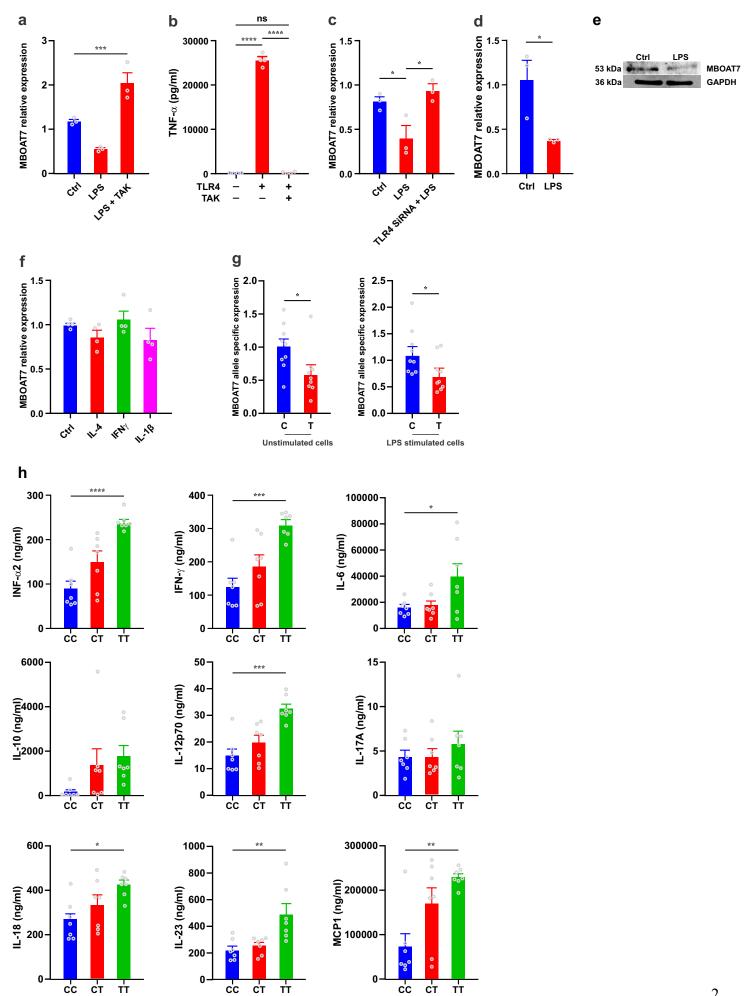
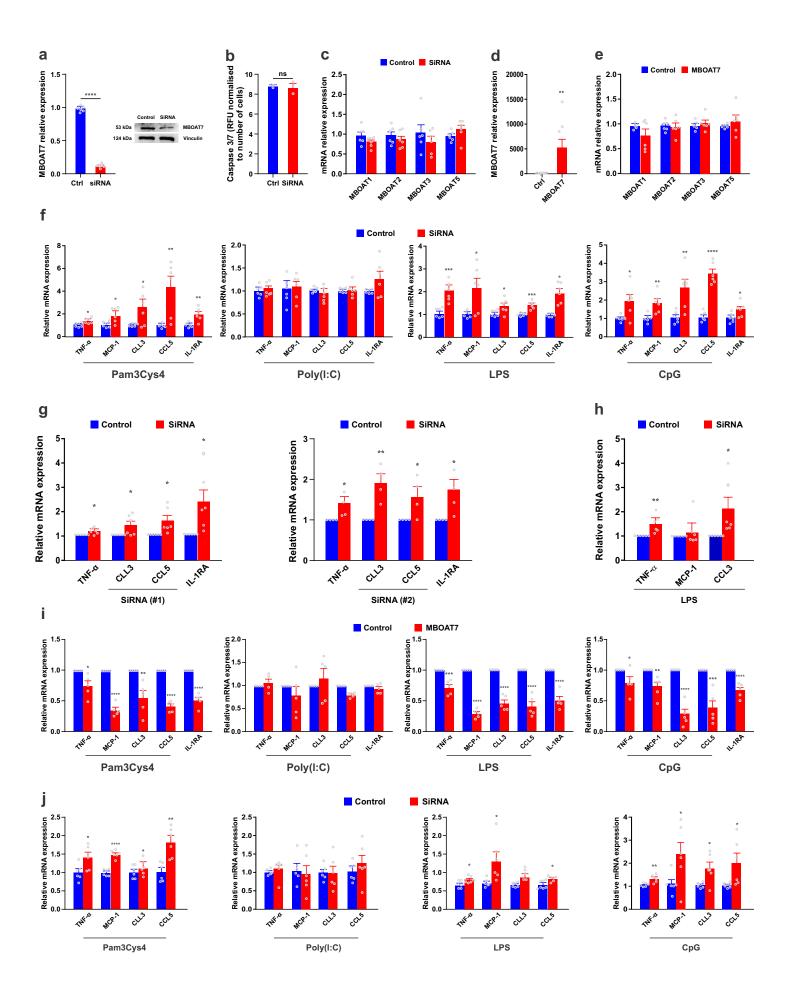
A metabolic associated fatty liver disease risk variant in MBOAT7 regulates toll like receptor induced outcomes

