

REVIEW

The expression and function of long noncoding RNAs in hepatocellular carcinoma

Jingli Du¹  | Yue Su¹ | Jianzhi Gao² | Yanhong Tai³

¹Senior Department of Tuberculosis, The 8th Medical Center of PLA General Hospital, Beijing, China

²Department of Oncology, Zhuozhou Hospital, Zhuozhou, Hebei, China

³Department of Pathology, The 5th Medical Center of PLA General Hospital, Beijing, China

Correspondence

Yanhong Tai, Department of Pathology, The 5th Medical Center of PLA General Hospital, Beijing 100071, China.
Email: taiyanhong29@163.com

Jianzhi Gao, Department of Oncology, Zhuozhou Hospital, Zhuozhou 072750, Hebei, China.
Email: gaoteng.bao@163.com

Funding information

None

Abstract

With the deepening of the genome project study, attention on noncoding RNAs is increasing. Long noncoding RNAs (lncRNAs) have become a new research hotspot. A growing number of studies have revealed that lncRNAs are involved in tumorigenesis and tumor suppressor pathways. Aberrant expressions of lncRNAs have been found in a variety of human tumors including hepatocellular carcinoma (HCC). In this review, we provide a brief introduction to lncRNA and highlight recent research on the functions and clinical significance of lncRNAs in HCC.

KEYWORDS

carcinogenesis, hepatocellular carcinoma, long noncoding RNA, noncoding RNAs, tumor suppressor

1 | BACKGROUND

Primary hepatocellular carcinoma (HCC) is a common human malignancy. The fatality rate of HCC ranks third among all malignancies worldwide [1], and HCC is one of the main causes of cancer-related death in China. With the development of medical technology in recent years, strategies for the diagnosis and treatment of HCC have made increasing progress. However, the recurrence, metastasis and mortality rates of HCC have not sufficiently improved and early detection of HCC remains challenging. The main reason for these obstacles is that the molecules and factors

involved in early symptoms and various pathological manifestations of primary HCC are still unclear. Therefore, the molecular mechanisms underlying the development and occurrence of HCC have become the focus of research studies in recent years.

A great deal of research has demonstrated that long noncoding RNAs (lncRNAs) are widely involved in physiological and pathological processes, and aberrations in lncRNAs have been found to be related to many human diseases, particularly cancer. Alterations in lncRNA expressions have been detected in a variety of human tumors, including HCC [2–6] and may be associated with tumor

Abbreviations: HCC, hepatocellular carcinoma; lncRNA, long noncoding RNA; MDR1, multidrug resistance 1 gene; ncRNA, noncoding RNA; PCG, protein-coding genes; pre-mRNA, precursor mRNA; T-DMR, tissue-dependent differentially methylated region; T-UCRs, transcribed ultra-conserved regions; UCRs, ultra-conserved regions.

Jingli Du and Yue Su contributed equally to this study and shared first authorship.

This is an open access article under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.

© 2023 The Authors. *Cancer Innovation* published by John Wiley & Sons Ltd on behalf of Tsinghua University Press.

occurrence and development. Furthermore, several lncRNAs have been shown to be sensitive and specific tumor markers. Together, these findings have led to a new understanding of the molecular mechanisms of HCC with implications for HCC diagnosis.

2 | LncRNAs

Early scientific research suggested that RNA was the only medium for transmitting information between DNA and protein. RNA was considered an intermediate for transmitting information for protein synthesis but was not thought to participate in the regulation of this process. However, with the development of technology, later studies revealed that less than 2% of genes encode protein and most DNA is transcribed into noncoding RNA (ncRNA) [7, 8]. A large number of ncRNAs constitute a huge molecular network that plays important regulatory roles in eukaryotes.

LncRNAs are RNAs more than 200 nt long, and most lack protein coding ability [9, 10]. The update defines lncRNAs as RNA molecules that may function as either primary or spliced transcripts and excluding known classes of small RNAs, such as miRNAs, small nucleolar RNAs, piwi-interacting RNAs, or into classes of structural RNAs including transfer RNAs, small nuclear RNAs, spliceosomal RNAs and so on [11].

