# **Role of pulmonary function testing in inflammatory bowel diseases (Review)**

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Abstract. Inflammatory bowel disease (IBD) is a term used to describe chronic inflammatory entities of the gastrointestinal system with an unclear etiology. Extra-intestinal manifestations beyond the involvement of the gastrointestinal tract can also occur. Several studies have investigated the alterations of pulmonary function tests (PFTs) in patients with IBD. To the best of our knowledge, the present review article is the first to summarize all the types of PFTs that have been performed in patients with IBD. Contradictory data exist regarding the association of PFT alterations with disease activity. PFT abnormalities can develop in individuals with IBD who have no clear clinical signs or radiological evidence, suggesting that PFTs may be useful in detecting latent respiratory involvement. The most prevalent finding in the PFTs of adults and children with IBD is an impairment in the diffusing capacity for carbon monoxide, although evidence on the other tests, particularly spirometric values, and their connection with disease activity is inconsistent.

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*Key words:* pulmonary function testing, inflammatory bowel disease, spirometry, diffusing capacity for carbon monoxide

## Contents

- 1. Introduction
- 2. Spirometry
- 3. Diffusing capacity for carbon monoxide
- 4. Lung volumes
- 5. Bronchoprovocation challenge testing
- 6. Exhaled nitric oxide measurement
- 7. Pulmonary function testing in children with IBD
- 8. Mechanisms responsible for alterations in PFTs in patients with IBDs
- 9. Conclusions and future perspectives

# 1. Introduction

Inflammatory bowel disease (IBD) is a term used to describe chronic entities characterized by inflammation, mainly affecting the gastrointestinal system. The underlying etiology of this condition remains unclear. Crohn's disease (CD) and ulcerative colitis (UC) represent the two main types of chronic IBDs (1). In spite of CD and UC being different entities, both may present with any of the following manifestations: Abdominal pain, tenesmus, diarrhea, steatorrhea, fever, rectal bleeding, severe cramps or muscle spasms in the pelvic area and weight loss (2).

Extra-intestinal manifestations or complications beyond the involvement of the gastrointestinal tract occur at an incidence of 21-41% (3). These manifestations can occur concurrently with intestinal inflammation or may develop independently of the intestinal activity. They can significantly affect the quality of life of patients, causing severe morbidity and involving several organs (4). Frequent extra-intestinal manifestations include anemia, osteoporosis, cutaneous lesions, as well as

ocular, liver and articular diseases (5). Ulcers in the oral cavity and thromboembolic disease can also occur (6). However, pulmonary involvement is relatively infrequent (7).

The spectrum of lung involvement in IBDs is broad, spanning from subclinical abnormal involvement to interstitial lung disease (ILD) (8). Other pulmonary manifestations are airway disease (panbronchiolitis, bronchiolitis obliterans organizing pneumonia and bronchiectasis), inflammatory tracheal stenosis, pulmonary vasculitis, thromboembolic disease, pleural disease, apical fibrosis, sarcoidosis, enteric-pulmonary fistulas, Langerhans cell histiocytosis, manifestations resembling Wegener's granulomatosis and adverse drug toxicities (9).

Lung abnormalities in IBDs can be present years following the onset of the disease, occurring commonly during active disease, infrequently independent of disease activity, and even in patients in the post-colectomy period (10). The underlying pathogenesis of lung abnormalities in IBDs may be associated with the fact that both the colonic and respiratory epithelial cells share a common embryonic origin. The respiratory and gastrointestinal systems contain submucosal lymphoid tissue and play a key role in host mucosal defense (11). The similarity in mucosal immunity leads to similar pathogenic alterations, which may be caused by epithelial cell exposure to common inhaled or ingested antigens, leading to lymphoid tissue sensitization and an inflammatory process (12). The activated inflammatory cells in the gastrointestinal tract are capable of producing numerous circulating cytokines that can regulate the endothelial cell adhesion molecules, modify leukocyte migration, enhance the production of toxic reactive oxygen metabolites and induce lung parenchyma damage (13). Recent research has demonstrated that there is a pathological connection between the lung and bowel with the longer course of the disease, the greater extent of the disease, and the more severe the pulmonary function damage is in individuals with UC (14). Animal experiments have also noted that rat models of UC exhibited lung pathological injury (14-16).

