



NK Cell Function Regulation by TGF-β-Induced Epigenetic Mechanisms

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TGF- β is a potent immunosuppressive cytokine that severely affects the function of NK cells. Tumor cells can take advantage of this ability, enriching their surrounding microenvironment with TGF- β . TGF- β can alter the expression of effector molecules and of activating and chemokine receptors, influence metabolism, induce the NK cell conversion toward the less cytolytic ILC1s. These and other changes possibly occur by the induction of complex gene expression programs, involving epigenetic mechanisms. While most of these programs are at present unexplored, the role of certain transcription factors, microRNAs and chromatin changes determined by TGF- β in NK cells start to be elucidated in human and/or mouse NK cells. The deep understanding of these mechanisms will be useful to design therapies contributing to restore the full NK function.

OPEN ACCESS

Edited by:

Laurent Brossay, Brown University, United States

Reviewed by:

Frank Momburg, German Cancer Research Center (DKFZ), Germany Marina Cella, Washington University School of Medicine in St. Louis, United States

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Specialty section:

This article was submitted to NK and Innate Lymphoid Cell Biology, a section of the journal Frontiers in Immunology

> Received: 13 September 2019 Accepted: 07 February 2020 Published: 25 February 2020

Citation:

Regis S, Dondero A, Caliendo F, Bottino C and Castriconi R (2020) NK Cell Function Regulation by TGF-β-Induced Epigenetic Mechanisms. Front. Immunol. 11:311. doi: 10.3389/fimmu.2020.00311 Keywords: NK cells, TGF- β , epigenetic, transcription factors, microRNAs

INTRODUCTION

NK cells are innate lymphocytes able to recognize and kill virus-infected and tumor cells. They are equipped with a set of inhibitory and activating receptors, whose integrated signaling selectively directs their activity (1). While inhibitory receptors bind molecules such as HLA class I or immune checkpoint ligands, activating receptors interact with stress molecules induced or upregulated by tumor transformation or viral infection. Several cytokines, like IL-2, IL-12, IL-15, and IL-18, influence NK cell activity. Stimulated NK cells can, in turn, produce other cytokines, including IFN-γ and TNF-α (1). Traditionally, two populations of NK cells are described, CD56^{bright}, prevailing in secondary lymphoid organs, producing large amounts of cytokines, and CD56^{dim}, abundant in peripheral blood, characterized by high cytotoxic potential. However, recent views consider the spectrum of NK cell diversity quite wider (2–4). Moreover, NK cells represent the cytolytic members of a large, innate lymphoid cell family (ILC) that includes ILC1, poor cytolytic cells sharing with NK cells the ability to produce IFN-γ, ILC2, secreting IL-5 and IL-13, and ILC3, producing IL-7 and IL-22 (5, 6).

The activity of NK cells can be strongly affected by TGF- β , an immunomodulatory cytokine with a prominent role in both innate and adaptive immune responses (7, 8). In particular, TGF- β is a negative regulator of IFN- γ production (9) and decreases the surface level of the activating receptors NKG2D and NKp30, reducing the cytotoxic ability of NK cells (10) and impairing their antitumor function. Accordingly, mice lacking the expression of the TGF- β receptor 2 (TGFBR2) in their NK cells show improved suppression of metastases compared to control mice (11). TGF- β ,

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also alters the surface expression of the chemokine receptors CXCR3, CXCR4, CX3CR1, with a possible impact on NK cell migration and recruitment (12). Surprisingly, some of the modulatory effects of TGF- β are potentiated by IL-18, via p38/MAPK pathway (13). The negative regulatory role of TGF- β also extends to different metabolic pathways including glycolysis and mitochondrial processes (14, 15).

TGF- β acts on target cells via specific receptors composed of TGFBR1 and TGFBR2 subunits, which can transduce the signal phosphorylating SMAD2 and SMAD3 proteins. These molecules bind to SMAD4 forming heterotrimeric transcriptional complexes that accumulate in the nucleus, where they activate or repress the transcription of sets of target genes. Other SMAD-independent non-canonical pathways proceed via PI3K and MAPK (16).

TGF- β signaling has been associated with epigenetic alterations (16), i.e., reversible and heritable changes not associated with DNA sequence mutation. These include DNA methylation, post-translational modification of chromatin histones as well as changes in the expression of transcription factors and noncoding RNAs (17).

