

Article



# A Comprehensive Analysis of Argonaute-CLIP Data Identifies Novel, Conserved and Species-Specific Targets of miR-21 in Human Liver and Hepatocellular Carcinoma

Aaron Balasingam Koenig <sup>1,2,†</sup>, Juan Martín Barajas <sup>1,†</sup>, María Jose Guerrero <sup>1</sup> and Kalpana Ghoshal <sup>1,\*</sup>

- <sup>1</sup> Department of Pathology, Comprehensive Cancer Center, The Ohio State University, Columbus, OH 43210, USA; aaron.koenig@osumc.edu (A.B.K.); barajas.10@osu.edu (J.M.B.); guerrero.41@buckeyemail.osu.edu (M.J.G.)
- <sup>2</sup> Medical Student Research Program, The Ohio State University College of Medicine, Columbus, OH 43210, USA
- \* Correspondence: kalpana.ghoshal@osumc.edu; Tel.: +1-614-292-8865
- + These authors contributed equally to this work.

Received: 24 January 2018; Accepted: 13 March 2018; Published: 14 March 2018

MicroRNAs are ~22 nucleotide RNAs that regulate gene expression at the Abstract: post-transcriptional level by binding messenger RNA transcripts. miR-21 is described as an oncomiR whose steady-state levels are commonly increased in many malignancies, including hepatocellular carcinoma (HCC). Methods known as cross-linking and immunoprecipitation of RNA followed by sequencing (CLIP-seq) have enabled transcriptome-wide identification of miRNA interactomes. In our study, we use a publicly available Argonaute-CLIP dataset (GSE97061), which contains nine HCC cases with matched benign livers, to characterize the miR-21 interactome in HCC. Argonaute-CLIP identified 580 miR-21 bound target sites on coding transcripts, of which 332 were located in the coding sequences, 214 in the 3'-untranslated region, and 34 in the 5'-untranslated region, introns, or downstream sequences. We compared the expression of miR-21 targets in 377 patients with liver cancer from the data generated by The Cancer Genome Atlas (TCGA) and found that mRNA levels of 402 miR-21 targets are altered in HCC. Expression of three novel predicted miR-21 targets (CAMSAP1, DDX1 and MARCKSL1) correlated with HCC patient survival. Analysis of RNA-seq data from SK-Hep1 cells treated with a miR-21 antisense oligonucleotide (GSE65892) identified RMND5A, an E3 ubiquitin ligase, as a strong miR-21 candidate target. Collectively, our analysis identified novel miR-21 targets that are likely to play a causal role in hepatocarcinogenesis.

Keywords: miRNA; miR-21; liver cancer; HCC

#### 1. Introduction

Liver cancer is the second leading cause of cancer-related death in the world [1]. Hepatocellular carcinoma (HCC) is a tumor of epithelial origin that typically arises in patients with underlying liver disease [2]. Risk factors for HCC are chronic infection with hepatitis B or C viruses, diabetes, alcohol and nonalcoholic steatohepatitis, exposure to carcinogens such as aflatoxin B<sub>1</sub> and genetic diseases such as hereditary hemochromatosis,  $\alpha_1$ -anti-trypsin deficiency or tyrosinemia [2]. Chronic liver inflammation, oxidative stress, and hypoxia associated with the underlying disease lead to the accumulation of oncogenic driver mutations eventually progressing to HCC [3–5].

MicroRNAs are small RNAs (18 to 25 nucleotides) that regulate gene expression at the post-transcriptional level typically by imperfectly base-pairing with the 3' untranslated region (3'UTR)

of mRNA in association with the RNA-induced silencing complex (RISC) [6]. This interaction represses overall protein levels by inducing mRNA destabilization or inhibiting its translation [6]. MicroRNA-21 (miR-21) is a conserved and ubiquitously expressed miRNA coded from the 3'UTR of *TMEM49* gene on chromosome 17 in humans [7]. However, transcription of primary miR-21 is regulated by a promoter distinct from that of *TMEM49*. Overexpression of miR-21 contributes to disease progression in many hematologic malignancies and solid tumors [8–10]. In primary HCC tissues, upregulation of miR-21 expression correlates with tumor grade [11]. In diseased liver, inflammatory cytokines stimulate transcription of *MIR21* through transcription factors STAT3 and AP-1 [12–14]. miR-21, in turn, suppresses the expression of tumor suppressors such as phosphatase and tensin homolog (PTEN) [15],

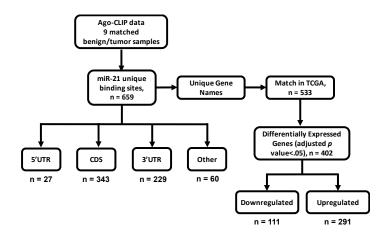
programmed cell death protein 4 (PDCD4) [16–18], reversion-inducing-cysteine-rich protein with kazal motifs (RECK), and metalloproteinase inhibitor 3 (TIMP3) by direct targeting [19]. miR-21 has been shown to be upregulated in primary human HCCs [15]. However, identity of miR-21 targets and their regulation in the transcriptome of tumors and host livers have not been described. Recently, our laboratory performed high-throughput sequencing of RNA isolated by cross-linking and immunoprecipitation of Argonaute (Ago-CLIP) [20] to identify microRNAs and their target mRNAs in mouse livers and human livers with their matching HCC tumor. We recently

reported significant enrichment of miR-21 and its targetome in HCCs compared to the benign livers [21]. Here, we expand on our previous findings and perform a transcriptome-wide characterization of the miR-21 interactome in primary human HCCs and benign livers using data from our previous Ago-CLIP analysis (GSE97061) [21]. Many novel targets were identified whose functional roles in hepatocarcinogenesis require further study.

# 2. Results

#### 2.1. miR-21 Binds Broadly within Liver and HCC Transcriptomes

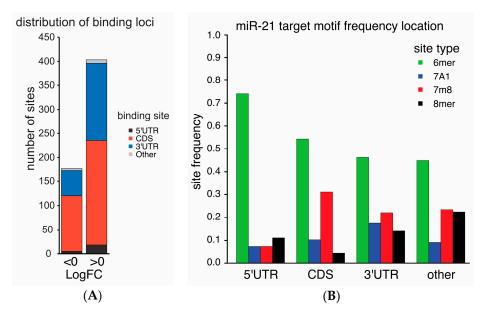
Recent Ago-CLIP analysis (GSE97061) of tumor and benign liver tissues from nine HCC patients identified miR-21 along with 659 of its targets associated with Argonaute. We classified 581 miR-21 binding sites in gene targets based on their enrichment in HCC and normal liver (Figure 1). Overall, 69.5% of miR-21 targets were enriched in HCCs. The majority of miR-21 binding sites enriched in HCC or in liver were located in either the coding sequence (CDS) (218/404 sites with logFC > 0 in HCC, 115/177 sites with logFC < 0 in HCC) or 3'UTR (161/404 sites with logFC > 0 in HCC, 53/177 sites with logFC < 0 in HCC, Figure 2A).



