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Personalizing first-line treatment in advanced colorectal cancer: Present status and future perspectives

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ABSTRACT

Background: Colorectal cancer is one of the most frequent neoplasms worldwide, and the majority of patients are diagnosed in advanced stages. Metastatic colorectal cancer (mCRC) harbors several mutations with different prognostic and predictive values; *KRAS*, *NRAS*, and *BRAF* mutations are the best known. Indeed, *RAS* and *BRAF* molecular status are associated with a different response to monoclonal antibodies (Anti-epidermal growth factor receptor and anti-vascular endothelial growth factor receptor agents), which are usually added to chemotherapy in first-line, and thus allow to select the optimal therapy for patients with mCRC. Furthermore, sidedness is an important predictive and prognostic factor in mCRC, which is explained by the different molecular profile of left and right-sided tumors. Recently, microsatellite instability-high has emerged as a predictive factor of response and survival from immune checkpoint inhibitors in mCRC. Finally, several other alterations have been described in lower frequencies, such as human epidermal growth factor receptor-2 overexpression/ amplification, *PIK3CA* pathway alterations, phosphatase and tension homolog loss, and hepatocyte growth factor/mesenchymal-epithelial transition factor pathway dysregulation, with several targeted therapies already demonstrating activity or being tested in currently ongoing clinical trials.

Aim: To review the importance of studying the predictive and prognostic roles of the molecular profile of mCRC, the changes occurred in recent years and how they would potentially change in the near future, to guide physicians in treatment decisions.

Relevance for Patients: Today, several different therapeutic options can be offered to patients in the first-line setting of mCRC. Therapies at present approved or under investigation in clinical trials will be thoroughly reviewed, with special emphasis on the molecular rationale behind them. Understanding the molecular status, resistance mechanisms and potential new druggable targets may allow physicians to choose the best therapeutic option in the first-line mCRC.

1. Introduction

In 2020, there were 1,931,590 new cases of colorectal cancer (CRC), accounting for 10% of all new cases of cancer worldwide, with the third and second most frequent incidences in men and women, respectively. Furthermore, CRC caused 935,173 deaths in 2020, making it the malignant neoplasm with the fifth highest mortality worldwide. This high mortality rate is explained because a majority of cases are diagnosed in an advanced stage [1].

CRC develops more frequently in patients over 50 years of age, especially in those with a history of smoking, alcoholism, obesity, heavy red meat consumption, and lack of

physical activity [2]. Although its exact origin is not yet known, CRC develops in the context of well-known acquired genetic aberrations, some of which have been shown to be prognostic while some others allow to predict the benefit from different biological agents [3,4].

RAS and *BRAF* mutational status and the analysis of microsatellite instability (MSI) are mandatory in patients with metastatic CRC (mCRC) for prognostic as well as therapeutic purposes. Moreover, some other genetic alterations are increasingly being tested to expand the array of druggable alterations in current daily practice in mCRC, and several agents against some other potentially targetable genetic aberrations are being tested in clinical trials [5,6]. Applying one of Sun Tzu's principles from his *Art of War* ("Know your enemy and know yourself, and you will be victorious in a thousand battles"), we aim to dissect the molecular biology and druggable mutational landscape of CRC to guide treatment decisions in the first-line setting, as well as its future perspectives.

2. Knowing Your Enemy: Molecular Pathways and Mutational Status in mCRC

The past 15 years saw the advent of the biological therapies for mCRC through the appearance of anti-vascular endothelial growth factor (VEGF) and anti-epidermal growth factor receptor (EGFR) agents. However, it was soon evidenced that not all mCRC patients benefited from anti-EGFR agents. First, KRAS exon 2 mutations were unveiled to confer resistance to cetuximab and panitumumab, and later KRAS exon 3 and 4, NRAS exons 2, 3, and 4, and BRAF mutations were also established as resistance mutations to EGFR blockade. Subsequently, it also became clear that patients with right-sided mCRC derived less benefit from anti-EGFR agents. More recently, other molecular alterations such as MSI, HER-2 amplification/overexpression, and NTRK fusions, among others, have been shown to be targetable in mCRC. Therefore, it is of utmost importance that clinicians are aware of the molecular biology of CRC and the biological rationale behind treatment decisions in mCRC.

2.1. The EGFR-related pathway

EGFR belongs to the erythroblastosis oncogene B (*ErbB*)/ human epidermal growth factor receptor (HER) family, which consists of four members: *ErbB1* (EGFR/HER1), *ErbB2* (Neu/ HER2), *ErbB3* (HER3), and *ErbB4* (HER4) [7,8]. Overexpression of EGFR has been observed in 25-77% of CRCs and might also associate with poor prognosis [9-11]. The typical *ErbB* receptor consists of 3 domains: a ligand-binding domain outside the cell, a transmembrane domain and an intracellular domain with distinct tyrosine residues in the C-terminal region where subsequent phosphorylation may take place on activation [12].

The union of the EGF ligand with the EGFR initiates the activation of the EGFR and its subsequent phosphorylation (pEGFR), allowing the formation of a coupling site for GRB2 and its union to SOS in the cytosol. The resulting complex (pEGFR united to SOS) promotes nucleotide exchange and

RAS activation [13-15]. Subsequent RAF activation leads to phosphorylation of mitogen-activated protein kinase (MAPK) and activation of extracellular signal-related kinase (ERK), which might then translocate inside the nucleus to regulate the expression of transcription factors and the activation of specific genes that stimulate cancer progression. Downstream intracellular signaling pathways, including the Phosphatidylinositol-4,5-bisphosphate 3-kinase (PI3K)/AKT and JAK/signal transducer and activator of transcription (STAT)3 pathways, are also triggered to regulate cell growth, survival, and migration [16-18].

2.2. RAS status: Cornerstone mutation of mCRC

RAS proteins are part of a large family of small guanosine triphosphate (GTP) nucleotide-binding proteins [19]. The human RAS superfamily consists of more than 100 members that can be divided into six subfamilies, the most characteristic being *HRAS*, *NRAS*, and *KRAS* [20,21]. *KRAS* mutations are the most common predictive mutations, occurring in 40-45% of all mCRC [22]. The vast majority of *KRAS* mutations (85-90%) occur in codons 12 and 13 of exon 2, while the rest are found in codons 61, 146, and other residues [23,24]. The patients with *KRAS* mutations are most often adult women with mucinous differentiation [25]. *HRAS* and *NRAS* mutations are found in <5% of patients [26]. Patients with *NRAS* mutations are also usually women with left sided tumors [27].

Among *RAS* family members, *KRAS* is the only one which is essential for normal development, as demonstrated by genetic studies in laboratory animals [13,28,29]. KRAS can be expressed as two different variants: 4A and 4B. Variant 4B is the dominant form, which is commonly known as *KRAS*, a cell membrane-bound GTPase that alternates between an active and an inactive form. GTPase activator proteins hydrolyze the nucleotide GTP leading to phosphate loss and formation of nucleotide guanosine diphosphate (GDP), while guanosine nucleotide exchange factors (GEF) facilitate the exchange of GDP to GTP. Both factors control the transition from the inactive form of *KRAS* to its active form [19,30,31].

Mutations in specific codons in *KRAS* alter the position of a glutamate residue at codon 61 [19,32]. *KRAS* activation occurs without the need for binding of the phosphorylated EGFR protein complex to GEF SOS, resulting in the reduction of the GTPase activity of *KRAS*, decreasing the hydrolysis rate of GTP approximately 3-9 times compared to the non-mutated *KRAS* [21,33,34]. The main effect of *RAS* signaling occurs through the RAF/MEK/ERK pathway and the secondary molecular cascade to the PI3K/AKT pathway, which control growth processes and cell survival [13]. This is achieved in part by activating transcription factors that promote ERK-regulated cell cycle progression and by AKT-mediated inactivation of apoptosis [35].

Various studies have shown that *RAS* mutations play a significant role in cell proliferation, suppression of apoptosis and in changing the tumor microenvironment that ultimately promote tumor cell survival and progression of cancer. Additional functions of *KRAS* have been described, such as regulation of cell

migration, endocytosis, cytoskeleton modification, and calcium signaling [36-38].

2.3. BRAF mutations: A particular event

BRAF mutations are found in 8%-10% of CRCs and do not usually overlap with RAS mutations, being considered mutually exclusive [39-41]. Two-thirds of the patients with BRAF mutations have primary tumors on the right side of the colon, being associated with a higher frequency of peritoneal involvement, lymph node metastases, and a lower frequency of pulmonary metastases [40]. Up to one-third of BRAF mutant tumors harbor a high MSI (MSI-H) and the same proportion of MSI-H tumors have BRAF mutations. BRAF appears to act through the dentate/methylating pathway and, indeed, BRAF-mutant tumors are characterized by the methylation of CpG islands that cause the epigenetic repression of tumor suppressor genes, known as CpG island methylating phenotype tumors [42-44]. The BRAF oncogene encodes a serine/ threonine kinase that acts in the MAPK pathway. BRAF mediates its effect through the activation of MAPK, thus promoting cell proliferation. BRAF V600E mutations account for 90% of BRAF mutations in CRC. Their occurrence has been associated with older adult women with a history of smoking [45]. The BRAFV600E mutation is the result of the transversion of thymidine to adenine at nucleotide 1799 in the kinase domain, resulting in a substitution of valine for glutamate leading to constitutive activation of MEK and uninhibited EGFR-independent cell proliferation [46,47]. The fact that BRAF and KRAS/NRAS are mutually exclusive mutations in CRC supports the hypothesis that BRAF is the main effector of KRAS/NRAS in the MAPK pathway and that both have similar effects on tumorigenesis [48,49].

2.4. The VEGF/VEFGR pathway

Angiogenesis, a physiological process by which new vessels form or reform from existing vessels, plays a key role in tumor initiation, growth, and metastasis. Angiogenesis is under a complex regulation involving various proangiogenic and antiangiogenic factors, such as VEGF [50-52]. The VEGF family consists of five members (VEGF-A, VEGF-B, VEGF-C, VEGF-D, and PIGF), which may bind to endothelial cells via tyrosine kinase VEGF receptors (VEGFRs). VEGF, VEGFR, VEGF-A, VEGF-B, and PIGF contribute predominantly to angiogenesis, while VEGF-C and VEGF-D tend to regulate lymphangiogenesis. VEGFRs are divided into three types, VEGFR-1, VEGFR-2, and VEGFR-3, along with the non-tyrosine kinase co-receptors neuropilin-1 and NP-2 [53-56].

VEGFR-1 regulates cell differentiation, migration and promotes differentiation of epithelial cells [57,58]. Meanwhile, VEGFR-3 mediates the differentiation, migration, proliferation, and survival of lymphatic endothelial cells [59]. VEGFR-2 is actively involved in vascular formation and is mostly expressed in blood and lymphatic epithelial cells [60]. VEGF-A and VEGF-B mainly bind to VEGFR-1 and VEGFR-2. VEGFR-1, VEGFR-2, and VEGFR-3 activation leads to phosphorylation of tyrosine residues and activation of various pathways, including the *RAS*/ RAF/ERK/MAPK pathways that promotes epithelial cell growth, and the PI3K/AKT pathway, by which cell apoptosis may be avoided and contributes to the differentiation, proliferation, migration, and apoptosis resistance of epithelial cells [52,56,59]. The proangiogenic effects of VEGF-VEGFR are important in local sites, favoring tumor progression and migration, as well as for neovascularization in metastatic sites to support cancer survival and growth [61].

