

Article

Synthesis of 2-(2-Aminopyrimidine)-2,2-difluoroethanols as Potential Bioisosters of Salicylidene Acylhydrazides

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Abstract: Salicylidene acylhydrazides are inhibitors of type III secretion in several Gramnegative pathogens. To further develop the salicylidene acylhydrazides, scaffold hopping was applied to replace the core fragment of the compounds. The novel 2-(2-amino-pyrimidine)-2,2-difluoroethanol scaffold was identified as a possible analog to the salicylidene acylhydrazide core structure. The synthesis of a library of 2-(2-amino-pyrimidine)-2,2-difluoro-ethanols is described in this paper.

Keywords: scaffold hopping; Reformatsky reaction; salicylidene acylhydrazide; type III secretion

1. Introduction

Type III secretion (T3S) is a virulence system found in several clinically important Gram-negative pathogens including *Yersinia* spp., *Chlamydia* spp., *Shigella* spp., *Salmonella* spp., *Pseudomonas aeruginosa*, and enteropathogenic and enterohaemorrhagic *Escherichia coli* [1]. The bacteria use the T3S machinery to secrete and translocate toxins into the cytoplasm of the eukaryotic target cell and thereby create a niche that allows bacterial survival and growth. The T3S system is essential for the bacteria to cause disease and compounds that inhibit T3S have the potential to be developed into novel antibacterial agents [2,3]. Importantly, the T3S machinery can be inhibited without affecting growth and this mode of action is distinct from clinically used antibiotics that all target bacterial growth.

Salicylidene acylhydrazides (Figure 1a) were identified in a screen for T3S inhibitors [4] and subsequently it was shown that this compound class blocks T3S in Y. pseudotuberculosis, Shigella *flexneri*, enterohaemorrhagic E. coli, Chlamydia spp., and Salmonella spp. [5-13]. While the compounds are interesting as chemical probes to study T3S and clearly indicate a possibility to develop novel antibacterial drugs, the salicylidene acylhydrazide core scaffold suffers from a number of disadvantages. We have found that the salicylidene acylhydrazide T3S inhibitors generally display limited solubility and modest potency. In addition, the scaffold suffers from poor patentability and decomposes rapidly in slightly acidic environments. Salicylidene acylhydrazides have been included in many commercially available screening libraries and as a result the compounds frequently appear in publications and patents. Recently, a strategy based on statistical molecular design was used to compute multivariate quantitative structure-activity relationship models for the salicylidene acylhydrazides [14] and a desirable continuation is to further develop the compounds by modifying the central scaffold. Compounds that mimic and replace the salicylidene acylhydrazide scaffold are of importance since the compound class has delivered biologically active molecules in a number of human and microbial systems as exemplified by several recent publications [15-26]. This paper describes the synthesis of a library of 2-(2-aminopyrimidine)-2,2-difluoroethanols, identified from a salicylidene acylhydrazide through scaffold hopping.

2. Results and Discussion

2.1. Scaffold Hopping from a Salicylidene Acylhydrazide

The global minimum conformation of a salicylidene acylhydrazide (Figure 1a) obtained with Macromodel [27] was used as query scaffold in a scaffold hopping search using SHOP [28].

Figure 1. a) The query scaffold of a salicylidene acylhydrazide used for scaffold hopping; b) The highest similarity ranked scaffold, identified in the scaffold hopping program; c) The structure used for a substructure search in SciFinder.



The conformation is planar with an intramolecular hydrogen bond between the phenolic proton and the methylidine nitrogen (Figure 1a). Since the target of the salicylidene acylhydrazides has not been identified, the search was performed at default settings (see Experimental section for details). The reference database used contained 10,556 scaffolds and a total of 124,317 conformers. 2-(2-Amino-6-phenylpyrimidin-4-yl)-2,2-difluoro-1-phenylethanol (**4a**, Figure 1b) was identified as the virtual compound with highest similarity to the query scaffold. A SciFinder substructure search of 2-(2-aminopyrimidin-4-yl)-2,2-difluoro-1-(methyl)ethanol (Figure 1c) (performed January 15, 2010) found

no records of compounds with this scaffold. Compound **4a** and a number of analogs were thus targeted for synthesis.

2.2. Synthesis of 2-(2-Amino-pyrimidine)-2,2-difluoro-ethanols

Retrosynthetic analysis identified alkynyl ketone **3** (Scheme 1) as a key intermediate, which upon cyclization with guanidine would yield the target compound **4a** following a published procedure [29]. The synthesis of enantiopure alkynyl ketone **3** had previously been described in the literature [30]. This synthetic approach would allow structural variation by selection of a set of benzaldehydes and alkynyl lithium reagents.





Reagents and Conditions: I) Et₂Zn, MeCN, Wilkinson's catalyst, r.t., 14 h (crude yields after aqueous work-up); II) NaOH (1 M), CH₂Cl₂, r.t., 20 min; III) Oxalylchloride, DMF, CH₂Cl₂, then MeNHOMe*HCl, TEA, r.t., 14 h; IV) PyBOP, DMF, TEA, then MeNHOMe*HCl, r.t., 14 h; V) Me₂AlCl, CH₂Cl₂, then MeNHOMe*HCl, r.t., 14 h; VI) THF, -78 °C, 2 h; VII) guanidine hydrochloride, base (Na₂CO₃ or K₂CO₃), MeCN, MWI, 120 °C, 20 min; VIII) Dess-Martin periodinane, CH₂Cl₂, r.t., 3 h.

A modified Reformatsky reaction of ethyl bromodifluoroacetate with benzaldehyde gave β -hydroxy ester **1a** in 80% crude yield, as described by Sato *et al.* [31]. The resulting crude β -hydroxy ester was hydrolyzed to the corresponding carboxylic acid and subsequently converted into an acid chloride. Using the acid chloride, the Weinreb amide **2a** was synthesized in 79% yield. Using commercially available lithiumphenylacetylide the Weinreb amide was alkynylated to give **3** in 43% yield following a literature procedure [30]. We unsuccessfully attempted to reproduce the reported yield [30] by first preparing the lithiumphenylacetylide from *n*-BuLi, then from freshly prepared LDA, and finally increasing the amount of lithiumphenylacetylide. Using the protocol published by Tomkinson and co-workers [29] **4a** was obtained in 61% yield (Scheme 1).