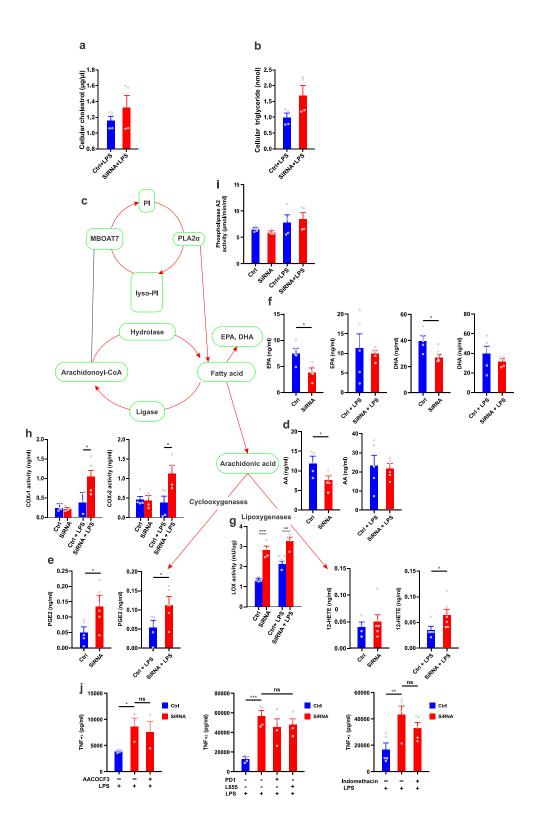
Jawaher Alharthi, Ali Bayoumi, Khaled Thabet, Ziyan Pan, Brian S. Gloss, Olivier Latchoumanin, Mischa Lundberg, Natalie A. Twine, Duncan McLeod, Shafi Alenizi, Leon A. Adams, Martin Weltman, Thomas Berg, Christopher Liddle, Jacob George, Mohammed Eslam.



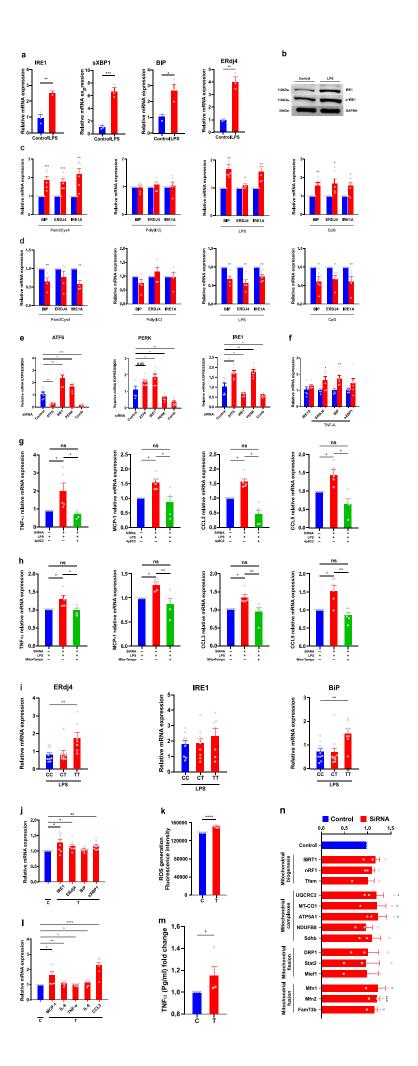
Supplementary Figure 1: a) Human MDMs from subjects heterozygous for rs8736 (n = 3) were stimulated with lipopolysaccharide (LPS) (TLR4) and / or a specific TLR4 inhibitor (TAK-242) for 24 hours and mRNA expression of MBOAT7 was analyzed by PCR and normalized to GAPDH in response to LPS stimulation. b) TNF-α levels in conditioned media measured by ELISA. c) Human MDMs from subjects heterozygous for rs8736 (n = 3) were stimulated with lipopolysaccharide (LPS) (TLR4) and / or a specific TLR4 SiRNA for 24 hours and mRNA expression of MBOAT7 was analyzed by PCR and normalized to GAPDH in response to LPS stimulation. d) Human hepatic Kupffer cells were treated with LPS for 24 hours and then examined for MBOAT7 relative mRNA expression by PCR and normalized to GAPDH, e) MBOAT7 protein expression by Western blot in MDMs. f) Human MDMs from subjects heterozygous for rs8736 were stimulated with IFN-γ, IL-4 or IL-1β for 24 hours and MBOAT7 mRNA expression was assessed by PCR and normalized to untreated MDMs and GAPDH. g) Allele-specific expression assay showing the ratio of rs8736 alleles in genomic DNA (gDNA) and complementary DNA (cDNA; synthesized from pre-mRNA) from MDMs of heterozygous individuals either unstimulated stimulated with LPS for 24 hours (n = 9). h) IFN-α2, IFN-γ, IL-6, IL-10, IL-12p70, IL-17A, IL-18, IL-23, and MCP-1 production by MDMs stratified by rs8736 genotype following 24 hours stimulation with Pam3Cys4 [TLR1/2] (n = 7 per genotype). P-value < 0.04 for all comparisons. Values are represented by vertical bars and are mean \pm sem; *P < 0.05, ** P < 0.01, ****P < 0.001. Statistical differences between groups were assessed by 2-tailed Student's t test or one-way ANOVA; multiple comparisons were corrected by Bonferroni correction. Source data are provided as a Source Data file.