2.1 | Classification of lncRNAs

The classification of lncRNA has not been well established. According to the position of lncRNA relative to host *protein-coding genes (PCG)*, the classifications of lncRNA include nature antisense RNA, intronic antisense RNA, bidirectional RNA, exon-sense overlapping RNA, intergenic RNA, intron-sense RNA and promoter- or enhancer-correlated RNA [10, 11]. Other researchers classified lncRNA into three groups: lincRNAs, which are transcribed from intergenic regions; lincRNAs, which are transcribed from ultra-conserved regions (UCRs); and other lncRNAs [12].

2.2 | Functions of lncRNAs

LncRNAs were initially believed to be a by-product of transcription or the “noise” generated from transcription of the genome with no biological function [13, 14]. However, research has confirmed that lncRNAs are involved in multiple regulatory roles and biological processes including genomic imprinting [15], X chromosome inactivation [16], chromatin structure [17], enhancer function [18, 19], transcriptional activation, transcriptional interference and

gene expression regulation by cis or trans regulatory mechanisms [17, 20]. LncRNAs play important biological roles in multiple levels of chromosome modification, transcription, and posttranscription [21].

LncRNA has been shown to regulate the expression of genes by various mechanisms, including transcriptional regulation, chromatin modification, post-transcriptional regulation, and so on (Figure 1). LncRNAs silence or activate gene expression by regulating DNA methylation or histone modification and chromatin remodeling. For example, Imamura et al. found that *lncRNA Khpsla* originates from a CpG island and overlaps with a tissue-dependent differentially methylated region (T-DMR) of *SPHK1*. Overexpression of two fragments of *Khps1* caused demethylation of CG sites in the T-DMR [22]. Silencing of the *Kcnq1ot1* gene was found to involve H3K9me2 and H3K27me3 histone modifications, which are partly caused by G9a and Ezh2 histone methyltransferases, resulting in cluster-wide repressive histone marks, gene silencing and DNA methylation of CpG islands of promoters [23]. A similar observation was reported with *Ari*, *Xist*, and *BACE1-AS*, which bind PRC2, TRX, and G9a and impart specific silencing of genomic loci both in cis and trans [17, 24-26]. In a study in leukemia, Yu et al. found that lncRNA antisense *P15* expression resulted in *p15* silencing through heterochromatin formation [27].

LncRNAs act as transcriptional regulators or coregulators to modulate gene expression; they can inhibit gene transcription via transcriptional interference [28]; and interact with transcription inhibiting complexes to influence the expression of target genes. LncRNAs interact with RNA polymerase II, interfere with the formation of the transcription initiation complex and inhibit transcription initiation, resulting in rapid changes in gene expression patterns. For example, *B2* RNA in mice and *Alu* RNA in humans repress mRNA transcription in response to heat shock; these lncRNAs inhibit the polymerase and DNA contacts by binding with RNA polymerase II and assemble into complexes on the promoter [29].

LncRNAs regulate mRNA splicing, transport, editing, translation, and degradation. Base pairing between sense and antisense RNAs masks the splice sites that trigger alternate splicing [30]. LncRNA stabilize or promote target gene translation by extended base pairing [31]. For example, the upregulation of *BACE1-AS* is linked to *BACE1* mRNA stabilization through extended base-pairing with *BACE1*, resulting in increased protein level [32]. Conversely, some base pairing facilitates mRNA decay or inhibits mRNA translation. For example, the cytoplasmic *1/2-sbsRNAs* promotes decay by partial base pairing with specific target mRNAs and recruitment of Staufen1 [31, 33]. Moreover, some lncRNAs form



FIGURE 1 Regulation of gene expression by long noncoding RNA through transcriptional regulation, chromatin modification and posttranscriptional regulation mechanisms.

complementary double-stranded with transcripts of protein-coding genes, and further produced endogenous siRNA under Dicer cleavage, resulting in mRNA degradation and influencing gene expression levels [34]. LncRNAs also bind with RNA binding proteins to suppress mRNA splicing and translation and can function as ceRNAs to increase expression of mRNA, the miRNA target [31]. For example, lncRNA *MALAT1* was implicated in splicing of precursor mRNA (pre-mRNA) by influencing the

distribution of serine/arginine-rich proteins [35]. LncRNA *BACE1-AS* competes with miR-485-5p to interact with *BACE1* mRNA, representing another mechanism by which *BACE1-AS* controls the stability of *BACE1* mRNA [36]. LncRNAs can act as a scaffold for various proteins to form ribonucleoprotein complexes [37].

