



The Role of Hydrogen Sulfide Regulation of Autophagy in Liver Disorders

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Abstract: Autophagy is a complex process of degradation of senescent or dysfunctional organelles in cells. Dysfunctional autophagy is associated with many diseases such as cancers, immune dysfunction, and aging. Hydrogen sulfide (H₂S) is considered to be the third gas signal molecule after nitrous oxide and carbon monoxide. In recent years, H₂S has been found to have a variety of important biological functions, and plays an important role in a variety of physiological and pathological processes. In this review, we review the recent role and mechanism of H₂S in regulating autophagy in liver disorders, in order to provide a basis for further research in the future.

Keywords: autophagy; hydrogen sulfide; liver disorders; nonalcoholic fatty liver disease; hepatic ischemia-reperfusion injury



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1. Introduction

Autophagy is a stable mechanism in eukaryotic cells. In this process, the abnormal proteins, pathogens, and organelles are wrapped by double membranes to form autophagosomes and then transferred to lysosomes for subsequent degradation [1-4]. It is well-known that autophagy is induced by various environmental stresses, such as hypoxia, nutrient deficiency, and growth factor deficiency, so as to eliminate the damage caused by these pressures and return to normal level after stress relief [5]. Autophagy is categorized into macroautophagy, microautophagy, and chaperone-mediated autophagy according to the differences in the target specificity, inducing signal, action time, and delivery pathway to lysosomes [6-8]. Macroautophagy is a conservative stress response process. The target substances, such as invading pathogens and damaged mitochondria, are isolated in double membrane vesicles called autophagosomes and transported to lysosomes for degradation [4,9]. Microautophagy refers to the direct phagocytosis of cargo in endoplasmic/lysosomal membrane invagination [10]. Chaperone-mediated autophagy is a type of selective autophagy, in which the proteins to be degraded in cells first bind to molecular chaperones, and then are degraded by lysosomal enzymes(Figure 1) [11,12]. Under physiological conditions, autophagy is usually maintained at a basic level. When stimulated by stressors, the significantly enhanced autophagy eliminates the abnormal proteins in cells to facilitate cell survival [13]. If autophagy is maintained at a high level for long time, it will cause cell death. Therefore, the effect of autophagy is a "double-edged sword" [14,15]. Autophagy plays an important role in many physiological and pathological processes, such as immune response, anti-aging, development, tumor inhibition [16–18], neurodegenerative diseases [19], cardiovascular diseases [20], infection and immunity [17]. However, the relevant mechanism is not completely clear.

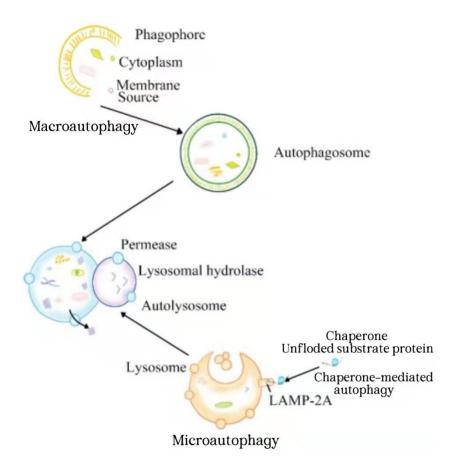


Figure 1. Process diagram of macroautophagy, microautophagy, and chaperone-mediated autophagy.

For decades, hydrogen sulfide(H₂S) has been regarded as a toxic gas. However, since the 1990s, more and more studies have demonstrated that H_2S , together with NO and CO, is gas transmission signal molecules [21,22]. There are three enzymes that catalyze the production of endogenous H₂S: cystine sulfide β -synthase (CBS), cystine sulfide γ -lyase (CSE), and 3-mercaptopyruvate sulfur transferase (3-MST). CBS catalyzes the β -substitution reaction of homocysteine with serine to produce L-cystathionine. The elimination of α , γ -cysteine of L-cystathionine is catalyzed by CSE to produce L-cystenine. On the one hand, L-cystine can be catalyzed by CSE/CBS through β elimination reaction to produce H₂S; on the other hand, it can also be catalyzed by cysteine aminotransferase (CAT) to transfer amine to α - ketoglutarate to form 3-mercaptopyruvate (3-MP). 3-MP is catalyzed by 3-MST to convert into H2S [23,24] (Figure 2). H₂S has been reported to contribute to many physiological processes, such as blood pressure reduction [25,26], anti-inflammation [27], antiapoptosis [28], anti-oxidative stress [29], cell proliferation/hypertrophy, cell survival/death, and cell differentiation [30]. PI3K/Akt/mTOR is an important pathway involved in the role of H_2S [31]. H_2S has been reported to act on autophagy through autophagy-related genes (such as Beclin1, ATG5) [32]. It has been reported recently that H_2S regulates autophagy in many physiological and pathological processes. In this review, we summarize the role and mechanism of H₂S regulating autophagy in liver disorders, hoping to provide theoretical reference for further related research in the future.

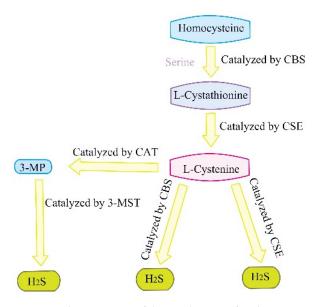


Figure 2. The summary of the production of endogenous H₂S. CBS: cystathionine-beta-synthase; CSE: cystathionine-gamma-lyase; 3-MST: 3-mercaptopyruvate thiotransferase; 3-MP: 3-mercaptopyruvate; CAT: cysteine aminotransferase.

2. Hydrogen Sulfide Plays a Protective Role by Regulating Autophagy in Nonalcoholic Fatty Liver Disease

Nonalcoholic fatty liver disease (NAFLD), affecting 25% of the adult population, is currently considered to be the most common liver disease in the world [33,34]. It includes a wide range of diseases, from simple steatosis to nonalcoholic steatohepatitis and fibrosis, and finally cirrhosis and hepatocellular carcinoma [35,36]. Many factors are considered to be related to NAFLD, including sedentary lifestyle, obesity, unhealthy diet, environmental factors, heredity, insulin resistance, and type 2 diabetes [37]. At present, no drugs have been approved for the treatment of NAFLD. The current treatment focuses on reducing disease-related risk factors, including obesity, dyslipidemia, insulin resistance, hyperglycemia, oxidative stress, and inflammation [38]. Hypertriglyceridemia (HTG) is a common metabolism disorder [39], and can lead to NAFLD [40-42]. Therefore, reducing blood triglyceride level can effectively improve NAFLD. The results of Li Sun et al. revealed that H₂S level in serum of patients with HTG and HTG model of C57BL/6 mice fed by high-fat diet (HFD) decreased. Sodium hydrosulfide (NaHS, a H₂S donor) could notably decrease the level of serum TG, liver weight, and liver free fatty acids (FFA) of HTG model of mice, which was counteracted by chloroquine (CQ, Washington, DC, USA, an inhibitor of autophagy). Further research showed that NaHS promoted autophagy by upregulating LC3BII/LC3BI ratio and downregulating p62 protein level in HFD-fed mice. Electron microscopy imaging revealed that NaHS treatment decreased the number of autophagosomes and lipid droplets, and some lipid droplets were obviously swallowed by autophagosomes in hepatocytes of HFD-fed mice, suggesting that autophagy might reduce lipid droplets in hepatocytes. Additionally, CQ abolished NaHS promotion of autophagy. It could be seen that exogenous H₂S decreased serum TG level to improve NAFLD through promotion of autophagy. Furthermore, the *p*-AMPK level significantly decreased in liver of HFD-fed mice, which was abolished by NaHS treatment. AMPK siRNA in L02 cells abolished NaHS promotion of autophagy; moreover, similar results were obtained in AMPK2⁻/ $^{-}$ mice, indicating that exogenous H₂S induced autophagy by activating AMPK/mTOR pathway. Summarizing, exogenous H₂S reduced serum TG level to improve NAFLD through autophagy promotion by activating AMPK/mTOR pathway [43]. Previous studies have shown that there are two triglyceride decomposition pathways in hepatocytes. One is autophagy decomposition pathway, which is called lipophagy; the other is cytoplasmic decomposition pathway [44]. Lipophagy is a selective