Encouraged by the successful synthesis of 4a, a small library of analogs was targeted for synthesis. Four different aldehydes were selected based on previous data obtained with salicylidene acylhydrazides [5,14] and used to synthesize the crude products **1a-d**. A fifth aldehyde, nicotinic aldehyde, was also used as starting material but in our hands the Reformatzky reaction failed to produce the target intermediate in acceptable yield. The acid chloride strategy used to prepare **2a** proved hard to reproduce. Using an alternative strategy **2a** and **2c** were synthesized from **1a** and **1c** using the coupling reagent benzotriazol-1-yloxy)tripyrrolidinophosphonium hexafluorophosphate (PyBOP) in 79% and 75% yield, respectively. The coupling reagent *O*-(7-azabenzotriazol-1-yl)-*N*,*N*,*N'*,*N'*-tetramethyluronium hexafluorophosphate (HATU) was found to be as effective as PyBOP (data not shown). These yields were reproducible but to avoid the two-step procedure of ester hydrolysis and amide formation an alternative method using dimethylaluminum chloride to activate the *N*,*O*-dimethylhydroxylamine, was used to prepare **2a**, **2b**, and **2d** in 78%, 68%, and 62% yield, respectively. The Weinreb amides **2a-d** were purified by chromatography and isolated from the corresponding aldehyde in 37–63% yield over two or three steps. Subsequently the Weinreb amides **2a-d** were reacted with five different lithium acetylides (Scheme 1, Table 1).

ID	R ₁	R ₂	\mathbf{Yield}^{\dagger}
4a	Н	Ph	26%
4b	Н	<i>n</i> -Propyl	19%
4c	Н	Cyclopropyl	11%
4d	Н	CH ₂ NME ₂	<1%
4e	Н	Н	8%
4f	<i>m</i> -OMe	Ph	25%
4g	<i>m</i> -OMe	<i>n</i> -Propyl	12%
4h	<i>m</i> -OMe	Cyclopropyl	11%
4i	<i>m</i> -CF ₃	Ph	42%
4j	<i>m</i> -CF ₃	<i>n</i> -Propyl	19%
4k	m-CF3	Cyclopropyl	24%
41	<i>m</i> -CF ₃	Н	28%
4m	<i>o</i> -F	Ph	9%
4n	<i>o</i> -F	<i>n</i> -Propyl	3%
40	<i>o</i> -F	Н	12%

Table 1. The final library of 2-(2-aminopyrimidine)-2,2-difluoroethanols.

[†]Yield starting from Weinreb amides **2a-2d**.

The products obtained using any lithiumacetylide other than lithiumphenylacetylide, rapidly decomposed, even when stored dry in a freezer under a nitrogen atmosphere. Slight decomposition of products was observed even for those products obtained using lithiumphenylacetylide. Therefore the syntheses of compounds **4b-n** (Scheme 1, Table 1) were all made from the crude products obtained after alkynylation of **2a-d**. For the final cyclization reaction with guanidine four different bases (Li₂CO₃, Na₂CO₃, K₂CO₃, and Cs₂CO₃) were tested. There was no significant difference in yield when Na₂CO₃ was exchanged with K₂CO₃. The cyclization reaction proceeded far slower when Li₂CO₃ was used, while reactions performed with Cs₂CO₃ resulted in formation of several byproducts. Cyclization with urea and thiourea, using the same conditions as for the guanidine cyclization, was attempted, but the desired oxygen and sulfur analogs could not be obtained. To access additional analogs oxidation of

the secondary alcohol, using Dess-Martin periodinane, was performed for some of the compounds (Scheme 1, Table 2). The resulting ketones were in equilibrium with the hydrates (Scheme 1) and purification and characterization proved to be problematic.

The yields were generally low in the final step, starting from the Weinreb amides (<1%–42%). The poor yields could be attributed to the instability of the alkynyl ketones and that the cyclization reactions had to be performed using crude products. The resulting mixtures after the cyclization reaction contained several impurities and generally required several purification steps, contributing to the poor yields. All target compounds (**4a-n**) had a purity of at least 98% as determined by reversed phase HPLC analysis and at least 95% as determined by ¹H-NMR spectroscopy.

The final library of products (**4a-o**) was tested in a T3S-linked luciferase reporter gene assay in *Y*. *pseudotuberculosis*, essentially as described previously [4]. Only two compounds (**4e** and **4o**) showed effect on the luciferase light emission with 40% inhibition at 1 mM and bacterial growth experiments using *Y*. *pseudotuberculosis* showed that **4e** and **4o** did not inhibit growth at this concentration.

	n	D	X7 4 - 1 - 1 [†]
ID	\mathbf{K}_1	\mathbf{K}_2	Y leid
5a	Н	Ph	98%
5b	Н	n-Propyl	73%
5c	Н	Cyclopropyl	68%
5d	Н	Н	27%
5e	<i>m</i> -CF ₃	Ph	54%
5f	<i>m</i> -CF ₃	<i>n</i> -Propyl	55%
5g	<i>m</i> -CF ₃	Cyclopropyl	38%
5h	<i>m</i> -CF ₃	Н	66%

Table 2. The final library of ketones 5a-h.

[†]Yield starting from the target compounds **4a-c**, **4e** and **4i-l**.

3. Experimental

3.1. General

Mass-spectra were recorded with an electrospray Waters Micromass ZG 2000 instrument using an XTerra® MS C₁₈, 5 µm particles, 4.6 × 50 mm column and an H₂O/acetonitrile/0.2% formic acid eluent system. Preparative reversed-phase HPLC was performed on a Beckman System Gold HPLC with a Supelco Discovery BIO Wide Pore C₁₈, 5 µm particles, 21.2 × 250 mm column, using an H₂O/acetonitrile eluent system with or without 0.1% trifluoroacetic acid and a flow rate of 10 mL min⁻¹ and detection at 254 nm. Microwave heated reactions were performed in EmrysTM process vials using a SmithCreatorTM microwave instrument from Biotage. HRMS analysis was performed using a Waters Micromass GCT with an electron impact ion source (EI+), direct inlet (20-400 deg.) or a Bruker microTOF with an ESI ion source in positive mode. ¹H-NMR and ¹³C-NMR spectra were recorded with Bruker DRX-400 (¹H-NMR: 400 MHz; ¹³C-NMR: 100 MHz) or DRX-500 (¹H-NMR: 500 MHz; ¹³C-NMR: 126 MHz) spectrometers. NMR experiments were conducted at 298 K in (CD₃)₂CO (residual solvent peak = 2.05 ppm ($\delta_{\rm H}$) and 29.84 ppm ($\delta_{\rm C}$)), MeOD (residual solvent peak = 3.31 ppm ($\delta_{\rm H}$) and 49.00 ppm ($\delta_{\rm C}$)), or CDCl₃ [residual solvent peak = 7.26 ppm ($\delta_{\rm H}$)

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and 77.16 ppm (δ_C)]. Peak assignments could be established from complementary HMBC, HSQC and COSY experiments. The secondary alcohol proton in the compounds **2a-d** and **4a-o**, and the amino group protons of compounds **4a-o**, and **5a-h** could not be identified in many cases due to exchange broadening with the solvent. Equilibrium between the ketones and their respective hydrates (**5a-h**) were observed and while the mixtures were pure according to HPLC analysis, interpretable ¹³C-NMR spectra could not be obtained. Therefore only ¹H-NMR data are given for these compounds (**5a-h**). ¹H-NMR, ¹³C-NMR, are given for all other compounds, except **4d**, that could not be fully characterized due to lack of material. Low resolution electrospray ionization mass spectrometry (ESIMS) data are given for compounds **4a-n** and **5a-h**. High resolution mass spectrometry (HRMS) data is given for **4o** and the Weinreb amides **2b-d**, which did not ionize using the ESIMS instrument.