Several epigenetic modifications contributing to the deep influence of TGF- β on the NK cell function, including chromatin remodeling, induction or repression of transcription factors, and post-transcriptional regulation of gene expression by miRNAs, have been described, and are here reviewed.

TRANSCRIPTION FACTORS AND CHROMATIN REMODELING

TGF- β affects NK cell function inducing relevant transcription programs, which can involve events of chromatin remodeling, through the action of various transcription factors; the landscape of these changes is presumably wide and mostly unexplored. Studies have been performed on the SMAD3-dependent regulation of T-bet and IFN- γ expression, on NK cell metabolism and conversion of NK cells to ILC1s (**Figure 1**).

T-bet-IFN-γ Pathway

Pro-inflammatory cytokines positively regulate the production of IFN-y and the expression of the transcription factor T-bet, which, as the other T-box protein Eomes, has a relevant role in development and function of NK cells (18). Acting as an immunosuppressive cytokine, TGF-\u03b3 antagonizes the effects of pro-inflammatory cytokines, down-regulating T-bet and IFN-y via SMAD3 (9, 19). In line with this evidence, SMAD3-deficient mouse NK cells produced more IFN- γ (9), and TGFBR2-deficient mouse NK cells stimulated with IL-15 showed increased Tbet expression (11). Interestingly, chromatin remodeling has been associated with the T-bet-dependent regulation of IFN- γ gene expression in CD4T cells polarizing toward $T_{\rm H}1$ cells (20). Moreover, it has been shown that T-bet, besides its ability to interact with the IFN-y promoter and with several enhancers, can recruit histone-modifying complexes inducing epigenetic modifications at the IFN- γ locus (20, 21). In response to activating receptor engagement, CD56^{dim} human NK cells showed increased expression of T-bet and IFN- γ , compared to CD56^{bright} NK cells. High IFN- γ production was accompanied by a reduced CpG methylation pattern and histone modifications of the IFN- γ gene promoter (22). Moreover, epigenetic remodeling of the *IFNG* conserved non-coding sequence (CNS) 1, located upstream of the human IFN- γ promoter, has been documented in NKG2C^{hi} "adaptive" NK cells, which expand in CMV seropositive individuals (23).

Interestingly, "TGF- β -imprinted" NK cells, produced by prolonged IL-2 and TGF- β stimulation of activated NK cells, contrarily to acutely TGF- β -treated NK cells, exhibited a proinflammatory phenotype, with abundant production of IFN- γ and TNF- α persisting after removal of TGF- β (24). The TGF- β pathway was altered in these cells, as indicated by the downregulation of SMAD3, whose locus showed reduced chromatin accessibility, and of T-bet.

Of note, TGF- β can inhibit IFN- γ expression both by a T-bet dependent and a T-bet independent manner (9). In the latter context, Tang et al. (25) reported that SMAD3 suppresses transcription of IFN- γ via E4BP4, which is an important transcription factor for the NK cell lineage commitment (26).

From a more general perspective, the intimate link between T-bet and chromatin modifications is underlined by a recent study exploring chromatin and gene expression changes associated with viral infection in mouse CD8 T cells. The authors reported that T-box and Runx genes have been shown to act as critical mediators of chromatin remodeling during CD8 T cell activation (27).

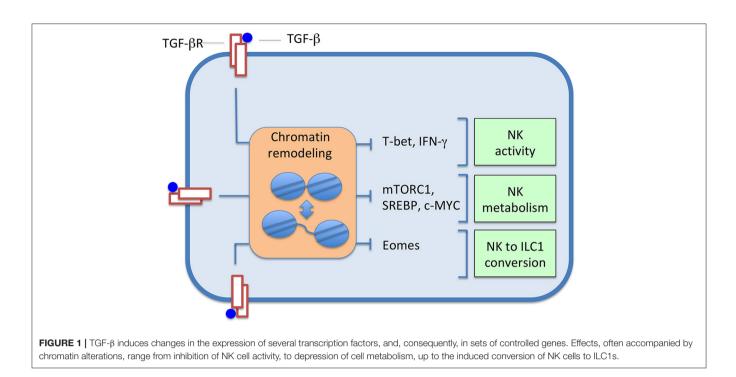
NK Cell Metabolism: mTORC1, SREBP, and c-MYC