autophagy against lipid droplets, which is the basic mechanism to maintain the stability of the internal environment of lipid droplets [45]. The above results, especially the electron microscopy imaging results, showed that H₂S promoted lipophagy to reduce plasma triglyceride levels [43]. Similarly, our previous study used GYY4137(a H₂S donor) to treat primary mouse hepatocytes stimulated by oleic acid. The results showed that H₂S promoted lipid autophagy [46], which further confirmed the above conclusion. Our previous results also showed that exogenous H₂S inhibited NLRP3-mediated inflammation by upregulating autophagy through activating the AMPK/mTOR pathway in L02 cells [47,48], which further confirmed that AMPK/mTOR pathway in L02 cells [47,48], which further showed that H₂S in autophagy in liver. Another study of Dongdong Wu et al. showed that H₂S improved HFD-induced NAFLD by inhibiting apoptosis and promoting autophagy by inhibiting reactive oxygen species (ROS)-mediated phosphatidylinositol 3-kinase (PI3K)/AKT/mammalian

target of rapamycin (mTOR) cascade in L02 cells induced by OA [49]. Sterol regulatory element-binding proteins (SREBP-1c) is a transcription factor responsible for activating genes involved in the synthesis of fatty acids and triglycerides [50]. It is highly expressed in liver [51,52]. The increasing evidence have shown that SREBP-1c contributes to NAFLD [53–55]. HFD significantly increased the expression level of SREBP-1c and its downstream lipid metabolism-related proteins, and downregulated autophagy in HFD-fed mice. Whereas, SREBP-1c deficiency ameliorated hepatic steatosis and promoted autophagy, suggesting that HFD-induced hepatic steatosis and autophagy inhibition were SREBP-1c dependent. SREBP-1c decreased cystathionine gamma-lyase (CSE) through miR-216a to reduce liver H₂S level and the subsequent sulfhydration-dependent activation of Unc-51-like autophagy-activating kinase 1 (ULK1). Moreover, ULK1 Cys951 Ser sulfhydration by H_2S promoted ULK1 kinase activity and autophagy. Whereas the mutation of Cys951Ser in ULK1 inhibited autolysosome formation and exacerbated hepatic lipid accumulation in HFD-fed mice. Summarizing, it indicated that ULK1 sulfhydration upregulated autophagy and improved liver lipid accumulation. Further research showed that CSE silence in HFD-fed mice with SREBP-1c deficiency increased liver lipids accumulation. Collectively, HFD-induced SREBP-1c reduced the level of CSE-dependent H_2S in liver through miR-216a, which led to the reduction of sulfur hydration-dependent autophagy of ULK1, thus resulting in excessive accumulation of liver lipids. It can be concluded from above that H₂S improved hepatic steatosis by enhancing ULK1 sulfhydration-mediated autophagy. While HFD promoted hepatic steatosis by inhibiting CSE /H₂S/autophagy pathway, which is one of the mechanisms by which HFD induces NAFLD. Additionally, in the above study, H₂S regulated autophagy by sulfurizing ULK1 mainly through promoting the fusion of autophagosome and lysosome via inducing the formation of UVRAG/Rubicon association and increasing ATG14 phosphorylation [56]. Whether ULK1 sulfhydration by H_2S can regulate autophagy through other mechanisms in liver remains to be further studied.

Lifestyle changes, including exercise training, have been reported to be an effective method for NAFLD treatment [57–59]. Exercise can improve insulin sensitivity [60], promote very low density lipoprotein clearance, enhance liver output of triglycerides [61], and ameliorate appetite control [62] and myopenia [63,64]. More and more evidence indicate that the regular exercise improves NAFLD by downregulating the content of fat in the liver and promoting fatty acids β -oxidation through autophagy [57,65,66]. However, the mechanism involved has not been fully studied. Bing Wang and colleagues fed male mice with HFD to construct NAFLD model that was given 24 h of moderate-intensity exercise. The follow-up results showed that in HFD-fed mice, exercise reduced weight gain, abated systemic insulin resistance and glucose tolerance, improved hepatic steatosis, hepatic fibrosis, and the mitochondrial function, and promoted mitochondrial oxidation, which significantly improved NAFDL. Research revealed that exercise increased the level of H₂S in plasma and liver, as well as the mRNA expression of CBS, CES, and 3-MST in the liver of HFD-fed mice. Exercise reduced p62 protein expression, but had no notable influence on LC3-II/LC3-I ratio in the liver of HFD-fed mice. The above indicated that exercise upregu-

lated H₂S bioavailability and autophagy in the liver of HFD-fed mice, which may help to improve HFD-induced NAFLD. In addition, exercise inhibited malondialdehyde formation, increased GSH/GSSG ratio, and reduced the expression of TNF- α and IL-6 in the liver of HFD-fed mice, indicating that exercise mitigated oxidative stress and inflammation [67]. Whether exercise improves NAFLD by H₂S regulation of autophagy remains to be studied.