**Figure 1.** Workflow for the selection and analysis of microRNA-21 (miR-21) targets. Unique miR-21 targets (n = 659) were identified using high-throughput sequencing of RNA isolated by cross-linking and immunoprecipitation of Argonaute (Ago-CLIP) data generated from matching benign liver and tumor tissues isolated from nine hepatocellular carcinoma (HCC) patients (GSE97061) [21]. Targets were sorted based on annotation of target loci: 5' untranslated region [5'UTR], coding sequence [CDS], 3' untranslated

region [3'UTR], or other. Other groups included were transposable elements, introns, etc. Unique targets were compared to RNA-seq data generated by The Cancer Genome Atlas (TCGA) after  $log_2$  transformation and differential expression analysis between benign tissue and tumor tissue using the limma package in R [22]. Significance for differentially expressed genes was defined as having an adjusted *p*-value < 0.05 (*n* = 402).

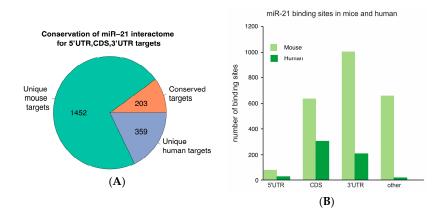
MicroRNA binding affinity varies by target motifs among 6mer, 7merA1, 7merM8, and 8mer sites by the degree of base pairing [23]. The distribution of miR-21 target motifs varied between coding sequences (CDS) and the 3'UTR ( $p = 2.1 \times 10^{-6}$ ), with the CDS having a paucity of 8mer binding sites compared to its binding sites in the 3'UTR (Figure 2B). Overall, 6mer sites were the most common motif.



**Figure 2.** Binding sites for miR-21 are most commonly in the CDS and 3'UTR of mRNA with variation in binding motif frequency. miR-21 binding sites identified by Ago-CLIP were classified by the sign of their log-fold enrichment in HCC and by the location of miR-21 binding on the target transcript: 5' untranslated region, CDS, 3'UTR or other (**A**). The binding sites of miR-21 were classified by the frequency of the target sequence motif (6mer, 7A1, 7m8, 8mer) at each annotated location (**B**). The distribution of sequence motifs varied between the CDS and the 3'UTR ( $p = 2.1 \times 10^{-6}$ ).

## 2.2. Conservation of miR-21 Targets between Mice and Humans

To determine the conservation of miR-21 binding sites, we compared gene identifiers from mouse liver transcripts identified by Ago-CLIP that interact with miR-21 (GSE97058) with human liver and HCC Ago-CLIP data. Among 562 genes with miR-21 binding sites in the 5' untranslated region (5'UTR), CDS or 3'UTR, 359 were unique to humans (Figure 3A). miR-21 binding sites in mouse livers were predominantly located in the 3'UTR whereas those in human livers or HCCs were more frequently found in the CDS (Figure 3B). An additional 1452 targets were identified in mouse livers (Supplementary Materials).



**Figure 3.** A low number of miR-21 sites are conserved among mice and humans. Targets in human livers or HCCs that precipitated miR-21 in Ago-CLIP (GSE97061) were compared to homolog gene identifiers from Ago-CLIP in mouse liver (GSE97058). Targets contained at least one binding site for miR-21 in the 5'UTR, CDS or 3'UTR (**A**). The number of binding sites at different locations in the mouse and human transcriptomes was compared (**B**).

#### 2.3. Certain Transcripts in the miR-21 Interactome Are Less Abundant in HCC

Matching of gene identifiers to RNA-seq data from 377 HCC patients and 59 normal livers in The Cancer Genome Atlas (TCGA) produced 533 gene matches with RNA-seq expression data. Of 402 miR-21 targets with significant change in expression in HCC, only 111 genes (27.6%) were downregulated, while 291 (72.4%) were upregulated (Supplementary Materials). Genes in the miR-21 interactome that were downregulated in HCC include *SPRY2* and *MAP2K3* that were previously validated as miR-21 targets (Table 1) [24–26]. Notably, several genes interacting with miR-21 were upregulated in HCC (Table 2).

**Table 1.** Association between miR-21 expression and components of miR-21 interactome dysregulatedafter anti-sense miR-21 treatment in SK-Hep-1 cells.

Gene Name	Full Name	miR-21 Expression	
		Correlation Coefficient	Adjusted <i>p</i> -Value
ANKRD46	Ankyrin repeat domain 46	-0.36509	$5.30  imes 10^{-12}$
ARL1	ADP ribosylation factor like GTPase 1	0.041021	1
ARRDC3	Arrestin-domain containing 3	-0.21111	$3.96 \times 10^{-3}$
CAMSAP2	Calmodulin-regulated spectrin-associated protein family member 2	Not defined *	Not defined *
CREB3L2	cAMP responsive element binding protein 3 like 2	0.061419	1
DDAH1	Dimethylarginine dimethylaminohydrolase 1	-0.340671	$3.32  imes 10^{-10}$
DDX1	DEAD-box helicase 1	0.090883	1
DSTYK	Dual serine/threonine and tyrosine protein kinase	0.237187	$3.07 imes10^{-4}$
EGLN1	Egl-9 family hypoxia inducible factor 1	0.12549213	1
GBP1	Guanylate-binding protein 1	-0.0315479	1
GGCX	Gamma-glutamyl carboxylase	-0.2833242	$1.38  imes 10^{-6}$
MARCKSL1	MARCKS like 1	0.27395541	$4.52 \times 10^{-6}$
PAG1	Phosphoprotein membrane anchor with glycosphingolipid microdomains 1	0.23283137	$4.86 imes10^{-4}$
PDZD8	PDZ domain containing 8	-0.0632384	1
PLEKHA1	Pleckstrin homology domain containing A1	-0.2021841	$8.88 imes10^{-3}$
RMND5A	Required for meiotic nuclear division 5 homolog A	-0.5028372	0
SLC46A3	Solute carrier family 46 member 3	-0.4672836	0
SMARCE1	SWI/SNF related, matrix associated, actin dependent regulator of chromatin, subfamily e, member 1	0.3145013	$1.88  imes 10^{-8}$
SPG20	Spastic paraplegia 20 (Troyer syndrome)	-0.2245667	$1.11  imes 10^{-3}$
TBC1D4	TBC1 domain family member 4	-0.0978456	1
TGFBR2	Transforming growth factor $\beta$ receptor 2	-0.1914894	0.0217
THBS1	Thrombospondin 1	-0.1524996	0.391
TIMP3	TIMP metallopeptidase inhibitor 3	-0.1720478	0.0994
TPRG1L	Tumor protein p63 regulated 1 like	-0.336802	$6.19 imes10^{-10}$

\* Gene identifier could not be matched in expression data.