TGF- β can sensibly reduce NK cell metabolism affecting glycolysis and oxidative phosphorylation, which are considerably increased in cytokine-stimulated NK cells and support effector functions (11). The serine/threonine kinase mTOR is part of the mTORC1 complex, a master regulator of various metabolic processes in different cell types. mTORC1 activity is important for activation-induced metabolic and functional responses in NK cells (14). TGF- β was found to inhibit mTOR-dependent activities in NK stimulated by IL-15 (11). Moreover, mouse TGFBR2-defective NK cells stimulated with IL-15 showed increased mTOR-dependent activities. mTORC1 also contributed to the regulation of the SREBP (28) and c-MYC (29) transcription factors, which are key controllers of glycolysis and oxidative phosphorylation in cytokine-stimulated NK cells (14).

In humans, it has been reported that TGF- β , acting by the canonical pathway, significantly decreases the rate of IL-2-induced mitochondrial metabolism, and inhibits the glycolysis in IL-2 stimulated human NK cells independently by mTORC1 (14, 30).

NK-ILC1 Cell Conversion

The potency of TGF- β action is further highlighted by its ability to induce the conversion of NK cells to ILC1s (31, 32). Although NK cells and ILC1s are both IFN- γ producers and functionally dependent on T-bet (5), they are developmentally distinct. ILC1s are not found in blood and lymphoid organs, having instead



tissue-resident features (33) and being weakly cytolytic (5). Moreover, while mature NK cells express Eomes, ILC1s do not.

Gao et al. (31) reported that, in the tumor microenvironment, TGF- β can induce the conversion of mouse NK cells to an NK-ILC1 intermediate cell type (intILC1s) and, finally, to ILC1s; the switch is accompanied by the down-regulation of the transcription factor Eomes. Compared to NK cells, intILC1s and ILC1s are unable to control tumor growth and metastasis. Therefore, TGF- β , inducing the conversion, inhibits cancer immunosurveillance by a new mechanism of immune evasion. Cortez et al. (32) generated SMAD4-deficient NK cells, which showed features of ILC1s, had impaired effector functions and were unable to control tumor metastasis, thus indicating that SMAD4 is a negative regulator of the NK-ILC1s conversion. In particular, SMAD4 was shown to hamper the conversion of NK cells to ILC1s inhibiting a non-canonical TGFBR1mediated pathway.

As stated by the RNAseq-based transcriptome data obtained by NKs, intILC1s, and ILC1s cell populations (31), the NK to ILC1 conversion induced by TGF- β involves sets of genes, presumably governed by still undefined, complex, transcriptional and epigenetic programs. Given the reduction of Eomes expression during cell conversion and its potential role as chromatin remodeler, we can speculate that it may have a relevant role in this event of cell plasticity. In a different context, similar considerations can be done for T-bet, which is reduced by TGF- β in human liver resident NK cells, this reduction being fundamental for the maintenance of the Eomes^{hi} T-bet^{lo} phenotype of this subtype of NK cells (4).

Notably, together with IL-23, TGF- β guides the conversion ILC1 toward ILC3, which can occur in mucosal tissues (34). TGF- β increases the expression of the transcription factors T-bet and

Aiolos. Interestingly, Aiolos, like the other IKZF members, can suppress target gene expression altering chromatin accessibility via histones deacetylation (35). By this mechanism, it suppresses the ILC3 specific genes IL-22 and ROR γ t, promoting the ILC3-ILC1 conversion (34).