3. Hydrogen Sulfide Plays a Protective Role by Regulating Autophagy in Hepatic Ischemia-Reperfusion Injury

Tissue ischemia is a significant cause of death and disability worldwide. After ischemia for a period of time, the restoration of blood supply further exacerbates the damage of tissues and organs, which is called ischemia-reperfusion(I/R) injury. Studies have shown that free radical induced cell injury plays an important role in I/R injury [68-70]. Hepatic I/R injury is a common clinical problem in liver surgery, which leads to a large part of early graft failure and organ rejection [71,72]. Therefore, it is particularly important to explore the mechanism and protective strategy of hepatic I/R injury. More and more evidence show that autophagy is an important target to improve hepatic I/R injury [73–75]. Previous studies have revealed that hepatic I/R overactivated autophagy and led to autophagy death. Therefore, blocking the autophagy-induced cell death could effectively improve hepatic I/R injury [76,77]. Ping Cheng and colleagues found that exogenous H_2S could ameliorate hepatic I/R injury by improving the serum levels of ALT and AST and pathological changes induced by hepatic I/R. H₂S also attenuated hepatocyte apoptosis and autophagy induced by hepatic I/R in vivo and in vitro. During hepatic I/R injury, the JNK signal pathway was overactivated, which was inhibited by exogenous H₂S. JNK1 inhibition with its inhibitor SP600125 potentiated H₂S improvement of hepatic I/R injury. Overall, exogenous H₂S alleviated hepatic I/R injury by suppressing autophagy and apoptosis through JNK pathway inhibition, which needed to be further confirmed. Further research revealed that further reducing autophagy with 3-MA (autophagy inhibitor) would mitigate H_2S protective effects of hepatic I/R injury, while rapamycin (autophagy enhancer) potentiated H_2S improvement of hepatic I/R injury [78]. This seems to contradict the previous results. Autophagy is an important protective mechanism against hepatic I/R injury. H₂S can inhibit autophagy to protect against liver ischemia-reperfusion injury. However, rapamycin can enhance this liver protection by reversing the autophagy inhibition of H_2S . This shows that the protection mechanism of H2S is multifaceted. Moderate suppression of autophagy by H₂S protects cells, whereas excessive suppression of autophagy by H₂S has the opposite result. Moreover, the reason for this contradiction may be related to the time of liver ischemia.

Scavenger receptor A (SRA) is a receptor contributing to the macrophage-mediated inflammation, and involved in I/R injury [79,80]. The evidence indicates that SRA activation inhibits autophagy in macrophages [81]. Exogenous H_2S ameliorated fatty liver I/R injury by alleviating the pathological changes of liver tissue and downregulating the levels of LDH, ALT, and AST. In-depth research revealed that exogenous H_2S enhanced autophagy in peritoneal macrophages through upregulating the level of LC3B and LC3-II/LC3-I ratio in fatty liver I/R injury. Additionally, exogenous H₂S also suppressed apoptosis, inflammation, and oxidative stress, and downregulated SRA protein expression in fatty liver I/R injury, indicating that exogenous H_2S improved fatty liver I/R injury through promoting autophagy by inhibiting SRA pathway, which required further research through the use of related inhibitors [82]. SRA may be an important target for H_2S to improve hepatic I/R injury by regulating autophagy. Activation of SRA has previously been shown to inhibit ERS-induced macrophage autophagy [81], suggesting that H₂S promotes autophagy by inhibiting the SRA pathway. In fatty liver I/R injury, exogenous H₂S also downregulated ERS [82], hence, whether ERS mediated autophagy need further study. It is worth studying that exogenous H₂S regulates the improvement of ERS-mediated autophagy on substance metabolism.

4. Hydrogen Sulfide Plays a Protective Role by Regulating Autophagy in Hepatocellular Carcinoma

Hepatocellular carcinoma is one of the common cancers in the world. Its incidence rate is rising, which is closely related to advanced liver disease [83–85]. Many factors can lead to liver cancer, including hepatitis B and C viruses, nonalcoholic fatty liver, and alcohol use [86]. About half of HCC cases are diagnosed early [87]. Although the treatment of liver cancer has made progress in recent years, the 5-year survival rate of patients with distant metastasis is still 2.4% [88]. The increasing evidence indicate that autophagy is involved in hepatocellular carcinoma [87,89,90]; however, the relevant mechanism is not completely clear, especially the role of H_2S in regulating autophagy in hepatocellular carcinoma has not been clarified. The results of Shanshan S Wang et al. showed that NaHS suppressed hepatoma cell proliferation, migration, and cell cycle progression to improve hepatocellular carcinoma. The in-depth research revealed that NaHS promoted autophagy by upregulating the expression of LC3-II and Atg5 and downregulated p62 expression in HepG2 and HLE cells. The combination of H₂S and rapamycin (an autophagy inducer) further notably upregulated LC3-II expression, suppressed the proliferation, migration, and cell cycle of hepatoma cells. Moreover, both rapamycin and NaHS notably suppressed the protein expression of *p*-PI3K, *p*-Akt, and mTOR in hepatoma cells, suggesting that H₂S promoted autophagy through the PI3K/AKT/mTOR signaling pathway inhibition. Overall, exogenous H₂S ameliorated hepatocellular carcinoma through promoting autophagy by inhibiting PI3K/AKT/mTOR pathway (Figure 3) [91]. Autophagy plays a double cast in cancer, including inhibiting tumor and promoting tumor, which indicate that autophagy plays a double-edged sword role in cancer cells [92]. Similarly, H₂S also has a dual role in cancer [93,94]. Therefore, whether the regulation of autophagy by H_2S plays a positive or negative role in cancer remains to be studied.

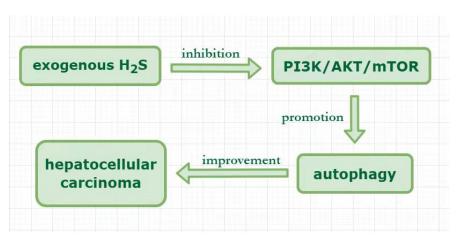


Figure 3. Hydrogen sulfide plays a protective role by regulating autophagy in hepatocellular carcinoma.

5. Hydrogen Sulfide Exposure Induces Oxidative Stress and Promotes Hepatocyte Autophagy to Lead Liver Injury

Oxidative stress plays an important regulatory role in autophagy in liver [95–97]. Studies have shown that high concentrations of H_2S might be cytotoxic and stimulate oxidative stress [98]. Thus, it can be deduced that high concentrations of H_2S regulates autophagy through oxidative stress. Jin Ming Guo and colleagues used one-day-old chickens as a model to assess the effects of high concentrations of H_2S and LPS on oxidative stress and autophagy. The results showed that compared with the control group, high concentrations of H_2S decreased the activity of antioxidant enzymes (superoxide dismutase, antioxidant glutathione, catalase and glutathione peroxidase) and increased the level of malondialdehyde. The in-depth studies showed that compared with the control group, high concentrations of H_2S inhibited expression of genes related to PI3/Akt/mTOR pathway and increased the expression of other autophagy-related genes (Beclin1, ATG5 and the

ratio of LC3-II/LC3-I), indicating that high concentrations of H_2S caused oxidative stress and induced autophagy in chicken hepatocytes by inhibiting PI3K/Akt/TOR pathway and leading liver injury. Moreover, high concentrations of H_2S aggravated oxidative stress and autophagy injury induced by LPS [99]. It has also been reported that H_2S can inhibit autophagy and play a cytoprotective role by inhibiting oxidative stress [95,100], which contradicts the above research. The reason may be the different concentration of H_2S and the different types of tissues and cells. Low concentrations of H_2S may often have a protective effect, while high concentrations of H_2S have the opposite effects.