Come Norme	Overall Survival		
Gene Name	Cox Coefficient	Adjusted <i>p</i> -Value	
DDX1	0.341	0.012	
MARCKSL1	0.324	0.0171	
CAMSAP2	0.282	0.0256	
PAG1	0.214	0.147	
ARRDC3	0.121	0.408	
DSTYK	0.109	0.48	
TBC1D4	0.097	0.543	
SPG20	0.05	0.802	
SMARCE1	0.04	0.836	
ARL1	0.033	0.871	
CREB3L2	0.017	0.936	
PLEKHA1	0.013	0.954	
DDAH1	-0.006	0.976	
EGLN1	-0.021	0.918	
TPRG1L	-0.037	0.866	
THBS1	-0.042	0.836	
GBP1	-0.069	0.687	
PDZD8	-0.075	0.645	
GGCX	-0.099	0.514	
ANKRD46	-0.11	0.505	
RMND5A	-0.104	0.493	
TGFBR2	-0.136	0.365	
TIMP3	-0.212	0.135	
SLC46A3	-0.214	0.151	

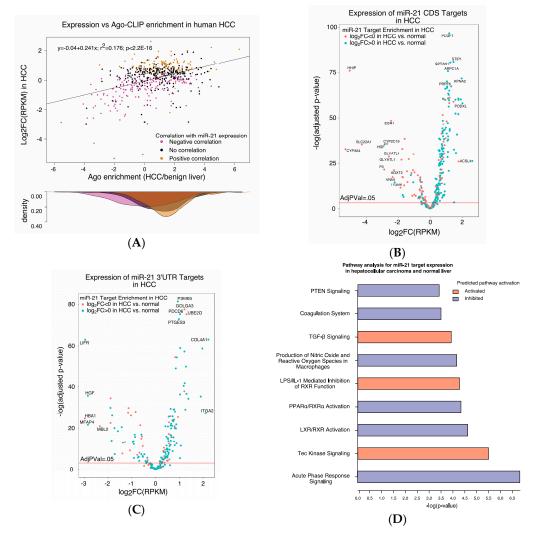
Table 2. Expression of the miR-21 interactome and overall survival in 300 HCC patients.

#### 2.4. Depletion of the miR-21 Interactome in Argonaute Complexes in HCC Is Associated with Repression

We then considered the outcome of miR-21 binding to the HCC transcriptome. We compared the Argonaute enrichment of miR-21 binding partners in HCC relative to normal tissue to their mRNA levels in HCC relative to normal tissue. The target enrichment score for Ago binding in HCC positively correlated with overexpression of the target in HCC relative to liver ( $p < 2.2 \times 10^{-16}$ ). In HCC, transcripts in the miR-21 interactome whose abundance in tumors directly correlated with miR-21 expression ( $p < 2.2 \times 10^{-16}$ ) (Figure 4A). Among the transcripts that interacted with miR-21, cytochrome 3A4, encoded by *CYP3A4*, and hedgehog inhibitory protein, encoded by *HHIP*, had an over 32-fold decrease in transcript abundance in HCC (Figure 4B,C). The expressions of miR-21 targets with binding sites outside of the CDS and 3'UTR are shown in Figure S1A,B, respectively.

## 2.5. The miR-21 Interactome Is Predicted to Regulate Tumor Metabolism

We subjected expression data of miR-21 targets from 377 HCC cases and 59 normal livers to Ingenuity Pathway Analysis (IPA). Signaling pathways predicted to be regulated in HCC by miR-21 targets include inhibition of acute phase response signaling, the liver X receptor/retinoid X receptor (LXR/RXR) pathway, the peroxisome proliferator-activated receptor alpha/retinoid X receptor alpha (PPAR $\alpha$ /RXR $\alpha$ ) pathway, phosphatase and tensin homolog (PTEN) signaling, and upregulation of Tec kinase signaling (Adjusted *p*-value < 0.0005, Figure 4D). The acute phase response is a systemic reaction to infection, neoplasm, tissue damage, or in inflammatory disorders that is associated with a change in hepatic metabolism [27]. Both PPAR $\alpha$ /RXR $\alpha$  and LXR/RXR are ligand-activated transcription factors involved in regulation of liver metabolism, including fatty acid metabolism [28]. PTEN modulates cell cycle progression and cell survival by dephosphorylating phosphatidylinositol (3,4,5)-triphosphate and acting as a negative regulator of Akt signaling [15]. Tec kinases are non-receptor tyrosine kinases



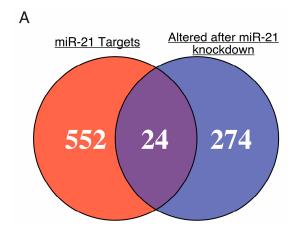
that regulate the development and activation of lymphoid and myeloid cells and play a role in liver regeneration [29–31].

**Figure 4.** The expression data of the miR-21 interactome in tumor (n = 377) and matched normal tissue (n = 59) in patients with liver cancer were extracted from TCGA and compared to the tumor enrichment score determined by Ago-CLIP (**A**). Log<sub>2</sub>-fold enrichment of miR-21 targets by Ago-CLIP in HCC positively correlated with log<sub>2</sub>-fold expression change in tumor ( $p < 2.2 \times 10^{-16}$ ). Genes whose expression in tumors negatively correlated with miR-21 expression were less enriched in HCC by Ago-CLIP than those with positive correlation ( $p < 2.2 \times 10^{-16}$ ). Among miR-21 CDS targets, 64 genes were significantly downregulated and 173 genes were significantly upregulated (**B**). Among miR-21 3'UTR targets, 48 genes were significantly downregulated and 173 genes were significantly upregulated (**C**). Expression data for miR-21 targets in HCC were subjected to Ingenuity Pathway Analysis (IPA). Pathways predicted to be activated or inhibited by the miR-21 targetome are shown (**D**). Pathway significance was defined as having an adjusted *p*-value < 0.0005.