In addition, LncRNA also directly interact with specific proteins to directly regulate their activity or alter their subcellular localization [38].

3 | LncRNAs RELATED TO HCC

Increasing research has shown that abnormal lncRNA expression is related to the occurrence and development of tumors, and lncRNAs have been shown to play important regulatory roles in these processes. Several studies have shown that some tumors did not harbor mutations in protein-encoding genes and instead exhibited abnormal expressions of lncRNAs [39]. Pan et al. [40] examined differentially expressed lncRNAs in a hepatoma carcinoma group and control group and found that the lncRNA expression profiles in the two groups were different, suggesting these differentially expressed lncRNAs may be involved in the molecular mechanisms of HCC. In the following section, we discuss the recent research related to lncRNAs in HCC (Table 1).

3.1 | LncRNAs promote the progression of HCC

Research has identified multiple lncRNAs that promote the occurrence or progression of HCC. The lncRNAs with oncogenic roles include *HULC*, *H19*, *MALAT1*, *HEIH*, *MVIH*, *HOTAIR*, and *HOTTIP/HOXA13*. Several lncRNAs associated with cell metabolism that promote HCC were recently reported, such as lncRNA *RP11-386G11.10* [41], *SNHG6* [42], *FASRL* [43], and *DACT3-AS1* [44].

3.1.1 | LncRNAs related to liver cancer cell proliferation

3.1.1.1 | Highly upregulated in liver cancer (*HULC*)

Various lncRNAs have been shown to regulate the proliferation of liver cancer cells, including *HULC*, *SNHG6*, *lncRNA RP11-386G11.10*, *MIAT*, and *ALKBH3-AS1*. Among these lncRNAs, *HULC* has been relatively well studied. *HULC* is located on chromosome 6p24.3 and has a length of approximately 500 nt. *HULC* is expressed at higher levels in human liver cancer cells compared with noncancerous liver cells. In contrast, *HULC* expression showed small differences or no differences in other tumor cells compared with the corresponding nontumor cells [45]. *HULC* is detectable in blood of HCC patients, and it is expected to be a potential biomarker of HCC [45]. Ruan et al. [46] found high expression of *HULC* in HCC, especially in HBV-related HCC. These studies suggest that *HULC* is a biological marker of HCC.

Knockdown of *HULC* in liver cancer cells resulted in changes in the expressions of tumor-related genes. Hepatitis B virus-related HCC was directly related with the

upregulation of the expression of *HULC* [46]. Subsequent studies demonstrated that *HULC* prevented the proliferation of hepatic cells induced by hepatitis B virus through the upregulation of p18. This would provide a treatment ideal for hepatitis B virus-positive patients with HCC [47]. These effects of *HULC* are achieved by its targeting and inhibiting miRNAs, like miR-372. One study showed that different genotypic variations of rs7763881 in *HULC* reduced the susceptibility of Han Chinese patients with long-term hepatitis B virus [48].

3.1.2 | LncRNAs related to drug resistance in HCC

Several differentially expressed lncRNAs in HCC are closely related to drug resistance of liver cancer cells such as *H19*, *lncRNA GAS5* [49], *lncRNA NIFK-AS1* [50], *lncRNA CRNDE* [51], *MALAT1*, and *lncRNA PCGEM1* [52]. *H19* was the first identified lncRNA and is derived from a paternal gene. *H19* is highly expressed in embryo somatic cells and is down-regulated in most tissues rapidly after birth [53]. Notably, *H19* was found to be highly expressed in different types of tumor tissues [54]. A study revealed that *H19* upregulation was related to HCC [55]. Methylated forms of *H19* were related to the overexpression of *H19* in HCC. Researchers found that there are three methylated forms of *H19* in HCC: hyper-, medium-, and hypomethylated *H19*; the hypo- and hyper-profiles were related to *H19* aberrant imprinting [56].