6. Conclusions

In this review, we summarized the role of H_2S regulation of autophagy in different types of liver disorders in recent years as follows:(1) H2S improved NAFLD via reducing serum TG level through autophagy promotion by activating AMPK/mTOR pathway; (2) H2S inhibited apoptosis and promoted autophagy by inhibiting ROS-mediated PI3K/AKT/mTOR to improve NAFLD; (3) H2S improved hepatic steatosis through promoting ULK1 sulfhydration-mediated autophagy; (4) exercise promoted H2S bioavailability and autophagy in the liver of HFD-fed mice to improve HFD-induced NAFLD; (5) H_2S improved hepatic I/R injury by reducing autophagy and apoptosis through inhibiting JNK pathway, which needed to be further confirmed; (6) H_2S improved fatty liver I/R injury by promoting autophagy through the inhibition of SRA pathway, which required to be further confirmed; (7) H₂S improved hepatocellular carcinoma by promoting autophagy through the inhibition of PI3K/AKT/mTOR pathway; (8) high concentrations of H_2S promoted autophagy in chicken hepatocytes by inhibiting PI3K/Akt/TOR pathway to induce liver injury (Table 1). It can be seen from the above that H_2S regulates autophagy, which can sometimes play a protective role in different liver disorders, and sometimes on the contrary. The reason may be related to the concentration of H_2S . In addition, sometimes H_2S can promote autophagy, while sometimes it can inhibit autophagy, which may be related to the different stages of the pathological processes and the basic level of autophagy. In addition, in liver disorder, the signaling pathway involved in H_2S regulating autophagy needs to be further studied.

The Type of Liver Disorder	The Role Hydrogen Sulfide Regulation of Autophagy	Experimental Model	References
Nonalcoholic fatty liver disease (NAFLD)	H ₂ S improved NAFLD through autophagy promotion by activating AMPK/mTOR pathway	male C57BL/6 mice model of NAFLD	[43]
NAFLD	H ₂ S promoted autophagy through the inhibition of reactive oxygen species (ROS)-mediated phosphatidylinositol 3-kinase (PI3K)/AKT/mammalian target of rapamycin (mTOR)	Mouse/mouse hepatocytes model of NAFLD	[49]
hepatic steatosis	H ₂ S improved hepatic steatosis through promotion of ULK1 sulfhydration-mediated autophagy	Mouse model of NAFLD	[56]
NAFLD	Exercise promoted H ₂ S bioavailability and autophagy to improve HFD-induced NAFLD	male C57BL/6J mice model of NAFLD	[67]
Hepatic ischemia-reperfusion injury(HIRI)	H ₂ S improved hepatic I/R injury through autophagy reduction by inhibiting JNK pathway, which needs to be further confirmed	Mouse/mouse hepatocytes model of HIRI	[78]

Table 1. The summary of the role of hydrogen sulfide regulation of autophagy in liver disorders.

The Type of Liver Disorder	The Role Hydrogen Sulfide Regulation of Autophagy	Experimental Model	References
HIRI	H ₂ S improved fatty liver I/R injury through autophagy promotion via the inhibition of SRA pathway, which requires to be further confirmed	Sprague Dawley rats model of HIRI	[82]
hepatocellular carcinoma	H ₂ S improved hepatocellular carcinoma through autophagy promotion via the inhibition of PI3K/AKT/mTOR pathway	hepatocellular carcinoma (HCC): HepG2 and HLE	[91]
liver injury	high concentrations of H ₂ S induced liver injury through autophagy promotionby inhibiting PI3K/Akt/TOR pathway	chicken hepatocytess	[99]

Table 1. Cont.

In recent years, the interaction between autophagy and NLRP3 inflammasome has been reported to be involved in many metabolic disorder-related diseases, including NAFLD [8]. Moreover, NLRP3 inflammasome can be used as a target of H_2S regulation in many diseases [21]. Therefore, whether H_2S can improve NAFLD by regulating autophagy/NLRP3 inflammasome is a topic worthy of study. With the deepening of the research, the role of H_2S in regulating autophagy may become an important therapeutic strategy for liver disorders.

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References

- 1. Wang, J.; Wu, D.; Wang, H. Hydrogen sulfide plays an important protective role by influencing autophagy in diseases. *Physiol. Res.* **2019**, *68*, 335–345. [CrossRef] [PubMed]
- Qiu, D.M.; Wang, G.L.; Chen, L.; Xu, Y.Y.; He, S.; Cao, X.L.; Qin, J.; Zhou, J.M.; Zhang, Y.X.; Qun, E. The expression of beclin-1, an autophagic gene, in hepatocellular carcinoma associated with clinical pathological and prognostic significance. *BMC Cancer* 2014, 14, 327. [CrossRef] [PubMed]
- Guo, Y.; Zhang, X.; Wu, T.; Hu, X.; Su, J.; Chen, X. Autophagy in Skin Diseases. *Dermatology* 2019, 235, 380–389. [CrossRef] [PubMed]
- 4. Galluzzi, L.; Baehrecke, E.H.; Ballabio, A.; Boya, P.; Bravo-San Pedro, J.M.; Cecconi, F.; Choi, A.M.; Chu, C.T.; Codogno, P.; Colombo, M.I.; et al. Molecular definitions of autophagy and related processes. *EMBO J.* **2017**, *36*, 1811–1836. [CrossRef]
- 5. Wang, Y.; Zhang, H. Regulation of Autophagy by mTOR Signaling Pathway. Adv. Exp. Med. Biol. 2019, 1206, 67–83. [CrossRef]
- 6. Ding, Y.; Fu, X.; Wang, Q.; Liu, H.; Wang, H.; Wu, D. The Complex Interplay between Autophagy and NLRP3 Inflammasome in Renal Diseases. *Int. J. Mol. Sci.* 2021, 22, 12766. [CrossRef]
- 7. Lv, S.; Wang, Z.; Wang, J.; Wang, H. Exogenous Hydrogen Sulfide Plays an Important Role Through Regulating Autophagy in Ischemia/Reperfusion Injury. *Front. Mol. Biosci.* **2021**, *8*, 681676. [CrossRef]
- 8. Lv, S.; Wang, H.; Li, X. The Role of the Interplay Between Autophagy and NLRP3 Inflammasome in Metabolic Disorders. *Front. Cell Dev. Biol.* **2021**, *9*, 634118. [CrossRef]
- 9. Levine, B.; Kroemer, G. Biological Functions of Autophagy Genes: A Disease Perspective. Cell 2019, 176, 11-42. [CrossRef]
- 10. Mijaljica, D.; Prescott, M.; Devenish, R.J. Microautophagy in mammalian cells: Revisiting a 40-year-old conundrum. *Autophagy* **2011**, *7*, 673–682. [CrossRef]