3.2. Scaffold Hopping

Conformational analysis of the salicylidene acylhydrazide was performed using the Macromodel program [27]. Scaffold hopping using the program SHOP [28] at default settings was performed using the global minimum conformation of the salicylidene acylhydrazide as query scaffold. The reference database used was previously built using Maybridge's building block collection from 2006 and using the virtual reaction manager as described in SHOP [32]. The virtual reaction utility program in SHOP assembles new core structures from fragments of building blocks [32]. The new compounds are expected to be synthetically accessible in five or less steps. The reference database used contained 10,556 scaffolds and a total of 124,317 conformers.

3.3. Synthesis

3.3.1. General procedure for the synthesis of Reformatsky adducts 1a-d

Wilkinson's catalyst (0.01 equivalents) was stirred in MeCN (20 mL) under N₂ atmosphere in an ice bath for 5 min. Ethyl bromodifluoroacetate (1.2 equivalents) and benzaldehyde (8–25 mmol, 1 equivalents) was added and the mixture was stirred for 5 min. Et₂Zn (1M, hexanes, 1.2 equivalents) was added in portions over 5 min and the reaction mixture was stirred overnight at r.t. The reaction mixture was quenched with excess HCl (1 M) and the organic solvents were evaporated under reduced pressure. The residue was dissolved in EtOAc and washed with HCl (1 M), NaHSO₃ (aq, sat.), and NaHCO₃ (aq, sat.). The organic phase was dried with anhydrous MgSO₄, filtered, and concentrated. The NMR spectrum of the crude product confirmed that the desired β -hydroxy ester had been obtained. The crude β -hydroxy ester was used directly in the next step.

3.3.2. Synthesis of Weinreb amide 2a via acid chloride formation in situ

NaOH (1 M, 40 mL) was added to a separation funnel containing the crude β -hydroxy ester **1a** (1.5 g, 6.5 mmol) in CH₂Cl₂ (10 mL), and the funnel was vigorously shaken for 10 min. The water phase was acidified with HCl (1 M) and extracted with EtOAc. The organic phase was dried with anhydrous MgSO₄, filtered, and concentrated. Most of the resulting crude acid (1.06 g, 5.24 mmol) was transferred to a round-bottom flask and CH₂Cl₂ (15 mL) was added. Under stirring in an ice-bath, oxalyl chloride (0.52 mL, 5.96 mmol) and a few drops of DMF was added. After 30 min, pyridine

(1.9 mL, 23.5 mmol) and *N*,*O*-dimethylhydroxylamine hydrochloride (0.62 g, 6.36 mmol) was added, and the reaction was stirred at r.t. overnight under a N_2 atmosphere. The reaction mixture was diluted with EtOAc, washed with HCl (1 M), and NaHCO₃ (aq, sat.). The aq. phases were extracted with EtOAc separately. The combined organic phases were dried with anhydrous MgSO₄, filtered, and concentrated. The resulting oil was purified by column chromatography (heptane:EtOAc 3:1) to give Weinreb amide **2a** in 79% yield (1.02 g).

3.3.3. Typical procedure for the synthesis of Weinreb amides **2a** and **2c**, using PyBOP, exemplified by the synthesis of **2c**

NaOH (2 M, 50 mL) was added to a stirred solution of β -hydroxy ester **1c** (4.38 g, 14.7 mmol) in EtOH (100 mL) at r.t. After 1.5 h, the mixture was acidified using excess HCl (1 M) and the solution was extracted with EtOAc. The organic phase was dried using anhydrous MgSO₄, filtered, and concentrated. The crude oil was co-evaporated with toluene and stored *in vacuo* before the resulting crude acid was used directly in the next reaction. To the crude acid, *N*,*O*-dimethylhydroxylamine hydrochloride (1.72 g, 17.6 mmol), PyBOP (9.17 g, 17.6 mmol), and pyridine (3.55 mL, 44 mmol) were added, and the mixture was stirred in DMF (70 mL) at r.t. over night. The reaction mixture was diluted with EtOAc and washed with HCl (1 M) and NaHCO₃ (aq. sat.). The aq. phases were extracted with EtOAc separately. The pooled organic phases were dried using anhydrous MgSO₄, filtered, and concentrated. The crude product was purified using column chromatography (heptane:EtOAc 1:1) to give Weinreb amide **2c** in 75% yield over two steps (4.58 g).

3.3.4. Typical procedure for the synthesis of Weinreb amides **2a**, **2b**, and **2d**, using dimethyl aluminium chloride, exemplified with the synthesis of **2d**

To a solution of *N*,*O*-dimethylhydroxylamine hydrochloride (1.8 g, 18.5 mmol) in CH₂Cl₂ at -78 °C under N₂ atmosphere, dimethyl aluminium chloride (22 mL, 0.9 M, heptane) was added and the mixture was stirred for 1 h. Crude β -hydroxy ester **1d** (1.41 g, 5.69 mmol) was added and the mixture was stirred at -78 °C for 30 min. The reaction mixture was then stirred at r.t. for 5.5 h at which time the β -hydroxy ester had been completely consumed according to TLC analysis. The reaction mixture was quenched with excess HCl (1 M) and diluted with EtOAc. The mixture was filtered through celite, washed with HCl (1 M), NaHCO₃ (aq, sat.), and extracted with EtOAc. The organic phase was dried with anhydrous MgSO₄, filtered, and concentrated. The crude product was purified using column chromatography (heptane:EtOAc 4:1 -> 2:1) to give Weinreb amide **2d** in 62% yield (0.927 g).

2,2-Difluoro-3-hydroxy-N-methoxy-N-methyl-3-phenyl-propionamide (2a). NMR data was in accordance with the data given in the literature [30].

2,2-Difluoro-3-hydroxy-N-methoxy-3-(3-methoxyphenyl)-N-methylpropionamide (**2b**). ¹H-NMR (400 MHz; acetone-d₆): δ 7.30–7.24 (m, 1H), 7.07 (s, 1H), 7.03 (d, J = 7.6 Hz, 1H), 6.90 (bd, J = 8.5 Hz, 1H), 5.43-5.33 (m, 1H), 5.23 (d, J = 5.3 Hz, 1H), 3.81 (s, 3H), 3.80 (s, 3H), 3.24 (s, 3H); ¹³C-NMR (126 MHz; acetone-d₆): δ 163.6, 160.5, 139.3 (d, ³J(C,F) = 2.7 Hz), 129.8, 121.1, 116.9 (dd,

 ${}^{1}J(C,F) = 258.9 \text{ Hz}, {}^{1}J(C,F) = 253.9 \text{ Hz}), 114.7, 114.5, 73.7 (dd, {}^{2}J(C,F) = 26.5 \text{ Hz}, {}^{2}J(C,F) = 23.0 \text{ Hz}), 62.3, 55.5, 33.7; ESI-HRMS [M+Na⁺]⁺ calcd. for [C₁₂H₁₅F₂NNaO₄]⁺:$ *m/z*: 298.0861; found: 298.0861.