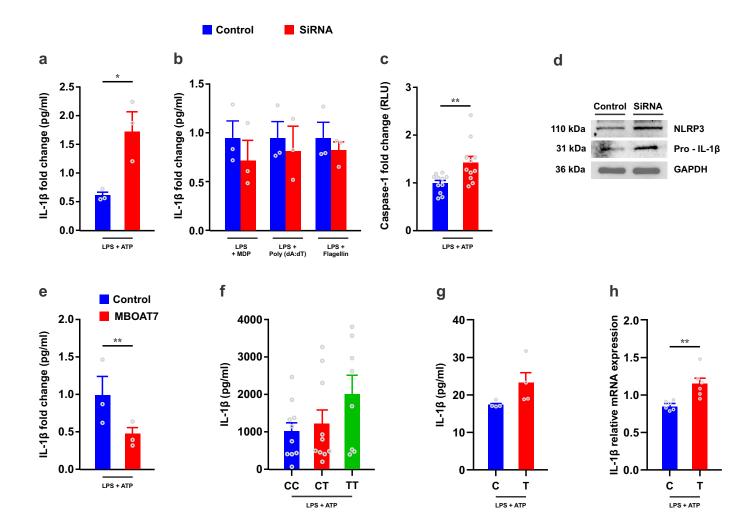
Supplementary Figure 2: a) Human MDMs were transfected with scrambled or MBOAT7 siRNA for 48 hours and MBOAT7 relative mRNA expression was assessed by RT-PCR and normalized to GAPDH and MBOAT7 protein by Western normalised to α-tubulin. b) Transfected cells were assessed for cell viability using Caspase 3/7 activity; shown is RFU normalised to number of cells (n=3). c) The expression of the MBOAT family were determined by RT-PCR and normalized to GAPDH. Human MDMs were transfected with control empty vector or an MBOAT7 vector for 48 hours and d) MBOAT7 relative mRNA expression was assessed by RT-PCR (n=4). e) The expression of MBOAT family members were determined by RT-PCR. f) Human MDMs were transfected with MBOAT7 siRNA or scramble as control and treated with the indicated TLRs ligands (Pam3Cys4 [TLR1/2], Poly(I:C) [TLR3], LPS [TLR4] and CpG [TLR9]) for 24 hours. Shown is relative mRNA expression of different cytokines assessed by RT-PCR and normalized to GAPDH. g) Relative mRNA expression of different cytokines in MDMs transfected with 2 additional MBOAT7 siRNAs or scramble as control and treated with LPS for 24 hours. h) Human Kupffer cells were transfected with MBOAT7 siRNA or scramble as control and treated with LPS for 24 hours. Shown is relative mRNA expression of different cytokines by RT-PCR. i) MDMs were transfected with MBOAT7 overexpression plasmid or the empty vector as control and treated with the indicated TLRs ligands for 24 hours. Shown is relative mRNA expression of different cytokines assessed by RT-PCR and normalized to GAPDH (n=3). i) Human MDMs were cultured with GM-CSF and transfected with MBOAT7 siRNA or scramble and treated with the indicated TLR ligands for 24 hours. Shown is relative mRNA expression of different cytokines assessed by RT-PCR and normalized to GAPDH. (n = 6 includes 3 biological replicates and 2 technical replicates in a, c, e, f, g, h and j). P-value < 0.04 for all comparisons. Values are represented by vertical bars and are mean \pm sem; *P < 0.05, ** P < 0.01, ****P < 0.001. Statistical differences between groups were assessed by 2-tailed Student's t test. Source data are provided as a Source Data file.



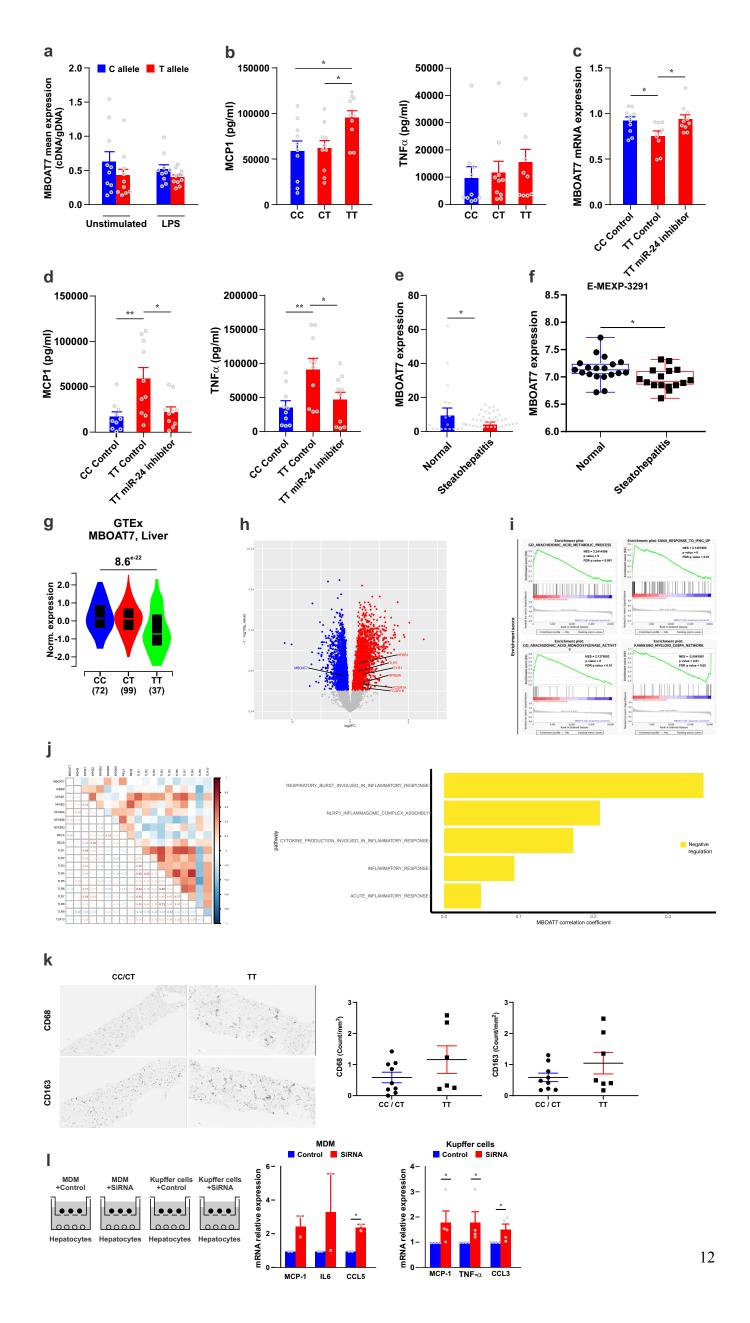
Supplementary Figure 3: MDMs transfected with MBOAT7 siRNA or scramble at base line and challenged with LPS for 24 h and **a**) cholesterol and **b**) triglycerides were measured (n = 4 includes 2 biological replicates from independent individuals and 2 technical replicates). **c**) Scheme illustrating the role of MBOAT7 in the Land's cycle pathway. MDMs were transfected with MBOAT7 siRNA or scramble as control (n=6); **d**) levels of Arachidonic Acid, **e**) products of the cyclooxygenase (PGE2) and lipoxygenase pathway (12-HETE), **f**) pro-resolving n-3 fatty acids eicosapentaenoic acid (EPA) and docosahexaenoic acid (DHA), **g**) LOX enzyme activity (n=4), **h**) COX-I and COX-II enzyme activity (n=4), and **i**) Phospholipase A2 enzyme activity(n=4). **j**) Effect of inhibition of key enzymes in the eicosanoids production pathway on TNF-α secretion assessed by ELISA in conditional media of MDMs (n=3-5). P-value < 0.04 for all comparisons. Values are represented by vertical bars and are mean ± sem; *P<0.05, ** P<0.01, ****P<0.001. Statistical differences between groups were assessed by 2-tailed Student's t test. Source data are provided as a Source Data file.