- 11. Kaushik, S.; Cuervo, A.M. The coming of age of chaperone-mediated autophagy. *Nat. Rev. Mol. Cell Biol.* **2018**, *19*, 365–381. [CrossRef] [PubMed]
- Fujiwara, Y.; Wada, K.; Kabuta, T. Lysosomal degradation of intracellular nucleic acids-multiple autophagic pathways. *J. Biochem.* 2017, 161, 145–154. [CrossRef] [PubMed]
- 13. Glick, D.; Barth, S.; Macleod, K.F. Autophagy: Cellular and molecular mechanisms. J. Pathol. 2010, 221, 3–12. [CrossRef] [PubMed]
- 14. Liu, Y.; Levine, B. Autosis and autophagic cell death: The dark side of autophagy. *Cell Death Differ.* **2015**, *22*, 367–376. [CrossRef] [PubMed]
- 15. Garcia-Huerta, P.; Troncoso-Escudero, P.; Jerez, C.; Hetz, C.; Vidal, R.L. The intersection between growth factors, autophagy and ER stress: A new target to treat neurodegenerative diseases? *Brain Res.* **2016**, *1649*, 173–180. [CrossRef]
- 16. Mizushima, N.; Yoshimori, T.; Ohsumi, Y. The role of Atg proteins in autophagosome formation. *Annu. Rev. Cell Dev. Biol.* **2011**, 27, 107–132. [CrossRef]
- 17. Deretic, V.; Saitoh, T.; Akira, S. Autophagy in infection, inflammation and immunity. *Nat. Rev. Immunol.* **2013**, *13*, 722–737. [CrossRef]
- Kuma, A.; Komatsu, M.; Mizushima, N. Autophagy-monitoring and autophagy-deficient mice. *Autophagy* 2017, 13, 1619–1628. [CrossRef]
- Menzies, F.M.; Fleming, A.; Rubinsztein, D.C. Compromised autophagy and neurodegenerative diseases. *Nat. Rev. Neurosci.* 2015, 16, 345–357. [CrossRef]
- 20. Shirakabe, A.; Ikeda, Y.; Sciarretta, S.; Zablocki, D.K.; Sadoshima, J. Aging and Autophagy in the Heart. *Circ. Res.* 2016, 118, 1563–1576. [CrossRef]
- Wang, H.; Shi, X.; Qiu, M.; Lv, S.; Zheng, H.; Niu, B.; Liu, H. Hydrogen Sulfide Plays an Important Role by Influencing NLRP3 inflammasome. *Int. J. Biol. Sci.* 2020, 16, 2752–2760. [CrossRef]
- Wen, Y.D.; Wang, H.; Zhu, Y.Z. The Drug Developments of Hydrogen Sulfide on Cardiovascular Disease. Oxid. Med. Cell. Longev. 2018, 2018, 4010395. [CrossRef] [PubMed]
- 23. Zhao, S.; Li, X.; Li, X.; Wei, X.; Wang, H. Hydrogen Sulfide Plays an Important Role in Diabetic Cardiomyopathy. *Front. Cell Dev. Biol.* **2021**, *9*, 627336. [CrossRef] [PubMed]
- 24. Lv, S.; Li, X.; Zhao, S.; Liu, H.; Wang, H. The Role of the Signaling Pathways Involved in the Protective Effect of Exogenous Hydrogen Sulfide on Myocardial Ischemia-Reperfusion Injury. *Front. Cell Dev. Biol.* **2021**, *9*, 723569. [CrossRef] [PubMed]
- Greaney, J.L.; Kutz, J.L.; Shank, S.W.; Jandu, S.; Santhanam, L.; Alexander, L.M. Impaired Hydrogen Sulfide-Mediated Vasodilation Contributes to Microvascular Endothelial Dysfunction in Hypertensive Adults. *Hypertension* 2017, 69, 902–909. [CrossRef]
- 26. Jin, S.; Teng, X.; Xiao, L.; Xue, H.; Guo, Q.; Duan, X.; Chen, Y.; Wu, Y. Hydrogen sulfide ameliorated L-NAME-induced hypertensive heart disease by the Akt/eNOS/NO pathway. *Exp. Biol. Med.* **2017**, 242, 1831–1841. [CrossRef]
- 27. Zhao, A.S.; Zou, D.; Wang, H.H.; Han, X.; Yang, P.; Huang, N. Hydrogen sulphide-releasing aspirin enhances cell capabilities of anti-oxidative lesions and anti-inflammation. *Med. Gas. Res.* **2019**, *9*, 145–152. [CrossRef] [PubMed]
- Li, J.; Yuan, Y.Q.; Zhang, L.; Zhang, H.; Zhang, S.W.; Zhang, Y.; Xuan, X.X.; Wang, M.J.; Zhang, J.Y. Exogenous hydrogen sulfide protects against high glucose-induced apoptosis and oxidative stress by inhibiting the STAT3/HIF-1alpha pathway in H9c2 cardiomyocytes. *Exp. Ther. Med.* 2019, *18*, 3948–3958. [CrossRef] [PubMed]
- 29. Tocmo, R.; Parkin, K. S-1-propenylmercaptocysteine protects murine hepatocytes against oxidative stress via persulfidation of Keap1 and activation of Nrf2. *Free Radic. Biol. Med.* **2019**, *143*, 164–175. [CrossRef]
- 30. Zhang, D.; Du, J.; Tang, C.; Huang, Y.; Jin, H. H2S-Induced Sulfhydration: Biological Function and Detection Methodology. *Front. Pharmacol.* **2017**, *8*, 608. [CrossRef]
- Nie, L.; Liu, M.; Chen, J.; Wu, Q.; Li, Y.; Yi, J.; Zheng, X.; Zhang, J.; Chu, C.; Yang, J. Hydrogen sulfide ameliorates doxorubicininduced myocardial fibrosis in rats via the PI3K/AKT/mTOR pathway. *Mol. Med. Rep.* 2021, 23, 1691–1700. [CrossRef] [PubMed]
- Iqbal, I.K.; Bajeli, S.; Sahu, S.; Bhat, S.A.; Kumar, A. Hydrogen sulfide-induced GAPDH sulfhydration disrupts the CCAR2-SIRT1 interaction to initiate autophagy. *Autophagy* 2021, 17, 3511–3529. [CrossRef] [PubMed]
- 33. Younossi, Z.M.; Koenig, A.B.; Abdelatif, D.; Fazel, Y.; Henry, L.; Wymer, M. Global epidemiology of nonalcoholic fatty liver disease-Meta-analytic assessment of prevalence, incidence, and outcomes. *Hepatology* **2016**, *64*, 73–84. [CrossRef] [PubMed]
- 34. Younossi, Z.; Anstee, Q.M.; Marietti, M.; Hardy, T.; Henry, L.; Eslam, M.; George, J.; Bugianesi, E. Global burden of NAFLD and NASH: Trends, predictions, risk factors and prevention. *Nat. Rev. Gastroenterol. Hepatol.* **2018**, *15*, 11–20. [CrossRef] [PubMed]
- Eslam, M.; Valenti, L.; Romeo, S. Genetics and epigenetics of NAFLD and NASH: Clinical impact. J. Hepatol. 2018, 68, 268–279. [CrossRef]
- Santhekadur, P.K.; Kumar, D.P.; Sanyal, A.J. Preclinical models of non-alcoholic fatty liver disease. J. Hepatol. 2018, 68, 230–237. [CrossRef] [PubMed]
- Buzzetti, E.; Pinzani, M.; Tsochatzis, E.A. The multiple-hit pathogenesis of non-alcoholic fatty liver disease (NAFLD). *Metabolism* 2016, 65, 1038–1048. [CrossRef] [PubMed]
- 38. Huang, P.L. A comprehensive definition for metabolic syndrome. Dis. Model. Mech. 2009, 2, 231–237. [CrossRef] [PubMed]
- 39. Simha, V. Management of hypertriglyceridemia. BMJ 2020, 371, m3109. [CrossRef] [PubMed]