#### 2.6. An Unbiased Approach Identified RMND5A as a Potential miR-21 Target

We then focused on genes in the miR-21 interactome for further validation. We compared the miR-21 target list from Ago-CLIP with aberrantly expressed genes from published microarray data (GSE65892) in SK-Hep-1, a human endothelial cell line derived from the ascites fluid of a patient with liver adenocarcinoma [32], depleted of miR-21 with anti-sense oligo [33]. We identified 24 genes in the miR-21 interactome among 298 genes aberrantly expressed in SK-Hep-1 cells after anti-miR-21

treatment (Figure 5). The identity of these genes and their correlation with miR-21 expression in HCC are listed in Table 1. Upregulation of expression after anti-miR-21 treatment was observed in 16 of 24 dysregulated miR-21 targets [33]. Ago-CLIP analysis in mouse liver identified miR-21 binding sites in murine homologs of 10 of these 24 miR-21 targets (Table S1).



**Figure 5.** Gene identifiers from transcripts in association with miR-21 identified by Argonaute-CLIP were matched to genes dysregulated in SK-Hep-1 cells after treatment with anti-miR-21 (GSE65892) [33]. The 24 common genes were considered potential miR-21 targets.

To investigate the functions of newly identified miR-21 targets, we queried the Ingenuity Knowledge Base for canonical pathway involvement and protein function (Table S2). Two genes identified by our analysis, *SMARCE1* and *TGFBR2*, shared function in glucocorticoid receptor signaling. B-cell receptor signaling and the downstream pathway of PI3K signaling were implicated by dysregulation of *PAG1* and *PLEKHA1*, respectively, in miR-21-depleted SK-Hep-1 cells.

Among potential miR-21 targets upregulated in miR-21-depleted SK-Hep-1 cells, abundance of *RMND5A*, transcript negatively correlated with miR-21 expression in HCC patients (Table 2). *RMND5A* bears a 7merA1 motif in the fifth CDS exon as its sole miR-21 binding site. *RMND5A*, also known as *CTLH*, encodes an E3 ubiquitin ligase in budding yeast [34].

#### 2.7. Elements of the miR-21 Interactome Predict Survival in HCC

Next, we conducted a survival analysis on potential targets of miR-21 in 369 patients with HCC. Results are shown in Table 2. Among miR-21 candidate targets, higher expression of *CAMSAP2*, *DDX1*, and *MARCKSL1* predicted shorter survival. Expression of *MARCKSL1* positively correlated with miR-21 expression (Table 1). Expression data for *CAMSAP2* were not available in the TCGA RNASeq data set.

#### 3. Discussion

Overexpression of miR-21 is a poor prognostic indicator in hepatocellular carcinoma and other solid tumors as well as hematologic malignancies [9,10,35]. Our analysis showed that the miR-21 interactome identified by Ago-CLIP modulates diverse pathways in HCC. The 562 gene transcripts that interact with miR-21 represent a substantial fraction of the liver transcriptome. We used these genes to investigate the functional consequences of miR-21 targeting in the setting of HCC.

We found that miR-21 targets whose expression varied inversely with miR-21 in HCC tended to be less bound to Argonaute. MicroRNAs can shorten the half-life of the messenger RNAs they bind to [36]. We expect that these transcripts in HCC are rapidly degraded following binding of miR-21, and would then be less enriched in Argonaute complex. It is likely that upregulation of miR-21 in HCC results in the repression of its targets. Surprisingly, we found that more genes in the miR-21

interactome were overexpressed than repressed in HCC. Both transcriptional and post-transcriptional mechanisms are involved in gene expression, and miR-21 represents one of the mechanisms that, in general, fine tune the expression of target genes [37]. However, we cannot rule out that miR-21 suppresses protein levels of some of these targets by blocking their translation [6].

Ago-CLIP analysis showed that a large number of functional miR-21 targets have binding sites in the CDS. MicroRNAs can mediate transcript depletion and/or translational inhibition by binding to CDS [38–41]. We found, however, that CDS miR-21 binding sites were less likely to be higher affinity sites, such as 8mer, than those in the 3'UTR. The extent of seed sequence complementarity may influence the ability of closely spaced microRNA binding sites to synergistically repress targets [23].

In our study, miR-21 was predicted to regulate multiple pathways in HCC, including the acute phase response, Tec kinase signaling, PTEN signaling, PPAR $\alpha$ /RXR $\alpha$  signaling, and LXR/RXR signaling. Positive acute phase response reactants, including C-reactive protein, fibrinogen and complement components, are overexpressed in liver and released to systemic circulation, while synthesis of negative acute phase response reactants, such as transthyretin and albumin, is curtailed [27]. Ferrín et al. reported that acute phase reactants fibrinogen and the C4a fragment of complement component 4 were found at higher levels in the serum of hepatitis C virus (HCV)-infected patients with HCC than HCV-infected patients with cirrhosis only [42]. Activation of the acute phase response in HCC may be a sign of metabolic reprogramming. Administration of lipopolysaccharide has acute effects on serum cholesterol, triacylglycerides, high-density lipoproteins, and very-low-density lipoproteins [43]. Interestingly, Beigneux et al. demonstrated that stimulation of hepatocytes with lipopolysaccharide reduces PPAR $\alpha$ -stimulated acyl-CoA synthetase expression [44]. Tec kinase signaling has an important role in liver regeneration. Protein kinase TEC has been demonstrated to activate ERK within the hepatocyte growth factor signaling pathway [45], while inhibition of MEK/ERK phosphorylation is partly responsible for the anti-tumor activity of the kinase inhibitor sorafenib [46].

Metabolic dysregulation is a common feature of cancer that may involve miR-21-mediated regulation [33,47]. HCC is dependent upon glycolysis rather than mitochondrial oxidation for energy [48]. We found that the miR-21 interactome is predicted to suppress the signaling of hepatic nuclear receptors PPAR $\alpha$ /RXR $\alpha$  and LXR/RXR. PPAR $\alpha$  activation during fasting stimulates  $\beta$ -oxidation of fatty acids and gluconeogenesis. Feeding reduces PPAR $\alpha$  signaling through mTORC1 activation and Akt signaling [49,50]. Meng et al. determined that silencing of PTEN by miR-21 contributes to activation of the Akt pathway in HCC [15]. Additionally, the LXR/RXR pathway activates transcription of lipogenic enzymes in liver as well as their upstream transcription factor SREBP1C in response to ligand binding [51]. However, Wu et al. reported that miR-21 mimic induced SREBP1C in HepG2 cells [47]. Effects of miR-21 on LXR/RXR signaling in primary tumors may be distinct from those in cell culture because of reduced oxygen levels and substrate availability.