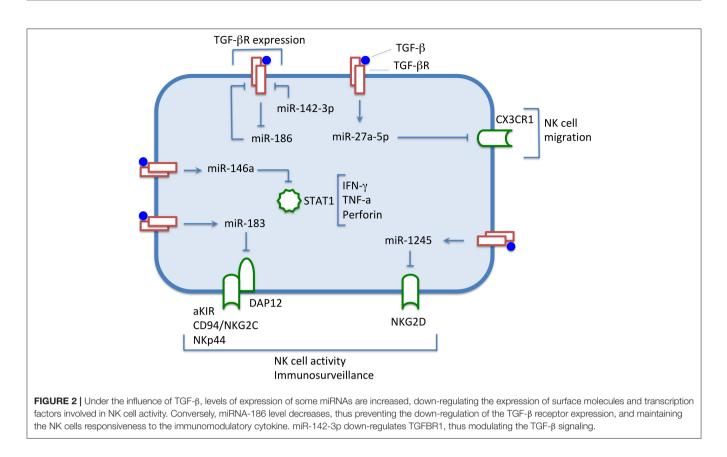
miRNAs

Besides chromatin remodeling and transcription factor manipulation, TGF- β can affect the NK cell function acting on the miRNA machinery, exploiting its capacity to influence gene expression at the post-transcriptional level. Distinct miRNA-containing pathways deregulating the expression of receptors or transcription factors have been described (**Figure 2**) (36).

miR-1245

NKG2D is a C-type lectin-like activating receptor expressed on NK and CD8T cells, which promotes the elimination of transformed and pathogen-infected cells. In humans, it recognizes stress-inducible ligands belonging to the MIC (MICA and MICB) and ULBP (ULBP1-6) families (37).

The surface expression of NKG2D is significantly downregulated in NK cells by TGF- β 1 or TGF- β 2 (10). Espinoza et al. (38) reported that TGF- β 1 causes an increase of miR-1245 in human NK cells. Overexpression of miR-1245 induced a down-regulation of NKG2D at the cell surface without affecting the other activating receptors NKp30, NKp44, and NKp46. Moreover, NK cells overexpressing miR-1245 had lower cytotoxicity against target cells expressing NKG2D ligands. MiR-1245 was shown to directly target and down-regulate the NKG2D mRNA. Interestingly, a polymorphism in the NKG2D 3'-UTR



region occurs in the region complementary to the miR-1245 seed region (39). NK cells carrying this polymorphism exhibit a reduced TGF- β -mediated modulation of NKG2D activity, which can be due to less efficient targeting of the miRNA. Importantly, the NKG2D variants seem to influence immunosurveillance capability and risk of cancer development (40).

miR-183

DAP12 (also known as KARAP or TYROBP) is a 12 kDa transmembrane protein containing an immune tyrosine-based activation motif (ITAM) in its cytoplasmic domain (41). DAP12 is a signaling molecule associating with different activating receptors, including the activating KIRs, CD94/NKG2C, and NKp44, the latter expressed by NK cells upon activation and by a subset of ILC3 (42).

Donatelli et al. (43) reported that TGF- β 1 down-regulates the DAP12 level in human NK cells. In particular, it up-regulated miR-183, which was shown to directly target DAP12 mRNA. The NK92 cell line overexpressing miR-183 had reduced DAP12, decreased NKp44 surface levels, and was less efficient in killing the Raji Burkitt lymphoma cell line.

Interestingly, NK cells infiltrating lung cancers were shown to have a diminished DAP12 expression (43). Recently, a mechanism of immune evasion has been described in human lung cancer cells (44). TGF- β appears to act simultaneously on NK cells and tumor cells via miR-183, with the result to inhibit the NK cells' anti-tumor activity by down-regulating DAP12, in NK cells, and NKG2D ligands (MICA and MICB), in cancer cells.

miR-27a-5p

 CX_3CR1 is a chemokine receptor expressed by different immune cell types, including NK cells, which binds CX_3CL1 (also known as fractalkine). CX_3CR1 drives, with other chemokine receptors, NK cells localization at a steady state in peripheral tissues, and promotes their migration under inflammatory conditions (45). Moreover, CX_3CR1 , CXCR4, and S1P5 regulate NK cells homing and egress from the bone marrow (46, 47).

