



Complete Genomic Sequence of an Australian Sindbis Virus Isolated 44 Years Ago Reveals Unique Indels in the E2 and nsP3 Proteins

Paul Pickering,^a John G. Aaskov,^{a,b} Wenjun Liu^a

^aAustralian Defence Force Malaria and Infectious Disease Institute, Enoggera, Australia ^bInstitute of Health and Biomedical Innovation, Queensland University of Technology, Brisbane, Australia

ABSTRACT The complete genome sequence of a Sindbis virus (SINV) strain (SINV_AUS_1975_18953) isolated in Australia in 1975 from *Culex annulirostris* mosquitoes revealed unique deletions in amino acid positions 182 to 184 and 201 to 228 of the E2 envelope protein and multiple indels in the nonstructural protein 3 (nsP3).

S*Alphavirus* (SINV) is a single-stranded positive-sense RNA virus of the genus *Alphavirus* in the *Togaviridae* family (1). The viral genome of the prototype strain AR339 contains 11,703 nucleotides (nt) with a 5' cap and a 3' poly(A) tail. The virus is maintained in nature in transmission cycles involving principally birds and *Culex* mosquitos, although it has been isolated from a variety of mosquito and vertebrate hosts (2). Humans can be infected by spillover of virus, resulting in disease with relatively benign symptoms, such as fever, rash, arthralgia, and myalgia (3–5).

SINV strain SINV_AUS_1975_18953 was isolated in suckling mice from a pool of Culex annulirostris mosquitoes collected in a forested area near Charleville (26.4°N, 146.2°E), Queensland, Australia, in 1975 (2). The mouse brain stock was used to infect cultures of C6/36 Aedes albopictus cells for 3 days at 30°C. The culture supernatant was recovered, and cell debris was removed by centrifugation at 5,000 \times q for 10 minutes. The resulting supernatant was stored at - 80°C and employed as a source of virus. Viral RNA was extracted from 140 μ l of this tissue culture supernatant using the QIAamp viral RNA minikit (Qiagen, USA) and reverse transcribed using random hexanucleotide primers (Promega, USA) and SuperScript III reverse transcriptase (Invitrogen, USA), according to the manufacturer's instructions. Overlapping fragments of 1.0 to 1.5 kb of the resultant cDNA were amplified by using Expand long template DNA polymerase (Roche) and virus-specific primers corresponding to the sequence of SINV strain MRE-16 (GenBank accession numbers AF492770 and U90536). The reverse transcription-PCR (RT-PCR) amplicons were sequenced by using dideoxy dye chain-termination Sanger sequencing technology at the Australian Genomic Research Facility. The set of Sanger sequences were read from ABI files and trimmed to remove the poor-quality reads of 5' and 3' ends by the quality value, and the trimmed reads were aligned to a reference SINV sequence (strain AR339, GenBank accession number J02363) to generate the whole-genome sequence with 100% coverage using the Geneious software package version 11.2.2. The nucleotide sequences in the 5' untranslated region (UTR) and the 3' UTR of SINV_AUS_1975_18953 were determined by Sanger sequencing of the reverse transcription and PCR amplicons using the rapid amplification of cDNA ends (RACE) strategy described previously (6).

The genome of SINV_AUS_1975_18953 contained 11,600 nt, with a G+C content of 51.8%. At the nucleotide level, the whole-genome sequence of SINV_AUS_1975_18953 shared 74.1% similarity with the prototype strain of SINV AR339 recovered from a pool of trapped mosquitoes in Sindbis Village, Egypt, in 1955 (7), 72.4% similarity with an

Citation Pickering P, Aaskov JG, Liu W. 2019. Complete genomic sequence of an Australian Sindbis virus isolated 44 years ago reveals unique indels in the E2 and nsP3 proteins. Microbiol Resour Announc 8:e00246-19. https://doi.org/10.1128/MRA.00246-19.

Editor Jelle Matthijnssens, KU Leuven Copyright © 2019 Pickering et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Wenjun Liu, Wenjun.liu@defence.gov.au.

Received 3 March 2019 **Accepted** 25 April 2019 **Published** 16 May 2019



FIG 1 The maximum clade credibility tree of the genetic relationships between SINV_AUS_1975_18953, MRE-16/ Malaysia/1975, and YN-222/China/2013 and other strains of SINV virus. The maximum clade credibility tree was generated using the Bayesian phylogenetic method in the Geneious software package version 11.1.2 for analysis of the selected whole-genome SINV sequences, employing the HKY85 substitution model, the gamma rate variation, and a chain length of 1,000,000 with a burn-in length of 10,000. Numbers at the nodes indicate posterior probabilities. The naming convention for the viruses is name of strain/country/year of isolation/GenBank accession number.

Australian strain of SINV, SW6562 (GenBank accession number AF429428), isolated in 1984, and 95.1% similarity with an isolate of SINV MRE-16 (GenBank accession numbers AF492770 and U90536) obtained in Malaysia from *Culex tritaeniorhynchus* mosquitoes in 1975 (8). It also shared 90.1% similarity with a SINV virus strain isolated in China in 2013 (YN_222/China/2013, GenBank accession number MH229928).

A maximum clade credibility phylogenetic analysis of the whole-genome sequences of SINV_AUS_1975_18953 and those in GenBank placed this virus in a distinct clade in SINV virus genotype II, along with SINV MRE-16/Malaysia/1975 and YN-222/China/2013 (Fig. 1).