We have identified several novel potential miR-21 targets. Among them, PLEKHA1 binds to phosphatidyl inositol (3,4)-diphosphate and interacts with PDZ domains, including PDZ-containing protein tyrosine phosphatase-like protein 1 (PTPL1), a negative regulator of Akt signaling [52,53]. The yeast homolog of *RMND5A*, *GID2*, was originally found to be required for the gluconeogenic to glycolytic metabolic switch that occurs upon replenishment of glucose in a medium containing a nonfermentable carbon source [54]. As a component of the Gid complex, Gid2 is required for polyubiquination of the gluconeogenic enzyme fructose-1,6-bisphosphatase and for degradation of phosphoenolpyruvate carboxykinase [34]. Further investigation is needed to determine whether *RMND5A* mediates the effects of miR-21 on tumor metabolism in a subset of HCC patients.

Our study shows that the expression of three genes, *CAMSAP2*, *DDX1* and *MARCKSL1* with miR-21 binding motifs may mediate decreased survival in HCC patients. The expression of all three of these genes negatively correlated with survival. Dead-box helicase 1 encoded by *DDX1* is an RNA helicase that participates in double-strand break repair [55]. *CAMSAP2* encodes calmodulin-regulated spectrin-associated protein family member 2, a microtubule-associated protein implicated in the

.

regulation of cell motility through microtubule stabilization [56]. The *MARCKSL1* gene encodes MARCKS-related protein, which regulates the actin cytoskeleton and cell migration [57,58]. While miR-21 is predicted to bind the transcript of *MARCKSL1*, its expression positively correlates with miR-21 expression. It is possible that miR-21 suppresses *MARCKSL1* by inhibiting its translation.

Our study is limited to miR-21 targets with known binding motifs for miR-21. In order to fully appreciate the functional role of the miR-21 targetome, it would be necessary to conduct Ago-CLIP and RNAseq analysis on a miR-21 knockout model. A similar analysis was recently performed for the miR-122 targetome in liver [21]. Finally, it would be helpful to validate our findings on the expression of the miR-21 interactome in an independent cohort of patients.

#### 4. Materials and Methods

## 4.1. Argonaute-CLIP Analysis

Ago-CLIP analysis for nine HCC patients and matched benign liver was performed by the Darnell lab as previously reported (GSE97061) [21]. Patient data are given in supplementary Table 1 of Luna et al., 2017 [21]. Ago-CLIP analysis in five mice was previously reported (GSE97058) [21]. This data were accessed through the Gene Expression Omnibus (GEO) data repository.

## 4.2. Gene Set Enrichment and Pathway Analysis

RNA-seq RPKM data from The Cancer Genome Atlas (cancergenome.nih.gov) were downloaded from firebrowse.org and log<sub>2</sub> transformed. Differential expression from RNA-seq data of The Cancer Genome Atlas was obtained using the limma package in R cite (cran.r-project.org). Expression data from HCC SK-Hep-1 treated with 20 nmol/L antago-miR-21 and RNA extracted sixteen hours after treatment were previously reported (GSE65892) [33].

Gene set enrichment and pathway analysis were performed using Ingenuity Pathway Analysis (IPA). Log-fold change in RNA-seq abundance in tumor versus normal and associated *p*-value were uploaded to IPA. Pathway analysis was set to consider only human and experimentally observed molecules and/or relationships. Other parameters used the default settings.

#### 4.3. Statistical Analysis

Ago-CLIP enrichment scores for miR-21 binding in HCC were categorized as enriched ( $\log FC > 0$ ) or depleted ( $\log FC < 0$ ). Comparison of categorical data was performed by Chi square. A two-sided Kolmogorov–Smirnov test was used for comparison of density distributions.

Best-fit linear models of quantitative data were produced by least squares regression and significance determined by *t*-test of Pearson's coefficient. The corr.test() function from psych package in R was used to correct *p*-values of multiple correlations by Holm's method. Significance was interpreted with  $\alpha = 0.05$  unless otherwise noted.

The OncoLnc package was used for survival analysis (www.oncolnc.org). Cox proportional hazards models were adjusted for age, sex and tumor grade. The significance of the Cox coefficient was corrected for the false-discovery rate as described [59].

#### Supplementary Materials: Supplementary materials can be found at www.mdpi.com/1422-0067/19/3/851/s1.

Acknowledgments: We would like to thank Lianbo Yu and other members of the Ghoshal lab for the thoughtful discussion. We would also like to acknowledge the individuals and institutions that have provided data for The Cancer Genome Atlas Liver Hepatocellular Carcinoma (TCGA-LIHC) dataset. This study was supported, in part, by grants R01CA193244 (Kalpana Ghoshal) and R01086978 (Kalpana Ghoshal and Samson T. Jacob) from the NIH. Aaron Balasingam Koenig was supported by The Ohio State University College of Medicine Roessler Medical Student Research Scholarship and Juan Martin Barajas is a Howard Hughes Medical Institute Gilliam Fellow.

**Author Contributions:** Juan Martin Barajas and Kalpana Ghoshal conceived the project and designed the experiments. Aaron Balasingam Koenig and Maria Jose Guerrero performed the experiments and analyzed the data. Aaron Balasingam Koenig and Juan Martin Barajas wrote the paper. All authors reviewed and edited the manuscript.

Conflicts of Interest: The authors declare no conflict of interest.

## Abbreviations

Ago-CLIP-seq	High-throughput sequencing of RNAs isolated by cross-linking and immunoprecipitation of Argonaute	
HCC	Hepatocellular carcinoma	
TCGA-LIHC	The cancer genome atlas liver hepatocellular carcinoma	
3'UTR	3' untranslated region	
5′UTR	5' untranslated region	
CDS	Coding DNA sequence	
IPA	Ingenuity Pathway Analysis	
PDCD4	Programmed cell death protein 4	
PTEN	Phosphatase and tensin homolog	
RECK	Reversion-inducing-cysteine-rich protein with kazal motifs	
TIMP3	Metalloproteinase inhibitor 3	
LXR/RXR	Liver X receptor/retinoid X receptor	
PPARa/RXRa	Peroxisome proliferator-activated receptor alpha/retinoid X receptor alpha	
HCV	Hepatitis C virus	
RNASeq	RNA sequencing	