Although pulmonary involvement in IBDs is rare, the prevalence of pulmonary function test (PFT) abnormalities in patients with IBDs has been reported to be 17-55%, indicating that occult pulmonary disease may be detected using PFT variables (17). Some studies have examined possible changes in pulmonary function parameters in patients with IBD and their association with disease activity (18,19). According to some reports, PFT alterations in patients with IBD are related to disease activity (20,21), whereas other authors have found that active disease does not affect the values of PFTs (22). Moreover, apart from traditional PFTs, additional PFTs, such as bronchoprovocation challenge and the measurement of exhaled nitric oxide (NO), have been performed in patients with IBDs and have been associated with disease activity (23,24). The present review article is the first, to the best of our knowledge, to summarize all the types of PFTs that have been performed in patients with IBD and to discuss the association of their results with the activity of the disease.

# 2. Spirometry

Spirometry calculates the maximal air volume that a patient can inspire and expire using maximal effort, estimating volume or flow as a function of time. The most common measurements are the forced vital capacity (FVC), which represents the air volume exhaled during a forceful and complete expiration and the forced expiratory volume in 1 sec (FEV1), which is the air volume exhaled in the first second during an FVC maneuver (25). Another variable that can be measured during the FVC maneuver is the mean forced expired flow as lung volume decreases from 75 to 25% of vital capacity  $FEF_{(25.75)}$ , which is associated with changes in the small airways (26).

Alterations in the values of FEV1, FVC and FEF(25.75) have been found in patients with IBDs (9,10,19,23,27-36). More specifically, obstructive dysfunction was observed in some studies (17,19,29,33,37), and obstructive and/or restrictive ventilatory defects were observed in others (10,21,22,27,35), and some investigators have demonstrated isolated reductions in the absolute values of FEV1, FVC and FEF(25.75) (9,21,28,3) 0-32,34,36). In some studies, these abnormalities were associated with disease activity (9,10,19,21,25,34,36), whereas other studies have mentioned no association between an impairment in spirometric values and the activity of IBDs (22,30-33,35). Of note, in the study by Ellrichmann et al (37), patients with active IBD presented with significant obstructive abnormalities in their PFTs, with obstruction being related to inflammatory activity. However, treatment with anti-tumor necrosis factor (TNF) antibody induced a significant improvement in obstructive dysfunction (37). On the other hand, some studies did not reveal alterations in the spirometric values of patients with IBDs compared to healthy controls (18,38-40).

### 3. Diffusing capacity for carbon monoxide

Diffusing capacity for carbon monoxide (DLCO) testing is used to differentiate patients with exertional dyspnea, spirometric obstruction and spirometric restriction, and to detect ILD, pulmonary vascular diseases, occupational pulmonary diseases and or pulmonary side-effects of radiation or drugs (41). Reduced DLCO values have been noted in the majority of studies investigating PFT performance in patients with IBD (9,10,13,18,19,32-36,38). In several studies, alterations in DLCO, consistent with ILD, have been observed and this lung involvement has been shown to be associated with IBD activity (9,10,13,19,34-36). In addition, in the study by Marvisi and Fornasari (13), there was a strong association between DLCO values and histopathological grading, suggesting that DLCO testing may reflect IBD activity.

### 4. Lung volumes

Total lung capacity (TLC) and residual volume (RV), which represents the air volume left in the respiratory system at the end of a maximal expiration, can be calculated either by gas dilution or whole-body plethysmography (42). The RV/TLC ratio reflects the resting pulmonary hyperinflation (43). Functional residual capacity (FRC) represents the air volume that remains in the respiratory tract after a normal exhalation. Increased lung volumes result in increased FRC (44).

The RV, TLC and RV/TLC ratio have all been found to be elevated in patients with IBD and to be related to disease activity (10,22,40). Additionally, FRC has been found to be greater in patients with IBDs compared with healthy controls and during the exacerbation of the IBDs than during the remission phase (24). Elevated lung volumes in patients with IBD may represent bronchial or bronchiolar inflammation, and an increase in RV/TLC may be a useful tool for investigating lung involvement in these patients (45).