3. *KRAS*, *NRAS*, and *BRAF* Status: Personalizing the First-line Treatment Today

Fluoropyrimidines are a main part of the backbone of combination regimes in mCRC. Randomized clinical trials have shown that fluoropyrimidine-based combinations with oxaliplatin or irinotecan (FOLFOX, FOLFIRI or XELOX) in the first-line significantly improve treatment efficacy, achieving a response rate of 34-55%, a time to progression of 7-8 months and a median overall survival (mOS) of 14-21 months. The triple therapy with FOLFOXIRI has been compared with FOLFOX or FOLFIRI, demonstrating superiority for FOLFOXIRI in terms of efficacy outcomes, notably with a 25% survival and a 30% increase in response rate. However, because of marked grade 3-4 toxicity, triple therapy is reserved for patients with mCRC with a good performance status, that are highly symptomatic and were the main therapy objective is response rate. In addition, in the past 15 years, monoclonal antibodies have been added to first-line chemotherapy regimens in mCRC [62-64]. Inhibition of the EGFR by panitumumab or cetuximab leads to KRAS becoming GDP-bound, which inhibits downstream signaling [39]. Cetuximab was approved by the FDA in mCRC in 2004, although it was not until 2012 that it was approved in the first-line setting. The OPUS and COIN trials demonstrated a higher objective response rate (ORR) with the firstline chemotherapy plus cetuximab in exon 2 KRAS wild-type mCRC patients in comparison with chemotherapy. However, no differences were reported in mOS and median progression-free survival (mPFS) [36,65]. The CRYSTAL study demonstrated numerically longer mOS (14.1 vs. 10.3 months, HR=0.91, P=0.7), and mPFS (8 vs. 5.6 months, HR=0.93, P=0.86), and a higher ORR (19.2% vs. 15.2%, P<0.0001) with FOLFIRI/Cetuximab in comparison with FOLFIRI alone in patients with KRAS wild-type/BRAF mutant. Benefit was superior with cetuximab regimen in patients with KRAS wild-type/ BRAF wild-type mCRC for mOS (25.1 vs. 21.6 months, HR=0.83, P=0.0549), mPFS (10.9 vs. 8.8 months, HR=0.68, P=0.0016) and ORR (61% vs. 42.6%, P<0.0001) [66]. The FIRE-3 trial evaluated the anti-EGFR cetuximab versus the anti-VEGF bevacizumab, both in combination with chemotherapy in patients with KRAS/NRAS/ BRAF wild-type mCRC. 8 months longer mOS was achieved with cetuximab/FOLFIRI in comparison with bevacizumab/FOLFIRI (33.1 vs. 25 months, HR=0.697, P=0.0059). In addition, ORR with cetuximab/FOLFIRI was higher (72% vs. 56.1%, OR 2.01, 0=0.003) [67]. Furthermore, anti-EGFR agents have also demonstrated its value in the neoadjuvant setting. The CELIM trial evaluated FOLFOX6/FOLFIRI and cetuximab in mCRC with unresectable liver metastasis, achieving a response rate of 70% in KRAS codon 12/13/61 wild-type patients, allowing liver resection in 93% of the studied population [68]. Likewise, the POCHER trial found a

similar response rate (79%) with chemotherapy/cetuximab in a similar population [69]. In mid-2021, the JACCRO trial achieved a median depth of response of 57.4% in *KRAS* wild-type mCRC with mFOLFOXIRI/Cetuximab, compared to a 46% with mFOLFOXIRI/Bevacizumab (P=0.001). Median depth of response was higher in the left vs. right-sided mCRC (60.3% vs. 46.1%, P=0.0007) [70].

Panitumumab was approved by the FDA for first-line mCRC in 2014. The PRIME trial evaluated panitumumab/FOLFOX4 versus FOLFOX4 in the first-line mCRC. This trial found superior mOS (23.9 vs. 19.7 months, HR=0.83, P=0.072), mPFS (9.6 vs. 8 months, HR=0.80, P=0.02) and ORR (57% vs. 48%, P=0.018) with panitumumab/FOLFOX4 in KRAS wild-type mCRC [34]. Further analysis reported modest benefit in mPFS (6.9 vs. 5.5 months, HR=0.68, P=0.006) and mOS (18.7 vs. 15.4 months, HR=0.83, P=0.15) with panitumumab/FOLFOX4 in patients with KRAS/NRAS/BRAF wild-type [71]. The PEAK trial evaluated panitumumab/chemotherapy versus bevacizumab/chemotherapy in KRAS/NRAS wild-type mCRC patients. mPFS with panitumumab/ chemotherapy was 13.1 months compared to 10.1 months (HR=0.61, P=0.0075) with bevacizumab/chemotherapy. mOS was also superior survival benefit (41.3 vs. 28.9 months, HR=0.63, P=0.058) [72]. In the neoadjuvant setting, the phase II VOLFI trial reported the highest ORR in mCRC with liver metastasis to date. FOLFOXIRI plus panitumumab achieved an ORR of 85.7% compared to 60.6% with FOLFOXIRI alone (OR=3.9; P=0.0098). In addition, ORR achieved 90.6% in left-sided and 86% in RAS/BRAF wild-type mCRC [73]. Anti-EGFR plus chemotherapy became the treatment of choice in *RAS* and *BRAF* wild-type mCRC [39].

The AVF 2107 phase III study evaluated the combination of bevacizumab/FOLFIRI versus FOLFIRI alone in first-line mCRC. mPFS (10.6 vs. 6.2 months, HR=0.54, P<0.0001), mOS (20.3 vs. 15.6 months, HR=0.66, P<0.001), and ORR (45% vs. 35%; P=0.004) were superior with bevacizumab/FOLFIRI [74]. Likewise, the NO16966 trial reported a longer mOS (21.2 m vs. 19.9 m, HR=0.89, 0.76-1.03, P=0.07) and mPFS (9.4 m vs. 8.0 m, HR=0.83, P=0.023) with bevacizumab/FOLFOX4 compared to FOLFOX4 alone [75]. In the phase III AVEX trial, bevacizumab remained relatively safe and effective when treating elderly patients with mCRC, achieving a mPFS of 9.1 months with capecitabine/bevacizumab and 5.1 months with capecitabine alone (HR=0.53, P<0.0001). Grade 3-4 adverse events were 40% in the combination group and 22% in the capecitabine-alone group [76]. A meta-analysis of 6 randomized trials including 3060 patients concluded that bevacizumab achieved a significantly longer PFS (HR=0.72, P<0.00001) and OS (HR=0.84, P<0.00001). Further investigation found that both KRAS mutant and KRAS wildtype mCRC may benefit from bevacizumab. A pooled analysis evaluated the efficacy and safety of bevacizumab in mCRC. This study included 3763 patients from different randomized trials and the addition of bevacizumab to chemotherapy was associated with statistically significant increases in OS (HR=0.80, 0.71-0.90) and PFS (HR=0.57, 0.46-0.71). The effects on OS and PFS across subgroups defined by the chemotherapy backbone, extent of disease, age, ECOG and KRAS status were consistent with the overall analysis. Interestingly, the benefit with bevacizumab was found even in mCRC patients with KRAS mutation [74,77-79].

In patients with mCRC who responded to bevacizumab plus chemotherapy, maintenance therapy with bevacizumab may be considered. In the prospective BRiTE study in patients with first-line mCRC, maintenance with bevacizumab dramatically improved mOS compared with no maintenance (31.8 vs. 19.9 months, HR=0.48, P<0.001) [80]. The MACRO trial reported that continuing with bevacizumab plus capecitabine or bevacizumab alone after bevacizumab plus CAPOX achieved a similar mOS (23 vs. 19 months, HR=1.09, P=0.38) [81]. In the phase III CAIRO3 trial, maintenance with bevacizumab plus capecitabine after bevacizumab/CAPOX achieved a mPFS2 of 11.7 months compared to 8.5 months in the capecitabine-alone group (HR=0.63, P<0.001) [82]. Maintaining bevacizumab beyond progression has also been evaluated in different trials. The ML18147 study reported that patients who continued bevacizumab plus chemotherapy after progression achieved a modest improvement in mOS compared to chemotherapy alone (11.2 vs. 9.8 months, HR=0.81, P=0.0062) [83]. However, in the BEBYP trial, the combination of bevacizumab plus chemotherapy beyond progression in the first-line setting achieved only a modest improvement in mPFS (6.8 vs. 5 months, HR=0.7, P=0.01) [84]. Finally, other antiangiogenic agents such as aflibercept and ramucirumab have been tested, respectively, within the VELOUR and RAISE studies in the second-line setting [85,86].

The most relevant studies of first-line therapy in mCRC will be discussed below and are summarized in Table 1.

4. Sidedness Matters in mCRC: Right Versus Left Colon Cancer

Primary tumor sidedness is a main clinical criterion that, combined with RAS and BRAF mutational status, is usually considered when choosing the best therapeutic option in mCRC [87]. It is well known that left and right colon cancers have a different embryological origin. Left colon develops from the hindgut, receiving irrigation from the inferior mesenteric artery, while the right colon develops from the midgut and is irrigated from the superior mesenteric artery. Right and left colon cancers have different molecular backgrounds and distinct clinical behaviors. Indeed, compared to left sided CRC, right colon cancer has a higher frequency of the BRAF, KRAS, PIK3CA mutations, more commonly harbors MSI-H, and more frequently shows mucinous differentiation. On the other hand, left colon cancer has a higher EREG expression, and more commonly shows18q loss, 20q gain, and EGFR and HER2 gains [88]. This particular molecular profile confers right colon cancer a worse prognosis and response to therapy. Left colon cancer usually has a less aggressive evolution and better prognosis. Furthermore, right colon cancer shows less benefit from anti-EGFR agents even in RAS and BRAF wild-type tumors [89,90]. Therefore, anti-VEGFR agents are the biologicals of choice to be combined with chemotherapy in right-sided mCRC independently of RAS and BRAF status and in left-sided mCRC with RAS or BRAF mutations, while anti-EGFR agents are the treatment of choice in the left-sided RAS/BRAF wildtype mCRC [91-93].