# References

- 1. Torre, L.A.; Bray, F.; Siegel, R.L.; Ferlay, J.; Lortet-Tieulent, J.; Jemal, A. Global cancer statistics, 2012. *CA Cancer J. Clin.* **2015**, 65, 87–108. [CrossRef] [PubMed]
- 2. Llovet, J.M.; Zucman-Rossi, J.; Pikarsky, E.; Sangro, B.; Schwartz, M.; Sherman, M.; Gores, G. Hepatocellular carcinoma. *Nat. Rev. Dis. Primer* **2016**, *2*, 16018. [CrossRef] [PubMed]
- Wang, K.; Lim, H.Y.; Shi, S.; Lee, J.; Deng, S.; Xie, T.; Zhu, Z.; Wang, Y.; Pocalyko, D.; Yang, W.J.; et al. Genomic landscape of copy number aberrations enables the identification of oncogenic drivers in hepatocellular carcinoma. *Hepatology* 2013, *58*, 706–717. [CrossRef] [PubMed]
- 4. Schulze, K.; Imbeaud, S.; Letouzé, E.; Alexandrov, L.B.; Calderaro, J.; Rebouissou, S.; Couchy, G.; Meiller, C.; Shinde, J.; Soysouvanh, F.; et al. Exome sequencing of hepatocellular carcinomas identifies new mutational signatures and potential therapeutic targets. *Nat. Genet.* **2015**, *47*, 505–511. [CrossRef] [PubMed]
- Niu, Z.S.; Niu, X.J.; Wang, W.H. Genetic alterations in hepatocellular carcinoma: An update. *World J. Gastroenterol.* 2016, 22, 9069–9095. [CrossRef] [PubMed]
- 6. Jonas, S.; Izaurralde, E. Towards a molecular understanding of microRNA-mediated gene silencing. *Nat. Rev. Genet.* **2015**, *16*, 421–433. [CrossRef] [PubMed]
- Lagos-Quintana, M.; Rauhut, R.; Lendeckel, W.; Tuschl, T. Identification of novel genes coding for small expressed RNAs. *Science* 2001, 294, 853–858. [CrossRef] [PubMed]
- 8. Volinia, S.; Calin, G.A.; Liu, C.G.; Ambs, S.; Cimmino, A.; Petrocca, F.; Visone, R.; Iorio, M.; Roldo, C.; Ferracin, M.; et al. A microRNA expression signature of human solid tumors defines cancer gene targets. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 2257–2261. [CrossRef] [PubMed]
- Schetter, A.J.; Leung, S.Y.; Sohn, J.J.; Zanetti, K.A.; Bowman, E.D.; Yanaihara, N.; Yuen, S.T.; Chan, T.L.; Kwong, D.L.W.; Au, G.K.H.; et al. MicroRNA expression profiles associated with prognosis and therapeutic outcome in colon adenocarcinoma. *JAMA* 2008, 299, 425–436. [CrossRef] [PubMed]
- 10. Rossi, S.; Shimizu, M.; Barbarotto, E.; Nicoloso, M.S.; Dimitri, F.; Sampath, D.; Fabbri, M.; Lerner, S.; Barron, L.L.; Rassenti, L.Z.; et al. microRNA fingerprinting of CLL patients with chromosome 17p deletion identify a miR-21 score that stratifies early survival. *Blood* **2010**, *116*, 945–952. [CrossRef] [PubMed]
- Shi, K.Q.; Lin, Z.; Chen, X.J.; Song, M.; Wang, Y.Q.; Cai, Y.J.; Yang, N.B.; Zheng, M.H.; Dong, J.Z.; Zhang, L.; et al. Hepatocellular carcinoma associated microRNA expression signature: Integrated bioinformatics analysis, experimental validation and clinical significance. *Oncotarget* 2015, *6*, 25093–25108. [CrossRef] [PubMed]

