

Figure S1. *Y. pestis* invasion into LCLs is inhibited by FBS and heritable. (**A**) Flow cytometric measurement of *Y. pestis* (KIM6+ +pMMB67GFP) invasion of LCLs at 4 hpi in RPMI media with either 10% FBS or 0.03% BSA after gentamicin protection assay. 2 experiments are grand mean normalized with 8 total replicates plotted for each condition. An unpaired t-test was performed to determine significance. (**B**) Parent-offspring linear regression of *Y. pestis* invasion into LCLs gives an estimated heritability of 0.15, a slope of 0.19 and a p-value = 0.014.



Figure S2. Locus zoom and genotypic mean plots of LCLs stratified by ancestry. Local Manhattan plots and genotypic mean plots stratified by African (GWD, YRI, ESN), Asian (JPT, CHB, KHV), European (CEU, IBS), or EurAsian (JPT, CHB, KHV, CEU, IBS) ancestries. A purple diamond denotes rs2282284 and LD with SNPs in the locus is shown by red >= 0.8, orange = 0.6-0.8, green = 0.4-0.6, light blue = 0.2-0.4, dark blue < 0.2, and grey has no LD data. LD was determined by all populations within the stratified group. - log₁₀(p) values at the rs2282284 locus was plotted and phenotypes were plotted by genotype. All populations demonstrate the C allele of rs2282284 is associated with decreased invasion.



Figure S3. No FCRL3 protein is detected in HeLa cells and intracellular *Y. pestis* is in LAMP+ vesicles at 4 hpi. (**A**) Bar graphs of expression in common cell lines from the Human Protein Atlas (Uhlen et al., 2015) show expression only in lymphoma, Calu-3, and DU145 cell lines and no expression in HeLa. (**B**) GFP^{high} cells have intracellular *Y. pestis* within LAMP1+ vacuoles. HeLa cells transfected with empty vector or *FCRL3* plasmid were infected with KIM6+ +pMMB67GFP *Y. pestis* for 1 hr, treated with gentamicin for 1 hr, and induced with IPTG for 2 hr prior to fixation with 4% paraformaldehyde. After

incubating for 30 minutes in block/perm, DNA was stained with 2.5µM DAPI, LAMP1 was stained red with a 1:20 dilution of H4A3-s mouse LAMP1 antibody (Developmental Studies Hybridoma Bank), and bacteria are shown in green at high exposure to include the GFP^{low} population. GFP^{low} bacteria are indicated with a teal arrow head and GFP^{high} bacteria with an orange arrow head. Images were taken on a Zeiss Observer Z1 inverted microscope with a 63x water objective. A 20µm scale bar is located on the merged image.

Table S1. (separate file) *Y. pestis* invasion measurement for 961 LCLs (3 replicates and mean)

GE	GUIDE 1	GUIDE 2	GUIDE 3	FWD primer	REV primer
NE					
CD4	GAGAAACAUGUC	AACUCGUAAGUC	UUGCUCCUUAGAG	TGCCTGGGTGAAT	TGTCAGAAACAGCAA
6	CAUAUAUA	CCAUUUGC	GAAAUAA	ATGAATCTT	GTAGTTTTG
FCR	AAUUUCCAGGCU	GAUACCAAUAUGU	CUGUGGACCAUG	TCTGCCTAGGATC	ACCCTGGTCCTGACT
L3	CUGUAAUU	GUCUCCC	GAGGAUUG	CCTGCAT	GGA

Table S2. sgRNAs and primers for CRISPR mutagenesis and validation

Table S3. Plasmids					
Plasmid backbone	insert	tag	Origin		
pCMV6		MYC/FLAG	Simon Gregory		
pCMV6	FCRL3	MYC/FLAG	Origene		
pCMV-SPORT6	CD31		Tim Wilson		
pFLAG-CMV-4	FCRL1	FLAG	Tim Wilson		
pFLAG-CMV-6	FCRL3	FLAG	Tim Wilson		
pFLAG-CMV-7	FCRL4	FLAG	Tim Wilson		
pFLAG-CMV-8	FCRL5	FLAG	Tim Wilson		
pFLAG-CMV-9	FCRL6	FLAG	Tim Wilson		
pD649-Hasp-COMP5AP	CD31	AviTag/9xHis	Addgene		
pD649-Hasp-COMP5AP	FCRL5	AviTag/9xHis	Addgene		

Table S4. Oligonucleotides for Quikchange mutagenesismutationForwardreverse

N721S	catgaagaagatgatgaagaaagctatgagaatgtaccacgtgta	tacacgtggtacattctcatagctttcttcatcatcttcttcatg
Y650F	ggagctggagccaatgttcagcaatgtaaatcctg	caggatttacattgctgaacattggctccagctcc
Y662F	ctggagatagcaacccgattttttcccagatctg	cagatctgggaaaaaatcgggttgctatctccag
Y692F	gaggaacttacagtcctcttttcagaactgaagaagaca	tgtcttcttcagttctgaaaagaggactgtaagttcctc
Y722F	cccatgaagaagatgatgaagaaaactttgagaatgtaccac	gtggtacattctcaaagttttcttcatcatcttcttcatggg