### 5. Bronchoprovocation challenge testing

Methacholine challenge testing is the most frequent type of bronchoprovocation testing using the acetylcholine derivative, methacholine, to cause bronchoconstriction. Methacholine has limited side-effects. Airway hyperreactivity is detected by a decrease in FEV1. The provocative dose (PD20) or concentration (PC20), which results in a 20% decrease in FEV1 in a positive test, is recorded. This test is used to aid with the diagnosis of asthma (45-47).

Increased bronchial responsiveness to the administration of methacholine has been described in patients with IBDs with no respiratory symptoms, no abnormal findings on chest computed tomography and normal FEV1 values (24,48,49). In addition, significantly increased bronchial responsiveness has been observed in patients with CD and extra-intestinal manifestations, and in patients with CD treated with azathioprine, possibly due to a side-effect of azathioprine (24). There is solid evidence in the existing literature of the association between airway inflammation and bronchial hyperresponsiveness (48). However, neither the decrease in FEV1 nor the PD20 values have been found to be associated with the disease activity or duration (24,48).

### 6. Exhaled nitric oxide measurement

The contribution of NO to the pathophysiology of the respiratory system has been extensively studied. There is conflicting evidence concerning the exact role of NO in respiratory diseases. In pathological conditions, NO is a pro-inflammatory factor with immunomodulatory properties, predisposing to the development of airway hyperresponsiveness. On the other hand, in physiological situations, NO weakly mediates smooth muscle relaxation and is protective against airway hyperresponsiveness. Exhaled NO originates from the airway epithelial cells. The clinical usefulness of NO measurement is most important in allergic airway disease (50).

Increased exhaled NO has been found in patients with IBDs (23,51-54). It has also been described that increased exhaled NO is positively associated with disease activity, as estimated by validated activity indexes (23,51,52). Exhaled NO levels have been found to have a fair association with systemic inflammatory biomarkers in patients with CD, with the exception of fecal calprorectin (51). Moreover, increased exhaled NO levels have been observed in patients with IBDs and lung involvement compared to those without pulmonary involvement, indicating that increased exhaled NO values may be used for identifying patients with IBDs who require the further evaluation of the respiratory system (52).

By contrast, Ikonomi *et al* (53) study found that patients with IBDs had almost the same exhaled NO levels as the controls, and this finding was attributed to the fact that gastro-intestinal mucosal activity was not reflected in the amount of exhaled NO.

### 7. Pulmonary function testing in children with IBD

Few studies have reported the performance of PFTs in children with IBDs. As regards the spirometric values, a significant deterioration during disease activity compared to remission status concerning FEV1, FVC and FEF<sub>(25.75)</sub> with a negative association between the disorder's activity and these parameters has been reported (55). As previously demonstrated, although expiratory flows were impaired, no important differences were found between the acute and quiescent phases (56). Moreover, in another study, maximum expiratory flows at 50 and 25% of vital capacity were mildly decreased in patients with CD (57). On the contrary, Welsh *et al* (58) mentioned spirometry measurements within normal limits and without significant associations with the duration of the disease and hospitalizations in children with IBDs.

DLCO is the most frequently reported abnormality in PFTs of children with IBDs, as it has been observed to be significantly impaired in all relevant studies regarding PFTs in these patients (55-58) and has been associated with disease activity (55,56). Lung volumes have been noted to be decreased (54) or normal in testing with plethysmography (58) and in one study, RV/TLC ratios were mildly increased in children with UC (57), while there was no association with the disease severity (55,57,58). Increased bronchial responsiveness with no association with treatment or the duration of the disease has been demonstrated in children with CD (59), while levels of exhaled NO have been found to be elevated in children with IBDs, with no association with intestinal disease activity or respiratory symptoms, indicating a latent lung involvement in the systemic disease (60).

The prolonged process of inflammation and lung damage caused by circulating factors and immune complexes, the common embryological origin of both gastrointestinal and respiratory epithelium, the similarity of the mucosal immune response and activated inflammatory cells in the gastrointestinal tract produce several circulating mediators capable of causing lung tissue damage, leading to affection of the alveolocapillary membrane and alveolar damage. These mechanisms can explain why lung involvement leads to abnormalities in the interstitium and airways that are presented early by abnormal PFTs even in asymptomatic children with IBDs (55). In the study by Furlano et al (57), the PFT changes that were observed did not last over a median of 34 months of follow-up. This suggests that lung involvement in children with IBDs is variable and changes over time (57). The findings of the performance of PFTs in IBDs are illustrated in Figs. 1-6.