Table 1. Most relevant trials of first-line therapy in biomarker-selected populations

Trial	Phase	Treatment	Target	mPFS	mOS
CRYSTAL	III	FOLFIRI+Cetuximab	EGFR	10.9 m	25.1 m
		VS.		8.8 m	21.6 m
		FOLFIRI		P=0.0549	P=0.0016
PRIME	III	FOLFOX+Panitumumab	EGFR	6.9 m	18.7 m
		VS.		5.5 m	15.4 m
		FOLFOX		P=0.006	P=0.15
PEAK	II	FOLFOX+Panitumumab	EGFR	13 m	41.3 m
		VS.		10.1 m	28.9 m
		FOLFOX+Bevacizumab		P=0.0075	P=0.058
FIRE-3	III	FOLFIRI+Cetuximab	EGFR	10 m	33.1 m
		vs.		10.3 m	25 m
		FOLFIRI+Bevacizumab		P=0.77	P=0.0059
AVF-2107	III	FOLFIRI+Bevacizumab	VEGFR	10.6 m	20.3 m
		VS.		6.2 m	15.6 m
		FOLFIRI		P<0.0001	P<0.001
NO16966	III	FOLFOX+Bevacizumab	VEGFR	9.4 m	21.2 m
		Vs.		8.0 m	19.9 m
		FOLFOX		P=0.023	P=0.07
CHECKMATE-142	II	Ipililumab/Nivolumab	PD-1/CTLA-4	NR	NR
KEYNOTE-177	III	Pembrolizumab	PD-1	54 m	NR
		VS.		24.9 m	36.7 m
		Chemotherapy		P<0.002	

EGFR: Epidermal growth factor receptor, VEGFR: Vascular endothelial growth factor receptor, PD-1: Programmed cell death protein-1, CTLA-4: Cytotoxic T-lymphocyte antigen 4, NR: Not reached, mPFS: Median progression free survival, mOS: Median overall survival. vs.: Versus

5. The New Weaponry: Immunotherapy in MSI-H CRC

The terminology regarding MSI is not homogeneous. However, MSI is commonly described as a hyper-mutable phenotype, resulting from a defective DNA mismatch repair (MMR) system [94]. The MMR system is responsible for correcting errors in DNA replication. Mutations in MMR genes lead to the accumulation of mutations favoring malignant transformation. Therefore, MSI-H tumors are associated with the production and accumulation of hundreds of somatic mutations, which lead to a high neoantigen exposure that favor the initiation of a robust antitumor immune response [22,23]. Response to immunotherapy has been studied in this particular population in recent years.

As when playing chess, and as depicted in Figure 1, immunotherapy has recently been added to the existing weaponry to combat mCRC.

The Keynote 028 and Keynote 164 trials demonstrated the efficacy of pembrolizumab, an anti-PD1 agent, in patients with heavily pretreated MSI-H mCRC. Furthermore, a whole exome sequencing study within Keynote 028, found that patients with DNA mismatch repair had a much higher mutational load than patients without DNA repair deficiency (1782 vs. 73, P=0.007). Checkmate-142, a multi-cohort phase II trial, evaluated the ORR with nivolumab, another anti PD-1, in heavily pretreated patients with MSI-H mCRC. ORR was 31.1% (95% CI 20.8-42.9) while the median duration of response (DOR) was not reached. Treatment with nivolumab achieved a 1-year PFS of 50.4% and a 1-year OS of 73.4%. Another cohort from Checkmate-142 evaluated the addition of ipilimumab, an anti CTLA-4 agent. The combination of ipilimumab/nivolumab in heavily pretreated mCRC achieved an ORR of 55% (95% CI, 45.2-63.8), and a 1-year OS and PFS of 71% and 85%, respectively; and the combination showed a favorable impact in quality of life, with grade 3-4 adverse events occuring in 32% of patients [95-99].

Another cohort from Checkmate-142 evaluated the combination of nivolumab/ipilimumab in the first line setting. ORR achieved 60% (95% CI, 49-78%) with a non-reached median DOR and a 1-year PFS, and 1-year OS of 77% and 83%, respectively [100]. Updated results after a median followup of 29 months, reported a 69% ORR and a 2-year PFS and OS of 74% and 79%, respectively, while median DOR, PFS and OS had not been reached yet. Of note, only 7% of patients developed grade 3-4 adverse events with this regimen, and the nivolumab/ipilimumab combination was finally FDA-approved as first-line therapy of MSI-H mCRC in July 2018 [101]. More recently, the Keynote-177 study evaluated pembrolizumab versus chemotherapy in the first-line MSI-H mCRC. Median PFS was twice longer with pembrolizumab than with chemotherapy (16.5 months vs. 8.2 months, HR=0.6 [95% CI 0.5-0.80], P<0.002). ORR was also higher with pembrolizumab (43% vs. 33%, P=0.275) and median DOR had not been reached in the immunotherapy arm. Toxicity with pembrolizumab was easily manageable. Notably, at 24-months follow-up, 48% of patients in the pembrolizumab arm remained free of disease progression compared to 19% in the chemotherapy arm [102]. Keynote-177 was considered a practice-changing trial and pembrolizumab was added to the therapeutic options for MSI-H mCRC (Figure 2),



Figure 1. Checkmating the king with the knight and bishop is one of the most complicated chess moves. The first monoclonal antibody approved by the FDA for the treatment of mCRC was bevacizumab in 2004. Subsequently, cetuximab and panitumumab joined the fight, with their corresponding approvals in 2009 and 2014, respectively. The recently FDA-approved pembrolizumab and nivolumab/ipilimumab add to the present weaponry against mCRC. VEGFR: Vascular endothelial growing factor receptor, EGFR: Epidermal growth factor receptor, mCRC: Metastatic colorectal cancer.

being FDA-approved for first-line mCRC in June 2020. In addition, after 36 months of follow-up, the pembrolizumab arm achieved a mPFS of 54 months, compared to 24.9 months in the chemotherapy arm. The 3-year PFS rate reached 60% in the pembrolizumab arm compared to 39% in the chemotherapy arm (HR=0.61, 0.44-0.83). ORR was also higher with pembrolizumab (45.1%) compared to chemotherapy (33.1%).

On the other hand, the anti PD-1/anti CTLA-4 combination, as already mentioned, achieved a much higher ORR (69%), although a significant benefit in OS is unclear yet. 3-year OS rate was higher with pembrolizumab compared with the chemotherapy arm, although without statistical significance (61% versus 50%, HR=0.74, 0.53-1.03). mOS was not reached with pembrolizumab, while patients in the chemotherapy arm achieved a 36.7 months mOS [103]. Further follow-up to determine the benefit in OS of the pembrolizumab arm is still needed.

6. Potential New Weapons: Future Perspectives in the First-line Treatment

All future potential treatment options are summarized in Table 2.

6.1. Targeting HER2 overexpression/amplification

HER2 overexpression/amplification is found in 1.3-6.3% of patients with CRC, especially those with *RAS* and *BRAF* wild-type left-sided tumors [104-108]. HER2 overexpression is most commonly analyzed by immunohistochemistry (IHC), while its amplification is usually determined by fluorescent *in situ* hybridization (FISH) [109]. Unlike other neoplasms, criteria for positivity have not yet been standardized in CRC. Recently, an international collaborative project established as criteria for HER2 positivity in CRC an IHC score of 3+ or 2+ associated with a FISH HER2/CEP17 ratio \geq 2.0 in >10% of tumor cells [110].

The HER2 oncogene is located in the 17q21 chromosome and encodes a transmembrane receptor tyrosine kinase. HER2 is a member of the human epidermoid receptor family that includes the EGFR (HER1), HER2, HER3, and HER4 receptors [111]. HER2 has no known ligand but can form heterodimers with EGFR, HER3, and sometimes HER4. Following dimerization, the intracellular tyrosine residues autophosphorylate and subsequently trigger a cascade of multiple important signaling pathways including *RAS*/ RAF/MEK/ERK, PI3K/AKT/mTOR, tyrosine Src kinase, and STAT pathways. Since HER2 overexpression activates almost constitutively, part of the downstream signaling that is shared by EGFR, this explains the resistance to anti-EGFR agents of HER2 positive mCRC [112-115].

Reports indicate that patients with with RAS/BRAF wild-type, HER2 amplified, mCRC have shorter PFS and OS than those without HER2 amplification. Results of anti-HER2 therapies in mCRC have been contradictory [116,117]. The HERACLES-A study found benefit (ORR 29.6%) with dual anti-HER2 blockade with trastuzumab and lapatinib in a cohort of patients with HER2-positive heavily pretreated RAS wild-type mCRC [118]. However, the HERACLES-B study did not find a positive impact on survival (PFS 4.1 months) or response rate (ORR 9.7%) with the combination of pertuzumab and T-DM1 in a similar population [119]. Treatment with trastuzumab and pertuzumab achieved a 32% ORR in patients with HER2-amplified mCRC enrolled within the MyPAthway program [120]. Recently, the DESTINY-CRC01 trial reported benefit with trastuzumab deruxtecan in pretreated patients with HER2 positive (IHC 3+ or IHC 2+/ISH+) mCRC. ORR was 45.3%, mPFS 6.9 months and mOS 15.5 months [121]. Siena et al. [122] coherently suggest the possible incorporation of anti-HER2 agents as firstline therapy in a near future, although stronger evidence is still needed. Trastuzumab and new anti-HER agents, such as pyrotinib and zanidatamab, are being currently studied in first-line clinical trials (NCT00003995, NCT03929666, NCT04380012, NCT03043313, NCT03365882).

6.2. Targeting PI3K pathway

PI3K is a key component of the PI3K/AKT1/MTOR pathway with an important role in CRC pathogenesis [123,124]. Gainof-function mutations in *PIK3CA* (PI3K catalytic subunit alpha gene) activate the p110a enzyme, the key catalytic subunit of



Figure 2. Molecular status and primary tumor sidedness are relevant predictive factors in mCRC. Pembrolizumab and Ipilimumab/nivolumab showed benefit in patients with MSI-H mCRC. Patients with the left sided wild-type *RAS/BRAF* mCRC are the most benefited with Cetuximab and Panitumumab (Anti-EGFR agents). If patients harboring any mutation (*NRAS, KRAS* or *BRAF*) and/or with a right-sided mCRC, bevacizumab (anti-VEGFR agent) is the best biological companion. CT: Chemotherapy, mCRC: Metastatic colorectal cancer, VEGFR: Vascular endothelial growing factor receptor, EGFR: Epidermal growth factor receptor, MSI-H: Microsatellite instability high.

Table 2. Randomized trials with new targeted therapies in mCRC

Target pathway	Trial	Phase	Treatment	Endpoint	Other results
HER2 Overexpression/Amplification	HERACLES-A	II	Trastuzumab/lapatinib	RR=29.6%	mPFS=21 w, mOS=46 w
	HERACLES-B	II	Pertuzumab/TDM-1	RR=9.7%	mPFS 4.1 m
	MyPAthway	IIa	Trastuzumab/Pertuzumab	RR=32%	mPFS=5.3 m, mPFS=14m
	DESTINY-CRC01	II	trastuzumab deruxtecan	RR=45.3%	mPFS=6.9 m, mOS=15.5 m
Targeting PI3K pathway	Rosen et al.	Ι	Apitolisib/CT	RR=3/30 pts	Safety=AEs in>20%
	Yang et al.	Ib	Buparlisib/FOLFOX	Safety	-
	Coleman et al.	Ι	Sapanisertib/Metformin	RR=0/2 pts	Safety
Loss of PTEN	Jansen et al.	Ι	Decitabine	Safety	-
	Garrido-Laguna et al.	I/II	Decitabine/Panitumumab	Safety	-
Targeting HGF/MET pathway	Van Cutsem et al.	I/II	Rilo or Ganitumab/Pani	RR=31%/22%	mPFS=5.2/13.8 m, mOS=5.3/10.6
	Bendell et al.	II	Onartuzumab/CT	Safety	m
	Eng et al.	I/II	Tivantinib/Cetu/CT	mPFS=8.3 m	mDOR=6.4 m
					Safety

mCRC: Metastatic colorectal cancer, HER2: Human epidermal growth factor receptor-2, RR: Response rate, mPFS: Median progression free survival, mOS: Median overall survival, mDOR: Median duration of response, CT: Chemotherapy, Rilo: Rilotumumab, PTEN: Phosphatase and tension homolog, HGF: Hepatocyte growth factor, MET: mesenchymal-epithelial transition

PI3K, stimulating the AKT-MTOR pathway and resulting in cancer growth and proliferation [125]. Mutations in the helicase and kinase domains of exons 9 and 20 of *PIK3CA* occur in 10-20% of CRC and are associated with other molecular alterations such as *BRAF* and *KRAS* mutations and high-grade CpG island methylator phenotype. *PIK3CA* mutation frequency increases from the rectum to the proximal colon and its prognostic value is controversial [126-131]. A number of studies indicate that anti-EGFR agents show no benefit in survival in patients with exon 20 *PIK3CA* mutations regardless of *RAS* and *BRAF* status [132,133].

