CASE REPORT

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A possible case of renal oxalate deposit reported in an African fruit bat (*Epomops franqueti*)

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ABSTRACT

We report a possible spontaneous case of oxalate nephrosis in an African fruit bat (*Epomops franqueti*), incidentally observed in Ibadan, South-West Nigeria, in an anatomical and serological survey of the species. Wild caught bats underwent sedation, intracardial perfusion, necropsy and histopathology. All 15 wild-caught African fruit bats were apparently healthy. However, light microscopy revealed mild oligofocal tubulonephrosis with intraluminal deposition of polarizing crystals interpreted as subclinical oxalate nephrosis in one case. In summary, we suggest a dietary aetiology, based on seasonal availability of high ascorbic acid or oxalate containing fruits. However, exposure to anthropogenic contaminants cannot be completely ruled out.

ARTICLE HISTORY

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1. Introduction

Renal dysfunctions have hardly been documented in fruit bats. Renal toxicosis has been reported after topical administration of Ivermectin in dog-faced fruit bats, Cynopterus brachyotis [1]. Another case of renal involvement was in an experimental hendra virus infection in pregnant guinea-pigs and fruit bats (Pteropus poliocephalus) [2]. Oxalate nephrosis is a renal dysfunction characterized by the deposition of calcium oxalate crystals in the renal tubules with pale renal corticomedullary streaks grossly. This condition arises from the imbalance between intratubular calcium oxalate deposition and the body's excretory capacity [3,4]. They may be a result of genetic defects in glyoxylate metabolism causing increased hepatic endogenous production of oxalate [3,5]. Endogenous deposition of oxalate also occurs following the normal degradation of glycine, an important constituent amino acid of collagen, elastin, hydroxyproline and serine [6] or increased absorption due to low intestinal calcium content [7]. Oxalate can also be produced endogenously as a metabolic by-product of ascorbic acid catabolism [3,4,6].

Various instances of primary oxalate nephrosis have been reported in humans [8–10]. Moreover, accidental ethylene glycol toxicity in humans, experimental exposure of domestic animals [11–13], as well as chronic renal disease [14,15] may cause these pathologic changes. A high percentage of Koala populations in parts of Australia have also been reported to come down with idiopathic oxalate nephrosis [16] and recently, oxalate-related renal disease was suggested as a potential cause of acute renal failure in a young cheetah [17].

This paper presents an incidental histological finding of a possible spontaneous renal oxalate deposition in an African fruit bat (*Epomops franqueti*), one of the samples caught in South-West Nigeria, in an anatomical and serological survey of the species.

2. Material and methods

A total of 15 African fruit bats (Epomops franqueti) were caught in University of Ibadan for an ongoing histological and serological study. Ethical approval was obtained by University of Ibadan - Animal Care and Use Research Ethics Committee (UI-ACUREC /App/2016/015). Bats were anaesthesized with ketamine (90 mg/kg) and xylazine (10 mg/kg) based on the work of Wirawati et al. [18] and then intracardially perfused with 10% neutral buffered formalin (NBF). Organ samples (including kidney, heart, brain, liver, spleen, pancreas, gastrointestinal tract, lungs blood vessels and lymph nodes) were fixed in 10% NBF and then embedded in paraffin, sectioned at 5 µm, and stained with Haematoxylin and Eosin based on routine method of Roulet [19] and Romeis [20] for light microscopy. Polarized light was used for confirmation of the oxalate crystals using a Zeiss Axio Scope.

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A1 and differential interference contrast illumination (Carl Zeiss Microscopy GmbH, Jena, Germany).

3. Histopathological findings

Only one of the 15 observed African fruit bats displayed mild, oligofocal tubular degeneration with intraluminal yellow/orange, translucent, fanshaped to irregular crystals (Figure 1(a)). Furthermore, polarized light microscopy revealed birefringence of the intraluminal crystals (Figure 1(b))

4. Discussion

This to the best of our knowledge is the first report of renal tubular degeneration with luminal birefringent crystal deposit suggestive of oxalate nephrosis in fruit bats in the tropics. There is a strong possibility that the condition has dietary origin, based on seasonal availability of fruits with high oxalate

content or its substrate, ascorbic acid. Oxalate nephrosis is a sequelae of precipitation of calcium oxalate crystals in the renal tubules. Ascorbic acid catabolism is one of the endogenous sources of oxalate deposition [3, 4, and 7]. Ascorbic acid intake through dietary sources of fruits and vegetables by wild primates and frugivorous bats in a tropical area was shown to be much greater than that of most human populations [21]. Consumption of plant-based diets and products containing high amounts of ascorbic acid have been discussed as the cause of oxalate nephrosis of herbivores and some other omnivores [22-25]. In an ecological study, of the same site studied, the wild fruits of heart-leaved fig (Ficus polita) and sponge gourd (Luffa cylindrica) were retrieved from captured fruit bats [26]. Moreover, sites of citrus species and watermelon (Citrullus lanatus) cultivation also exist within the vicinity. However, exposure to anthropogenic contaminants such as glycol, which is commonly used as antifreeze in



Figure 1. (a) Bright field light microscopy of H&E section of the kidney of male adult *Epomops franqueti* showing intraluminal yellow/orange, translucent, variably shaped crystals (arrows) (Bar: 50 μm). (b) Polarized light microscopy of the same section showing rhomboid and highly refractive crystals (arrows), without a definite arrangement in the clusters (Bar: 50 μm).

the cooling systems of combustion engines, cannot be ruled out.