Supplementary Figure 4: MDMs were treated with LPS or control for 24h (n=3). Shown is a) Relative mRNA expression of different ER stress markers assessed by RT-PCR and b) IRE1 and p-IRE1 protein expression by Western blot. Relative mRNA expression of ER stress markers by RT-PCR in MDMs transfected with c) MBOAT7 siRNA or scramble or d) MBOAT7 overexpression plasmid or the empty vector after 24h of treatment with the indicated TLRs ligands (n = 5-6; 3 biological replicates and 1-2 technical replicates). e) Relative mRNA expression of ATF6, PERK and IRE1 by RT-PCR in MDMs transfected with scramble, ATF6 siRNA, IRE1 siRNA, PERK siRNA or combination of the three siRNAs for 48h and treated with LPS for 24h. f) Relative mRNA expression of different ER stress markers by RT-PCR in MDMs treated with TNF- α (n = 6; 3 biological replicates and 2 technical replicates). Relative mRNA expression of different cytokines by RT-PCR in MDMs transfected with scramble or MBOAT7 siRNA for 48h and treated with LPS only or in combination with g) 4µ8C (IREA1 inhibitor) or with **h)** Mito-TEMPO (mitochondria-targeted antioxidant) for 24h (n = 5; 3 biological replicates and 1-2 technical replicates). i) Relative mRNA expression of ER stress markers in MDMs from subjects of all genotypes for rs8736 and challenged with LPS for 24h (n=7-10/genotype). HeLa cells were transfected with full-length 3'UTRs of MBOAT7 containing either the C or the T allele (n=3) and analysed for; j) Relative mRNA expression of different ER stress markers, k) ROS generation, l) Relative mRNA expression of different cytokines (n = 8, 5, and 9; 3 biological replicates and 2-3 technical replicates, respectively) and m) Secreted TNF-α levels measured by ELISA (n=3). n) MDMs were transfected with scramble or MBOAT7 siRNA for 48h then examined for the relative mRNA expression levels of several regulators of mitochondrial dynamics(n=3). P-value < 0.04 for all comparisons. Values are represented by vertical bars and are mean \pm sem; *P < 0.05, ** P < 0.01, ****P < 0.001. Statistical differences between groups were assessed by 2-tailed Student's t test or one-way ANOVA; multiple comparisons were corrected by Bonferroni correction. Source data are provided as a Source Data file.

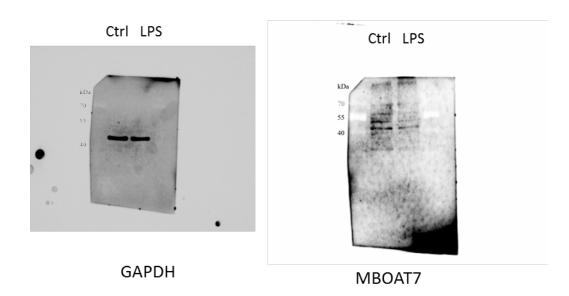


Supplementary Figure 5: MDMs were transfected with MBOAT7 siRNA or scramble as control and incubated with LPS (500 ng/ml, 4 h) (n=3) and then a) treated with NLRP3 inflammasome activator ATP (5 mM, 30 min), b) NLRP1 inflammasome activator muramyl dipeptide (MDP) (200 ng/ml, 8h), the NLRC4 inflammasome activator flagellin ((200 ng/ml, 8h), or the AIM2 inflammasome activator poly(dA:dT) (1 ug/ml, 8h) and quantitative ELISA of the conditional media measuring IL-1β levels is shown. c) MBOAT7 silencing led to higher levels of active caspase-1(n = 11 includes 3 biological replicates from independent individuals)and 3-4 technical replicates), d) and protein levels of pro- IL-1β and NLRP3 upon NLRP3 inflammasome activation e) MDMs were transfected with MBOAT7 overexpression plasmid or the empty vector as control and incubated with LPS (500 ng/ml, 4 h) (n=3) and then treated with NLRP3 inflammasome activator ATP (5 mM, 30 min) and quantitative ELISA of the conditional media measuring IL-1 β levels is shown. f) MDMs (n = 10/genotype) were treated with LPS (500 ng/ml, 4 h) and then treated with NLRP3 inflammasome activator ATP (5 mM, 30 min) and the secreted IL-1β levels were measured by quantitative ELISA. Human HeLa cells were transfected with full-length 3'UTRs of MBOAT7 containing either the C or the T allele and incubated with LPS (500 ng/ml, 4 h) and then treated with NLRP3 inflammasome activator ATP (5 mM, 30 min) (n=4-6) analysed for g) IL-1β secretion using quantitative ELISA and h) IL-B mRNA expression using RT-PCR. Data was normalised to GAPDH. Pvalue < 0.02 for all comparisons. Values are represented by vertical bars and are mean \pm sem; *P < 0.05, ** P < 0.01, ****P < 0.001. Statistical differences between groups were assessed by 2-tailed Student's t test or one-way ANOVA; multiple comparisons were corrected by Bonferroni correction. Source data are provided as a Source Data file.

