

# Draft Genome Sequence of *Porphyromonas gingivalis* Strain Ando Expressing a 53-Kilodalton-Type Fimbrilin Variant of Mfa1 Fimbriae

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**Periodontopathic *Porphyromonas gingivalis* strain Ando abundantly expresses a 53-kDa-type Mfa1 fimbria. Here, we report the draft genome sequence of Ando, with a size of 2,229,994 bp, average G+C content of 48.4%, and 1,755 predicted protein-coding sequences.**

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*Porphyromonas gingivalis*, a Gram-negative anaerobe, is a major contributor to periodontal diseases (1, 2). *P. gingivalis* expresses two types of fimbriae, FimA and Mfa1 (3), and it is known that there are six genotypes in genes encoding the major fimbrilin of FimA fimbriae (4). Recently, we reported that there were variants even in the major fimbrilin of Mfa1 fimbriae, the Mfa1 (75-kDa) and 53-kDa types (5). The published complete genome sequences of *P. gingivalis* strains, including W83 (6), ATCC 33277 (7), TDC60 (8), and HG66 (9), show that they all possess a DNA sequence corresponding to the gene encoding a 75-kDa fimbrilin. Here, we report the draft genome sequence of *P. gingivalis* strain Ando, which abundantly expresses a 53-kDa-type Mfa1 fimbria.

The genomic DNA of Ando was sequenced using the Illumina HiSeq 2000 (90- or 100-bp paired-end reads, with an average 186-bp insert size). After the raw sequences were trimmed and their quality filtered (Sanger QV,  $\geq 10$ ), the remaining 5,247,742 reads, with approximately 224-fold genome coverage, were assembled *de novo* using Velvet 1.2.08 (the best *k*-mer, 91 bp). The final draft assembly consists of 112 contigs ( $>180$  bp), for a total length of 2,229,994 bp ( $N_{50}$ , 55,724 bp) and a G+C content of 48.4%. Protein-coding sequences (CDSs), tRNA genes, and clustered regularly interspaced short palindromic repeats (CRISPRs) were predicted by MetaGeneAnnotator (10), tRNAscan-SE 1.23 (11), and CRISPRFinder (12), respectively. Functional annotation of CDSs comes from BLASTP searches against NCBI's nonredundant (NR) protein database (<ftp://ftp.ncbi.nlm.nih.gov/blast/db/FASTA/nr.gz>).

A total of 1,755 CDSs, 47 tRNA genes, 4 rRNA genes, and 3 CRISPRs were predicted in the Ando genome. PHAST (13) did not detect any prophage region. A comparative analysis of the CDSs among Ando and the other four *P. gingivalis* strains using CD-HIT 4.6.4 (14) (cutoff, 90% sequence similarity, 90 to 110% length coverage) showed significant similarities, with 87.7% similarity to TDC60, 86.2% similarity to HG66, 85.1% similarity to W83, and 85.1% similarity to ATCC 33277. A reciprocal best hit analysis of Ando chromosomal CDSs against W83 chromosomal

CDSs (6) using BLASTP (*E* value cutoff,  $10^{-10}$ ) predicted potential virulence genes in Ando (described as numbers in Ando out of numbers in W83): 1 of 1 hemolysin, 9 of 13 adhesin (e.g., hemagglutinin), 62 of 67 evasion proteins (e.g., glycosyltransferase), 10 of 10 invasion proteins, 16 of 16 stress response proteins, 5 of 5 antibiotic resistance proteins, and 40 of 41 peptidases. The gene encoding the 53-kDa fimbrilin (PGANDO\_1061) in Ando was not found in the other four *P. gingivalis* strains. However, the PGANDO\_1061 gene was located at the same locus of the *mfa1* gene in other strains. We found several other genes that were specifically detected in Ando (e.g., plasmid-related proteins, CRISPR-associated proteins, and virulence-associated protein E). Further comparative genomic and functional analyses with other strains that express 53-kDa fimbrilins will help understand the pathogenic mechanism of *P. gingivalis*.

**Nucleotide sequence accession numbers.** This genome sequence and raw sequence reads have been deposited, respectively, in DDBJ/ENA/GenBank and DDBJ Sequence Read Archive under the accession numbers BCBV01000001 to BCBV01000112 and DRA003978 (BioProject PRJDB4201).