- Iliopoulos, D.; Jaeger, S.A.; Hirsch, H.A.; Bulyk, M.L.; Struhl, K. STAT3 activation of miR-21 and miR-181b-1 via PTEN and CYLD are part of the epigenetic switch linking inflammation to cancer. *Mol. Cell* 2010, *39*, 493–506. [CrossRef] [PubMed]
- Zhu, Q.; Wang, Z.; Hu, Y.; Li, J.; Li, X.; Zhou, L.; Huang, Y. miR-21 promotes migration and invasion by the miR-21-PDCD4-AP-1 feedback loop in human hepatocellular carcinoma. *Oncol. Rep.* 2012, 27, 1660–1668. [CrossRef] [PubMed]
- 14. Zhang, N.; Duan, W.D.; Leng, J.J.; Zhou, L.; Wang, X.; Xu, Y.Z.; Wang, X.D.; Zhang, A.Q.; Dong, J.H. STAT3 regulates the migration and invasion of a stem-like subpopulation through microRNA-21 and multiple targets in hepatocellular carcinoma. *Oncol. Rep.* **2015**, *33*, 1493–1498. [CrossRef] [PubMed]
- Meng, F.; Henson, R.; Wehbe-Janek, H.; Ghoshal, K.; Jacob, S.T.; Patel, T. MicroRNA-21 regulates expression of the PTEN tumor suppressor gene in human hepatocellular cancer. *Gastroenterology* 2007, 133, 647–658. [CrossRef] [PubMed]
- Asangani, I.A.; Rasheed, S.A.K.; Nikolova, D.A.; Leupold, J.H.; Colburn, N.H.; Post, S.; Allgayer, H. MicroRNA-21 (miR-21) post-transcriptionally downregulates tumor suppressor Pdcd4 and stimulates invasion, intravasation and metastasis in colorectal cancer. *Oncogene* 2008, 27, 2128–2136. [CrossRef] [PubMed]
- Frankel, L.B.; Christoffersen, N.R.; Jacobsen, A.; Lindow, M.; Krogh, A.; Lund, A.H. Programmed cell death 4 (PDCD4) is an important functional target of the microRNA miR-21 in breast cancer cells. *J. Biol. Chem.* 2008, 283, 1026–1033. [CrossRef] [PubMed]
- Lu, Z.; Liu, M.; Stribinskis, V.; Klinge, C.M.; Ramos, K.S.; Colburn, N.H.; Li, Y. MicroRNA-21 promotes cell transformation by targeting the programmed cell death 4 gene. *Oncogene* 2008, 27, 4373–4379. [CrossRef] [PubMed]
- Gabriely, G.; Wurdinger, T.; Kesari, S.; Esau, C.C.; Burchard, J.; Linsley, P.S.; Krichevsky, A.M. MicroRNA 21 promotes glioma invasion by targeting matrix metalloproteinase regulators. *Mol. Cell. Biol.* 2008, 28, 5369–5380. [CrossRef] [PubMed]
- Chi, S.W.; Zang, J.B.; Mele, A.; Darnell, R.B. Argonaute HITS-CLIP decodes microRNA-mRNA interaction maps. *Nature* 2009, 460, 479–486. [CrossRef] [PubMed]
- 21. Luna, J.M.; Barajas, J.M.; Teng, K.Y.; Sun, H.L.; Moore, M.J.; Rice, C.M.; Darnell, R.B.; Ghoshal, K. Argonaute CLIP defines a deregulated miR-122-bound transcriptome that correlates with patient survival in human liver cancer. *Mol. Cell* **2017**, *67*, 400–410.e7. [CrossRef] [PubMed]
- 22. Ritchie, M.E.; Phipson, B.; Wu, D.; Hu, Y.; Law, C.W.; Shi, W.; Smyth, G.K. limma powers differential expression analyses for RNA-sequencing and microarray studies. *Nucleic Acids Res.* **2015**, *43*, e47. [CrossRef] [PubMed]
- 23. Fang, Z.; Rajewsky, N. The impact of miRNA target sites in coding sequences and in 3'UTRs. *PLoS ONE* **2011**, *6*, e18067. [CrossRef] [PubMed]
- 24. Kwak, H.J.; Kim, Y.J.; Chun, K.R.; Woo, Y.M.; Park, S.J.; Jeong, J.A.; Jo, S.H.; Kim, T.H.; Min, H.S.; Chae, J.S.; et al. Downregulation of Spry2 by miR-21 triggers malignancy in human gliomas. *Oncogene* **2011**, *30*, 2433–2442. [CrossRef] [PubMed]
- 25. Zhao, J.; Tang, N.; Wu, K.; Dai, W.; Ye, C.; Shi, J.; Zhang, J.; Ning, B.; Zeng, X.; Lin, Y. MiR-21 simultaneously regulates ERK1 signaling in HSC activation and hepatocyte EMT in hepatic fibrosis. *PLoS ONE* **2014**, *9*, e108005. [CrossRef] [PubMed]
- Xu, G.; Zhang, Y.; Wei, J.; Jia, W.; Ge, Z.; Zhang, Z.; Liu, X. MicroRNA-21 promotes hepatocellular carcinoma HepG2 cell proliferation through repression of mitogen-activated protein kinase-kinase 3. *BMC Cancer* 2013, 13, 469. [CrossRef] [PubMed]
- 27. Gruys, E.; Toussaint, M.J.M.; Niewold, T.A.; Koopmans, S.J. Acute phase reaction and acute phase proteins. *J. Zhejiang Univ. Sci. B* 2005, *6*, 1045–1056. [CrossRef] [PubMed]
- 28. Oberkofler, H.; Schraml, E.; Krempler, F.; Patsch, W. Restoration of sterol-regulatory-element-binding protein-1c gene expression in HepG2 cells by peroxisome-proliferator-activated receptor-gamma co-activator-1alpha. *Biochem. J.* **2004**, *381 Pt 2*, 357–363. [CrossRef] [PubMed]
- 29. Bradshaw, J.M. The Src, Syk, and Tec family kinases: Distinct types of molecular switches. *Cell. Signal.* **2010**, 22, 1175–1184. [CrossRef] [PubMed]

- Melcher, M.; Unger, B.; Schmidt, U.; Rajantie, I.A.; Alitalo, K.; Ellmeier, W. Essential roles for the Tec family kinases Tec and Btk in M-CSF receptor signaling pathways that regulate macrophage survival. *J. Immunol.* 2008, 180, 8048–8056. [CrossRef] [PubMed]
- 31. Wang, S.Y.; Li, F.F.; Zheng, H.; Yu, K.K.; Ni, F.; Yang, X.M.; Qu, C.K.; Li, J. Rapid induction and activation of Tec tyrosine kinase in liver regeneration. *J. Gastroenterol. Hepatol.* **2006**, *21*, 668–673. [CrossRef] [PubMed]
- 32. Heffelfinger, S.C.; Hawkins, H.H.; Barrish, J.; Taylor, L.; Darlington, G.J. SK HEP-1: A human cell line of endothelial origin. *In Vitro Cell. Dev. Biol.* **1992**, *28A*, 136–142. [CrossRef] [PubMed]
- 33. Wagenaar, T.R.; Zabludoff, S.; Ahn, S.M.; Allerson, C.; Arlt, H.; Baffa, R.; Cao, H.; Davis, S.; Garcia-Echeverria, C.; Gaur, R.; et al. Anti–miR-21 suppresses hepatocellular carcinoma growth via broad transcriptional network deregulation. *Mol. Cancer Res.* **2015**, *13*, 1009–1021. [CrossRef] [PubMed]
- 34. Santt, O.; Pfirrmann, T.; Braun, B.; Juretschke, J.; Kimming, P.; Scheel, H.; Hofmann, K.; Thumm, M.; Wolf, D.H. The yeast GID complex, a novel ubiquitin ligase (E3) involved in the regulation of carbohydrate metabolism. *Mol. Biol. Cell* **2008**, *19*, 3323–3333. [CrossRef] [PubMed]
- Wang, W.Y.; Zhang, H.F.; Wang, L.; Ma, Y.P.; Gao, F.; Zhang, S.J.; Wang, L.C. miR-21 expression predicts prognosis in hepatocellular carcinoma. *Clin. Res. Hepatol. Gastroenterol.* 2014, *38*, 715–719. [CrossRef] [PubMed]
- Hayes, J.; Peruzzi, P.P.; Lawler, S. MicroRNAs in cancer: Biomarkers, functions and therapy. *Trends Mol. Med.* 2014, 20, 460–469. [CrossRef] [PubMed]
- Ally, A.; Balasundaram, M.; Carlsen, R.; Chuah, E.; Clarke, A.; Dhalla, N.; Holt, R.A.; Jones, S.J.M.; Lee, D.; Ma, Y.; et al. Comprehensive and integrative genomic characterization of hepatocellular carcinoma. *Cell* 2017, *169*, 1327–1341.e23. [CrossRef] [PubMed]
- Saxena, S.; Jónsson, Z.O.; Dutta, A. Small RNAs with imperfect match to endogenous mRNA repress translation. Implications for off-target activity of small inhibitory RNA in mammalian cells. *J. Biol. Chem.* 2003, 278, 44312–44319. [CrossRef] [PubMed]
- 39. Duursma, A.M.; Kedde, M.; Schrier, M.; le Sage, C.; Agami, R. miR-148 targets human DNMT3b protein coding region. *RNA* **2008**, *14*, 872–877. [CrossRef] [PubMed]
- 40. Tay, Y.; Zhang, J.; Thomson, A.M.; Lim, B.; Rigoutsos, I. MicroRNAs to Nanog, Oct4 and Sox2 coding regions modulate embryonic stem cell differentiation. *Nature* **2008**, 455, 1124–1128. [CrossRef] [PubMed]
- 41. Forman, J.J.; Legesse-Miller, A.; Coller, H.A. A search for conserved sequences in coding regions reveals that the let-7 microRNA targets Dicer within its coding sequence. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 14879–14884. [CrossRef] [PubMed]
- 42. Ferrín, G.; Rodríguez-Perálvarez, M.; Aguilar-Melero, P.; Ranchal, I.; Llamoza, C.; Linares, C.I.; González-Rubio, S.; Muntané, J.; Briceño, J.; López-Cillero, P.; et al. Plasma protein biomarkers of hepatocellular carcinoma in HCV-infected alcoholic patients with cirrhosis. *PLoS ONE* **2015**, *10*, e0118527. [CrossRef] [PubMed]
- Hardardóttir, I.; Grunfeld, C.; Feingold, K.R. Effects of endotoxin on lipid metabolism. *Biochem. Soc. Trans.* 1995, 23, 1013–1018. [CrossRef] [PubMed]
- 44. Beigneux, A.P.; Moser, A.H.; Shigenaga, J.K.; Grunfeld, C.; Feingold, K.R. The acute phase response is associated with retinoid X receptor repression in rodent liver. *J. Biol. Chem.* **2000**, 275, 16390–16399. [CrossRef] [PubMed]
- 45. Li, F.; Jiang, Y.; Zheng, Q.; Yang, X.; Wang, S. TEC protein tyrosine kinase is involved in the Erk signaling pathway induced by HGF. *Biochem. Biophys. Res. Commun.* **2011**, *404*, 79–85. [CrossRef] [PubMed]
- 46. Liu, L.; Cao, Y.; Chen, C.; Zhang, X.; McNabola, A.; Wilkie, D.; Wilhelm, S.; Lynch, M.; Carter, C. Sorafenib blocks the RAF/MEK/ERK pathway, inhibits tumor angiogenesis, and induces tumor cell apoptosis in hepatocellular carcinoma model PLC/PRF/5. *Cancer Res.* **2006**, *66*, 11851–11858. [CrossRef] [PubMed]
- 47. Wu, H.; Ng, R.; Chen, X.; Steer, C.J.; Song, G. MicroRNA-21 is a potential link between non-alcoholic fatty liver disease and hepatocellular carcinoma via modulation of the HBP1-p53-Srebp1c pathway. *Gut* **2016**, *65*, 1850–1860. [CrossRef] [PubMed]
- 48. Beyoğlu, D.; Imbeaud, S.; Maurhofer, O.; Bioulac-Sage, P.; Zucman-Rossi, J.; Dufour, J.F.; Idle, J.R. Tissue metabolomics of hepatocellular carcinoma: Tumor energy metabolism and the role of transcriptomic classification. *Hepatolology* **2013**, *58*, 229–238. [CrossRef] [PubMed]
- 49. Sengupta, S.; Peterson, T.R.; Laplante, M.; Oh, S.; Sabatini, D.M. mTORC1 controls fasting-induced ketogenesis and its modulation by ageing. *Nature* **2010**, *468*, 1100–1104. [CrossRef] [PubMed]