5. Conclusion

We observed a mild, oligofocal tubulonephrosis with crystal deposition in an African fruit bat, suggestive of a mild oxalate nephrosis. Further studies are needed to get more knowledge in this regard.

Disclosure statement

No potential conflict of interest was reported by the authors.

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References

- DeMarco JH, Heard DJ, Fleming GJ, et al. Ivermectin toxicosis after topical administration in dog-faced fruit bats (Cynopterus brachyotis) [J]. J Zoo Wildl Med. 2002;33(2):147–150.
- [2] Williamson MM, Hooper PT, and Selleck PW, et al. Experimental hendra virus infection in pregnant guinea-pigs and fruit bats (Pteropus poliocephalus)
 [J]. J Comp Pathol. 2000;122(2–3):201–207.
- [3] Bhasin B, Urekli HM, Atta MG, et al. Primary and secondary hyperoxaluria: understanding the enigma[J]. World J Nephrol. 2015;4(2):235–244.
- [4] Cianciolo RE, Mohr FC. Urinary system. In: Maxie M, editor. Jubb, Kennedy and Palmers pathology of domestic animals. 6th ed. St Louis, MO: Elsevier, Inc; 2016. p. 425–426.
- [5] Chaplin AJ. Histopathological occurrence and characterisation of calcium oxalate: a review [J]. J Clin Pathol. 1977;30(9):800-811.
- [6] Matson E, Reginato AM. Crystalline disorders associated with renal disease including oxalate arthropathy. In Terkeltaub: Gout & Crystal-Induced Arthropathies. Elsevier, Saunders; 2012. p. 282–292.
- [7] Holmes RP, Goodman HO, Assimos DG, et al. Contribution of dietary oxalate to urinary oxalate excretion [J]. Kidney Int. 2001;59(1):270–276.
- [8] Greer KE, Cooper PH, Campbell F, et al. Primary oxalosis with livedo reticularis [J]. Arch Dermatol. 1980;116(2):213-214.

- [9] Broyer M, Brunner FP, Brynger H, et al. Kidney transplantation in primary oxalosis: data from the EDTA Registry [J]. Nephrol Dialysis Transplantation. 1990;5(5):332–336.
- [10] Harambat J, van Stralen KJ, Espinosa L, et al. Characteristics and outcomes of children with primary oxalosis requiring renal replacement therapy [J]. Clin J Am Soc Nephrol. 2012;7(3):458–465.
- [11] Egbert PA, Abraham K. Ethylene glycol intoxication: pathophysiology, diagnosis, and emergency management [J]. Nephrol Nurs J. 1999;26(3):295.
- [12] Frape DL. Ethylene glycol toxicity [J]. Equine Vet Educ. 2002;14(5):238–239.
- [13] Schweighauser A, Francey T. Ethylene glycol poisoning in three dogs: importance of early diagnosis and role of hemodialysis as a treatment option [J]. Schweiz Arch Tierheilkd. 2016;158(2):109–114.
- [14] Morgan SH, Maher ER, Purkiss P, et al. Oxalate metabolism in end-stage renal disease: the effect of ascorbic acid and pyridoxine [J]. Nephrol Dialysis Transplantation. 1988;3(1):28-32.
- [15] Brown CA, Jeong KS, Poppenga RH, et al. Outbreaks of renal failure associated with melamine and cyanuric acid in dogs and cats in 2004 and 2007 [J]. J Vet Diagn Invest. 2007;19(5):525–531.
- [16] Speight KN, Boardman W, Breed WG, et al. Pathological features of oxalate nephrosis in a population of koalas (Phascolarctos cinereus) in South Australia [J]. Vet Pathol. 2013;50(2):299–307.
- [17] Mitchell EP, Church ME, Nemser SM, et al. Pathology and epidemiology of oxalate nephrosis in cheetahs [J]. Vet Pathol. 2017;54(6):977–985.
- [18] Wirawati V, Widiati NDA, Gunawan G, et al. The distribution of serotonergic nerve on the hippocampus of the fruit bats (Rousettus amplexicaudatus) [J]. Vet World. 2019;12(9):1460–1466.
- [19] Roulet F. Methoden der pathologischen Histologie. Wien: Springer Verlag; 1948.
- [20] Romeis B. Mikroskopische Technik. 17. Aufl.Verl. München-Wien-Baltimore: Urban u. Schwarzenberg; 1989.
- [21] Milton K, Jenness R. Ascorbic acid content of neotropical plant parts available to wild monkeys and bats [J]. Experientia. 1987;43(3):339–342.
- [22] Waltner-Toews D, Meadows DH. Urolithiasis in a herd of beef cattle associated with oxalate ingestion[J]. Can Vet J. 1980;21(2):61.
- [23] Ellis TM, Copland MD, Gaynor WT, et al. Oxalate toxicity in a scaly-tailed possum, a Patagonian cavy and a swamp wallaby [J]. J Wildl Dis. 1983;19 (3):290-293.
- [24] Botha CJ, Truter M, Bredell T, et al. Putative Aspergillus niger-induced oxalate nephrosis in sheep: clinical communication [J]. J S Afr Vet Assoc. 2009;80(1):50-53.
- [25] Vanselow BA, Pines MK, Bruhl JJ, et al. Oxalate nephropathy in a laboratory colony of common marmoset monkeys (Callithrix jacchus) following the ingestion of Eucalyptus viminalis [J]. Vet Record-English Edition. 2011;169(4):100.
- [26] Adeyanju TE, Adeyanju AT. Species richness and diets of bats from two sites in South-West, Nigeria[J]. Ethiopian J Environ Stud Manage. 2018;11(2):215 230.