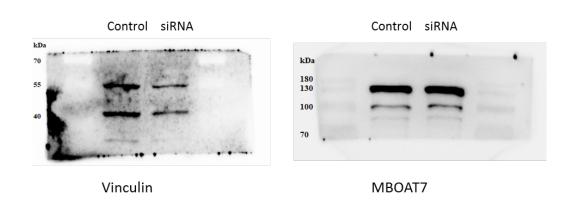


Supplementary Figure 6: Human MDMs isolated from patients with MAFLD and challenged with LPS for 24 hours (n= 10/group); a) Allele-specific expression assay showing the ratio of rs8736 alleles in genomic DNA (gDNA) and complementary DNA (cDNA) synthesized from pre-mRNA, b) Secreted MCP-1 and TNF-α levels measured by ELISA. MDMs from patients with MAFLD harbouring the CC genotype for rs8736 and transfected with miRNA control compared to subjects harbouring the TT genotype and transfected with either miRNA-24 inhibitor or control (n = 10/group); c) MBOAT7 relative mRNA expression assessed by RT-PCR, d) Secreted TNF-α and MCP-1 levels measured by ELISA. MBOAT7 relative mRNA expression e) by RT-PCR in control subjects (n=25) and patients with MAFLD related metabolic steatohepatitis (n=38), **f)** ArrayExpress public repository data (E-MEXP-3291) control subjects (n=19) and patients with MAFLD related metabolic steatohepatitis (n=16). g) MBOAT7 gene expression in the liver based on the Genotype-Tissue Expression (GTEx) dataset. h) Volcano plot and i) Gene Set Enrichment Analysis of inflammation pathways gene expression between the low and high expressed MBOAT7 samples in ArrayExpress public repository data (E-MEXP-3291). j) Heatmap of correlation (Pearson correlation) between hepatic MBOAT7 expression and inflammatory markers in E-MEXP-3291 dataset. Correlation between hepatic MBOAT7 expression and GSVA scores for inflammatory biological processes. k) Immunohistochemistry analysis of liver sections from patients with MAFLD harbouring CC/CT (n=9) or TT (n=6) genotype stained for sCD163 and CD68. Shown are representative immunohistochemistry images and summarized data. I) Diagram depicting the co-culture of primary human hepatocytes with MDMs or human Kupffer cells transfected with scramble or MBOAT7 siRNA for 48 hours and treated with LPS for 24 hours and relative mRNA expression of different cytokines in hepatocytes cultured with MDMs and with Kupffer cells assessed by q-PCR (n = 2 and 4 includes 2 biological replicates and 1-2 technical replicates respectively). P-value < 0.02 for all comparisons. Values are represented by vertical bars and are mean \pm sem; *P < 0.05, ** P < 0.01, ****P < 0.001. Statistical differences between groups were assessed by 2-tailed Student's t test or one-way ANOVA; multiple comparisons were corrected by Bonferroni correction. Source data are provided as a Source Data file.

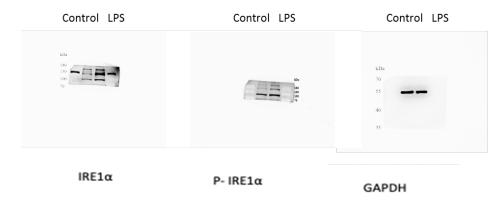
Supplementary 1 E:



Supplementary 2 A:



Supplementary 4 B:



Supplementary 5 D:

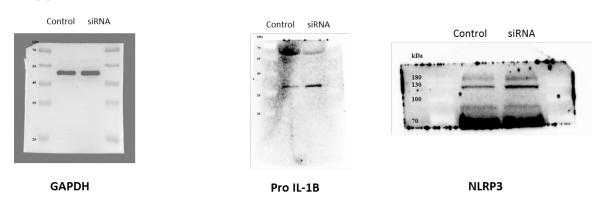


Figure 4A

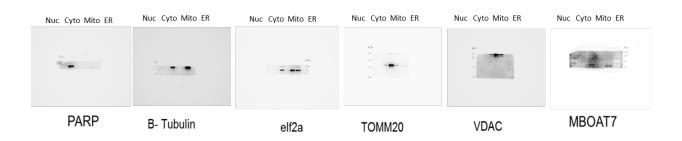


Figure 4B

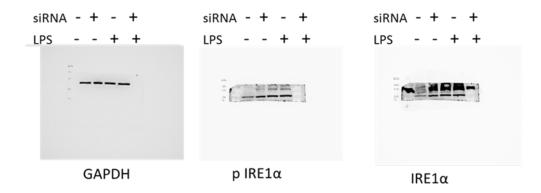


Figure 7H

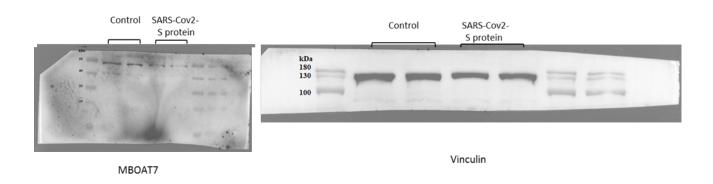
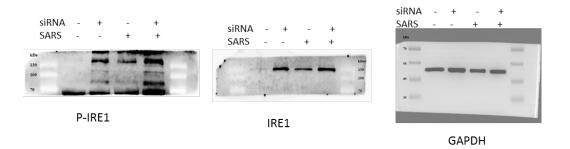