2,2-Difluoro-3-hydroxy-N-methoxy-N-methyl-3-(3-trifluoromethylphenyl)-propionamide (2c). The alcohol proton was exchange broadened. ¹H-NMR (500 MHz; acetone-d₆): δ 7.84 (s, 1H), 7.78 (d, J = 7.6 Hz, 1H), 7.71 (d, J = 7.7 Hz, 1H), 7.63 (t, J = 7.7 Hz, 1H), 5.59 (dd, J = 17.2, J = 7.6 Hz, 1H), 3.82 (s, 3H), 3.26 (s, 3H); ¹³C-NMR (126 MHz; acetone-d₆): δ 163.1 (bs), 139.1, 132.8, 130.7 (q, ²J(C,F) = 32.0 Hz), 129.8, 126.0 (q, ³J(C,F) = 3.7 Hz), 125.4 (q, ³J(C,F) = 3.6 Hz), 125.3 (q, ¹J(C,F) = 271.2 Hz), 116.6 (dd, ¹J(C,F) = 259.1 Hz, ¹J(C,F) = 254.7 Hz), 73.0 (dd, ²J(C,F) = 27.1 Hz, ²J(C,F) = 23.7 Hz), 62.4, 33.4; ESI-HRMS [M+Na⁺]⁺ calcd. for [C₁₂H₁₂F₅NNaO₃]⁺: *m/z*: 336.0630; found: 336.0630.

2,2-Difluoro-3-(2-fluorophenyl)-3-hydroxy-N-methoxy-N-methylpropionamide (2d). ¹H-NMR (400 MHz, CDCl₃): δ 7.70–7.55 (m, 1H), 7.40–7.28 (m, 1H), 7.24–7.13 (m, 1H), 7.10–6.99 (m, 1H), 5.71 (bd, J = 19.4 Hz, 1H), 3.84 (bs, 1H), 3.75 (s, 3H), 3.26 (s, 3H); ¹³C-NMR (126 MHz; acetone-d₆): δ 163.3, 161.3 (d, ¹J(C,F) = 245.8 Hz), 131.1 (d, ^{3*}J(C,F) = 8.5 Hz), 130.9 (d, ^{3*}J(C,F) = 3.8 Hz), 125.0–(d, ^{4*}J(C,F) = 3.8 Hz), 125.0–124.8 (m, 1C) 116.6 (dd, ¹J(C,F) = 259.1 Hz, ¹J(C,F) = 254.4 Hz), 115.6 (d, ²J(C,F) = 22.6 Hz), 67.1-66.4 (m, 1C), 62.4, 33.6; ESI-HRMS [M+Na⁺]⁺ calcd. for [C₁₁H₁₂F₃NNaO₃]⁺: *m/z*: 286.0661; found: 286.0662. *Tentative assignment.

3.3.5. Typical procedure for the synthesis of pyrimidines 4a-o, exemplified by the synthesis of 4j

n-BuLi (1.6 M, 3.0 mL, 4.8 mmol) was added dropwise to a stirred solution of diisopropylamine (0.78 mL, 5.6 mmol) in THF at -78 °C under a stream of N₂. After 30 min, 1-pentyne (435 mg, 6.39 mmol) was added dropwise. After 1.5 h, a solution of Weinreb amide **2c** (500 mg, 1.60 mmol) in THF (5 mL) was added dropwise at -78 °C. The reaction was quenched with NH₄Cl (aq, sat., 5 mL) followed by addition of excess NaHCO₃ (aq, sat.). The THF was removed under reduced pressure. The remaining aqueous mixture was extracted with EtOAc and the organic phase was subsequently washed with brine, dried with MgSO₄, filtered, and concentrated. The resulting crude alkynyl ketone was heated using microwave irradiation (120 °C, 20 + 30 min) with guanidine hydrochloride (457 mg, 4.78 mmol) and Na₂CO₃ (677 mg, 6.39 mmol) in MeCN (15 mL). The reaction was filtered and purified by column chromatography (heptane:EtOAc 2:1 -> 1:1) to give pyrimidine **4j** in 19% yield (110 mg) over two steps.

4,4-Difluoro-5-hydroxy-1,5-diphenylpent-1-yn-3-one (**3**). NMR data was in accordance with the data given in the literature [30].

2-(2-Amino-6-phenylpyrimidin-4-yl)-2,2-difluoro-1-phenylethanol (**4a**). ¹H-NMR (400 MHz, MeOD): δ 8.17–8.08 (m, 2H), 7.57–7.45 (m, 5H), 7.40–7.28 (m, 4H), 6.49 (s, 2H), 5.49–5.33 (m, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 167.1, 164.7, 164.5 (dd, ²*J*(C,F) = 30.7 Hz, ²*J*(C,F) = 26.7 Hz), 138.3, 137.8, 131.8, 129.6, 129.1, 129.0, 128.6, 127.9, 119.1 (dd, ¹*J*(C,F) = 250.4 Hz, ¹*J*(C,F) = 246.1 Hz), 103.4 (t, ³*J*(C,F) = 4.6 Hz), 74.5 (t, ²*J*(C,F) = 26.8 Hz); ESIMS *m*/*z* calcd. [M+H⁺]⁺: 328; found: 328. 2-(2-Amino-6-propylpyrimidin-4-yl)-2,2-difluoro-1-phenylethanol (**4b**). The alcohol and amine protons were exchange broadened. ¹H-NMR (500 MHz; acetone-d₆): δ 7.48–7.44 (m, 2H), 7.39–7.32 (m, 3H), 6.90 (s, 1H), 5.34 (dd, J = 17.5, 7.3 Hz, 1H), 2.77–2.68 (m, 2H), 1.74 (app. sextet, J = 7.5 Hz, 2H), 0.94 (t, J = 7.4 Hz, 3H); ¹³C-NMR (126 MHz; acetone-d₆): δ 170.6, 167.2 (dd, ²*J*(C,F) = 31.3 Hz, ²*J*(C,F) = 27.9 Hz), 161.1, 137.7, 129.3, 129.0, 128.8, 118.8 (dd, ¹*J*(C,F) = 251.2 Hz, ¹*J*(C,F) = 247.3 Hz), 107.1 (t, ³*J*(C,F) = 4.2 Hz), 74.4 (dd, ²*J*(C,F) = 29.4 Hz, ²*J*(C,F) = 24.5 Hz), 37.6, 22.4, 13.7; ESIMS *m*/z calcd. [M+H⁺]⁺: 294; found: 295.