- 40. Paiva, A.A.; Raposo, H.F.; Wanschel, A.C.; Nardelli, T.R.; Oliveira, H.C. Apolipoprotein CIII Overexpression-Induced Hypertriglyceridemia Increases Nonalcoholic Fatty Liver Disease in Association with Inflammation and Cell Death. *Oxid. Med. Cell. Longev.* **2017**, 2017, 1838679. [CrossRef]
- Esteve-Luque, V.; Padro-Miquel, A.; Fanlo-Maresma, M.; Corbella, E.; Corbella, X.; Pinto, X.; Candas-Estebanez, B. Implication between Genetic Variants from APOA5 and ZPR1 and NAFLD Severity in Patients with Hypertriglyceridemia. *Nutrients* 2021, 13, 552. [CrossRef] [PubMed]
- Chalasani, N.; Younossi, Z.; Lavine, J.E.; Charlton, M.; Cusi, K.; Rinella, M.; Harrison, S.A.; Brunt, E.M.; Sanyal, A.J. The diagnosis and management of nonalcoholic fatty liver disease: Practice guidance from the American Association for the Study of Liver Diseases. *Hepatology* 2018, 67, 328–357. [CrossRef] [PubMed]
- 43. Sun, L.; Zhang, S.; Yu, C.; Pan, Z.; Liu, Y.; Zhao, J.; Wang, X.; Yun, F.; Zhao, H.; Yan, S.; et al. Hydrogen sulfide reduces serum triglyceride by activating liver autophagy via the AMPK-mTOR pathway. *Am. J. Physiol. Endocrinol. Metab.* **2015**, *309*, E925–E935. [CrossRef] [PubMed]
- 44. Fu, X.M.; Xie, J.; Hu, Z.W. Mycelium of Hirsutella hepiali Chen et Shen activates autophagy and protects against metabolic syndrome in mice fed with high fat diet. *Yao Xue Xue Bao* **2014**, *49*, 615–621. [PubMed]
- 45. Shin, D.W. Lipophagy: Molecular Mechanisms and Implications in Metabolic Disorders. Mol. Cells 2020, 43, 686–693. [CrossRef]
- Wang, H.-G.; Zhang, Y.-J.; Huang, C.-S.; Chen, L.M.; Wang, J.; Wu, D.D.; Xie, X.Z.; Li, Z.I.; Ji, L.A. Effect of exogenous hydrogen sulfide on lipophagy in mouse primary hepatocytes. Chin. J. Pathophysiol. 2017, 33, 1901–1905.
- Wu, D.; Zhong, P.; Wang, J.; Wang, H. Exogenous hydrogen sulfide mitigates LPS + ATP-induced inflammation by inhibiting NLRP3 inflammasome activation and promoting autophagy in L02 cells. *Mol. Cell. Biochem.* 2019, 457, 145–156. [CrossRef] [PubMed]
- 48. Wang, H.; Zhong, P.; Sun, L. Exogenous hydrogen sulfide mitigates NLRP3 inflammasome-mediated inflammation through promoting autophagy via the AMPK-mTOR pathway. *Biol. Open* **2019**, *8*, bio043653. [CrossRef] [PubMed]
- Wu, D.; Zhong, P.; Wang, Y.; Zhang, Q.; Li, J.; Liu, Z.; Ji, A.; Li, Y. Hydrogen Sulfide Attenuates High-Fat Diet-Induced Non-Alcoholic Fatty Liver Disease by Inhibiting Apoptosis and Promoting Autophagy via Reactive Oxygen Species/Phosphatidylinositol 3-Kinase/AKT/Mammalian Target of Rapamycin Signaling Pathway. *Front. Pharmacol.* 2020, 11, 585860. [CrossRef] [PubMed]
- 50. Ferre, P.; Phan, F.; Foufelle, F. SREBP-1c and lipogenesis in the liver: An update1. Biochem. J. 2021, 478, 3723–3739. [CrossRef]
- Foretz, M.; Pacot, C.; Dugail, I.; Lemarchand, P.; Guichard, C.; Le Liepvre, X.; Berthelier-Lubrano, C.; Spiegelman, B.; Kim, J.B.; Ferre, P.; et al. ADD1/SREBP-1c is required in the activation of hepatic lipogenic gene expression by glucose. *Mol. Cell. Biol.* 1999, 19, 3760–3768. [CrossRef] [PubMed]
- Shimomura, I.; Bashmakov, Y.; Ikemoto, S.; Horton, J.D.; Brown, M.S.; Goldstein, J.L. Insulin selectively increases SREBP-1c mRNA in the livers of rats with streptozotocin-induced diabetes. *Proc. Natl. Acad. Sci. USA* 1999, 96, 13656–13661. [CrossRef] [PubMed]
- 53. Zhu, X.; Bian, H.; Wang, L.; Sun, X.; Xu, X.; Yan, H.; Xia, M.; Chang, X.; Lu, Y.; Li, Y.; et al. Berberine attenuates nonalcoholic hepatic steatosis through the AMPK-SREBP-1c-SCD1 pathway. *Free Radic. Biol. Med.* **2019**, *141*, 192–204. [CrossRef]
- Ziamajidi, N.; Khaghani, S.; Hassanzadeh, G.; Vardasbi, S.; Ahmadian, S.; Nowrouzi, A.; Ghaffari, S.M.; Abdirad, A. Amelioration by chicory seed extract of diabetes- and oleic acid-induced non-alcoholic fatty liver disease (NAFLD)/non-alcoholic steatohepatitis (NASH) via modulation of PPARalpha and SREBP-1. *Food Chem. Toxicol.* 2013, *58*, 198–209. [CrossRef] [PubMed]
- 55. Saleh Al-Maamari, J.N.; Rahmadi, M.; Panggono, S.M.; Prameswari, D.A.; Pratiwi, E.D.; Ardianto, C.; Balan, S.S.; Suprapti, B. The effects of quercetin on the expression of SREBP-1c mRNA in high-fat diet-induced NAFLD in mice. *J. Basic. Clin. Physiol. Pharmacol.* **2021**, *32*, 637–644. [CrossRef] [PubMed]
- Nguyen, T.T.P.; Kim, D.Y.; Lee, Y.G.; Lee, Y.S.; Truong, X.T.; Lee, J.H.; Song, D.K.; Kwon, T.K.; Park, S.H.; Jung, C.H.; et al. SREBP-1c impairs ULK1 sulfhydration-mediated autophagic flux to promote hepatic steatosis in high-fat-diet-fed mice. *Mol. Cell* 2021, *81*, 3820–3832.e7. [CrossRef] [PubMed]
- Farzanegi, P.; Dana, A.; Ebrahimpoor, Z.; Asadi, M.; Azarbayjani, M.A. Mechanisms of beneficial effects of exercise training on non-alcoholic fatty liver disease (NAFLD): Roles of oxidative stress and inflammation. *Eur. J. Sport Sci.* 2019, 19, 994–1003. [CrossRef]
- Van der Windt, D.J.; Sud, V.; Zhang, H.; Tsung, A.; Huang, H. The Effects of Physical Exercise on Fatty Liver Disease. *Gene Expr.* 2018, 18, 89–101. [CrossRef] [PubMed]
- 59. Romero-Gomez, M.; Zelber-Sagi, S.; Trenell, M. Treatment of NAFLD with diet, physical activity and exercise. *J. Hepatol.* **2017**, *67*, 829–846. [CrossRef]
- 60. Cuthbertson, D.J.; Shojaee-Moradie, F.; Sprung, V.S.; Jones, H.; Pugh, C.J.; Richardson, P.; Kemp, G.J.; Barrett, M.; Jackson, N.C.; Thomas, E.L.; et al. Dissociation between exercise-induced reduction in liver fat and changes in hepatic and peripheral glucose homoeostasis in obese patients with non-alcoholic fatty liver disease. *Clin. Sci.* **2016**, *130*, 93–104. [CrossRef]
- Shojaee-Moradie, F.; Cuthbertson, D.J.; Barrett, M.; Jackson, N.C.; Herring, R.; Thomas, E.L.; Bell, J.; Kemp, G.J.; Wright, J.; Umpleby, A.M. Exercise Training Reduces Liver Fat and Increases Rates of VLDL Clearance But Not VLDL Production in NAFLD. J. Clin. Endocrinol. Metab. 2016, 101, 4219–4228. [CrossRef] [PubMed]
- Martins, C.; Morgan, L.; Truby, H. A review of the effects of exercise on appetite regulation: An obesity perspective. *Int. J. Obes.* 2008, 32, 1337–1347. [CrossRef] [PubMed]