Some PI3K inhibitors have been developed and evaluated in phase I trials. Copanlisib prevents the growth of malignant cells through the induction of apoptosis via protein p53 upregulated modulator of apoptosis (PUMA) [134]. A phase I study demonstrated a manageable safety profile and a 40% disease control rate (DCR) with copanlisib in a cohort of patients with solid tumors, including mCRC [135]. Dactolisib, a dual PI3K

and mTOR inhibitor, binds to the ATP-binding cleft of PI3K and mTOR kinase, inhibiting their catalytic activities [136]. Dactolisib effectively inhibits the growth of human colon cancer cells (SW480) by targeting the PI3K/mTOR signaling pathway and inducing apoptosis [137]. Another phase Ib dose-escalation study evaluated apitolisib, another PI3K inhibitor, in combination with capecitabine (Arm A: 19 patients) or mFOLFOX6 + bevacizumab (Arm B: 11 patients) in advanced solid tumors, including CRC. Partial response was observed in only one mCRC patient with mutations in PIK3CA and KRAS. Further evaluation in the CRC expansion cohort, found that 2 additional patients achieved partial responses. In general, treatment was well tolerated; the most common grade 3 or higher adverse event was hyperglycemia (40%), followed by stomatitis, hypophosphatemia and neutropenia [138]. Another phase I trial evaluating the safety of PI3K inhibitors in patients with advanced solid tumors, reported that buparlisib in combination with mFOLFOX6 significantly increased toxicity compared to buparlisib or mFOLFOX6 alone, and therefore buparlisib is not being developed further in CRC [139]. However, PI3K inhibitors are still being studied. In mid-2021, a phase I study evaluated sapanisertib in combination with metformin in patients with mTOR/AKT/PI3K pathway alterations and heavily pre-treated advanced solid malignancies. Thirty patients were included (only 2 patients with mCRC) and PI3KCA was the most common genomic alteration (27%). Disease control rate was 60% with the combination, although patients with mCRC were not among responders [140]. The complexity of the PI3K/AKT/mTOR signaling network involves numerous feedback loops and extensive crosstalk nodes with other signaling pathways and compensatory pathways, and therefore, unfortunately intrinsic and acquired resistance currently limits the therapeutic efficacy of PI3K inhibitors in mCRC [141]. These agents, alone or in combination, are being studied in the first line setting in ongoing trials (NCT04495621, NCT04753203, NCT02861300, NCT03711058, NCT04753203).

6.3. Targeting phosphatase and tension homolog (PTEN) loss

PTEN is a multifunctional suppressor protein of the PI3K/AKT pathway [142]. This protein dephosphorylates PI3K products by counteracting the PI3K/AKT signaling cascade. PTEN controls cell proliferation, promotes apoptosis, regulates cell migration/ adhesion and the formation of new vasculature [143,144]. Loss of *PTEN* results in the development of cancer due to the activation of the PI3K/AKT pathway and is found in 20-40% of patients with mCRC [145]. *PTEN* alterations seem to be more frequently correlated with right-sided tumors, MSI, *BRAF* mutations, lymph node metastases, and a higher tumor stage [146]. Loss of *PTEN* may be associated with resistance to anti-EGFR treatment, but clinical studies have shown conflicting results [147].

In a recent review, Salvatore et al. [146], discuss potential ways of targeting PTEN in CRC. Potentiating PTEN transcription by removing an epigenetic block or modifying the exposure to activating or inhibitory transcription factors is a means of increasing PTEN function [148]. Decitabine, a DNA methyltransferase inhibitor, significantly decreased cell proliferation, induced apoptosis and cell cycle arrest of a colon carcinoma cell line in vitro [149]. The safety of decitabine through hepatic arterial infusion was investigated in patients with unresectable liver metastases from solid tumors in a dose escalation phase I clinical trial. Decitabine was administered at 3 different dose levels as a 1-h hepatic arterial infusion in 9 patients (4 with mCRC). Decitabine infusion was safe, with grade 1-2 hematological toxicity being the most frequent treatmentrelated adverse event with no treatment-limiting adverse events. However, there were no objective tumor responses [150]. DNA methyltransferase inhibitors remove methyl groups from DNA, causing the demethylation of DNA. The combination of decitabine and panitumumab was well tolerated and showed activity in KRAS wild-type mCRC patients previously treated with cetuximab in a phase I/II trial [151]. Some of the transcription factors can be pharmacologically stimulated: PPARy (via rosiglitazone),

EGR-1 (via irradiation), and NFAT (through butyrate, a fatty acid produced by colonic microbiota fermentation) [152]. At the post-transcriptional level, PTEN expression can be impaired by microRNAs (miRNAs) or RNA-binding protein (RBP). miRNAs bind mRNAs causing loss of PTEN expression and activation of the PI3K/AKT signaling cascade. Modulation of those regulatory RNAs and RNA-RBPs represent a therapeutic strategy aiming at restoring PTEN translation and expression, exploiting its antitumor activity, and increasing cellular drug sensitivity [153]. Some PTEN isoforms originating from different start codon translations have been identified. Of those, PTEN-L was shown to counteract the PI3K/AKT pathway, leading to cell death, both in vitro and in vivo [154]. Finally, post-translational modifications at specific aminoacidic residues can directly modulate PTEN catalytic or binding activity subsequently impacting on PTEN function [155]. Reverting those post-translational modifications or targeting the enzymes involved could be effective at restoring PTEN function in PTEN positive neoplasms [156]. The long noncoding RNA Linc02023 specifically binds to PTEN and blocks its ubiquitination, promoting CRC cell proliferation and survival. Thus, Linc02023 may serve as a novel therapeutic target for restoring PTEN tumor suppressor activity [157].

6.4. Targeting the hepatocyte growth factor (HGF)/ mesenchymal-epithelial transition (MET) pathway

The HGF and the tyrosine kinase receptor known as MET factor play an important role in proliferation, survival, metastasis, and acquired resistance to cancer treatment [158]. HGF is produced primarily by mesenchymal tissue and is the only known ligand for MET. Patients with CRC have an elevated serum HGF at diagnosis. MET is a member of the transmembrane surface receptor family expressed on endothelial cells and both normal and malignant epithelial cells [87]. Tissue and serum expression of HGF and elevated levels of MET protein and mRNA associate with a poor prognosis in CRC [159,160]. MET mutations and amplifications represent, respectively, 2-5% and 0.5-2% of all mutations in CRC. Overexpression of HGF/MET mRNA and HGF/MET protein occur in 70% and 50% of CRC tissue samples, respectively. HGFinduced translocation of metastasis-associated in colon cancer 1 from plasma to nucleus and its binding to the MET promoter initiates transcription in the MET pathway [161]. The activation of MET signaling starts with the binding of HGF to the MET receptor at the cell membrane level, triggering the formation of a multifunctional intracellular coupling site from two tyrosine residues that bind to subsequent substrates. Activation of the HGF/ MET pathway initiates signaling pathways, including MAPK/ ERK, PI3K/AKT, and STAT/JAK, the nuclear factor kB complex, regulates hematopoiesis, and promotes organ regeneration and wound healing [162]. Subsequent studies supported the theory that MET over-activation promotes HGF transcription and expression, leading to subsequent MET activation and expression in a loop manner that can be increased via paracrine HGF produced by reactive stromal cells in the tumor microenvironment or in situations such as hypoxia or inflammation [163].

Unfortunately, clinical trials with HGF and MET inhibitors have shown negative results. A randomized phase I/II trial evaluating panitumumab in combination with rilotumumab, ganitumab, or placebo in patients with KRAS-wild type mCRC reported ORRs of 31%, 22%, and 21% respectively, while mPFS and mOS were 5.2 and 13.8 months, 5.3 and 10.6 months, and 3.7 and 11.6 months, respectively. Exploratory biomarker analyzes, including MET and IGF-related protein expression, failed to demonstrate a clear predictive value [164]. A phase II trial of onartuzumab combined with mFOLFOX-6 and bevacizumab did not improve survival in previously untreated MET IHC-positive mCRC and MET expression by IHC was not predictive of response [165]. Another phase I/II trial investigated the addition of the oral MET inhibitor, tivantinib to cetuximab/irinotecan (CETIRI). The combination of tivantinib and CETIRI was well tolerated but did not significantly improve PFS in previously treated KRAS-wild type mCRC [166]. Finally, in a phase II study enrolling patients with MET-high-amplified, *KRAS* wild-type mCRC, treated with ≥ 1 prior systemic therapy and showing tumor progression on cetuximab or panitumumab within 3 months before enrollment, treatment with tivantinib plus cetuximab showed only modest results in ORR and PFS [167]. Future trials will evaluate the role of HGF and MET inhibitors in mCRC (NCT03592641, NCT02205398, NCT04515394).

7. Conclusions

mCRC harbors molecular alterations that besides being prognostic, also allow physicians to make the most adequate treatment decisions for each patient. *RAS* and *BRAF* mutational status and MSI are at present mandatory determinations in all newly-diagnosed advanced CRCs to choose between anti-EGFR, anti-VEGF, and immune checkpoint inhibitory agents. However, other more recently described alterations such as those in HER2, PIK3, PTEN, and others, have been shown to be targetable and constitute promising therapeutic options in the first-line setting. It is, therefore, of utmost importance that physicians are aware of the rapidly-evolving molecular biology and therapeutic advances in advanced CRC to offer the most appropriate and individualized management approaches for these patients.

Conflict of Interest

The authors declare no conflicts of interest.

References

- [1] Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, Bray F. Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. CA Cancer J Clin 2021;71:209-49.
- [2] Haggar FA, Boushey RP. Colorectal Cancer Epidemiology: Incidence, Mortality, Survival, and Risk Factors. Clin Colon Rectal Surg 2009;22:191-7.
- [3] Bogaert J, Prenen H. Molecular Genetics of Colorectal Cancer. Ann Gastroenterol 2014;27:9-14.