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## REFERENCES

- Darveau RP, Hajishengallis G, Curtis MA. 2012. *Porphyromonas gingivalis* as a potential community activist for disease. *J Dent Res* 91:816–820. <http://dx.doi.org/10.1177/0022034512453589>.
- Socransky SS, Haffajee AD. 2005. Periodontal microbial ecology. *Periodontol* 2000 38:135–187. <http://dx.doi.org/10.1111/j.1600-0757.2005.00107.x>.
- Yoshimura F, Murakami Y, Nishikawa K, Hasegawa Y, Kawaminami S. 2009. Surface components of *Porphyromonas gingivalis*. *J Periodontol Res* 44:1–12. <http://dx.doi.org/10.1111/j.1600-0765.2008.01135.x>.

4. Amano A, Nakagawa I, Okahashi N, Hamada N. 2004. Variations of *Porphyromonas gingivalis* fimbriae in relation to microbial pathogenesis. *J Periodontol Res* 39:136–142. <http://dx.doi.org/10.1111/j.1600-0765.2004.00719.x>.
5. Nagano K, Hasegawa Y, Yoshida Y, Yoshimura F. 2015. A major fimbriin variant of Mfa1 fimbriae in *Porphyromonas gingivalis*. *J Dent Res* 94:1143–1148. <http://dx.doi.org/10.1177/0022034515588275>.
6. Nelson KE, Fleischmann RD, DeBoy RT, Paulsen IT, Fouts DE, Eisen JA, Daugherty SC, Dodson RJ, Durkin AS, Gwinn M, Haft DH, Kolonay JF, Nelson WC, Mason T, Tallon L, Gray J, Granger D, Tettelin H, Dong H, Galvin JL, Duncan MJ, Dewhirst FE, Fraser CM. 2003. Complete genome sequence of the oral pathogenic bacterium *Porphyromonas gingivalis* strain W83. *J Bacteriol* 185:5591–5601. <http://dx.doi.org/10.1128/JB.185.18.5591-5601.2003>.
7. Naito M, Hirakawa H, Yamashita A, Ohara N, Shoji M, Yukitake H, Nakayama K, Toh H, Yoshimura F, Kuhara S, Hattori M, Hayashi T, Nakayama K. 2008. Determination of the genome sequence of *Porphyromonas gingivalis* strain ATCC 33277 and genomic comparison with strain W83 revealed extensive genome rearrangements in *P. gingivalis*. *DNA Res* 15:215–225. <http://dx.doi.org/10.1093/dnares/dsn013>.
8. Watanabe T, Maruyama F, Nozawa T, Aoki A, Okano S, Shibata Y, Oshima K, Kurokawa K, Hattori M, Nakagawa I, Abiko Y. 2011. Complete genome sequence of the bacterium *Porphyromonas gingivalis* TDC60, which causes periodontal disease. *J Bacteriol* 193:4259–4260. <http://dx.doi.org/10.1128/JB.05269-11>.
9. Siddiqui H, Yoder-Himes DR, Mizgalska D, Nguyen K-, Potempa J, Olsen I. 2014. Genome sequence of *Porphyromonas gingivalis* strain HG66 (DSM 28984). *Genome Announc* 2(5):e00947-14. <http://dx.doi.org/10.1128/genomeA.00947-14>.
10. Noguchi H, Taniguchi T, Itoh T. 2008. MetaGeneAnnotator: detecting species-specific patterns of ribosomal binding site for precise gene prediction in anonymous prokaryotic and phage genomes. *DNA Res* 15:387–396. <http://dx.doi.org/10.1093/dnares/dsn027>.
11. Lowe TM, Eddy SR. 1997. tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence. *Nucleic Acids Res* 25:955–964. <http://dx.doi.org/10.1093/nar/25.5.0955>.
12. Grissa I, Vergnaud G, Pourcel C. 2007. CRISPRFinder: a Web tool to identify clustered regularly interspaced short palindromic repeats. *Nucleic Acids Res* 35:W52–W57. <http://dx.doi.org/10.1093/nar/gkm360>.
13. Zhou Y, Liang Y, Lynch KH, Dennis JJ, Wishart DS. 2011. PHAST: a fast phage search tool. *Nucleic Acids Res* 39:W347–W352. <http://dx.doi.org/10.1093/nar/gkr485>.
14. Li W, Godzik A. 2006. CD-HIT: a fast program for clustering and comparing large sets of protein or nucleotide sequences. *Bioinformatics* 22:1658–1659. <http://dx.doi.org/10.1093/bioinformatics/btl158>.