# 8. Mechanisms responsible for alterations in PFTs in patients with IBDs

In addition to the mechanisms responsible for the spirometry measurements alterations described above, an elevated percentage of alveolar lymphocytes has been mentioned in bronchoalveolar lavage obtained from asymptomatic patients with CD, indicating a shift in the proportions of alveolar cells compatible with alveolitis (39). This finding was significantly associated with a decrease in spirometry parameters (39). Another theory suggests that lymphocytes that are sensitized from the gastrointestinal tract may lead to



Figure 1. Findings from spirometry in inflammatory bowel diseases. IBDs, inflammatory bowel diseases; FEV1, forced expiratory volume in 1 sec; FVC, forced vital capacity; FEF<sub>(25.75%)</sub>, mean forced expired flow as lung volume decreases from 75 to 25% of vital capacity.



IBDs Bronchoprovocation Challenge Testing No association with disease activity

Figure 2. Findings from measurement of diffusing capacity for carbon monoxide in inflammatory bowel diseases. IBDs, inflammatory bowel diseases; DLCO, diffusing capacity for carbon monoxide.



Figure 4. Findings from bronchoprovocation challenging testing in inflammatory bowel diseases. IBDs, inflammatory bowel diseases.

Bronchial



Figure 3. Findings from measurement of lung volumes in inflammatory bowel diseases. IBDs, inflammatory bowel diseases; FRC, functional residual capacity; RV, residual volume; TLC, total lung capacity.

inflammation in the mucosa of other tissues. According to a study by Mohamed-Hussein *et al* sputum lymphocytosis in patients with UC, was significantly associated with a decrease in spirometry parameters (21). Moreover, the loss of body proteins and the reduction in body mass index in patients with IBD have been found to be related to reductions in spirometry parameters, indicating a poor nutritional status as a possible factor resulting in abnormalities in spirometric values (21,61).

Figure 5. Findings from the measurement of exhaled NO in inflammatory bowel diseases. IBDs, inflammatory bowel diseases; NO, nitric oxide.

It has been indicated that subclinical alveolar inflammation or ILD may be present in patients with active IBD, since a reduced DLCO of the lungs is an early manifestation of ILD, and supports the hypothesis that DLCO testing may be utilized as a non-invasive indicator of gastrointestinal inflammation in patients with IBD (13). As regards the question of whether the lungs are target organs in IBDs, the reply seems to be positive. The pathogenesis of IBD causing ILD is unclear.



Figure 6. Findings from pulmonary function test in children with inflammatory bowel diseases. IBDs, inflammatory bowel diseases; DLCO, diffusing capacity for carbon monoxide; NO, nitric oxide; RV, residual volume; TLC, total lung capacity.

However, similarities in both morphology and development exist between the colonic and the bronchial epithelium (13). The activated inflammatory cells in bowel tissues produce circulating cytokines, such as interleukin (IL)-1, IL-2, IL-6 and TNF- $\alpha$ , which mediate inflammatory processes, resulting in lung tissue damage (13). This hypothesis is also supported by alveolar lymphocytosis in the bronchoalveolar lavage of asymptomatic patients (39). Furthermore, in a series of cases by Marvisi and Fornasari (13), a low incidence of positive p-ANCA tests was observed, and the researchers suggested the possible pathogenetic contribution of these antibodies in the context of neutrophil enzymatic release and lung damage.

Increased bronchial responsiveness can also be explained by the inflammation of the small airways induced by the recruitment of lymphocytes in bronchi, activated in the gastrointestinal tract (24). Abnormal findings from bronchoprovocation challenge testing have also been noted in patients with IBD receiving azathioprine (24). Azathioprine is the nitroimidazole derivative of 6-mercaptopurine and the nitroimidazole moiety is responsible for hypersensitivity reactions (62). Therefore, a side-effect of azathioprine cannot be excluded as a mechanism of increased bronchial responsiveness in these patients (24).