- [4] Morano F, Corallo S, Lonardi S, Raimondi A, Cremolini C, Rimassa L, et al. Negative Hyperselection of Patients With RAS and BRAF Wild-Type Metastatic Colorectal Cancer Who Received Panitumumab-Based Maintenance Therapy. J Clin Oncol 2019;37:3099-110.
- [5] Huang Y, Liu N, Liu J, Liu Y, Zhang C, Long S, Luo G, Zhang L, Zhang Y. Mutant p53 Drives Cancer Chemotherapy Resistance Due to Loss of Function on Activating Transcription of PUMA. Cell Cycle 2019;18:3442-55.
- [6] Goetz LH, Schork NJ. Personalized Medicine: Motivation, Challenges, and Progress. Fertil Steril 2018;109:952-63.
- [7] Arteaga CL, Engelman JA. ERBB Receptors: From Oncogene Discovery to Basic Science to Mechanism-Based Cancer Therapeutics. Cancer Cell 2014;25:282-303.
- [8] Tebbutt N, Pedersen MW, Johns TG. Targeting the ERBB Family in Cancer: Couples Therapy. Nat Rev Cancer 2013;13:663-73.
- [9] Hsu JL, Hung MC. The Role of HER2, EGFR, and other Receptor Tyrosine Kinases in Breast Cancer. Cancer Metastasis Rev 2016;35:575-88.
- [10] Rotow J, Bivona TG. Understanding and Targeting Resistance Mechanisms in NSCLC. Nat Rev Cancer 2017;17:637-58.
- [11] Roskoski R Jr. The ErbB/HER Family of Protein-tyrosine Kinases and Cancer. Pharmacol Res 2014;79:34-74.
- [12] Ishibashi K, Fukumoto Y, Hasegawa H, Abe K, Kubota S, Aoyama K, et al. Nuclear ErbB4 Signaling through H3K9me3 is Antagonized by EGFR-Activated c-Src. J Cell Sci 2013;126 Pt 2:625-37.
- [13] Downward J. Targeting RAS Signalling Pathways in Cancer Therapy. Nat Rev Cancer 2003;3:11-22.
- [14] Ray S. The Cell: A Molecular Approach. Yale J Biol Med 2014;87:603-4.
- [15] Buday L, Downward J. Epidermal Growth Factor Regulates p21ras through the Formation of a Complex of Receptor, Grb2 Adapter Protein, and Sos Nucleotide Exchange Factor. Cell 1993;73:611-20.
- [16] Roskoski R Jr. Small Molecule Inhibitors Targeting the EGFR/ErbB Family of Protein-tyrosine Kinases in Human Cancers. Pharmacol Res 2019;139:395-411.
- [17] Vecchione L, Jacobs B, Normanno N, Ciardiello F, Tejpar S. EGFR-targeted Therapy. Exp Cell Res 2011;317:2765-71.
- [18] Wang Z. ErbB Receptors and Cancer. Methods Mol Biol 2017;1652:3-35.
- [19] Pylayeva-Gupta Y, Grabocka E, Bar-Sagi D. RAS Oncogenes: Weaving a Tumorigenic Web. Nat Rev Cancer 2011;11:761-74.
- [20] Giehl K. Oncogenic Ras in Tumour Progression and Metastasis. Biol Chem 2005;386:193-205.
- [21] Lowy DR, Willumsen BM. Function and Regulation of Ras. Annu Rev Biochem 1993;62:851-91.

- [22] Martini G, Dienstmann R, Ros J, Baraibar I, Cuadra-Urteaga JL, Salva F, et al. Molecular Subtypes and the Evolution of Treatment Management in Metastatic Colorectal Cancer. Ther Adv Med Oncol 2020;12:1758835920936089.
- [23] Ünlü M, Uzun E, Bengi G, Sağol Ö, Sarıoğlu S. Molecular Characteristics of Colorectal Hyperplastic Polyp Subgroups. Turk J Gastroenterol 2020;31:573-80.
- [24] Fearon ER, Vogelstein B. A Genetic Model for Colorectal Tumorigenesis. Cell 1990;61:759-67.
- [25] Li W, Qiu T, Zhi W, Shi S, Zou S, Ling Y, et al. Colorectal Carcinomas with KRAS Codon 12 Mutation are Associated with more Advanced Tumor Stages. BMC Cancer 2015;15:340.
- [26] Goldberg RM. Cetuximab. Nat Rev Drug Discov 2005;Suppl: S10-1.
- [27] Irahara N, Baba Y, Nosho K, Shima K, Yan L, Dias-Santagata D, et al. NRAS Mutations are Rare in Colorectal Cancer. Diagn Mol Pathol 2010;19:157-63.
- [28] Johnson L, Greenbaum D, Cichowski K, Mercer K, Murphy E, Schmitt E, *et al.* K-ras is an Essential Gene in the Mouse with Partial Functional Overlap with N-ras. Genes Dev 1997;11:2468-81.
- [29] Malumbres M, Barbacid M. RAS Oncogenes: The First 30 Years. Nat Rev Cancer 2003;3:459-65.
- [30] Brand TM, Wheeler DL. KRAS Mutant Colorectal Tumors: Past and Present. Small GTPases 2012;3:34-9.
- [31] Jancík S, Drábek J, Radzioch D, Hajdúch M. Clinical Relevance of KRAS in Human Cancers. J Biomed Biotechnol 2010;2010:150960.
- [32] Karnoub AE, Weinberg RA. Ras Oncogenes: Split Personalities. Nat Rev Mol Cell Biol 2008;9:517-31.
- [33] Shields JM, Pruitt K, McFall A, Shaub A, Der CJ. Understanding Ras: "It ain't Over til it's Over". Trends Cell Biol 2000;10:147-54.
- [34] Pruitt K, Der CJ. Ras and Rho Regulation of the Cell Cycle and Oncogenesis. Cancer Lett 2001;171:1-10.
- [35] Jänicke RU, Sohn D, Schulze-Osthoff K. The Dark Side of a Tumor Suppressor: Anti-Apoptotic p53. Cell Death Differ 2008;15:959-76.
- [36] Maughan TS, Adams RA, Smith CG, Meade AM, Seymour MT, Wilson RH, *et al.* Addition of Cetuximab to Oxaliplatin-based First-line Combination Chemotherapy for Treatment of Advanced Colorectal Cancer: Results of the Randomised Phase 3 MRC COIN trial. Lancet 2011;377:2103-14.
- [37] Esteller M, González S, Risques RA, Marcuello E, Mangues R, Germà JR, *et al.* K-ras and p16 Aberrations Confer Poor Prognosis in Human Colorectal Cancer. J Clin Oncol 2001;19:299-304.
- [38] Westra JL, Schaapveld M, Hollema H, de Boer JP, Kraak MM, de Jong D, *et al.* Determination of TP53

Mutation is more Relevant than Microsatellite Instability Status for the Prediction of Disease-free Survival in Adjuvant-treated Stage III Colon Cancer Patients. J Clin Oncol 2005;23:5635-43.

- [39] Siddiqui AD, Piperdi B. KRAS Mutation in Colon Cancer: A Marker of Resistance to EGFR-I Therapy. Ann Surg Oncol 2010;17:1168-76.
- [40] Tran B, Kopetz S, Tie J, Gibbs P, Jiang ZQ, Lieu CH, et al. Impact of BRAF Mutation and Microsatellite Instability on the Pattern of Metastatic Spread and Prognosis in Metastatic Colorectal Cancer. Cancer 2011;117:4623-32.
- [41] Tveit KM, Guren T, Glimelius B, Pfeiffer P, Sorbye H, Pyrhonen S, et al. Phase III Trial of Cetuximab with Continuous or Intermittent Fluorouracil, Leucovorin, and Oxaliplatin (Nordic FLOX) Versus FLOX Alone in First-line Treatment of Metastatic Colorectal Cancer: The NORDIC-VII Study. J Clin Oncol 2012;30:1755-62.
- [42] Issa JP. CpG Island Methylator Phenotype in Cancer. Nat Rev Cancer 2004;4:988-93.
- [43] Toyota M, Ahuja N, Ohe-Toyota M, Herman JG, Baylin SB, Issa JP. CpG Island Methylator Phenotype in Colorectal Cancer. Proc Natl Acad Sci U S A 1999;96:8681-6.
- [44] Weisenberger DJ, Siegmund KD, Campan M, Young J, Long TI, Faasse MA, et al. CpG Island Methylator Phenotype Underlies Sporadic Microsatellite Instability and is Tightly Associated with BRAF Mutation in Colorectal Cancer. Nat Genet 2006;38:787-93.
- [45] Tol J, Nagtegaal ID, Punt CJ. BRAF Mutation in Metastatic Colorectal Cancer. N Engl J Med 2009;361:98-9.
- [46] Davies H, Bignell GR, Cox C, Stephens P, Edkins S, Clegg S, et al. Mutations of the BRAF Gene in Human Cancer. Nature 2002;417:949-54.
- [47] Roberts PJ, Der CJ. Targeting the Raf-MEK-ERK Mitogenactivated Protein Kinase Cascade for the Treatment of Cancer. Oncogene 2007;26:3291-310.
- [48] Rajagopalan H, Bardelli A, Lengauer C, Kinzler KW, Vogelstein B, Velculescu VE. Tumorigenesis: RAF/ RAS Oncogenes and Mismatch-repair Status. Nature 2002;418:934.
- [49] Saharinen P, Eklund L, Pulkki K, Bono P, Alitalo K. VEGF and Angiopoietin Signaling in Tumor Angiogenesis and Metastasis. Trends Mol Med 2011;17:347-62.
- [50] Goel HL, Mercurio AM. VEGF Targets the Tumour Cell. Nat Rev Cancer 2013;13:871-82.
- [51] Ferrara N, Gerber HP, LeCouter J. The Biology of VEGF and its Receptors. Nat Med 2003;9:669-76.
- [52] Falcon BL, Chintharlapalli S, Uhlik MT, Pytowski B. Antagonist Antibodies to Vascular Endothelial Growth Factor Receptor 2 (VEGFR-2) as Anti-angiogenic Agents. Pharmacol Ther 2016;164:204-25.
- [53] Karaman S, Leppänen VM, Alitalo K. Vascular Endothelial Growth Factor Signaling in Development and Disease.

Development 2018;145:dev151019.