- 63. Lee, Y.H.; Jung, K.S.; Kim, S.U.; Yoon, H.J.; Yun, Y.J.; Lee, B.W.; Kang, E.S.; Han, K.H.; Lee, H.C.; Cha, B.S. Sarcopaenia is associated with NAFLD independently of obesity and insulin resistance: Nationwide surveys (KNHANES 2008-2011). *J. Hepatol.* 2015, 63, 486–493. [CrossRef] [PubMed]
- 64. Koo, B.K.; Kim, D.; Joo, S.K.; Kim, J.H.; Chang, M.S.; Kim, B.G.; Lee, K.L.; Kim, W. Sarcopenia is an independent risk factor for non-alcoholic steatohepatitis and significant fibrosis. *J. Hepatol.* **2017**, *66*, 123–131. [CrossRef] [PubMed]
- Gao, Y.; Zhang, W.; Zeng, L.Q.; Bai, H.; Li, J.; Zhou, J.; Zhou, G.Y.; Fang, C.W.; Wang, F.; Qin, X.J. Exercise and dietary intervention ameliorate high-fat diet-induced NAFLD and liver aging by inducing lipophagy. *Redox Biol.* 2020, 36, 101635. [CrossRef] [PubMed]
- Pi, H.; Liu, M.; Xi, Y.; Chen, M.; Tian, L.; Xie, J.; Chen, M.; Wang, Z.; Yang, M.; Yu, Z.; et al. Long-term exercise prevents hepatic steatosis: A novel role of FABP1 in regulation of autophagy-lysosomal machinery. *FASEB J.* 2019, 33, 11870–11883. [CrossRef] [PubMed]
- 67. Wang, B.; Zeng, J.; Gu, Q. Exercise restores bioavailability of hydrogen sulfide and promotes autophagy influx in livers of mice fed with high-fat diet. *Can. J. Physiol. Pharmacol.* **2017**, *95*, 667–674. [CrossRef] [PubMed]
- Temiz, C.; Solmaz, I.; Tehli, O.; Kaya, S.; Onguru, O.; Arslan, E.; Izci, Y. The effects of splenectomy on lipid peroxidation and neuronal loss in experimental spinal cord ischemia/reperfusion injury. *Turk. Neurosurg.* 2013, 23, 67–74. [CrossRef] [PubMed]
- 69. Usul, H.; Cakir, E.; Cobanoglu, U.; Alver, A.; Peksoylu, B.; Topbas, M.; Baykal, S. The effects of tyrphostine Ag 556 on experimental spinal cord ischemia reperfusion injury. *Surg. Neurol.* **2004**, *61*, 45–54; discussion 54. [CrossRef]
- Musayeva, A.; Unkrig, J.C.; Zhutdieva, M.B.; Manicam, C.; Ruan, Y.; Laspas, P.; Chronopoulos, P.; Gobel, M.L.; Pfeiffer, N.; Brochhausen, C.; et al. Betulinic Acid Protects from Ischemia-Reperfusion Injury in the Mouse Retina. *Cells* 2021, *10*, 2240. [CrossRef] [PubMed]
- Zhou, J.; Hu, M.; He, M.; Wang, X.; Sun, D.; Huang, Y.; Cheng, X.; Fu, J.; Cai, J.; Ma, T.; et al. TNFAIP3 Interacting Protein 3 Is an Activator of Hippo-YAP Signaling Protecting Against Hepatic Ischemia/Reperfusion Injury. *Hepatology* 2021, 74, 2133–2153. [CrossRef] [PubMed]
- Christou, C.D.; Tsoulfas, G. The Role of microRNA in Hepatic Ischemia/Reperfusion Injury. *Microrna* 2020, 9, 248–254. [CrossRef] [PubMed]
- Dusabimana, T.; Kim, S.R.; Kim, H.J.; Park, S.W.; Kim, H. Nobiletin ameliorates hepatic ischemia and reperfusion injury through the activation of SIRT-1/FOXO3a-mediated autophagy and mitochondrial biogenesis. *Exp. Mol. Med.* 2019, *51*, 1–16. [CrossRef] [PubMed]
- Xu, Y.; Tang, Y.; Lu, J.; Zhang, W.; Zhu, Y.; Zhang, S.; Ma, G.; Jiang, P.; Zhang, W. PINK1-mediated mitophagy protects against hepatic ischemia/reperfusion injury by restraining NLRP3 inflammasome activation. *Free Radic. Biol. Med.* 2020, 160, 871–886. [CrossRef] [PubMed]
- 75. Rezq, S.; Hassan, R.; Mahmoud, M.F. Rimonabant ameliorates hepatic ischemia/reperfusion injury in rats: Involvement of autophagy via modulating ERK- and PI3K/AKT-mTOR pathways. *Int. Immunopharmacol.* **2021**, *100*, 108140. [CrossRef] [PubMed]
- 76. Chen, M.; Du, Y.; Qui, M.; Wang, M.; Chen, K.; Huang, Z.; Jiang, M.; Xiong, F.; Chen, J.; Zhou, J.; et al. Ophiopogonin B-induced autophagy in non-small cell lung cancer cells via inhibition of the PI3K/Akt signaling pathway. Oncol. Rep. 2013, 29, 430–436. [CrossRef]
- 77. Shen, M.; Lu, J.; Dai, W.; Wang, F.; Xu, L.; Chen, K.; He, L.; Cheng, P.; Zhang, Y.; Wang, C.; et al. Ethyl pyruvate ameliorates hepatic ischemia-reperfusion injury by inhibiting intrinsic pathway of apoptosis and autophagy. *Mediat. Inflamm.* 2013, 2013, 461536. [CrossRef]
- 78. Cheng, P.; Wang, F.; Chen, K.; Shen, M.; Dai, W.; Xu, L.; Zhang, Y.; Wang, C.; Li, J.; Yang, J.; et al. Hydrogen sulfide ameliorates ischemia/reperfusion-induced hepatitis by inhibiting apoptosis and autophagy pathways. *Mediat. Inflamm.* 2014, 2014, 935251. [CrossRef]
- Ohnishi, K.; Komohara, Y.; Fujiwara, Y.; Takemura, K.; Lei, X.; Nakagawa, T.; Sakashita, N.; Takeya, M. Suppression of TLR4mediated inflammatory response by macrophage class A scavenger receptor (CD204). *Biochem. Biophys. Res. Commun.* 2011, 411, 516–522. [CrossRef]
- 80. Xu, Y.; Qian, L.; Zong, G.; Ma, K.; Zhu, X.; Zhang, H.; Li, N.; Yang, Q.; Bai, H.; Ben, J.; et al. Class A scavenger receptor promotes cerebral ischemic injury by pivoting microglia/macrophage polarization. *Neuroscience* **2012**, *218*, 35–48. [CrossRef]
- 81. Huang, H.; Li, X.; Zhuang, Y.; Li, N.; Zhu, X.; Hu, J.; Ben, J.; Yang, Q.; Bai, H.; Chen, Q. Class A scavenger receptor activation inhibits endoplasmic reticulum stress-induced autophagy in macrophage. *J. Biomed. Res.* **2014**, *28*, 213–221. [CrossRef]
- Ruan, Z.; Liang, M.; Deng, X.; Lai, M.; Shang, L.; Su, X. Exogenous hydrogen sulfide protects fatty liver against ischemiareperfusion injury by regulating endoplasmic reticulum stress-induced autophagy in macrophage through mediating the class A scavenger receptor pathway in rats. *Cell Biol. Int.* 2019, 44, 306–316. [CrossRef] [PubMed]
- Hartke, J.; Johnson, M.; Ghabril, M. The diagnosis and treatment of hepatocellular carcinoma. *Semin. Diagn. Pathol.* 2017, 34, 153–159. [CrossRef] [PubMed]
- 84. Sim, H.W.; Knox, J. Hepatocellular carcinoma in the era of immunotherapy. Curr. Probl. Cancer 2018, 42, 40–48. [CrossRef]
- Clark, T.; Maximin, S.; Meier, J.; Pokharel, S.; Bhargava, P. Hepatocellular Carcinoma: Review of Epidemiology, Screening, Imaging Diagnosis, Response Assessment, and Treatment. *Curr. Probl. Diagn. Radiol.* 2015, 44, 479–486. [CrossRef]
- Kulik, L.; El-Serag, H.B. Epidemiology and Management of Hepatocellular Carcinoma. *Gastroenterology* 2019, 156, 477–491.e1. [CrossRef] [PubMed]