Active IBD is associated with the increased activity of inducible NO synthase (iNOS), which results in an increase in mucosal and plasma NO levels. Pro-inflammatory mediators and products of bacteria are present in increased amounts in tissues with inflammation in patients with IBD. Due to increased intestinal permeability, the release of these mediators into the systemic circulation is facilitated, inducing the expression of iNOS in other organs, including the lungs, leading to airway inflammation and increased exhaled NO levels (63). This fact is in accordance with the presence of increased counts of airway eosinophils and lymphocytes in patients with IBD (64).

# 9. Conclusions and future perspectives

PFT abnormalities can occur in patients with IBDs without any obvious clinical manifestations or radiological findings, reflecting the possible importance of PFTs in the detection of latent respiratory involvement. An impairment in DLCO is the most common finding in the PFTs of adults and children with IBDs, while there are conflicting data regarding other tests, particularly spirometric values, and their association with disease activity. However, according to the existing literature, there is no evidence to support the use of PFTs for monitoring IBD activity. Moreover, according to the existing literature, there is no difference between UC and CD as regards the association with PFTs. In addition, the existing literature refers to the comparison of variables of PFTs in remission and activity of the diseases and not in different stages. Thus, larger prospective studies are required to clarify the role of PFTs as diagnostic tools for recognizing subclinical pulmonary damage and determining the activation of IBDs.

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CD, KT, PP and DM conceptualized the study. VEG, DAS, AG, SC, PS, NT and DM analyzed the data from the literature for inclusion in the review and wrote and prepared the draft of the manuscript. DAS and DM provided critical revisions. All authors contributed to manuscript revision and have read and approved the final version of the manuscript. Data authentication is not applicable.

# Ethics approval and consent to participate

Not applicable.

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Not applicable.

# **Competing interests**

The authors declare that they have no competing interests.

### References

- 1. Baumgart DC and Carding SR: Inflammatory bowel disease: Cause and immunobiology. Lancet 369: 1627-1640, 2007.
- Wang GF, Ren JA, Liu S, Chen J, Gu GS, Wang XB, Fan CG and Li JS: Clinical characteristics of non-perianal fistulating Crohn's disease in China: A single-center experience of 184 cases. Chin Med J (Engl) 125: 2405-2410, 2012.
- 3. Su CG, Judge TA and Lichtenstein GR: Extraintestinal manifestations of inflammatory bowel disease. Gastroenterol Clin North Am 31: 307-327, 2002.
- 4. Williams H, Walker D and Orchard TR: Extraintestinal manifestations of inflammatory bowel disease. Curr Gastroenterol Rep 10: 597-605, 2008.
- Veloso FT, Carvalho J and Magro F: Immune-related systemic manifestations of inflammatory bowel disease. A prospective study of 792 patients. J Clin Gastroenterol 23: 29-34, 1996.