Figure 7 N



Supplementary Table 1: Characteristics of the patient cohort with MAFLD

Variables	NAFLD Cohort (n=30)	
Age (years)	58.5 (44.7-63)	
Male/ Female (%)	17 (56.7%)/ 13 (43.3%)	
BMI (Kg/m^2)	31.3 (25.5-36.5)	
Patients with diabetes' (%)	17 (56.7)	
ALT (IU/L)	51 (40.2-82.5)	
AST (IU/L)	36 (19.7-58.5)	
GGT (IU/L)	57.5 (30.7-177)	
Platelets (x10 ⁹ /L)	214 (180-246)	

^{*}Data are given as median \pm interquartile range or as %.

Supplementary Table 2: A list of the used reagents

REAGENT or RESOURCE	Source / Identifier
Antibodies	upplier name/ Catalog number / Clone name / Dilution
anti-GAPDH	Abcam (ab181602), monoclonal, 1/10000
anti-Vinculin	Abcam (ab129002), monoclonal, 1/20000
anti-PARP	Cell Signaling Technology (9542), monoclonal, 1/1000
anti-β -Tublin	Cell Signaling Technology (2128), monoclonal, 1/1000
anti-EIF2A	Abcam (ab169528), monoclonal, 1/1000
anti-TOMM20	Abcam (ab186735), monoclonal, 1/1000
anti-VDAC	Abcam (ab14734), monoclonal, 1 μg/ml
anti-IL-1β	Cell Signaling (12242), monoclonal, 1/1000
anti-NLRP3	AdipoGen (AG-20B-0014), monoclonal, 1µg/ml
anti-pIRE1 alpha [p ser 724]	Novus Biologicals [NB100-2323], polyclonal, 1/1000
anti-IRE1 alpha	Novus Biologicals [NB100-2324], polyclonal, 1/1000
anti-MBOAT7	Sapphire Bioscience (ARP49810), polyclonal, 1/500
anti-CD163	Cell Marque, MRQ-26, monoclonal, 1/50
anti-CD68	Abcam, ab955, monoclonal, 1/500
Bacterial and Virus Strains	
NA	
Chemicals, Peptides, and Recombinant Proteins	
qScript cDNA SuperMix	Genescript
SYBR Green PCR Master Mix	Invitrogen
Bovine serum albumin (BSA)	Sigma Aldrich
RIPA buffer	Sigma Aldrich
Trypsin	Sigma Aldrich
Protease inhibitors	Sigma Aldrich
Dimethyl sulfoxide (DMSO)	Sigma Aldrich
β-mercaptoethanol,	Sigma Aldrich
EDTA (Ethylenediaminetetraacetic acid)	Sigma Aldrich
Actinomycin D	Sigma Aldrich
IL-β protein	Thermo fisher (A42508)

Penicillin	Gibco BRL through Life Technologies
Streptomycin	Gibco BRL through Life Technologies
Lipofectamine RNAi-MAX	Gibco BRL through Life Technologies
Transfection Reagent	Gibco BRL through Life Technologies
Opti-MEM medium	Gibco BRL through Life Technologies
Dulbecco's Modified Eagle Medium (DMEM)	Lonza
RPMI 1640 Medium	Lonza
Phosphate buffered saline (PBS)	Lonza
Recombinant Human M-CSF	Lonza
TheraPEAKTM X-VIVOTM-15 Serum-free	Lonza
Hematopoietic Cell Medium,	Lonza
Ethanol, absolute	POCD Healthcare
Ficoll-Paque PLUS	GE Healthcare
Seahorse XF RPMI medium	In Vitro Technologies
Glutamine solution	In Vitro Technologies
Glucose solution	In Vitro Technologies
Pyruvate solution	In Vitro Technologies
Cryo Human Kupffer cells	Life Technologies
TaqMan TM Fast Advanced Master Mix	Thermo Fisher Scientific
Critical Commercial Assays	
Detergent Compatible (DC) Protein Assay Kit I	Bio-Rad (5000111)
FavorPrep Tissue Total RNA Purification Mini Kit	Favorgen Biotech Corp (FATRK001-2)
EasySep Human CD14 Positive Selection Kit II	STEMCELL Technologies (17858)
Human TLR1-9 Agonist Kit	InvivoGen (tlrl-kit1hw)
QIAGEN Plasmid Maxi Kit (25)	QIAGEN (12163)
QIAGEN miRNeasy Mini Kit	QIAGEN (217004)
DCFDA Cellular ROS Detection Assay Kit	Abcam (ab113851)
Human TNF-α ELISA Kit	Abcam (ab46087)
Human MCP-1 ELISA Kit	Abcam (ab179886)
Human IL-6 ELISA Kit	Abcam (ab178013)
Human IL-1β ELISA Kit	Abcam (ab214025)
Seahorse XF Cell Mito Stress Test Kit	In Vitro Technologies (SEA103015100)
Dual Luciferase (Firefly-Renilla) Assay System	JOMAR BIOSCIENCE (60683-2)