2-(2-Amino-6-cyclopropylpyrimidin-4-yl)-2,2-difluoro-1-phenylethanol (4c). The alcohol and amine protons were exchange broadened. ¹H-NMR (500 MHz; acetone-d₆): δ 7.50–7.46 (m, 2H, Ph*H*), 7.39–7.31 (m, 3H, Ph*H*), 6.74 (s, 1H, pyrimidine-C*H*), 5.35 (dd, *J* = 18.1, 6.6 Hz, 1H, benzylic C*H*), 2.23–2.16 (m, 1H, cyclopropyl-C*H*), 1.34–1.27 (m, 2H, cyclopropyl-C*H*₂), 1.26-1.16 (m, 2H, cyclopropyl-C*H*2); ¹³C-NMR (126 MHz; acetone-d₆): δ 174.4, 164.8 (dd, ²*J*(C,F) = 31.9 Hz, ²*J*(C,F) = 26.9 Hz), 160.2, 137.6, 129.3, 129.0, 128.8, 118.6 (dd, ¹*J*(C,F)= 251.8 Hz, ¹*J*(C,F)= 247.4 Hz), 104.1 (bs), 74.3 (dd, ²*J*(C,F)= 29.7, ²*J*(C,F)= 24.3 Hz), 16.4, 13.1, 13.0; ESIMS *m/z* calcd. [M+H⁺]⁺: 292; found: 292.

2-(2-Amino-6-dimethylaminomethylpyrimidin-4-yl)-2,2-difluoro-1-phenylethanol (4d). The alcohol and amine protons were exchange broadened. ¹H-NMR (500 MHz; acetone-d₆): δ 7.46 (d, *J* = 7.1 Hz, 2H, Ph*H*), 7.37–7.29 (m, 3H, Ph*H*), 6.96 (s, 1H, pyrimidine-C*H*), 5.33 (dd, *J* = 18.0, 7.2 Hz, 1H, benzylic C*H*), 4.37 (s, 2H, N-C*H*₂), 2.99 (s, 6H, N-C*H*₃); ESIMS *m*/z calcd. [M+H⁺]⁺: 309; found: 309.

2-(2-Aminopyrimidin-4-yl)-2,2-difluoro-1-phenylethanol (4e). The alcohol and amine protons were exchange broadened. ¹H-NMR (500 MHz; acetone-d₆): δ 8.44 (d, J = 5.3 Hz, 1H), 7.46 (d, J = 7.1 Hz, 2H), 7.38-7.30 (m, 3H), 6.90 (d, J = 5.3 Hz, 1H), 5.36 (dd, J = 17.6, 7.3 Hz, 1H);¹³C-NMR (126 MHz; acetone-d₆): δ 165.4–165.0 (m, 1C), 163.0, 158.4, 138.0, 129.1, 129.1, 128.7, 118.8 (dd, ¹*J*(C,F) = 250.6 Hz, ¹*J*(C,F) = 246.3 Hz), 107.8 (t, ³*J*(C,F) = 4.1 Hz), 74.4 (dd, ²*J*(C,F) = 29.5, ²*J*(C,F) = 24.4 Hz); ESIMS *m/z* calcd. [M+H⁺]⁺: 252; found: 252.

2-(2-Amino-6-phenylpyrimidin-4-yl)-2,2-difluoro-1-(3-methoxyphenyl)ethanol (**4f**). The amine and alcohol protons were exchange broadened. ¹H-NMR (500 MHz; acetone-d₆): δ 8.12 (bd, J = 6.6 Hz, 2H), 7.56-7.48 (m, 3H), 7.30 (s, 1H), 7.28-7.21 (m, 1H), 7.09-7.02 (m, 2H), 6.90-6.84 (m, 1H), 5.43–5.35 (m, 1H), 3.77 (s, 3H); ¹³C-NMR (126 MHz; acetone-d₆): δ 167.2, 164.6, 164.5 (dd, ²*J*(C,F) = 30.2 Hz, ²*J*(C,F) = 26.6 Hz), 160.4, 139.8 (d, ³*J*(C,F) = 1.8 Hz), 137.8, 131.9, 129.6, 129.6, 128.0, 121.4, 119.1 (dd, ¹*J*(C,F) = 250.1 Hz, ¹*J*(C,F) = 246.7 Hz), 114.7, 114.5, 103.5 (t, ³*J*(C,F) = 4.5 Hz), 74.5 (dd, ²*J*(C,F) = 29.3 Hz, ²*J*(C,F) = 24.7 Hz), 55.5; ESIMS *m/z* calcd. [M+H⁺]⁺: 358; found: 358

2-(2-Amino-6-propylpyrimidin-4-yl)-2,2-difluoro-1-(3-methoxyphenyl)ethanol (4g). The amine and alcohol protons were exchange broadened. ¹H-NMR (400 MHz; acetone-d₆): δ 7.27 (t, J = 7.9 Hz, 1H), 7.06–6.99 (m, 2H), 6.95 (s, 1H), 6.91 (ddd, J = 8.3 Hz, J = 5.6 Hz, J = 0.9 Hz, 1H), 5.31 (dd, J = 17.3 Hz, J = 7.4 Hz, 1H), 3.79 (s, 3H), 2.83-2.73 (m, 2H), 1.81-1.69 (m, 2H), 0.95 (t, J = 7.3 Hz,

3H); ¹³C NMR (126 MHz; acetone-d₆): δ 169.9, 167.9 (dd, ²*J*(C,F) = 31.6 Hz, ²*J*(C,F) = 26.8 Hz), 160.5, 160.4, 139.0 (d, ³*J*(C,F) = 1.8 Hz), 129.8, 121.3, 118.7 (dd, ¹*J*(C,F) = 251.3 Hz, ¹*J*(C,F) = 247.8 Hz), 114.7, 114.6, 107.1 (dd, ³*J*(C,F) = 4.9 Hz, ³*J*(C,F) = 3.2 Hz), 74.4 (dd, ²*J*(C,F) = 29.4 Hz, ²*J*(C,F) = 24.7 Hz), 55.5, 37.0, 22.5, 13.7;ESIMS *m/z* calcd. [M+H⁺]⁺: 324; found: 324

