- Mattheyse FJ, van Heerden K, Mattheyse M, et al. Reduced activation of peripheral blood neutrophils after late-phase asthmatic responses but not in mild stable asthma. *Respiration*. 2001;68(5):471-479.
- 9. PelikanZ. Delayed type of asthmatic response to allergen challenge and cytokines in the peripheral blood. *Respiration*. 2012;84(5):385-395.

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SUPPORTING INFORMATION

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Factors associated with hyperresponsiveness to adenosine 5'-monophosphate in healthy subjects

LETTER TO THE EDITORTo the editor

Airway hyperresponsiveness (AHR) is used to diagnose asthma. AHR can be assessed with a direct (eg, methacholine) or an indirect (eg, adenosine 5'-monophosphate (AMP)) test. Direct tests induce airway constriction by stimulating airway smooth muscle contraction. Indirect tests provoke mast cells and other inflammatory cells to release mediators, that cascade into airway constriction. AHR can be defined as the concentration of the stimulus that causes a 20% drop in forced expiratory volume in the first second (FEV₁)(PC₂₀). Although healthy subjects are expected to be unresponsive to AMP,¹ in a group of 108 healthy controls we observed 8 (7.4%) subjects with AHR to AMP (PC₂₀AMP ≤160 mg/mL). We therefore investigated which factors associate with AHR to AMP in these healthy subjects.

The NORM database (approved by the ethical committee of our hospital, NCT00848406, see Data S1) consists of healthy never or current smokers (>10 cigarettes per day). Subjects had no respiratory complaints (ie, cough or dyspnoea) nor any doctor's respiratory diagnosis, in the past or present. They presented with normal lung function (FEV₁ >80%_{predicted}, FEV₁/FVC >70%, and <10% increase in FEV₁%_{predicted} after 400 µg salbutamol) and the absence of AHR to methacholine (PC₂₀methacholine >19.8 mg/mL; Table 1). To investigate which factors associate with AHR to AMP, we compared subjects with AHR to AMP to those without (<160 mg/mL vs >160 mg/mL, Table 1) with a chi-square or Mann-Whitney U test. From each group of variables, we entered in a multivariate logistic regression model the variable which increased the model's adjusted- R^2 most (univariate P < 0.01 and mutual Spearman's correlation <|0.7|; Table 2, SPSS version 22).

We expected a relation with allergies based on a skin prick test (P = 0.59) and self-reported allergic rhinitis (P = 0.91), but this relation was absent. Two samples had sputum eosinophils above the commonly used threshold of 3%. Therefore, we analyzed the percentage of sputum eosinophils categorized as <1%, ≥1%, and missing (only 74 subjects were able to expectorate sputum). AHR was significantly associated with sputum eosinophils (Table 1). The multivariate model (Table 2) shows that AHR is more likely in subjects with ≥1% sputum eosinophils, but

equally likely in subjects with a missing sputum sample compared to those with <1% sputum eosinophils. This indicates that ≥1% sputum eosinophils in a healthy subject associates with AHR to AMP. This aligns with earlier observations in asthma and allergic rhinitis that sputum eosinophils predict AHR to AMP.² We now show the association in healthy subjects.

Strikingly, all subjects with AHR to AMP were current smokers. Smoking status was significantly different (P < 0.01) between subjects with and without AHR and smokers with AHR smoked more cigarettes per day than smokers without AHR (P < 0.01). Previously reported healthy subjects with AHR to AMP were also all current smokers.³ Furthermore, smoking is associated with AMP sensitivity in patients with chronic obstructive pulmonary disease (COPD),³ as COPD smokers had a significantly lower PC₂₀ AMP compared to COPD nonsmokers. Based on these findings, we speculate that AHR to AMP in healthy subjects is affected by cigarette smoke-induced presence of mast cells, the primary target for AMP. Cigarette smoke irritates epithelial cells, initiating the release of pro-inflammatory cytokines.⁴ These trigger mast cell infiltration and proliferation, resulting in increased numbers of mast cells in the airway's submucosa.⁵ Cigarette smoke also causes the reduction of epithelial integrity which facilitates the cigarette smoke to reach the mast cells.⁶ As AMP provocation initiates bronchoconstriction through mediators released by mast cells,⁷ healthy smokers may have AHR to AMP.