- [54] Koch S, Claesson-Welsh L. Signal Transduction by Vascular Endothelial Growth Factor Receptors. Cold Spring Harb Perspect Med 2012;2:a006502.
- [55] Peng K, Bai Y, Zhu Q, Hu B, Xu Y. Targeting VEGFneuropilin Interactions: A Promising Antitumor Strategy. Drug Discov Today 2019;24:656-64.
- [56] Simons M, Gordon E, Claesson-Welsh L. Mechanisms and Regulation of Endothelial VEGF Receptor Signalling. Nat Rev Mol Cell Biol 2016;17:611-25.
- [57] Takahashi H, Shibuya M. The Vascular Endothelial Growth Factor (VEGF)/VEGF Receptor System and its Role under Physiological and Pathological Conditions. Clin Sci (Lond) 2005;109:227-41.
- [58] Shibuya M. Vascular Endothelial Growth Factor (VEGF) and its Receptor (VEGFR) Signaling in Angiogenesis: A Crucial Target for Anti-and Pro-Angiogenic Therapies. Genes Cancer 2011;2:1097-105.
- [59] Cébe-Suarez S, Zehnder-Fjällman A, Ballmer-Hofer K. The Role of VEGF Receptors in Angiogenesis; Complex Partnerships. Cell Mol Life Sci 2006;63:601-15.
- [60] Tarnawski AS, Ahluwalia A, Jones MK. Angiogenesis in Gastric Mucosa: An Important Component of Gastric Erosion and Ulcer Healing and its Impairment in Aging. J Gastroenterol Hepatol 2014;29 Suppl 4:112-23.
- [61] Fransén K, Klintenäs M, Osterström A, Dimberg J, Monstein HJ, Söderkvist P. Mutation Analysis of the BRAF, ARAF and RAF-1 Genes in Human Colorectal Adenocarcinomas. Carcinogenesis 2004;25:527-33.
- [62] Aparicio J, Esposito F, Serrano S, Falco E, Escudero P, Ruiz-Casado A, *et al.* Metastatic Colorectal Cancer. First Line Therapy for Unresectable Disease. J Clin Med 2020;9:3889.
- [63] Marques RP, Duarte GS, Sterrantino C, Pais HL, Quintela A, Martins AP, et al. Triplet (FOLFOXIRI) Versus Doublet (FOLFOX or FOLFIRI) Backbone Chemotherapy as First-line Treatment of Metastatic Colorectal Cancer: A Systematic Review and Meta-analysis. Crit Rev Oncol Hematol 2017;118:54-62.
- [64] Jindal V, Gupta R, Sahu KK, Rahi MS, Stender MJ, Jaiyesimi IA. Doublet (FOLFOX or FOLFIRI) Versus Triplet (FOLFOXIRI) Backbone Chemotherapy Regimen as First-line Treatment of Metastatic Colorectal Cancer: A Meta-analysis and Systematic Review. J Clin Oncol 2021;39 Suppl 15:3593-3.
- [65] Bokemeyer C, Bondarenko I, Hartmann JT, de Braud F, Schuch G, Zubel A, *et al.* Efficacy According to Biomarker Status of Cetuximab Plus FOLFOX-4 as First-line Treatment for Metastatic Colorectal Cancer: The OPUS Study. Ann Oncol 2011;22:1535-46.
- [66] Bokemeyer C, Van Cutsem E, Rougier P, Ciardiello F, Heeger S, Schlichting M, *et al.* Addition of Cetuximab to

Chemotherapy as First-line Treatment for KRAS Wildtype Metastatic Colorectal Cancer: Pooled Analysis of the CRYSTAL and OPUS Randomised Clinical Trials. Eur J Cancer 2012;48:1466-75.

- [67] Stintzing S, Modest DP, Rossius L, Lerch MM, von Weikersthal LF, Decker T, et al. FOLFIRI Plus Cetuximab Versus FOLFIRI Plus Bevacizumab for Metastatic Colorectal Cancer (FIRE-3): A Post-hoc Analysis of Tumour Dynamics in the Final RAS Wild-type Subgroup of this Randomised Open-label Phase 3 Trial. Lancet Oncol 2016;17:1426-34.
- [68] Folprecht G, Gruenberger T, Bechstein W, Raab HR, Weitz J, Lordick F, et al. Survival of Patients with Initially Unresectable Colorectal Liver Metastases Treated with FOLFOX/cetuximab or FOLFIRI/cetuximab in a Multidisciplinary Concept (CELIM Study). Ann Oncol 2014;25:1018-25.
- [69] Garufi C, Torsello A, Tumolo S, Ettorre GM, Zeuli M, Campanella C, et al. Cetuximab Plus Chronomodulated Irinotecan, 5-Fluorouracil, Leucovorin and Oxaliplatin as Neoadjuvant Chemotherapy in Colorectal Liver Metastases: POCHER Trial. Br J Cancer 2010;103:1542-7.
- [70] Ohori H, Yamaguchi T, Matsuura M, Nishioka A, Makiyama A, Noura S, *et al.* The RANDOMIZED Phase II Study of FOLFOXIRI Plus Cetuximab versus FOLFOXIRI plus Bevacizumabas the First-Line Treatment in Metastatic Colorectal Cancer with RAS Wild-Type Tumors: The Deeper Trial (JACRRO CC-13). ASCO Annual Meeting; 2021.
- [71] Douillard JY, Siena S, Cassidy J, Tabernero J, Burkes R, Barugel M, et al. Final Results from PRIME: Randomized Phase III Study of Panitumumab with FOLFOX4 for Firstline Treatment of Metastatic Colorectal Cancer. Ann Oncol 2014;25:1346-55.
- [72] Rivera F, Karthaus M, Hecht JR, Sevilla I, Forget F, Fasola G, et al. Final Analysis of the Randomised PEAK Trial: Overall Survival and Tumour Responses during Firstline Treatment with mFOLFOX6 Plus Either Panitumumab or Bevacizumab in Patients with Metastatic Colorectal Carcinoma. Int J Colorectal Dis 2017;32:1179-90.
- [73] Kurreck A, Geissler M, Martens UM, Riera-Knorrenschild J, Greeve J, Florschütz A, et al. Dynamics in Treatment Response and Disease Progression of Metastatic Colorectal Cancer (mCRC) Patients with Focus on BRAF Status and Primary Tumor Location: Analysis of Untreated RAS-wild-type mCRC Patients Receiving FOLFOXIRI either with or without Panitumumab in the VOLFI Trial (AIO KRK0109). J Cancer Res Clin Oncol 2020;146:2681-91.
- [74] Hurwitz HI, Tebbutt NC, Kabbinavar F, Giantonio BJ, Guan ZZ, Mitchell L, et al. Efficacy and Safety of Bevacizumab in Metastatic Colorectal Cancer: Pooled Analysis from Seven Randomized Controlled Trials.

Oncologist 2013;18:1004-12.

- [75] Cassidy J, Clarke S, Díaz-Rubio E, Scheithauer W, Figer A, Wong R, et al. XELOX vs FOLFOX-4 as Firstline Therapy for Metastatic Colorectal Cancer: NO16966 Updated Results. Br J Cancer 2011;105:58-64.
- [76] Cunningham D, Lang I, Marcuello E, Lorusso V, Ocvirk J, Shin DB, et al. Bevacizumab Plus Capecitabine Versus Capecitabine Alone in Elderly Patients with Previously Untreated Metastatic Colorectal Cancer (AVEX): An Open-label, Randomised Phase 3 Trial. Lancet Oncol 2013;14:1077-85.
- [77] Welch S, Spithoff K, Rumble RB, Maroun J; Gastrointestinal Cancer Disease Site Group. Bevacizumab Combined with Chemotherapy for Patients with Advanced Colorectal Cancer: A Systematic Review. Ann Oncol 2010;21:1152-62.
- [78] Luo HY, Xu RH. Predictive and Prognostic Biomarkers with Therapeutic Targets in Advanced Colorectal Cancer. World J Gastroenterol 2014;20:3858-74.
- [79] Pathak S, Sushmitha S, Banerjee A, Marotta F, Gopinath M, Murugesan R, et al. Review on Comparative Efficacy of Bevacizumab, Panitumumab and Cetuximab Antibody Therapy with Combination of FOLFOX-4 in KRAS-mutated Colorectal Cancer Patients. Oncotarget 2017;9:7739-48.
- [80] Grothey A, Sugrue MM, Purdie DM, Dong W, Sargent D, Hedrick E, et al. Bevacizumab beyond First Progression is Associated with Prolonged Overall Survival in Metastatic Colorectal Cancer: Results from a Large Observational Cohort Study (BRiTE). J Clin Oncol 2008;26:5326-34.
- [81] Díaz-Rubio E, Gómez-España A, Massutí B, Sastre J, Abad A, Valladares M, et al. First-line XELOX Plus Bevacizumab Followed by XELOX Plus Bevacizumab or Single-agent Bevacizumab as Maintenance Therapy in Patients with Metastatic Colorectal Cancer: The Phase III MACRO TTD Study. Oncologist 2012;17:15-25.
- [82] Simkens LH, van Tinteren H, May A, ten Tije AJ, Creemers GJ, Loosveld OJ, et al. Maintenance Treatment with Capecitabine and Bevacizumab in Metastatic Colorectal Cancer (CAIRO3): A Phase 3 Randomised Controlled Trial of the Dutch Colorectal Cancer Group. Lancet 2015;385:1843-52.
- [83] Bennouna J, Sastre J, Arnold D, Österlund P, Greil R, Van Cutsem E, et al. Continuation of Bevacizumab after First Progression in Metastatic Colorectal Cancer (ML18147): A Randomised Phase 3 Trial. Lancet Oncol 2013;14:29-37.
- [84] Masi G, Salvatore L, Boni L, Loupakis F, Cremolini C, Fornaro L, et al. Continuation or Reintroduction of Bevacizumab beyond Progression to First-line Therapy in Metastatic Colorectal Cancer: Final Results of the Randomized BEBYP Trial. Ann Oncol 2015;26:724-30.
- [85] Tabernero J, Van Cutsem E, Lakomý R, Prausová J, Ruff P, van Hazel GA, et al. Aflibercept Versus Placebo in Combination with Fluorouracil, Leucovorin and Irinotecan

in the Treatment of Previously Treated Metastatic Colorectal Cancer: Prespecified Subgroup Analyses from the VELOUR Trial. Eur J Cancer 2014;50:320-31.

- [86] Tabernero J, Yoshino T, Cohn AL, Obermannova R, Bodoky G, Garcia-Carbonero R, et al. Ramucirumab Versus Placebo in Combination with Second-line FOLFIRI in Patients with Metastatic Colorectal Carcinoma that Progressed during or after First-line Therapy with Bevacizumab, Oxaliplatin, and a Fluoropyrimidine (RAISE): A Randomised, Double-blind, Multicentre, Phase 3 Study. Lancet Oncol 2015;16:499-508.
- [87] Cremolini C, Schirripa M, Antoniotti C, Moretto R, Salvatore L, Masi G, *et al.* First-line Chemotherapy for mCRC a Review and Evidence-based Algorithm. Nat Rev Clin Oncol 2015;12:607-19.
- [88] Missiaglia E, Jacobs B, D'Ario G, Di Narzo AF, Soneson C, Budinska E, et al. Distal and Proximal Colon Cancers Differ in Terms of Molecular, Pathological, and Clinical Features. Ann Oncol 2014;25:1995-2001.
- [89] Loupakis F, Yang D, Yau L, Feng S, Cremolini C, Zhang W, et al. Primary Tumor Location as a Prognostic Factor in Metastatic Colorectal Cancer. J Natl Cancer Inst 2015;107:dju427.
- [90] Arnold D, Lueza B, Douillard JY, Peeters M, Lenz HJ, Venook A, et al. Prognostic and Predictive Value of Primary Tumour Side in Patients with RAS Wild-Type Metastatic Colorectal Cancer Treated with Chemotherapy and EGFR Directed Antibodies in Six Randomized Trials. Ann Oncol 2017;28:1713-29.
- [91] Modest DP, Stintzing S, von Weikersthal LF, Decker T, Kiani A, Vehling-Kaiser U, *et al.* Exploring the Effect of Primary Tumor Sidedness on Therapeutic Efficacy Across Treatment Lines in Patients with Metastatic Colorectal Cancer: Analysis of FIRE-3 (AIOKRK0306). Oncotarget 2017;8:105749-60.
- [92] Rosen LS, Jacobs IA, Burkes RL. Bevacizumab in Colorectal Cancer: Current Role in Treatment and the Potential of Biosimilars. Target Oncol 2017;12:599-610.
- [93] Elez E, Argilés G, Tabernero J. First-Line Treatment of Metastatic Colorectal Cancer: Interpreting FIRE-3, PEAK, and CALGB/SWOG 80405. Curr Treat Options Oncol 2015;16:52.
- [94] Le DT, Uram JN, Wang H, Bartlett BR, Kemberling H, Eyring AD, et al. PD-1 Blockade in Tumors with Mismatch-Repair Deficiency. N Engl J Med 2015;372:2509-20.
- [95] O'Neil BH, Wallmark JM, Lorente D, Elez E, Raimbourg J, Gomez-Roca C, et al. Safety and Antitumor Activity of the Anti-PD-1 Antibody Pembrolizumab in Patients with Advanced Colorectal Carcinoma. PLoS One 2017;12:e0189848.
- [96] Le DT, Durham JN, Smith KN, Wang H, Bartlett BR, Aulakh LK, et al. Mismatch Repair Deficiency Predicts Response of Solid Tumors to PD-1 Blockade. Science

2017;357:409-13.