2-(2-*Amino-6-cyclopropyl-pyrimidin-4-yl*)-2,2-*difluoro-1-(3-methoxy-phenyl*)-*ethanol* (**4h**). ¹H-NMR (400 MHz; acetone-d₆): δ 7.24 (t, J = 7.5 Hz, 1H, Ph*H*), 7.06–6.99 (m, 2H, Ph*H*), 6.91–6.83 (m, 1H, Ph*H*), 6.72 (s, 1H, pyridimidine-C*H*), 6.21 (s, 2H, N*H*₂), 5.46–5.40 (m, 1H, O*H*), 5.38–5.28 (m, 1H, benzylic-C*H*), 3.77 (s, 3H, methoxy-C*H*₃), 1.99–1.89 (m, 1H, cyclopropyl-C*H*), 1.04–0.92 (m, 4H, cyclopropyl-C*H*₂); ¹³C NMR (126 MHz; acetone-d₆; for C atom numbering used see Figure 2) δ 174.8 (C5), 164.1 (C3, dd, ²*J*(C,F) = 31.4 Hz, ²*J*(C,F) = 26.8 Hz), 161.5 (C6), 160.4 (C9), 139.3 (C7, d, ³*J*(C,F) = 1.8 Hz), 129.7 (C11), 121.3 (C10*), 118.7 (C2, dd, ¹*J*(C,F) = 251.4 Hz, ¹*J*(C,F) = 247.1 Hz), 114.7 (C8*), 114.6 (C12*), 104.6 (C4, t, ³*J*(C,F) = 4.4 Hz), 74.3 (C1, dd, ²*J*(C,F) = 29.4 Hz, ²*J*(C,F) = 24.0 Hz), 55.5 (C13), 16.7 (C14), 12.4 (C15*), 12.3 (C16*); ESIMS *m/z* calcd. [M+H⁺]⁺: *m/z*: 322; found: 322. *Tentative assignment.

Figure 2. Numbering of carbon atoms of compound 4h, used for assignment of ¹³C-NMR peaks.



2-(2-Amino-6-phenylpyrimidin-4-yl)-2,2-difluoro-1-(3-trifluoromethylphenyl)ethanol (**4i**). The alcohol and amine protons were exchange broadened. ¹H-NMR (500 MHz; acetone-d₆): δ 8.17–8.12 (m, 2H), 7.86 (s, 1H), 7.79 (d, *J* = 7.6 Hz, 1H), 7.69 (d, *J* = 7.8 Hz, 1H), 7.62 (t, *J* = 7.7 Hz, 1H), 7.55–7.49 (m, 3H), 7.35 (s, 1H), 5.58 (dd, *J* = 17.8, 6.5 Hz, 1H); ¹³C-NMR (126 MHz; acetone-d₆): δ 167.4, 164.7, 164.0 (dd, ²*J*(C,F) = 30.6 Hz, ²*J*(C,F) = 26.3 Hz), 139.7 (d, ³*J*(C,F) = 1.2 Hz), 137.7, 133.0, 131.9, 130.5 (q, ²*J*(C,F) = 31.9 Hz), 129.6, 129.6, 128.0, 125.8 (q, ³*J*(C,F) = 3.7 Hz), 125.7 (q, ³*J*(C,F) = 3.8 Hz), 125.4 (q, ¹*J*(C,F) = 271.6 Hz),119.0 (dd, ¹*J*(C,F) = 250.6 Hz, ¹*J*(C,F) = 246.1 Hz), 103.4 (t, ³*J*(C,F) = 4.4 Hz), 73.8 (dd, ²*J*(C,F) = 30.0 Hz, ²*J*(C,F) = 24.4 Hz); ESIMS *m*/z calcd. [M+H⁺]⁺: 396; found: 396.

2-(2-Amino-6-propylpyrimidin-4-yl)-2,2-difluoro-1-(3-trifluoromethylphenyl)ethanol (**4j**). The alcohol and amine protons were exchange broadened. ¹H-NMR (500 MHz; acetone-d₆): δ 7.80 (s, 1H), 7.75 (d, J = 7.4 Hz, 1H), 7.69 (d, J = 7.5 Hz, 1H), 7.61 (t, J = 7.6 Hz, 1H), 6.78 (s, 1H), 5.50 (dd, J = 17.3, 6.8 Hz, 1H), 2.62 (t, J = 7.6 Hz, 2H), 1.74-1.65 (m, 2H), 0.92 (t, J = 7.3 Hz, 3H).¹³C-NMR (126 MHz; acetone-d₆): δ 173.3, 164.0 (dd, ²J(C,F) = 30.6 Hz, ²J(C,F) = 26.7 Hz), 163.3, 139.5, 133.0, 130.6 (q, ²J(C,F) = 32.0 Hz), 129.7, 125.9 (q, ¹J(C,F) = 3.7 Hz), 125.6 (q, ³J(C,F) = 3.8 Hz), 125.4 (q, ¹J(C,F) = 271.3 Hz), 118.8 (dd, ¹J(C,F) = 250.4 Hz, ¹J(C,F) = 246.4 Hz), 106.8 (t, ³J(C,F) = 4.2 Hz),

73.8 (dd, ${}^{2}J(C,F) = 29.7$ Hz, ${}^{2}J(C,F) = 24.5$ Hz), 39.4, 22.4, 13.9.ESIMS m/z calcd. $[M+H^{+}]^{+}$: 362; found: 362.

2-(2-Amino-6-cyclopropylpyrimidin-4-yl)-2,2-difluoro-1-(3-trifluoromethylphenyl)ethanol (**4k**). The alcohol and amine protons were exchange broadened. ¹H-NMR (500 MHz; acetone-d₆): δ 7.82 (s, 1H), 7.76 (d, J = 7.6 Hz, 1H), 7.70 (d, J = 7.8 Hz, 1H), 7.62 (t, J = 7.7 Hz, 1H), 6.78 (s, 1H), 5.51 (dd, J = 18.0, 6.2 Hz, 1H), 2.10-2.06 (m, 1H), 1.16-1.07 (m, 4H); ¹³C-NMR (126 MHz; acetone-d₆): δ 175.4, 163.2-162.6 (m, 2C), 139.5, 133.0, 130.7 (q, ²*J*(C,F) = 31.9 Hz), 129.7, 125.9 (q, ³*J*(C,F) = 3.7 Hz), 125.7 (q, ³*J*(C,F) = 3.6 Hz), 125.4 (q, ¹*J*(C,F) = 271.3 Hz), 118.6 (dd, ¹*J*(C,F) = 251.0 Hz, ¹*J*(C,F) = 246.5 Hz), 105.1, 73.7 (dd, ²*J*(C,F) = 30.0 Hz, ²*J*(C,F) = 24.4 Hz), 17.1, 11.8; ESIMS *m/z* calcd. [M+H⁺]⁺: 360; found: 360.

