancet Infect. Dis.* 17, 153–163. doi: 10.1016/S1473-3099(16)30257-2
- Iovleva, A., and Doi, Y. (2017). Carbapenem-resistant *Enterobacteriaceae*. Clin. Lab. Med. 37, 303–315. doi: 10.1016/j.cll.2017.01.005
- Li, J., Yu, T., Tao, X. Y., Hu, Y. M., Wang, H. C., Liu, J. L., et al. (2020). Emergence of an NDM-5-producing *Escherichia coli* sequence type 410 clone in infants in a children's hospital in China. *Infect. Drug Resist.* 13, 703–710. doi: 10.2147/IDR.S244874
- Liu, Y. Y., Wang, Y., Walsh, T. R., Yi, L. X., Zhang, R., Spencer, J., et al. (2016). Emergence of plasmid-mediated colistin resistance mechanism MCR-1 in animals and human beings in China: a microbiological and molecular biological study. *Lancet Infect. Dis.* 16, 161–168. doi: 10.1016/S1473-3099(15)00424-7
- Nordmann, P., Dortet, L., and Poirel, L. (2012). Carbapenem resistance in *Enterobacteriaceae*: here is the storm!. *Trends Mol. Med.* 18, 263–272. doi: 10.1016/j.molmed.2012.03.003
- Perez, F., and Villegas, M. V. (2015). The role of surveillance systems in confronting the global crisis of antibiotic-resistant bacteria. *Curr. Opin. Infect. Dis.* 28, 375–383. doi: 10.1097/QCO.0000000000 00182
- Poirel, L., Walsh, T. R., Cuvillier, V., and Nordmann, P. (2011). Multiplex PCR for detection of acquired carbapenemase genes. *Diagn. Micr. Infec. Dis.* 70, 119–123. doi: 10.1016/j.diagmicrobio.2010.12.002
- Shu, L., Dong, N., Lu, J., Zheng, Z., Hu, J., Zeng, W., et al. (2019). Emergence of OXA-232 carbapenemase-producing *Klebsiella pneumoniae* that carries a pLVPK-like virulence plasmid among elderly patients in China. *Antimicrob. Agents Chemother.* 63:e02246-18. doi: 10.1128/AAC. 02246-18
- Tian, D., Pan, F., Wang, C., Sun, Y., and Zhang, H. (2018). Resistance phenotype and clinical molecular epidemiology of carbapenem-resistant *Klebsiella pneumoniae* among pediatric patients in Shanghai. *Infect Drug Resist.* 11, 1935–1943. doi: 10.2147/IDR.S175584
- van Duin, D., and Doi, Y. (2016). The global epidemiology of carbapenemase-producing *Enterobacteriaceae*. *Virulence* 8, 460–469. doi: 10.1080/21505594.2016.1222343
- Villegas, M. V., Pallares, C. J., Escandón-Vargas, K., Hernández-Gómez, C., Correa, A., Álvarez, C., et al. (2016). Characterization and clinical impact of bloodstream infection caused by carbapenemase-producing

*Enterobacteriaceae* in seven latin American Countries. *PLoS ONE* 11:e0154092. doi: 10.1371/journal.pone.0154092

- Wang, Q., Wang, X., Wang, H., Ouyang, P., Jin, C., Wang, R., et al. (2018). Phenotypic and genotypic characterization of carbapenem-resistant *Enterobacteriaceae* data from a longitudinal large-scale CRE study in China (2012–2016). *Clin. Infect. Dis.* 67, S196–S205. doi: 10.1093/cid/ciy660
- Yin, D., Dong, D., Li, K., Zhang, L., Liang, J., Yang, Y., et al. (2017). Clonal dissemination of OXA-232 carbapenemase-producing *Klebsiella pneumoniae* in neonates. *Antimicrob. Agents Chemother.* 61:e00385–17. doi: 10.1128/AAC.00385-17
- Yin, D., Zhang, L., Wang, A., He, L., Cao, Y., Hu, F., et al. (2018). Clinical and molecular epidemiologic characteristics of carbapenem-resistant *Klebsiella pneumoniae* infection/colonization among neonates in China. J. Hosp. Infect. 100, 21–28. doi: 10.1016/j.jhin.2018.05.005
- Yong, D., Toleman, M. A., Giske, C. G., Cho, H. S., Sundman, K., Lee, K., et al. (2009). Characterization of a new metallo-beta-lactamase gene, bla(NDM-1), and a novel erythromycin esterase gene carried on a unique genetic structure in *Klebsiella pneumoniae* sequence type 14 from India. *Antimicrob. Agents Chemother.* 53, 5046–5054. doi: 10.1128/AAC.00774-09
- Zhang, R., Liu, L., Zhou, H., Chan, E. W., Li, J., Fang, Y., et al. (2017). Nationwide surveillance of clinical carbapenem-resistant *Enterobacteriaceae* (CRE) strains in China. *EBioMed.* 19, 98–106. doi: 10.1016/j.ebiom.2017.04.032
- Zhang, X., Chen, D., Xu, G., Huang, W., and Wang, X. (2018). Molecular epidemiology and drug resistant mechanism in carbapenem-resistant *Klebsiella pneumoniae* isolated from pediatric patients in Shanghai, China. *PLoS ONE* 13:e0194000. doi: 10.1371/journal.pone.0194000
- Zhou, Y., Tao, M., Feng, Y., Yang, R., Liao, X., Liu, Y., et al. (2017). Increased activity of colistin in combination with amikacin against *Escherichia coli* co-producing NDM-5 and MCR-1. *J. Antimicrob. Chemoth.* 72, 1723–1730. doi: 10.1093/jac/dkx038

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

The reviewer YY declared a shared affiliation with one of the authors RZ to the handling editor at time of review.

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