- [97] Le DT, Kim TW, Van Cutsem E, Geva R, Jäger D, Hara H, et al. Phase II Open-Label Study of Pembrolizumab in Treatment-Refractory, Microsatellite Instability-High/ Mismatch Repair-Deficient Metastatic Colorectal Cancer: KEYNOTE-164. J Clin Oncol 2020;38:11-9.
- [98] Brahmer JR, Tykodi SS, Chow LQ, Hwu WJ, Topalian SL, Hwu P, et al. Safety and Activity of Anti-PD-L1 Antibody in Patients with Advanced Cancer. N Engl J Med 2012;366:2455-65.
- [99] Overman MJ, McDermott R, Leach JL, Lonardi S, Lenz HJ, Morse MA, et al. Nivolumab in Patients with Metastatic DNA Mismatch Repair-deficient or Microsatellite Instability-high Colorectal Cancer (CheckMate 142): An Open-label, Multicentre, Phase 2 Study. Lancet Oncol 2017;18:1182-91.
- [100] Overman MJ, Lonardi S, Wong KY, Lenz HJ, Gelsomino F, Aglietta M, et al. Durable Clinical Benefit with Nivolumab Plus Ipilimumab in DNA Mismatch Repair-Deficient/ Microsatellite Instability-High Metastatic Colorectal Cancer. J Clin Oncol 2018;36:773-9.
- [101] Lenz HJ, Lonardi S, Zagonel V, Van Cutsem E, Limon ML, Wong KY, et al. Nivolumab Plus Low-dose Ipilimumab as First-line Therapy in Microsatellite Instability-high/DNA Mismatch Repair Deficient Metastatic Colorectal Cancer: Clinical Update. J Clin Oncol 2021;38:JCO2101015.
- [102] André T, Shiu KK, Kim TW, Jensen BV, Jensen LH, Punt C, et al. Pembrolizumab in Microsatellite-Instability-High Advanced Colorectal Cancer. N Engl J Med 2020;383:2207-18.
- [103] Andre T, Shiu K, Kim TW, Jensen BV, Jensen LH, Punt C, et al. Final Overall Survival for the Phase 3 KN177 Study: Pembrolizumab Versus Chemotherapy in Microsatellite Instaability-High/Mismatch Repair Deficient (MSI-H/ dMMR) Metastatic Colorectal Cancer (mCRC). J Clin Oncol 2021;39 Suppl 15:3500.
- [104] Bertotti A, Migliardi G, Galimi F, Sassi F, Torti D, Isella C, et al. A Molecularly Annotated Platform of Patientderived Xenografts ("Xenopatients") Identifies HER2 as an Effective Therapeutic Target in Cetuximab-resistant Colorectal Cancer. Cancer Discov 2011;1:508-23.
- [105] Heppner BI, Behrens HM, Balschun K, Haag J, Krüger S, Becker T, et al. HER2/neu Testing in Primary Colorectal Carcinoma. Br J Cancer 2014;111:1977-84.
- [106] Richman SD, Southward K, Chambers P, Cross D, Barrett J, Hemmings G, et al. HER2 Overexpression and Amplification as a Potential Therapeutic Target in Colorectal Cancer: Analysis of 3256 Patients Enrolled in the QUASAR, FOCUS and PICCOLO Colorectal Cancer Trials. J Pathol 2016;238:562-70.
- [107] Sawada K, Nakamura Y, Yamanaka T, Kuboki Y, Yamaguchi D, Yuki S, et al. Prognostic and Predictive Value of HER2 Amplification in Patients with

Metastatic Colorectal Cancer. Clin Colorectal Cancer 2018;17:198-205.

- [108] Cancer Genome Atlas Network. Comprehensive Molecular Characterization of Human Colon and Rectal Cancer. Nature 2012;487:330-7.
- [109] Seo AN, Kwak Y, Kim DW, Kang SB, Choe G, Kim WH, et al. HER2 Status in Colorectal Cancer: Its Clinical Significance and the Relationship between HER2 Gene Amplification and Expression. PLoS One 2014;9:e98528.
- [110] Yeh YM, Lee CH, Chen SH, Lee CT, Chen YL, Lin BW, et al. Comprehensive Assessment of HER2 Alteration in a Colorectal Cancer Cohort: From Next-generation Sequencing to Clinical Significance. Cancer Manag Res 2019;11:7867-75.
- [111] Bosman FT, Delorenzi M, Tejpar S. Distal and Proximal Colon Cancers Differ in Terms of Molecular, Pathological, and Clinical Features. Ann Oncol 2014;25:1995-2001.
- [112] De Cuyper A, Van Den Eynde M, Machiels JP. HER2 as a Predictive Biomarker and Treatment Target in Colorectal Cancer. Clin Colorectal Cancer 2020;19:65-72.
- [113] Greally M, Kelly CM, Cercek A. HER2: An Emerging Target in Colorectal Cancer. Curr Probl Cancer 2018;42:560-71.
- [114] Sergina NV, Moasser MM. The HER Family and Cancer: Emerging Molecular Mechanisms and Therapeutic Targets. Trends Mol Med 2007;13:527-34.
- [115] Moasser MM. The Oncogene HER2: It Signaling and Transforming Functions and its Role in Human Cancer Pathogenesis. Oncogene 2007;26:6469-87.
- [116] Jeong JH, Kim J, Hong YS, Kim D, Kim JE, Kim SY, et al. HER2 Amplification and Cetuximab Efficacy in Patients with Metastatic Colorectal Cancer Harboring Wild-type RAS and BRAF. Clin Colorectal Cancer 2017;16:e147-52.
- [117] Martin V, Landi L, Molinari F, Fountzilas G, Geva R, Riva A, et al. HER2 Gene Copy Number Status may Influence Clinical Efficacy to Anti-EGFR Monoclonal Antibodies in Metastatic Colorectal Cancer Patients. Br J Cancer 2013;108:668-75.
- [118] Sartore-Bianchi A, Trusolino L, Martino C, Bencardino K, Lonardi S, Bergamo F, *et al.* Dual-targeted Therapy with Trastuzumab and Lapatinib in Treatment-refractory, KRAS Codon 12/13 Wild-type, HER2-positive Metastatic Colorectal Cancer (HERACLES): A Proof-of-concept, Multicentre, Open-label, Phase 2 Trial. Lancet Oncol 2016;17:738-46.
- [119] Sartore-Bianchi A, Lonardi S, Martino C, Fenocchio E, Tosi F, Ghezzi S, *et al.* Pertuzumab and Trastuzumab Emtansine in Patients with HER2-amplified Metastatic Colorectal Cancer: The Phase II HERACLES-B Trial. ESMO Open 2020;5:e000911.
- [120] Meric-Bernstam F, Hurwitz H, Raghav KP, McWilliams RR, Fakih M, VanderWalde A, et al.

Pertuzumab Plus Trastuzumab for HER2-amplified Metastatic Colorectal Cancer (MyPathway): An Updated Report from a Multicentre, Open-label, Phase 2a, Multiple Basket Study. Lancet Oncol 2019;20:518-30.

- [121] Yoshino T. Trastuzumab Deruxtecan (T-DXd; DS-8201) in Patients with HER2-expressing Metastatic Colorectal Cancer: Final Results from a Phase 2, Multicenter, Openlabel Study (DESTINY-CRC01). National Cancer Center Hospital East, Kashiwa, Japan June, 2021. ASCO Annual Meeting; 2021.
- [122] Siena S, Sartore-Bianchi A, Marsoni S, Hurwitz HI, McCall SJ, Penault-Llorca F, *et al.* Targeting the Human Epidermal Growth Factor Receptor 2 (HER2) Oncogene in Colorectal Cancer. Ann Oncol 2018;29:1108-19.
- [123] Samuels Y, Diaz LA Jr., Schmidt-Kittler O, Cummins JM, Delong L, Cheong I, *et al.* Mutant PIK3CA Promotes Cell Growth and Invasion of Human Cancer Cells. Cancer Cell 2005;7:561-73.
- [124] Shaw RJ, Cantley LC. Ras, PI(3)K and mTOR Signalling Controls Tumour Cell Growth. Nature 2006;441:424-30.
- [125] Ogino S, Lochhead P, Giovannucci E, Meyerhardt JA, Fuchs CS, Chan AT. Discovery of Colorectal Cancer PIK3CAMutation as Potential Predictive Biomarker: Power and Promise of Molecular Pathological Epidemiology. Oncogene 2014;33:2949-55.
- [126] Liao X, Lochhead P, Nishihara R, Morikawa T, Kuchiba A, Yamauchi M, et al. Aspirin Use, Tumor PIK3CA Mutation, and Colorectal-cancer Survival. N Engl J Med 2012;367:1596-606.
- [127] Zhu K, Yan H, Wang R, Zhu H, Meng X, Xu X, et al. Mutations of KRAS and PIK3CA as Independent Predictors of Distant Metastases in Colorectal Cancer. Med Oncol 2014;31:16.
- [128] He Y, Van't Veer LJ, Mikolajewska-Hanclich I, van Velthuysen ML, Zeestraten EC, Nagtegaal ID, et al. PIK3CA Mutations Predict Local Recurrences in Rectal Cancer Patients. Clin Cancer Res 2009;15:6956-62.
- [129] Liao X, Morikawa T, Lochhead P, Imamura Y, Kuchiba A, Yamauchi M, et al. Prognostic Role of PIK3CA Mutation in Colorectal Cancer: Cohort Study and Literature Review. Clin Cancer Res 2012;18:2257-68.
- [130] Yamauchi M, Morikawa T, Kuchiba A, Imamura Y, Qian ZR, Nishihara R, et al. Assessment of Colorectal Cancer Molecular Features along Bowel Subsites Challenges the Conception of Distinct Dichotomy of Proximal Versus Distal Colorectum. Gut 2012;61:847-54.
- [131] Jin J, Shi Y, Zhang S, Yang S. PIK3CA Mutation and Clinicopathological Features of Colorectal Cancer: A Systematic Review and Meta-Analysis. Acta Oncol 2020;59:66-74.
- [132] Mao C, Yang ZY, Hu XF, Chen Q, Tang JL. PIK3CA Exon 20 Mutations as a Potential Biomarker for Resistance to Anti-EGFR Monoclonal Antibodies in KRAS Wild-

type Metastatic Colorectal Cancer: A Systematic Review and Meta-analysis. Ann Oncol 2012;23:1518-25.