- Camus P, Piard F, Ashcroft T, Gal AA and Colby TV: The lung in inflammatory bowel disease. Medicine (Baltimore) 72: 151-183, 1993.
- Hoffmann RM and Kruis W: Rare extraintestinal manifestations of inflammatory bowel disease. Inflamm Bowel Dis 10: 140-147, 2004.
- Storch I, Sachar D and Katz S: Pulmonary manifestations of inflammatory bowel disease. Inflamm Bowel Dis 9: 104-115, 2003.
- 9. Ji XQ, Wang LX and Lu DG: Pulmonary manifestations of inflammatory bowel disease. World J Gastroenterol 20: 13501-13511, 2014.
- Songür N, Songür Y, Tüzün M, Doğan I, Tüzün D, Ensari A and Hekimoglu B: Pulmonary function tests and high-resolution CT in the detection of pulmonary involvement in inflammatory bowel disease. J Clin Gastroenterol 37: 292-298, 2003.
- Higenbottam T, Cochrane GM, Clark TJ, Turner D, Millis R and Seymour W: Bronchial disease in ulcerative colitis. Thorax 35: 581-585, 1980.
- 12. Black H, Mendoza M and Murin S: Thoracic manifestations of inflammatory bowel disease. Chest 131: 524-532, 2007.
- Marvisi M and Fornasari G: Is the lung a target organ in inflammatory bowel disease? Recenti Prog Med 92: 774-777, 2001 (In Italian).
- 14. Wang JY, Wang XY, Wu HY, Sun HY, Liu DM, Zhang W, Jin CX and Wang SR: The association between pulmonary function impairment and colon inflammation in ulcerative colitis patients: A scientific basis for exterior-interior correlation between lung and large intestine. Chin J Integr Med 22: 894-901, 2016.
- 15. Ni L, Jing S, Zhu L, Yang X, Wang X and Tu S: The immune change of the lung and bowel in an ulcerative colitis rat model and the protective effect of sodium houttuyfonate combined with matrine. Front Immunol 13: 888918, 2022.
- 16. Yan X, Yu X, Jiang C, Cao Y, Zhu L, Du C and Jia Y: Tonifying-Qi-and-detoxification decoction attenuated injuries of colon and lung tissues in ulcerative colitis rat model via regulating NF-κB and p38MAPK pathway. Ann Transl Med 10: 455, 2022.
- Gupta SJ, Gupta VL, Kothari HG, Samarth AR, Gaikwad NR and Parmar SM: Assessment of occult pulmonary involvement in ulcerative colitis. Inflamm Intest Dis 5: 144-150, 2020.
- Neilly JB, Main AN, McSharry C, Murray J, Russell RI and Moran F: Pulmonary abnormalities in Crohn's disease. Respir Med 83: 487-491, 1989.
- Yilmaz A, Yilmaz Demirci N, Hoşgün D, Uner E, Erdoğan Y, Gökçek A and Cağlar A: Pulmonary involvement in inflammatory bowel disease. World J Gastroenterol 16: 4952-4957, 2010.
- 20. Pasquis P, Colin R, Denis P, Baptiste P, Galmiche JP and Hecketsweiler P: Transient pulmonary impairment during attacks of Crohn's disease. Respiration 41: 56-59, 1981.
- Mohamed-Hussein AA, Mohamed NA and Ibrahim ME: Changes in pulmonary function in patients with ulcerative colitis. Respir Med 101: 977-982, 2007.
- 22. Tunc B, Filik L, Bilgic F, Arda K and Ulker A: Pulmonary function tests, high-resolution computed tomography findings and inflammatory bowel disease. Acta Gastroenterol Belg 69: 255-260, 2006.
- 23. Koek GH, Verleden GM, Evenepoel P and Rutgeerts P: Activity related increase of exhaled nitric oxide in Crohn's disease and ulcerative colitis: A manifestation of systemic involvement? Respir Med 96: 530-535, 2002.
- 24. Louis E, Louis R, Drion V, Bonnet V, Lamproye A, Radermecker M and Belaiche J: Increased frequency of bronchial hyperresponsiveness in patients with inflammatory bowel disease. Allergy 50: 729-733, 1995.
- 25. Graham BL, Steenbruggen I, Miller MR, Barjaktarevic IZ, Cooper BG, Hall GL, Hallstrand TS, Kaminsky DA, McCarthy K, McCormack MC, *et al*: Standardization of spirometry 2019 update. an official American thoracic society and european respiratory society technical statement. Am J Respir Crit Care Med 200: e70-e88, 2019.
- Riley CM, Wenzel SE, Castro M, Erzurum SC, Chung KF, Fitzpatrick AM, Gaston B, Israel E, Moore WC, Bleecker ER, *et al*: Clinical implications of having reduced mid forced expiratory flow rates (FEF25-75), independently of FEV1, in adult patients with asthma. PLoS One 10: e0145476, 2015.
   Goyal A, Ghoshal UC, Nath A, Jindal S and Mohindra S:
- 27. Goyal A, Ghoshal UC, Nath A, Jindal S and Mohindra S: Pulmonary function in patients with ulcerative colitis and its relationship with disease severity. JGH Open 1: 32-37, 2017.