TMRE-Mitochondrial Membrane Potential Assay Kit	Abcam ab113852
MitoSOX TM Red Mitochondrial Superoxide Indicator	Thermo Fisher Scientific (M36008)
OneStep qMethyl-PCR Kit	ZYMO Research (D5310)
GSK1210151A	Sigma-Aldrich (SML0666-5MG)
Human Mitochondrial DNA Monitoring primer Set	Takara (7246)
Cytosolic Phospholipase A2 Assay Kit (ab133090)	Abcam (ab133090)
ATAC-Seq Kit	Active Motif (53150)
CUT&Tag-IT™ Assay Kit	Active Motif (53160)
Histone H3K27me3 antibody (pAb)	Active Motif (39055)
Histone H3K4me1 antibody (pAb)	Active Motif (39498)
ATP Colorimetric/Fluorometric Assay Kit	Biovision (K354-100)
Deproteinizing Sample Preparation Kit	Biovision (K808-200)
Lactate Colorimetric/Fluorometric Assay Kit	Biovision (K607-100)
Cyclooxygenase (COX) Activity Assay Kit (Fluorometric)	Abcam (ab204699)
Protease Inhibitor Cocktail (EDTA-free)	Abcam (ab201111)
Lipoxygenase Assay Kit	Abcam (ab241038)
Cytosolic Phospholipase A2 Assay Kit	Abcam (ab133090)
Fluo-8 Calcium Flux Assay Kit	Abcam (ab112129)
Caspase-1 Assay Kit	Promega Corporation (G9951)
Triglyceride Assay quantitation Kit	Abcam (ab65336)
Cholesterol/Cholesteryl Ester quantitation Kit	Abcam (ab65359)
Tocris AACOCF3	Invitro Tec. RDS (14625)
Indomethacin	Sigma-Aldrich (I7378-5G)
Actinomycin D	Sigma-Aldrich (A1410-2MG)
SARS-CoV-2 (2019-nCoV) Spike S1-His Recombiant Protein (HPLC-verified)	Jomar Life Research (40591-V08H-100UG)
Flagellin	Abcam (AB201366)
ATP	Sigma (A2383-1G)
Poly(dA:dT)	Jomar Life Research
MDP	Jomar Life Research
Mito-TEMPO	Sigma-Aldrich (SML0737)
Carbonyl cyanide 3-chlorophenylhydrazone (CCCP)	Sigma-Aldrich
Potassium chloride solution	Sigma-Aldrich

BAPTA-AM	Sigma-Aldrich
Experimental Models: Organisms/Strains	
NA	
Oligonucleotides	
Assay ID: Hs02786624_g1 / GAPDH	Thermo Fisher Scientific (4448489)
Assay ID: Hs04270764_pri, hsa-mir-23a	Thermo Fisher Scientific (4427012)
SNP ID: MBOAT7rs8736 C_2916339_10	Thermo Fisher Scientific (4351376)
mirVana™ miRNA Inhibitor, Negative Control	Thermo Fisher Scientific (4464076)
hsa-miR-24-2-5p mirVana® miRNA inhibitor	Thermo Fisher Scientific (4464084)
hsa-miR-24-2-5p mirVana® miRNA mimic	Thermo Fisher Scientific (4464066)
Other	
LENG4(MBOAT7) Human Tagged ORF Clone	OriGene
pCMV6-AC-GFP Tagged Cloning Vector	OriGene
siGENOME SMARTpool human MBOAT7	Dharmacon Cat. No. M-010084-01-0005
On-TARGET plus Non-Targeting pool	Dharmacon Cat. No. D-001810-10-05
LENG4 (MBOAT7) Human siRNA Oligo Duplex (Locus ID 79143)	ORIGENE Cat. No. SR312359
s18102 human PERK Silencer select	Thermo fisher (4427038)
s200430 human IRE1α Silencer select	Thermo fisher (4427038)
s223543 human ATF6 Silencer select	Thermo fisher (4427037)
TLR4 siRNA	Thermo fisher (4427038)
s13563 human STIM1 Silencer® Select	Thermo fisher (4427037)
s228396 human ORAI1 Silencer® Select	Thermo fisher (4427037)
Silencer TM Select Negative Control No. 1 siRNA	Thermo fisher 4390843
Software and Algorithms	
ImageJ bundled with 64-bit Java 8	https://wsr.imagej.net/distros/win/ij153-win-java8.zip
GraphPad Prism 9	http://www.graphpad.com/scientificsoftware/prism/
StepOne Software v2.3	https://www.thermofisher.com/hk/en/home/technical-resources/software-downloads/StepOne-and-StepOnePlus-Real-Time-PCR-System.html
R version 3.6.1	https://cran.r-project.org/bin/windows/base/old/3.6.1/
STAR version 2.7.2b (Dobin, 2013 #573)	https://www.ncbi.nlm.nih.gov/pubmed/23104886
Cuffdiff2 version 2.2.1 (Trapnell, 2013 #572)	https://bioinformaticshome.com/tools/rna-seq/descriptions/Cuffdiff-2.html

Ingenuity Pathway Analysis (IPA)	https://www.qiagen.com/us/products/discovery-and-translational-research/next-generation-sequencing/informatics-and-data/interpretation-content-databases/ingenuity-pathway-analysis/
Bowtie2 version 2.4.4	https://bowtie-bio.sourceforge.net/bowtie2/index.shtml
Homer version 4.11	http://homer.ucsd.edu/homer/
deepTools(version 3.5.1)	https://deeptools.readthedocs.io/en/develop/content/inst allation.html