Next to smoking, AHR to AMP strongly associated with small airways function. Univariately, AHR to AMP associated with lower forced expiratory flow at 50%, 75%, and 25%-75% of the forced vital capacity (FVC) (FEF₅₀, FEF₇₅, and FEF₂₅₋₇₅, respectively), a higher resistance in the small airways (difference in resistance to 5 Hz and 20 Hz (R₅-R₂₀)), and lower lung compliance (increased resonance frequency (F_{res}) and reactance area (AX); all P < 0.03, Table 1). Multivariately, small airways function, in terms of R₅-R₂₀, was independently associated with AHR (Table 2). Previously, small airways function has been suggested to explain why some patients present with respiratory complaints, despite a normal FEV₁. It is possible that small airways anomalies are already important before respiratory complaints arise, as AHR to AMP might occur earlier than AHR to methacholine. Michils *et al*⁸ proposed that a discrepancy in the TABLE 1 Univariate analysis comparing subject with and without airway hyperresponsiveness to AMP

	AMP > 160 mg/mL (n = 100)	AMP ≤ 160 mg/mL (n = 8)	P-value
Gender (male-female)	57-43	4-4	7.0 × 10 ⁻¹
Age (years)	40.50 (22.0; 55.0)	43.00 (25.5; 57.5)	5.4×10^{-1}
Positive skin prick test (no-yes)	66-34	6-2	5.9×10^{-1}
Allergic rhinitis (self-reported) (no-yes)	86-14	7-1	9.1 × 10 ⁻¹
Smoking status (never-current)	55-45	0-8	$2.8 \times 10^{-3^*}$
Packyears (years)	0.3 (0.0; 10.5)	22.55 (7.0; 46.8)	$6.0 \times 10^{-4^*}$
Smokers - Packyears (years)	13.3 (2.9; 25.0)	22.55 (7.0; 46.8)	1.6×10^{-1}
Smokers - Equivalent number of cigarettes/day	14.0 (10.0; 18.0)	19.5 (17.3; 23.8)	9.5 × 10 ^{-3*}
Blood eosinophils (%)	2.4 (1.4; 3.4)	2.6 (2.0; 5.1)	2.7×10^{-1}
Preprovocation sputum eosinophils (categorized) (absent (<1%)−present (≥1%)−no sample)	60-7-33	3-4-1	$1.0 \times 10^{-3^*}$
Preprovocation sputum eosinophils (%) ^a	0.0 (0.0; 0.4)	1.0 (0.7; 2.9)	$2.7 \times 10^{-3^*}$
FEV _{1%} predicted	109 (101; 114)	100 (90; 110)	7.2 × 10 ⁻²
FEF _{25%} predicted	102 (92; 118)	104 (94; 119)	6.4×10^{-1}
FEV ₁ /FVC (%)	79.33 (76.1; 84.1)	74.98 (71.7; 83.6)	1.5×10^{-1}
R ₅ (kPa/sL)	0.30 (0.2; 0.4)	0.36 (0.3; 0.5)	9.1 × 10 ⁻²
R ₂₀ (kPa/sL)	0.28 (0.2; 0.3)	0.28 (0.2; 0.3)	8.8×10^{-1}
FEF _{50%} predicted	89 (77; 105)	72 (67; 83)	2.3×10^{-2}
FEF _{75%} predicted	73 (59; 94)	52 (42; 64)	$7.7 \times 10^{-3^*}$
FEF _{25-75%} predicted	88 (74; 103)	68 (63; 79)	1.1×10^{-2}
R ₅ -R ₂₀ (kPa/sL)	0.02 (0.0; 0.0)	0.11 (0.0; 0.1)	$2.7 \times 10^{-3^*}$
AX (kPa/L)	0.15 (0.1; 0.2)	0.35 (0.2; 1.1)	$2.8 \times 10^{-3^*}$
F _{res} (L/s ⁻¹)	9.77 (8.39; 11.42)	14.91 (10.71; 19.65)	$4.8 \times 10^{-3^*}$
TL _{CO} %predicted	92 (83; 101)	81 (75; 86)	$7.4 \times 10^{-3^*}$
K _{CO} %predicted	97 (89; 107)	92 (88; 94)	9.9 × 10 ⁻²
COPD Control Questionnaire (CCQ)	0.20 (0.1; 0.4)	0.60 (0.4;1.2)	$4.7 \times 10^{-3^*}$
Asthma Control Questionnaire (ACQ)	0.00 (0.0; 0.0)	0.00 (0.0;0.1)	2.7×10^{-2}
St. George Respiratory Questionnaire (SGRQ)	1.93 (1.1; 4.4)	7.65 (2.3; 16.7)	$1.0 \times 10^{-2^*}$