- [133] De Roock W, Claes B, Bernasconi D, De Schutter J, Biesmans B, Fountzilas G, *et al.* Effects of KRAS, BRAF, NRAS, and PIK3CA Mutations on the Efficacy of Cetuximab Plus Chemotherapy in Chemotherapyrefractory Metastatic Colorectal Cancer: A Retrospective Consortium Analysis. Lancet Oncol 2010;11:753-62.
- [134] Yan J, Yang S, Tian H, Zhang Y, Zhao H. Copanlisib Promotes Growth Inhibition and Apoptosis by Modulating the AKT/FoxO3a/PUMA Axis in Colorectal Cancer. Cell Death Dis 2020;11:943.
- [135] Doi T, Fuse N, Yoshino T, Kojima T, Bando H, Miyamoto H, et al. A Phase I Study of Intravenous PI3K Inhibitor Copanlisib in Japanese Patients with Advanced or Refractory Solid Tumors. Cancer Chemother Pharmacol 2017;79:89-98.
- [136] Haddadi N, Lin Y, Travis G, Simpson AM, Nassif NT, McGowan EM. PTEN/PTENP1: "Regulating the Regulator of RTK-dependent PI3K/Akt Signalling", New Targets for Cancer Therapy. Mol Cancer 2018;17:37.
- [137] Furnari FB, Huang HJ, Cavenee WK. The Phosphoinositol Phosphatase Activity of PTEN Mediates a Serumsensitive G1 Growth Arrest in Glioma Cells. Cancer Res 1998;58:5002-8.
- [138] Rosen L, Goldman J, Hubbard JM, Roos M, Capdevila J, Maynes J, et al. 382 Phase Ib Study of Oral Dual-PI3K/ mTOR Inhibitor GDC-0980 in Combination with Capecitabine and mFOLFOX6 + Bevacizumab in Patients with Advanced Solid Tumors and Colorectal Cancer. Eur J Cancer 2014;50:122-3.
- [139] McRee AJ, Sanoff HK, Carlson C, Ivanova A, O'Neil BH. A Phase I Trial of mFOLFOX6 Combined with the Oral PI3K Inhibitor BKM120 in Patients with Advanced Refractory Solid Tumors. Invest New Drugs 2015;33:1225-31.
- [140] Coleman N, Naing A, Zhang S, Piha-Paul SA, Tsimberidou AM, Janku F, et al. Phase I Study of mTORC1/2 Inhibitor Sapanisertib (TAK-228) in Combination with Metformin in Patients (PTS) with mTOR/AKT/PI3K Pathway Alterations and Advanced Solid Malignancies. ASCO Annual Meeting; 2021.
- [141] Yang J, Nie J, Ma X, Wei Y, Peng Y, Wei X. Targeting PI3K in Cancer: Mechanisms and Advances in Clinical Trials. Mol Cancer 2019;18:26.
- [142] Kotelevets L, van Hengel J, Bruyneel E, Mareel M, van Roy F, Chastre E. The Lipid Phosphatase Activity of PTEN is Critical for Stabilizing Intercellular Junctions and Reverting Invasiveness. J Cell Biol 2001;155:1129-35.
- [143] Szado T, Vanderheyden V, Parys JB, De Smedt H, Rietdorf K, Kotelevets L, *et al.* Phosphorylation of Inositol 1,4,5-Trisphosphate Receptors by Protein Kinase B/Akt Inhibits Ca²⁺ Release and Apoptosis. Proc Natl Acad Sci U S A 2008;105:2427-32.

- [144] Siena S, Sartore-Bianchi A, Di Nicolantonio F, Balfour J, Bardelli A. Biomarkers Predicting Clinical Outcome of Epidermal Growth Factor Receptor-targeted Therapy in Metastatic Colorectal Cancer. J Natl Cancer Inst 2009;101:1308-24.
- [145] Laurent-Puig P, Cayre A, Manceau G, Buc E, Bachet JB, Lecomte T, et al. Analysis of PTEN, BRAF, and EGFR Status in Determining Benefit from Cetuximab Therapy in Wild-type KRAS Metastatic Colon Cancer. J Clin Oncol 2009;27:5924-30.
- [146] Salvatore L, Calegari MA, Loupakis F, Fassan M, Di Stefano B, Bensi M, *et al.* PTEN in Colorectal Cancer: Shedding Light on Its Role as Predictor and Target. Cancers (Basel) 2019;11:1765.
- [147] Yang X, Niu B, Wang L, Chen M, Kang X, Wang L, et al. Autophagy Inhibition Enhances Colorectal Cancer Apoptosis Induced by Dual Phosphatidylinositol 3-Kinase/ Mammalian Target of Rapamycin Inhibitor NVP-BEZ235. Oncol Lett 2016;12:102-6.
- [148] Kotelevets L, Scott MGH, Chastre E. Targeting PTEN in Colorectal Cancers. Adv Exp Med Biol 2018;1110:55-73.
- [149] Abou Najem S, Khawaja G, Hodroj MH, Rizk S. Synergistic Effect of Epigenetic Inhibitors Decitabine and Suberoylanilide Hydroxamic Acid on Colorectal Cancer In Vitro. Curr Mol Pharmacol 2019;12:281-300.
- [150] Jansen YJ, Verset G, Schats K, Van Dam PJ, Seremet T, Kockx M, et al. Phase I Clinical Trial of Decitabine (5-aza-2'-deoxycytidine) Administered by Hepatic Arterial Infusion in Patients with Unresectable Liver-predominant Metastases. ESMO Open 2019;4:e000464.
- [151] Garrido-Laguna I, McGregor KA, Wade M, Weis J, Gilcrease W, Burr L, et al. A Phase I/II Study of Decitabine in Combination with Panitumumab in Patients with Wildtype (wt) KRAS Metastatic Colorectal Cancer. Invest New Drugs 2013;31:1257-64.
- [152] Xue Q, Sun K, Deng HJ, Lei ST, Dong JQ, Li GX. AntimiRNA-221 Sensitizes Human Colorectal Carcinoma Cells to Radiation by Upregulating PTEN. World J Gastroenterol 2013;19:9307-17.
- [153] Hopkins BD, Fine B, Steinbach N, Dendy M, Rapp Z, Shaw J, et al. A Secreted PTEN Phosphatase that Enters Cells to Alter Signaling and Survival. Science 2013;341:399-402.
- [154] Dillon LM, Miller TW. Therapeutic Targeting of Cancers with Loss of PTEN Function. Curr Drug Targets 2014;15:65-79.
- [155] Wang Q, Feng Y, Peng W, Ji D, Zhang Z, Qian W, et al. Long Noncoding RNA Linc02023 Regulates PTEN Stability and Suppresses Tumorigenesis of Colorectal Cancer in a PTEN-dependent Pathway. Cancer Lett 2019;451:68-78.
- [156] Wang Q, Yang S, Wang K, Sun SY. MET Inhibitors for

Targeted Therapy of EGFR TKI-resistant Lung Cancer. J Hematol Oncol 2019;12:63.

- [157] Matsumoto K, Umitsu M, De Silva DM, Roy A, Bottaro DP. Hepatocyte Growth Factor/MET in Cancer Progression and Biomarker Discovery. Cancer Sci 2017;108:296-307.
- [158] Gao H, Guan M, Sun Z, Bai C. High c-Met Expression is a Negative Prognostic Marker for Colorectal Cancer: A Meta-analysis. Tumour Biol 2015;36:515-20.
- [159] Stein U, Walther W, Arlt F, Schwabe H, Smith J, Fichtner I, et al. MACC1, a Newly Identified Key Regulator of HGF-MET Signaling, Predicts Colon Cancer Metastasis. Nat Med 2009;15:59-67.
- [160] Mo HN, Liu P. Targeting MET in Cancer Therapy. Chronic Dis Transl Med 2017;3:148-53.
- [161] Bigatto V, De Bacco F, Casanova E, Reato G, Lanzetti L, Isella C, *et al.* TNF-α Promotes Invasive Growth through the MET Signaling Pathway. Mol Oncol 2015;9:377-88.
- [162] Ye M, Hu D, Tu L, Zhou X, Lu F, Wen B, et al. Involvement of PI3K/Akt Signaling Pathway in Hepatocyte Growth Factor-induced Migration of Uveal Melanoma Cells. Invest Ophthalmol Vis Sci 2008;49:497-504.
- [163] Ide T, Kitajima Y, Miyoshi A, Ohtsuka T, Mitsuno M, Ohtaka K, et al. The Hypoxic Environment in Tumorstromal Cells Accelerates Pancreatic Cancer Progression Via the Activation of Paracrine Hepatocyte Growth Factor/ c-Met Signaling. Ann Surg Oncol 2007;14:2600-7.
- [164] Van Cutsem E, Eng C, Nowara E, Swieboda-Sadlej A, Tebbutt NC, Mitchell E, *et al.* Randomized Phase Ib/II Trial of Rilotumumab or Ganitumab with Panitumumab Versus Panitumumab Alone in Patients with Wildtype KRAS Metastatic Colorectal Cancer. Clin Cancer Res 2014;20:4240-50.
- [165] Bendell JC, Hochster H, Hart LL, Firdaus I, Mace JR, McFarlane JJ, et al. A Phase II Randomized Trial (GO27827) of First-Line FOLFOX Plus Bevacizumab with or without the MET Inhibitor Onartuzumab in Patients with Metastatic Colorectal Cancer. Oncologist 2017;22:264-71.
- [166] Eng C, Bessudo A, Hart LL, Severtsev A, Gladkov O, Müller L, et al. A Randomized, Placebo-controlled, Phase 1/2 Study of Tivantinib (ARQ 197) in Combination with Irinotecan and Cetuximab in Patients with Metastatic Colorectal Cancer with Wild-type KRAS who have Received First-line Systemic Therapy. Int J Cancer 2016;139:177-86.
- [167] Rimassa L, Bozzarelli S, Pietrantonio F, Cordio S, Lonardi S, Toppo L, *et al.* Phase II Study of Tivantinib and Cetuximab in Patients with KRAS Wild-type Metastatic Colorectal Cancer with Acquired Resistance to EGFR Inhibitors and Emergence of MET Overexpression: Lesson Learned for Future Trials With EGFR/MET Dual Inhibition. Clin Colorectal Cancer 2019;18:125-32.e2.