Supplementary Table 3: A list of the sequence of primers

Primers	
GAPDH	For: AAGGTGAAGGTCGGAGTCAAG
	Rev: GGGGTCATTGATGGCAACAATA
TNF-α	For: TCTCTAATCAGCCCT
	Rev: TACAACATGGGCTAC
IL-6	For: TGCAATAACCACCCTGACC
	Rev: ATTTGCCGAAGAGCCCTCAG
MCP-1	For: TGC CGC CCT TCT GTG CCT G
	Rev: ACA GCA GGT GAC TGG GGC AT
CCL3	For: TGCAACCAGTTCTCTGCATC
	Rev: AATCTGCCGGGAGGTGTA
CCL5	For: CAACCCAGCAGTCGTCTTTG
	Rev: TCCCGAACCCATTTCTTCTC
IL1RA	For: GTGCTTTGGTACAGGGATTCCTG
	Rev: CACAGTCAGAGGTAGACCCTTC
MBOAT1	For: GTTTCCACAGCTTGCCAGA
	Rev: AGGTGATGCCCAACTTGTGT
MBOAT2	For: TCTGGTGAAAATGGAAAAGAAGA Rev: GCTTCTGAACAACCGCAGTA
MDO 4 T2	
MBOAT3	For: TGACCTTTGATCGCTTCCAT
	Rev: TCACCCTCGCGTCTCACT
MBOAT5	For: TGACATACCAGGAAAGATACCAAA Rev: GGTAGAAAAGGCCCAGACTCA
1.000	
MBOAT7	For: CATGCGGTACTGGAACATGA
	Rev: CCAGTAGGCGCTCAGCAG
BIP	For: TGTTCAACCAATTATCAGCAAACTC
	Rev: TTCTGCTGTATCCTCTTCACCAGT
ERDJ4	For: GCTACTCCCCAGTCAATTTTCA
	Rev: CCGATTTTGGCACACCTAAGAT
IREA1	For: GAAGATCCAGTCCTGCAGGTC
	Rev: AGAAGAGGGTTGATGGGCAG
	l

sXBP-1	For: CTGCCAGAGATCGAAAGAAGGC
	Rev: CTCCTGGTTCTCAACTACAAGGC
Affymetrix MBOAT7	For: AATATTGCAGACTTGGAAGG
	Rev: CAACACTTTATTGGGAAAGATT
Zymo MBOAT7	For: TTCTACGTGGCCTGGATTG
	Rev: GTTGCGGATGGTCTCATAGT
DRP1	For: CAAAGCAGTTTGCCTGTGGA
	Rev: TCTTGGAGGACTATGGCAGC
STAT2	For: CCGGGACATTCAGCCCTTTT
	Rev: CTCATGTTGCTGGCTCTCCA
MIEF1	For: AGGATGACAATGGCATTGGC
	Rev: CCGATCGTACATCCGCTTAAC
PRKN	For: AACATCAGTAGCTTTGCACCTG
	Rev: GGGGGAGTGATGCTATTTTT
NRF1	For: GGAGGAGTTCAATGAACTGCTGTC
	Rev: CTCTGGACCTTCTGCTTCATCTGT
PGC-1α	For: AGCCTCTTTGCCCAGATCTT
	Rev: GGCAATCCGTCTTCATCCAC
TFAM	For: GGCAAGTTGTCCAAAGAAACC
	Rev: GCATCTGGGTTCTGAGCTTTA
UQCRC2	For: AATTTCGTCGTTGGGAAGTAGC
	Rev: ATGAGTCTGCGGATTCTGAAAG
MT-CO1	For: GAGCTGCTGTTCGGTGTCC
	Rev: TGCCAGTGGTAGAGATGGTTG
ATP5A1	For: ATGACGACTTATCCAAACAGGC
	Rev: CGGGAGTGTAGGTAGAACACAT
SOD2	For: CTGCTGGGGATTGATGTGTGG
	Rev: TGCAAGCCATGTATC TTTCAG T
MFN1	For: CCTGGCATCCAGGAGTTAGA
	Rev: TGGTTCCAGCAATGCGATTT
MFN2	For: TGCAGGTGTAAGGGACGATT
	Rev: GAGGCTCTGCAAATGGGATG
FAM73B	For: CTCCTGCAGGTGGTAGGC

Rev: CAGAGACTGCATCAGAGCCA
For: TGGCAAAGGAGCAGATTAGTAGG
Rev: CTGCCACAAGAACTAGAGGATAAGA
For: GTCCCAAGGCTTTGGAATCTGTC
Rev: CCTACCAAGACAGGAGTTCTGG
For: CAGACAGTACCAACGCTTATGCC
Rev: GCAGAACTCCAGGTGCTTGAAG
For: AATCAGAAGGCAGTCCTCCC
Rev: TCGGGGAGCACAGAGTGTAC
For: GCTCCCTGACCGCTCACAGC
Rev: TGCCAGTGCATCGGTTCACCC
For: GACACCAACCTCAATAAGGTCTC
Rev: GGCTCAATGGATTTGTACTGTGC
For: CCTATGACTCAGAGCAGGTG
Rev: TCTTCGTAGTAGCAACG
For: ACCCAGCCAGCACTATGTG
Rev: TTGCCTTCATCTGACAGAACC
For: CGAGGGAACCAGCTGATAGA
Rev: CATGGGAACACTGAGCTTCA
For: 5- CACACTCTTTGGCACCTTCC-3'
Rev: 5-TGACAATCTGGAAGCCACAG-3'
For: 5-ACTGGATCGGCCAGAGTTAC-3'
Rev: 5-CGGCTCAAGTAGAGCTTGC-3'
For: 5'-CCACAGACCTTCCAGGAGAATG-3'
Rev: 5'-GTGCAGTTCAGTGATCGTACAGG-3'