- 87. Singal, A.G.; El-Serag, H.B. Hepatocellular Carcinoma From Epidemiology to Prevention: Translating Knowledge into Practice. *Clin. Gastroenterol. Hepatol.* **2015**, *13*, 2140–2151. [CrossRef] [PubMed]
- Howlader, N.; Noone, A.M.; Krapcho, M.; Garshell, J.; Miller, D.; Altekruse, S.F.; Kosary, C.L.; Yu, M.; Ruhl, J.; Tatalovich, Z.; et al. (Eds.) SEER Cancer Statistics Review, 1975–2010; National Cancer Institute: Bethesda, MD, USA, 2013.
- Li, Q.; Ni, Y.; Zhang, L.; Jiang, R.; Xu, J.; Yang, H.; Hu, Y.; Qiu, J.; Pu, L.; Tang, J.; et al. HIF-1alpha-induced expression of m6A reader YTHDF1 drives hypoxia-induced autophagy and malignancy of hepatocellular carcinoma by promoting ATG2A and ATG14 translation. *Signal Transduct. Target. Ther.* 2021, *6*, 76. [CrossRef]
- Sun, R.; Zhai, R.; Ma, C.; Miao, W. Combination of aloin and metformin enhances the antitumor effect by inhibiting the growth and invasion and inducing apoptosis and autophagy in hepatocellular carcinoma through PI3K/AKT/mTOR pathway. *Cancer Med.* 2020, 9, 1141–1151. [CrossRef]
- Wang, S.S.; Chen, Y.H.; Chen, N.; Wang, L.J.; Chen, D.X.; Weng, H.L.; Dooley, S.; Ding, H.G. Hydrogen sulfide promotes autophagy of hepatocellular carcinoma cells through the PI3K/Akt/mTOR signaling pathway. *Cell Death Dis.* 2017, *8*, e2688. [CrossRef] [PubMed]
- Huang, F.; Wang, B.R.; Wang, Y.G. Role of autophagy in tumorigenesis, metastasis, targeted therapy and drug resistance of hepatocellular carcinoma. *World. J. Gastroenterol.* 2018, 24, 4643–4651. [CrossRef] [PubMed]
- 93. Hellmich, M.R.; Szabo, C. Hydrogen Sulfide and Cancer. Handb. Exp. Pharmacol. 2015, 230, 233–241. [CrossRef] [PubMed]
- 94. Wu, D.; Si, W.; Wang, M.; Lv, S.; Ji, A.; Li, Y. Hydrogen sulfide in cancer: Friend or foe? Nitric Oxide 2015, 50, 38–45. [CrossRef]
- Kong, D.; Zhang, Z.; Chen, L.; Huang, W.; Zhang, F.; Wang, L.; Wang, Y.; Cao, P.; Zheng, S. Curcumin blunts epithelialmesenchymal transition of hepatocytes to alleviate hepatic fibrosis through regulating oxidative stress and autophagy. *Redox Biol.* 2020, 36, 101600. [CrossRef]
- 96. Xie, L.; Yu, S.; Yang, K.; Li, C.; Liang, Y. Hydrogen Sulfide Inhibits Autophagic Neuronal Cell Death by Reducing Oxidative Stress in Spinal Cord Ischemia Reperfusion Injury. *Oxid. Med. Cell. Longev.* **2017**, 2017, 8640284. [CrossRef]
- Honma, Y.; Sato-Morita, M.; Katsuki, Y.; Mihara, H.; Baba, R.; Hino, K.; Kawashima, A.; Ariyasu, T.; Harada, M. Trehalose alleviates oxidative stress-mediated liver injury and Mallor-Denk body formation via activating autophagy in mice. *Med. Mol. Morphol.* 2021, 54, 41–51. [CrossRef]
- Greabu, M.; Totan, A.; Miricescu, D.; Radulescu, R.; Virlan, J.; Calenic, B. Hydrogen Sulfide, Oxidative Stress and Periodontal Diseases: A Concise Review. *Antioxidants* 2016, 5, 3. [CrossRef]
- Guo, J.M.; Xing, H.J.; Cai, J.Z.; Zhang, H.F.; Xu, S.W. H2S exposure-induced oxidative stress promotes LPS-mediated hepatocyte autophagy through the PI3K/AKT/TOR pathway. *Ecotoxicol. Environ. Saf.* 2021, 209, 111801. [CrossRef] [PubMed]
- 100. Ge, X.; Sun, J.; Fei, A.; Gao, C.; Pan, S.; Wu, Z. Hydrogen sulfide treatment alleviated ventilator-induced lung injury through regulation of autophagy and endoplasmic reticulum stress. *Int. J. Biol. Sci.* **2019**, *15*, 2872–2884. [CrossRef] [PubMed]