2-(2-Aminopyrimidin-4-yl)-2,2-difluoro-1-(3-trifluoromethylphenyl)ethanol (41). The alcohol and amine protons were exchange broadened. ¹H-NMR (400 MHz; acetone-d₆): δ 8.50 (d, J = 5.4 Hz, 1H), 7.82 (s, 1H), 7.76 (d, J = 7.6 Hz, 1H), 7.71 (d, J = 7.8 Hz, 1H), 7.62 (t, J = 7.7 Hz, 1H), 6.98 (d, J = 5.3 Hz, 1H), 5.52 (dd, J = 17.8, 6.5 Hz, 1H); ¹³C-NMR (100 MHz; acetone-d₆): δ 165.5 (dd, ²*J*(C,F) = 32.0 Hz, ²*J*(C,F) = 26.8 Hz), 162.4, 157.6, 139.3 (d, ³*J*(C,F) = 1.5 Hz), 133.0, 130.6 (q, ²*J*(C,F) = 32.1 Hz), 129.7, 126.0 (q, ³*J*(C,F) = 3.8 Hz), 125.6 (q, ³*J*(C,F) = 3.7 Hz), 125.3 (q, ¹*J*(C,F) = 271.2), 118.6 (dd, ¹*J*(C,F) = 251.1 Hz, ¹*J*(C,F) = 246.6 Hz), 107.8 (dd, ³*J*(C,F) = 5.3 Hz; ³*J*(C,F) = 3.4 Hz), 73.6 (dd, ²*J*(C,F) = 30.0 Hz, ²*J*(C,F) = 24.3 Hz); ESIMS *m/z* calcd. [M+H⁺]⁺: 320; found: 320.

2-(2-Amino-6-phenylpyrimidin-4-yl)-2,2-difluoro-1-(2-fluorophenyl)ethanol (4m). The amine and alcohol protons were exchange broadened. ¹H-NMR (400 MHz; acetone-d₆): δ 8.18–8.11 (m, 2H), 7.77–7.71 (m, 1H), 7.55–7.48 (m, 3H), 7.43–7.35 (m, 2H), 7.29–7.23 (m, 1H), 7.15–7.09 (m, 1H), 5.82 (dd, J = 19.5 Hz, J = 4.5 Hz, 1H); ¹³C-NMR (126 MHz; acetone-d₆): δ 167.2, 164.9, 164.2 (ddd, ²J(C,F) = 31.6 Hz, ²J(C,F) = 25.3 Hz, ³J(C,F) = 1.9 Hz), 161.5 (d, ¹J(C,F) = 245.4 Hz), 137.9 (d, ³J(C,F) = 1.8 Hz), 131.8 (d, ³J(C,F) = 1.8 Hz), 131.0 (d, ³J(C,F) = 1.8 Hz), 131.0 (d, ³J(C,F) = 1.8 Hz), 129.6 (d, ³J(C,F) = 1.8 Hz), 128.0 (d, ³J(C,F) = 1.8 Hz), 125.5 (d, ²J(C,F) = 1.8 Hz), 103.4, 67.5 (dd, ²J(C,F) = 32.8 Hz, ²J(C,F) = 23.7 Hz); ESIMS *m/z* calcd. [M+H⁺]⁺: *m/z*: 346; found: 346

2-(2-Amino-6-propylpyrimidin-4-yl)-2,2-difluoro-1-(2-fluorophenyl)ethanol (**4n**). The alcohol proton was exchange broadened. ¹H-NMR (400 MHz; acetone-d₆): δ 7.70 (bt, J = 7.4 Hz, 1H), 7.43–7.36 (m, 1H), 7.24 (td, J = 7.5 Hz, J = 1.0 Hz, 1H), 7.11 (ddd, J = 10.5 Hz, J = 8.3 Hz, J = 1.1 Hz, 1H), 6.76 (s, 1H), 6.53 (bs, 2H), 5.75 (dd, J = 19.1 Hz, J = 4.8 Hz, 1H), 2.62–2.57 (m, 2H), 1.70 (h, J = 7.5 Hz, 2H), 0.93 (t, J = 7.4 Hz, 3H); ¹³C-NMR (126 MHz; acetone-d₆): δ 174.3, 164.5, 163.0 (dd, ²*J*(C,F) = 31.6 Hz, ²*J*(C,F) = 26.1 Hz), 161.4 (dd, ¹*J*(C,F) = 245.8 Hz, ³*J*(C,F) = 2.7 Hz) 130.9, 131.0, 125.5 (d, ²*J*(C,F) = 13.5 Hz), 124.9 (t, ³*J*(C,F) = 2.7 Hz), 119.0 (t, ¹*J*(C,F) = 248.2 Hz), 115.6 (dd, ²*J*(C,F) = 22.7 Hz, ³*J*(C,F) 2 Hz), 106.6 (t, ³*J*(C,F) = 2.7 Hz), 67.4 (t, ²*J*(C,F) = 28.3 Hz), 40.4, 22.4, 14.0; ESIMS *m/z* calcd. [M+H⁺]⁺: *m/z*: 312; found: 312

2-(2-Aminopyrimidin-4-yl)-2,2-difluoro-1-(2-fluorophenyl)ethanol (40). ¹H-NMR (400 MHz; acetone-d₆): δ 8.42 (d, J = 4.9 Hz, 1H), 7.72 (t, J = 7.4 Hz, 1H), 7.44–7.34 (m, 1H), 7.25 (t, J = 7.4 Hz, 1H), 7.16–7.06 (m, 1H), 6.85 (d, J = 4.9 Hz, 1H), 6.47 (s, 2H), 5.79 (bd, J = 19.6 Hz, 1H), 5.50 (s, 1H); ¹³C-NMR (100 MHz; acetone-d₆): δ 164.7-164.5 (m, 1C), 163.1 (dd, ²J(C,F) = 32.6 Hz, ²J(C,F) = 25.7 Hz), 161.3 (d, ¹J(C,F) = 245.7 Hz), 160.8, 131.0 (d, ³J(C,F) = 8.6 Hz), 130.9–130.8 (m, 1C), 125.4 (bd, ²J(C,F) = 13.7 Hz), 124.9 (d, ³J(C,F) = 3.5 Hz), 118.9 (dd, ¹J(C,F) = 250.0 Hz, ¹J(C,F) = 245.4 Hz), 115.6 (d, ²J(C,F) = 22.3 Hz), 107.7-107.6 (m, 1C), 67.6-66.9 (m, 1C); EI-HRMS [M+H]⁺ calcd. for [C₁₂H₁₀N₃OF₃]: *m/z*: 269.0776; found: 269.0773

3.3.6. Typical procedure for the synthesis of ketones 5a-h, exemplified by the synthesis of 5f

To a stirred solution of 2-amino pyrimidine **4j** in CH_2Cl_2 (1 mL), Dess-Martin periodinane (47 mg, 110 µmol) was added at r.t. After 3 h, the reaction was concentrated and purified by column chromatography (heptane:EtOAc 4:1 -> 2:1) followed by preparative HPLC to give ketone **5f** in 55% yield (11 mg).