- 50. Preidis, G.A.; Kim, K.H.; Moore, D.D. Nutrient-sensing nuclear receptors PPARα and FXR control liver energy balance. *J. Clin. Investig.* **2017**, 127, 1193–1201. [CrossRef] [PubMed]
- Bindesbøll, C.; Fan, Q.; Nørgaard, R.C.; MacPherson, L.; Ruan, H.B.; Wu, J.; Pedersen, T.Å.; Steffensen, K.R.; Yang, X.; Matthews, J.; et al. Liver X receptor regulates hepatic nuclear O-GlcNAc signaling and carbohydrate responsive element-binding protein activity. *J. Lipid Res.* 2015, *56*, 771–785. [CrossRef] [PubMed]
- 52. Kimber, W.A.; Deak, M.; Prescott, A.R.; Alessi, D.R. Interaction of the protein tyrosine phosphatase PTPL1 with the PtdIns (3,4) P2-binding adaptor protein TAPP1. *Biochem. J.* **2003**, *376*, 525–535. [CrossRef] [PubMed]
- 53. Li, H.; Marshall, A.J. Phosphatidylinositol (3,4) bisphosphate-specific phosphatases and effector proteins: A distinct branch of PI3K signaling. *Cell. Signal.* **2015**, *27*, 1789–1798. [CrossRef] [PubMed]
- Regelmann, J.; Schüle, T.; Josupeit, F.S.; Horak, J.; Rose, M.; Entian, K.D.; Thumm, M.; Wolf, D.H. Catabolite degradation of fructose-1,6-bisphosphatase in the yeast *Saccharomyces cerevisiae*: A genome-wide screen identifies eight novel GID genes and indicates the existence of two degradation pathways. *Mol. Biol. Cell* 2003, 14, 1652–1663. [CrossRef] [PubMed]
- 55. Gustafson, E.A.; Wessel, G.M. DEAD-box helicases: Posttranslational regulation and function. *Biochem. Biophys. Res. Commun.* **2010**, 395, 1–6. [CrossRef] [PubMed]
- 56. Jiang, K.; Hua, S.; Mohan, R.; Grigoriev, I.; Yau, K.W.; Liu, Q.; Katrukha, E.A.; Maarten Altelaar, A.F.; Heck, A.J.R.; Hoogenraad, C.C.; et al. Microtubule minus-end stabilization by polymerization-driven CAMSAP deposition. *Dev. Cell* **2014**, *28*, 295–309. [CrossRef] [PubMed]
- 57. Chen, J.; Chang, S.; Duncan, S.A.; Okano, H.J.; Fishell, G.; Aderem, A. Disruption of the *MacMARCKS* gene prevents cranial neural tube closure and results in anencephaly. *Proc. Natl. Acad. Sci. USA* **1996**, *93*, 6275–6279. [CrossRef] [PubMed]
- 58. Finlayson, A.E.; Freeman, K.W. A cell motility screen reveals role for MARCKS-related protein in adherens junction formation and tumorigenesis. *PLoS ONE* **2009**, *4*, e7833. [CrossRef] [PubMed]
- 59. Anaya, J. OncoLnc: Linking TCGA survival data to mRNAs, miRNAs, and lncRNAs. *PeerJ Comput. Sci.* 2016, 2, e67. [CrossRef]



© 2018 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).