- Godet PG, Cowie R, Woodman RC and Sutherland LR: Pulmonary function abnormalities in patients with ulcerative colitis. Am J Gastroenterol 92: 1154-1156, 1997.
- Karadag F, Ozhan MH, Akçiçek E, Günel O, Alper H and Veral A: Is it possible to detect ulcerative colitis-related respiratory syndrome early? Respirology 6: 341-346, 2001.
- Dierkes-Globisch A and Mohr H: Pulmonary function abnormalities in respiratory asymptomatic patients with inflammatory bowel disease. Eur J Intern Med 13: 385, 2002.
- Zhao Y, Wang J, Liu Z, Lin H, Shi Y and Sun X: Pulmonary dysfunction in 114 patients with inflammatory bowel disease. Medicine (Baltimore) 96: e6808, 2017.
- 32. Heatley RV, Thomas P, Prokipchuk EJ, Gauldie J, Sieniewicz DJ and Bienenstock J: Pulmonary function abnormalities in patients with inflammatory bowel disease. Q J Med 51: 241-250, 1982.
- Douglas JG, McDonald CF, Leslie MJ, Gillon J, Crompton GK and McHardy GJ: Respiratory impairment in inflammatory bowel disease: Does it vary with disease activity? Respir Med 83: 389-394, 1989.
- 34. Herrlinger KR, Noftz MK, Dalhoff K, Ludwig D, Stange EF and Fellermann K: Alterations in pulmonary function in inflammatory bowel disease are frequent and persist during remission. Am J Gastroenterol 97: 377-381, 2002.
- Sethy PK, Dutta U, Aggrawal AN, Das R, Gulati M, Sinha SK and Singh K: Pulmonary and hematological alterations in idiopathic ulcerative colitis. Indian J Gastroenterol 22: 176-179, 2003.
- 36. Ateş F, Karincaoğlu M, Hacievlıyagıl SS, Yalniz M and Seçkın Y: Alterations in the pulmonary function tests of inflammatory bowel diseases. Turk J Gastroenterol 22: 293-299, 2011.
- 37. Ellrichmann M, Bethge J, Boesenkoetter J, Conrad C, Noth R, Bahmer T, Nikolaus S, Aden K, Zeissig S and Schreiber S: Subclinical pulmonary involvement in active IBD responds to biologic therapy. J Crohns Colitis 15: 1339-1345, 2021.
- Tzanakis N, Bouros D, Samiou M, Panagou P, Mouzas J, Manousos O and Siafakas N: Lung function in patients with inflammatory bowel disease. Respir Med 92: 516-522, 1998.
- Johnson NM, Mee AS, Jewell DP and Clarke SW: Pulmonary function in inflammatory bowel disease. Digestion 18: 416-418, 1978.
- Wallaert B, Colombel JF, Tonnel AB, Bonniere P, Cortot A, Paris JC and Voisin C: Evidence of lymphocyte alveolitis in Crohn's disease. Chest 87: 363-367, 1985.
- Enright Md P: Office-based DLCO tests help pulmonologists to make important clinical decisions. Respir Investig 54: 305-311, 2016.
- 42. O'Donnell CR, Bankier AA, Stiebellehner L, Reilly JJ, Brown R and Loring SH: Comparison of plethysmographic and helium dilution lung volumes: Which is best for COPD? Chest 137: 1108-1115, 2010.
- 43. Shin TR, Oh YM, Park JH, Lee KS, Oh S, Kang DR, Sheen S, Seo JB, Yoo KH, Lee JH, *et al*: The prognostic value of residual volume/total lung capacity in patients with chronic obstructive pulmonary disease. J Korean Med Sci 30: 1459-1465, 2015.
- 44. Hopkins E and Sharma S: Physiology, functional residual capacity. In: StatPearls [Internet]. StatPearls Publishing, Treasure Island, FL, 2020. https://www.ncbi.nlm.nih.gov/books/NBK500007. Updated September 28, 2020.)
- Lee MK, Yoon HK, Kim SW, Kim TH, Park SJ and Lee YM: Nonspecific bronchoprovocation test. Tuberc Respir Dis (Seoul) 80: 344-350, 2017.
- Davis BE, Blais CM and Cockcroft DW: Methacholine challenge testing: Comparative pharmacology. J Asthma Allergy 11: 89-99, 2018.
- 47. Bohadana AB, Rokach A, Wild P and Izbicki G: Asthma-like symptoms induced by the methacholine challenge test: Do they predict a negative-to-positive switch in the test result?-case report. J Thorac Dis 10: E716-E720, 2018.
- Ceyhan BB, Karakurt S, Cevik H and Sungur M: Bronchial hyperreactivity and allergic status in inflammatory bowel disease. Respiration 70: 60-66, 2003.