2-(2-Amino-6-phenylpyrimidin-4-yl)-2,2-difluoro-1-phenylethanone (**5a**). ¹H-NMR (400 MHz; acetone-d₆): δ 8.25–8.21 (m, 2H), 8.06–8.03 (m, 2H), 7.71–7.66 (m, 2H), 7.57–7.51 (m, 5H); ESIMS *m/z* calcd. [M+H⁺]⁺: 326; found: 326.

2-(2-Amino-6-propylpyrimidin-4-yl)-2,2-difluoro-1-phenylethanone (**5b**). ¹H-NMR (500 MHz; acetone-d₆): δ 8.00 (d, J = 7.5 Hz, 2H), 7.69 (tt, J = 7.5, 1.2 Hz, 1H), 7.53 (br t, J = 8.0 Hz, 2H), 7.06 (s, 1H), 2.67 (t, J = 7.6 Hz, 2H), 1.79-1.71 (m, 2H), 0.96 (td, J = 7.4, 0.7 Hz, 3H); ESIMS *m*/*z* calcd. [M+H⁺]⁺: 292; found: 292.

2-(2-Amino-6-cyclopropylpyrimidin-4-yl)-2,2-difluoro-1-phenylethanone (**5c**). ¹H-NMR (500 MHz; acetone-d₆): δ 8.01 (d, J = 7.7 Hz, 2H), 7.68 (t, J = 7.4 Hz, 1H), 7.53 (t, J = 7.9 Hz, 2H), 7.07 (s, 1H), 2.10–2.08 (m, 2H), 1.10–1.05 (m, J = 2.7 Hz, 4H); ESIMS *m*/*z* calcd. [M+H⁺]⁺: 290; found: 290.

2-(2-Aminopyrimidin-4-yl)-2,2-difluoro-1-phenylethanone (5d). ¹H-NMR (500 MHz; acetone-d₆): δ 8.55 (d, J = 4.9 Hz, 1H), 8.00 (d, J = 7.6 Hz, 2H), 7.71–7.68 (tt, J = 7.5, 1.2 Hz, 1H), 7.54 (t, J = 7.9 Hz, 2H), 7.12 (d, J = 4.9 Hz, 1H); ESIMS *m/z* calcd. [M+H⁺]⁺: 250; found: 250.

2-(2-Amino-6-phenylpyrimidin-4-yl)-2,2-difluoro-1-(3-trifluoromethylphenyl)ethanone (**5e**). ¹H-NMR (400 MHz; acetone-d₆): δ 8.34–8.30 (m, 2H), 8.26–8.21 (m, 2H), 8.08–8.02 (m, 1H), 7.82 (t, J = 8.0 Hz, 1H), 7.71 (s, 1H), 7.64–7.61 (m, 1H), 7.58–7.51 (m, 2H); ESIMS *m/z* calcd. [M+H⁺]⁺: 394; found: 394.

2-(2-Amino-6-propylpyrimidin-4-yl)-2,2-difluoro-1-(3-trifluoromethylphenyl)ethanone (**5f**). ¹H-NMR (400 MHz; acetone-d₆): δ 8.30–8.27 (m, 2H), 8.05 (d, J = 7.9 Hz, 1H), 7.81 (t, J = 7.8 Hz, 1H), 7.05 (s, 1H), 6.33 (s, 2H), 2.65 (t, J = 7.6 Hz, 2H), 1.80-1.68 (m, 2H), 0.95 (t, J = 7.4 Hz, 3H); ESIMS *m*/*z* calcd. [M+H⁺]⁺: 360; found: 360.

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2-(2-Amino-6-cyclopropylpyrimidin-4-yl)-2,2-difluoro-1-(3-trifluoromethylphenyl)ethanone (**5g**). ¹H-NMR (400 MHz; acetone-d₆): δ 8.31–8.26 (m, 2H), 8.05 (d, J = 8.1 Hz, 1H), 7.82 (t, J = 8.2 Hz, 1H), 7.09 (s, 1H), 2.11–2.07 (m, 1H), 1.08–1.02 (m, 4H); ESIMS *m*/*z* calcd. [M+H⁺]⁺: 358; found: 358.

2-(2-Aminopyrimidin-4-yl)-2,2-difluoro-1-(3-trifluoromethylphenyl)ethanone (5h). ¹H-NMR (500 MHz; acetone-d₆): δ 8.59 (d, J = 5.0 Hz, 1H), 8.32–8.29 (m, 2H), 8.06 (d, J = 7.9 Hz, 1H), 7.84–7.81 (m, 1H), 7.16 (d, J = 5.0 Hz, 1H); ESIMS *m*/*z* calcd. [M+H⁺]⁺: 318; found: 318.

4. Conclusions

In conclusion, we have demonstrated the synthesis of a small library of 2-(2-amino-pyrimidin-4-yl)-2,2-difluoro-1-(phenyl)ethanols. While the synthetic procedure give low yields in the last step, the yields are generally sufficient to produce enough material for biological screening and evaluation.

The target compounds **4a-o** did not inhibit T3S at concentrations that warrant further investigation. A possible explanation to the lack of activity might be due to that the compounds do not mimick crucial properties of the salicylidene acylhydrazides. The bioactive conformation of the salicylidene acylhydrazides is unknown and different weights could have been applied to the parameters in SHOP if information about the binding had been available. Although the compound databases contained more than 10,556 compounds in around 124,317 conformations, the chemical space sampled might have been insufficient in order to find biologically active salicylidene acylhydrazide analogs. The compounds were evaluated in cell-based assays and even if the compounds mimic the bioactive conformation of the salicylidene acylhydrazides and can bind to the target protein(s), a number of factors, such as low membrane permeability, bacterial efflux, metabolism, or poor distribution of the compounds could still render them inactive. Another factor that might play a role is the metal chelating capacity of the salicylidene acylhydrazides [9], which may be part of the mode of action. The metal chelating ability of **4a-o** is likely to be substantially different.

The synthesized target compounds **4a-o** belong to a structure class that to our knowledge has not been published to date. The compounds have high aqueous solubility compared to the salicylidene acylhydrazides. No precipitation was detected at 200 μ M for the entire library and at 1 mM for **4e** and **4o**, whereas many of the active T3S inhibiting salicylidene acylhydrazides start to precipitate at concentrations above 100 μ M. The logarithm of solubility in water was calculated, using the program MOE [33], to be around -3 for **4e** and **4o** while as low as -6 for **4i**. The compounds **4a-o** all fulfill Lipinski's rules (molecular weight 251-395, two donor atoms, 4-5 acceptor atoms, and logP 1.6-4.5) and the compounds do not decompose under acidic workup (HCl, 1 M).

The 2-(2-amino-pyrimidin-4-yl)-2,2-difluoro-1-(methyl)ethanol scaffold is novel and it can be of general interest to include compounds such as the ones presented in high throughput screening collections. Additionally the scaffold can be of special interest in projects where salicylidene acylhydrazides have shown biological activity in mammalian and microbial systems [15-26].

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Sample Availability: Please contact the corresponding author.

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