It has been shown that TGF- β 1 released by neuroblastoma tumor cells down-regulated the surface expression of CX₃CR1 in human NK cells (12). Afterward, TGF- β 1 was found to upregulate in NK cells the miR-23a-27a-24-2 cluster, encoding, among others, miR-27a-5p, that has been demonstrated to target CX₃CR1 (48). In agreement with this finding (12), NK cells cultured in the presence of neuroblastoma cells exhibited a significant inverse correlation between miR-27a-5p and CX₃CR1 mRNA expression (48). Notably, an unusual CX₃CR1^{low/neg} phenotype has been described in circulating CD56^{dim} NK cells and in NKp46⁺ bone marrow resident NK cells from high-risk NB patients (12).

miR-186

Neviani et al. (49) reported that miR-186 down-regulates TGFBR1 and TGFBR2 preventing the TGF- β -dependent inhibition of human NK cells. MiR-186 was found to be down-regulated in TGF- β -treated NK cells as well as in neuroblastoma cells. Importantly, by receiving miR-186, carried by NK exosomes, neuroblastoma cells down-regulated the oncogenic

proteins MYCN and AURKA. Therefore, delivery of miR-186 to neuroblastoma and NK cells may hamper both tumorigenic potential and inhibition of NK cells.

miR-142

MiR-142 is a critical regulator of ILC biology. It is induced by IL-15 and mediates modulation of TGFBR1. miR-142^{-/-} knockout mice showed increased TGF- β signaling compared to normal controls, as indicated by the augmented expression of TGFBR1 and functionally related genes in NK/ILC1 cells from spleen and bone marrow (50). Interestingly, miR-142^{-/-} mice presented an altered number and functionality of NK cells, with increased susceptibility to cytomegalovirus infection. On the contrary, an increase of ILC1-like cells was observed, possibly related to the increased TGF- β signaling and NK-ILC1 conversion.

miR-146a

Xu et al. (51) reported that human miR-146a, which is upregulated by TGF- β in NK cells, targets STAT1, a transcription factor involved in interferons signal transduction, which contributes to NK cell cytotoxicity (52). miR-146 overexpression lowered the production of effector molecules as IFN- γ , TNF- α , and perforin. Moreover, the expression level of miR-146a negatively correlated with NK cell-mediated cytotoxicity (51).

CONCLUDING REMARKS

TGF- β can exert its immunosuppressive action influencing NK cells at different levels. Acting by transcription factors, TGF- β can hamper NK cell function inhibiting the T-bet-IFN- γ pathway and negatively influences cell metabolism, thus hampering cell activation. Using miRNAs, TGF- β down-regulates activating receptors, receptor-associated signaling molecules, and chemokine receptors, thus inhibiting NK cell cytotoxicity and recruitment in tissues. Acting by still undefined combined transcriptional and epigenetic programs, TGF- β converts NK cells to ILC1, which are less cytolytic against tumor cells. Moreover, we can speculate that long non-coding RNA (lncRNAs), which are known to be involved

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in TGF- β -driven fibrosis, epithelial-mesenchymal transition and cancer progression (53, 54), may represent additional mediators of still undefined TGF- β -dependent immune-modulatory functions on NK cells.

Studying how TGF-B alters the NK/ILC biology through epigenetic mechanisms could be an important area of investigation to understand its functions in the tumor microenvironment and in other diseases containing TGF-B rich milieus, such as fibrosis. In this context, it will be also important the development of molecular approaches aimed to control the TGF-B signaling. Several strategies have been recently elaborated for the therapeutic inhibition of the TGF- β pathway, particularly to improve the antitumor immune responses. Combination therapy with an inhibitor of TGF^β type I receptor (TGFβR1) kinase, galunisertib, anti-GD2 antibody and ex vivo activated NK cells, has been used by Tran et al. (55) in a mouse model of neuroblastoma with positive results. Genetically modified SMAD3-silenced human NK-92 cells inhibited cancer progression in two xenograft mouse models with human hepatoma and melanoma (56). NK cells genetically modified to express a truncated TGFBR2 receptor fused to the DAP12 activation domain exhibited higher cytotoxic activity against neuroblastoma (57). A deeper knowledge of the mechanisms by which TGF-B acts on NK cells will be useful to improve therapeutic strategies aimed to efficiently restore NK cell activity or to increase the NK cell pool for an effective antitumor response.

AUTHOR CONTRIBUTIONS

All authors contributed to the writing of the manuscript.

FUNDING

This work from our laboratories has been supported by Associazione Italiana per la Ricerca sul Cancro (A.I.R.C.), Ministero dell'Istruzione, dell'Università e della Ricerca (M.I.U.R), and 100008-2018-SS-DIP_001 project (Department of Experimental Medicine, School of Medical and Pharmaceutical Science, University of Genova).

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Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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