- 49. Bartholo RM, Zaltman C, Elia C, Cardoso AP, Flores V, Lago P, Cassabian L, Dorileo FC and Lapa-e-Silva JR: Bronchial hyperresponsiveness and analysis of induced sputum cells in Crohn's disease. Braz J Med Biol Res 38: 197-203, 2005.
- disease. Braz J Med Biol Res 38: 197-203, 2005.
  50. Taylor DR, Pijnenburg MW, Smith AD and De Jongste JC: Exhaled nitric oxide measurements: Clinical application and interpretation. Thorax 61: 817-827, 2006.
- Quenon L, Hindryckx P, De Vos M, De Looze D, Joos G, Brusselle G and Peeters H: Hand-held fractional exhaled nitric oxide measurements as a non-invasive indicator of systemic inflammation in Crohn's disease. J Crohns Colitis 7: 644-648, 2013.
- 52. Ozyilmaz E, Yildirim B, Erbas G, Akten S, Oguzulgen IK, Tunc B, Tuncer C and Turktas H: Value of fractional exhaled nitric oxide (FE NO) for the diagnosis of pulmonary involvement due to inflammatory bowel disease. Inflamm Bowel Dis 16: 670-676, 2010.
- 53. Ikonomi E, Rothstein RD, Ehrlich AC and Friedenberg FK: Measurement of fractional exhaled nitric oxide as a marker of disease activity in inflammatory bowel disease. J Gastroenterol Pancreatol Liver Disord 3: 10.15226/2374-815X/3/1/00146, 2016.
- 54. Protopapas AA, Vradelis S, Karampitsakos T, Steiropoulos P, Chatzimichael A and Paraskakis E: Elevated levels of alveolar nitric oxide may indicate presence of small airway inflammation in patients with inflammatory bowel disease. Lung 197: 663-670, 2019.
- 55. El Amrousy DM, Hassan S, El-Ashry H, Yousef M and Sharshar R: Pulmonary function tests abnormalities in children with inflammatory bowel disease: Is it common? J Pediatr Gastroenterol Nutr 67: 346-350, 2018.
- 56. Munck A, Murciano D, Pariente R, Cezard JP and Navarro J: Latent pulmonary function abnormalities in children with Crohn's disease. Eur Respir J 8: 377-380, 1995.
- 57. Furlano RI, Basek P, Müller P, Bieli C, Braegger CP, Barben J, Hammer J, Moeller A and Trachsel D: Pulmonary function test abnormalities in pediatric inflammatory bowel disease. Respiration 90: 279-286, 2015.
- Welsh L, Haller W, King LE, Soto-Martinez M, Oliver M, Catto-Smith A and Robinson PJ: Pulmonary function abnormalities in children with active Crohn's disease. Am J Respir Crit Care Med 186: 1060-1061, 2012.
- Mansi A, Cucchiara S, Greco L, Sarnelli P, Pisanti C, Franco MT and Santamaria F: Bronchial hyperresponsiveness in children and adolescents with Crohn's disease. Am J Respir Crit Care Med 161: 1051-1054, 2000.
- 60. Gut G, Ben-Tov A, Lahad A, Soferman R, Cohen S, Tauman R and Sivan Y: Pulmonary functions in children with inflammatory bowel disease. Eur J Gastroenterol Hepatol 28: 708-713, 2016.
- Christie PM and Hill GL: Effect of intravenous nutrition on nutrition and function in acute attacks of inflammatory bowel disease. Gastroenterology 90: 730-736, 1990.
- Stetter M, Schmidl M and Krapf R: Azathioprine hypersensitivity mimicking Goodpasture's syndrome. Am J Kidney Dis 23: 874-877, 1994.
- 63. Hollander D, Vadheim CM, Brettholz E, Petersen GM, Delahunty T and Rotter JI: Increased intestinal permeability in patients with Crohn's disease and their relatives. A possible etiologic factor. Ann Intern Med 105: 883-885, 1986.
- 64. Fireman Z, Osipov A, Kivity S, Kopelman Y, Sternberg A, Lazarov E and Fireman E: The use of induced sputum in the assessment of pulmonary involvement in Crohn's disease. Am J Gastroenterol 95: 730-734, 2000.

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