Note: Data are presented as count or median (inter quartile range (IQR)). Binominal data were analyzed with a chi-square test, while continuous data were analyzed with a Mann-Whitney U test.

Abbreviations: AMP= adenosine 5'-monophosphate; FEF₂₅, forced expiratory flow at 25% of FVC; FEF₅₀, forced expiratory flow at 50% of FVC; FEF₇₅, forced expiratory flow at 25% to 75% of FVC; R₅, resistance to 5 Hz; R₂₀, resistance to 20 Hz; R₅-R₂₀, difference in resistance to 5 Hz and 20 Hz; AX, reactance area; F_{res}, resonance frequency; TL_{C0}, transfer factor for carbon monoxide; K_{co}, TL_{C0} corrected for the alveolar volume; FEV₁, forced expiratory volume in the first second; FVC, forced vital capacity.

^an = 74, 67 without AHR and 7 with AHR.

 $^*P \leq 0.01.$

response to methacholine and AMP arises from the different areas the provocative agents trigger. They showed that AHR to methacholine originates more centrally in the lungs, where the targets for methacholine, the muscarinic receptors, have a higher density. By contrast, AHR to AMP originates from the most peripheral lung zone (pre-acinar airways), where in its submucosa mast cells are predominantly localized. Involvement of the most peripheral lung zones is furthermore indicated by the association between PC₂₀AMP <160 mg/mL and a lower diffusion capacity (T_{L,CO}), in both univariate and multivariate analysis (Table 1, 2). Diffusion capacity indirectly reflects the small airways via its association with acinar airways dysfunction (increased S_{acin}).⁹

Finally, AHR to AMP associated with a lower quality of life. Univariately, hyperresponsive subjects scored higher on the St. George respiratory questionnaire (SGRQ) and asthma and COPD control questionnaires (ACQ and CCQ, respectively; Table 1). Nevertheless, all scores were below the clinically used thresholds of ≤ 25 , ≤ 1.5 , and ≤ 1 , respectively, indicating that our subjects have few symptoms. Multivariately, the SGRQ was independently associated with AHR to AMP (Table 2) indicating an association with lower quality of life. From this, we speculate that AHR to AMP in healthy

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TABLE 2 Multivariate logistic regression model predicting

 airway hyperresponsiveness to AMP

	В	Standard error	P-value		
Equivalent number of cigarettes/day ^a	0.224	0.134	9.6 × 10 ⁻²		
R ₅ -R ₂₀ ^b	0.299	0.147	4.1×10^{-2}		
TL _{CO} %predicted	-0.060	0.059	3.1×10^{-1}		
Preprovocation sputum eosinophils					
Absent(<1%)	-	-	-		
Present (≥1%)	3.016	1.655	6.8×10^{-2}		
Missing (no sample)	-4.084	3.074	1.8×10^{-1}		
St. George Respiratory Questionnaire (SGRQ)	0.544	0.256	3.4 × 10 ⁻²		