The deduced amino acid sequences of the open reading frames of SINV_AUS_ 1975_18953 contained two significant deletions in the E2 protein between amino acids 182 and 184 and between 201 and 228, as well as indels in the nonstructural protein 3 (nsP3) compared to the prototype strain AR339 (Table 1). These deletions in domain B of E2 are the putative cell receptor recognition site involving virus entry (9). Similar but slightly different deletions in the E2 protein were observed in a small-plaque variant of the SINV MRE-16/Malaysia/1975 strain with a deletion between amino acids 200 and 229 and in an SINV strain of YN-222/China/2013 with a deletion between amino acids 199 and 228. The deletion of an amino acid sequence between amino acids 200 to 229 of the E2 protein in MRE-16/Malaysia/1975 virus significantly reduced its ability to infect *Aedes aegypti* mosquitoes by the oral route compared to its E2-undeleted parental virus (8).

It is unclear how or why these deletions occurred in the E2 protein of SINV strains sampled over decades and from locations thousands of kilometers apart. It also is unclear whether SINV_AUS_1975_18953 contained only this deletion mutant, rather

TABLE 1 Amino	acid indels	in Sindbis	virus strain	SINV_A	AUS_1975_	18953	compared	with
the prototype s	train AR339	(GenBank	accession n	o. <mark>J023</mark>	63)			

	Amino acid	Amino acid sequences				
Gene	positions	Insertion(s)	Deletions			
nsP3	329–330 353–373 395 431 464–465 481–507 512–513	VTDVSLDVEGGHVAANRSEVHSE V Q PS	PE GGVSMSLGSIFDGETARQAAVQPLATG PM			
	571	Q				
E2	182–184 201–228		ESS CKCGDYKTGTVSTRTEITGCTAIKQCVA			

than it being a minor subpopulation, as was the case with the Malaysian SINV MRE-16, because the original isolation had been made in the neurological tissues of mice. The significance of the indels in the nsP3 protein also remains to be determined. Duplication of elements in this region of the nsP3 protein of another alphavirus, Ross River virus, was associated with fitness gain and an explosive outbreak of infection in the Pacific regions (10).

Data availability. The SINV genome sequence in this announcement is available in GenBank as accession number MG182396.

ACKNOWLEDGMENTS

This work was supported by the Joint Health Command of the Australian Defence Force. The funder had no role in the study design, data collection and analysis, decision to publish, or preparation of the manuscript.

The opinions expressed by authors contributing to this article do not necessarily reflect the opinions of the institutions with which they are affiliated.

REFERENCES

- Adouchief S, Smura T, Sane J, Vapalahti O, Kurkela S. 2016. Sindbis virus as a human pathogen—epidemiology, clinical picture and pathogenesis. Rev Med Virol 26:221–241. https://doi.org/10.1002/rmv.1876.
- Doherty RL, Carley JG, Filippich C, Kay BH, Gorman BM, Rajapaksa N. 1977. Isolation of Sindbis (alphavirus) and Leanyer viruses from mosquitoes collected in the Northern Territory of Australia, 1974. Aust J Exp Biol Med Sci 55:485–489. https://doi.org/10.1038/icb.1977.47.
- Doherty RL, Bodey AS, Carew JS. 1969. Sindbis virus infection in Australia. Med J Aust 2:1016–1017.
- Guard RW, McAuliffe MJ, Stallman ND, Bramston BA. 1982. Haemorrhagic manifestations with Sindbis infection. Case report. Pathology 14:89–90. https://doi.org/10.3109/00313028209069049.
- Bergqvist J, Forsman O, Larsson P, Naslund J, Lilja T, Engdahl C, Lindstrom A, Gylfe A, Ahlm C, Evander M, Bucht G. 2015. Detection and isolation of Sindbis virus from mosquitoes captured during an outbreak in Sweden, 2013. Vector Borne Zoonotic Dis 15:133–140. https://doi.org/ 10.1089/vbz.2014.1717.
- 6. Liu W, Pickering P, Duchene S, Holmes EC, Aaskov JG. 2016. Highly

divergent Dengue virus type 2 in traveler returning from Borneo to Australia. Emerg Infect Dis 22:2146–2148. https://doi.org/10.3201/eid2212.160813.

- Taylor RM, Hurlbut HS, Work TH, Kingston JR, Frothingham TE. 1955. Sindbis virus: a newly recognized arthropodtransmitted virus. Am J Trop Med Hyg 4:844–862. https://doi.org/10.4269/ajtmh.1955.4.844.
- Myles KM, Pierro DJ, Olson KE. 2003. Deletions in the putative cell receptor-binding domain of Sindbis virus strain MRE16 E2 glycoprotein reduce midgut infectivity in Aedes aegypti. J Virol 77:8872–8881. https:// doi.org/10.1128/JVI.77.16.8872-8881.2003.
- Li L, Jose J, Xiang Y, Kuhn RJ, Rossmann MG. 2010. Structural changes of envelope proteins during alphavirus fusion. Nature 468:705–708. https:// doi.org/10.1038/nature09546.
- Aaskov J, Jones A, Choi W, Lowry K, Stewart E. 2011. Lineage replacement accompanying duplication and rapid fixation of an RNA element in the nsP3 gene in a species of alphavirus. Virology 410:353–359. https:// doi.org/10.1016/j.virol.2010.11.025.