Note: The logistic regression model was obtained by including the variable which improved the model's adjusted- R^2 most (univariate $P \le 0.01$ and mutual Spearman's correlations <|0.7|), from each group of variables in Table 1. This model has a Cox & Snellen R^2 of 30.4% and 96.3% correctly predicted (SPSS version 22).

Abbreviations: R_5 - R_{20} , difference in resistance to 5 and 20 Hz; TL_{CO} , transfer factor for carbon monoxide.

^aBased on all subject (never and current smokers); current and never smokers.

^bincorporated in the multivariate regression model on 0.01 scale.

subjects may be an early predictor for development of pulmonary complaints.

In conclusion, in a group of healthy never and current smokers, eight subjects (7.4%) expressed AHR to AMP ($PC_{20}AMP \le 160 \text{ mg/}$ mL). These subjects were current smokers with a higher cigarette consumption compared to subjects without AHR. Furthermore, AHR to AMP associated with a reduced small airways function, higher sputum eosinophil levels, and a lower quality of life. Further research should concentrate on whether AHR to AMP in healthy smokers indicates the onset of respiratory disease development.

CONFLICT OF INTEREST

C.A. Cox, J.M. Vonk, and H.A.M. Kerstjens have nothing to disclose. Maarten van den Berge reports grants paid to the University from Astra Zeneca, TEVA, GSK, Chiesi, outside the submitted work.

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REFERENCES

- Cushley MJ, Tattersfield AE, Holgate ST. Inhaled adenosine and guanosine on airway resistance in normal and asthmatic subjects. Br J Clin Pharmacol. 1983;15:161-165.
- Van den Berge M, Meijer RJJ, Kerstjens HAM, de Reus DM, Koëter GHH, Kaufmann HF, et al. PC 20 Adenosine 5 ' -Monophosphate Is More Closely Associated with Airway Inflammation in Asthma Than PC 20 Methacholine. *Am J Respir Crit Care Med*. 2001;163:1546-1550.
- Oosterhoff Y, de Jong JW, Jansen MA, Koëter GH, Postma DS. Airway responsiveness to adenosine 5'-monophosphate in chronic obstructive pulmonary disease is determined by smoking. *Am Rev Respir Dis.* 1993;147:553-558.
- Wei XM, Kim HS, Kumar RK, Heywood GJ, Hunt JE, McNeil HP, et al. Effects of cigarette smoke on degranulation and NO production by mast cells and epithelial cells. *Respir Res.* 2005;6:108.
- 5. Pesci A, Rossi GA, Bertorelli G, Aufiero A, Zanon P, Olivieri D. Mast cells in the airway lumen and bronchial mucosa of patients with chronic bronchitis. *Am J Respir Crit Care Med*. 1994;149:1311-1316.
- Heijink IH, Brandenburg SM, Postma DS, van Oosterhout AJM. Cigarette smoke impairs airway epithelial barrier function and cellcell contact recovery. *Eur Respir J.* 2012;39:419-428.
- Joos GF, O'Connor B, Anderson SD, Chung F, Cockcroft DW, Dahlén B, et al. Indirect airway challenges. *Eur Respir J.* 2003;21:1050-1068.
- Michils A, Elkrim Y, Haccuria A, Van Muylem A. Adenosine 5'-monophosphate challenge elicits a more peripheral airway response than methacholine challenge. J Appl Physiol. 2011;110:1241-1247.
- Jarenbäck L, Ankerst J, Bjermer L, Tufvesson E. Acinar ventilation heterogeneity in COPD relates to diffusion capacity, resistance and reactance. *Respir Med.* 2016;110:28-33.

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