H19 influences gene expression and affects cancer occurrence and development, but the mechanism is not completely understood. *H19* is believed to induce P-glycoprotein expression and *multidrug resistance 1 gene* (*MDR1*)-associated drug resistance through regulation of *MDR1* promoter methylation in HCC cells, which is involved in drug resistance, and thus knockdown of *H19* may be a target in HCC chemotherapy [57]. Knockdown of *H19* in HCC and gastric carcinoma cells prevented the anchoring growth of cancer cells after restoration from hypoxia [58]. Other research results also showed that *H19* prevents the development and growth of cells under hypoxia [59], which has specific implications for tumor tissues with high oxygen consumption, as tumor cells are in the hypoxic state because of the rapid growth of tumor tissues.

3.1.3 | LncRNAs related to liver cancer cell invasion, metastasis, and apoptosis

Multiple lncRNAs such as *lncRNA HOXD-AS1* [52], *MALAT1*, *CEBPA-DT* [60], *lnc-CTHCC* [61], and *HOTAIR* are associated with the metastasis of HCC. Below

TABLE 1 Long noncoding RNA (lncRNA) related to hepatocellular carcinoma (HCC).

	Function	Genes	Result
LncRNAs promotes the progression of HCC	Liver cancer cell proliferation	<i>HULC</i>	<ol style="list-style-type: none"> (1) <i>HULC</i> is not only that it is a biological mark of HCC, but more importantly, it has also been determined that it can be proven to promote HCC cells proliferation. (2) <i>HULC</i> could prevent the proliferation of hepatic cells induced by Hepatitis B virus through the upregulation of P18, which is one of the treatment methods for the Hepatitis B virus positive patients with HCC. (3) Different genotypic variations of rs7763881 in <i>HULC</i> can reduce the sensitive susceptibility degree of the patients with long-term carrying of Hepatitis B virus within the Han Chinese population. (4) <i>HULC</i> could be expected to be a novel biomarker of HCC.
	Drug resistance in HCC	<i>H19</i>	<ol style="list-style-type: none"> (1) <i>H19</i> could methylate the promoter of the multidrug resistance 1 gene (<i>MDR1</i> gene), by which the HCC cells produce the corresponding drug resistance, and could be used as a target for chemotherapy of HCC. (2) Knockdown of <i>H19</i> in HCC and gastric carcinoma cells could prevent the anchoring growth of cancer cells after their restoration from the hypoxia state. (3) <i>H19</i> could prevent the development and growth of cells under hypoxia, which had specific meaning for the tumor tissues with high oxygen consumption.
	Liver cancer cell invasion, metastasis, apoptosis	<i>MALAT1</i>	<ol style="list-style-type: none"> (1) The reduction of the expression of <i>MALAT1</i> in HCC cells can not only effectively reduce the invasion and metastasis of HCC cells, also promote apoptosis of cancer cells. (2) <i>MALAT1</i> may be a potential biomarker for predicting the metastasis of HCC and therapeutic targets. (3) The silencing and suppressing of the gene <i>MALAT1</i> by shRNA could block the cell growth cycle and invasive properties of many malignant tumors. (4) The inhibition of <i>MALAT1</i> could prevent the tumor cells proliferation and invasion in HCC and bladder cancer, also could promote tumor cell apoptosis.
		<i>HOTAIR</i>	<ol style="list-style-type: none"> (1) The inhibition of <i>HOTAIR</i> in HCC cells can decrease MMP9 production, as well as the vascular endothelial growth factor 68, which were found to be very important for the movement and transfer of the cells. (2) Decreasing its expression is expected to become a new type of treatment for HCC.
	HBV infection	<i>HEIH</i>	<ol style="list-style-type: none"> (1) The expression levels of <i>HEIH</i> could be as a biomarker for predicting the survival time. (2) lncRNA-<i>HEIH</i> participates in regulating cell cycles, which could recruit PRC2 might by combining with EZH2, and thereby inhibiting the expression of the downstream target genes, as well as affecting its regulation function.
	Liver cancer prognosis	<i>MVIH</i>	<ol style="list-style-type: none"> (1) The high expression of <i>MVIH</i> was closely related to microvascular invasion, intrahepatic metastasis, and poor prognosis. (2) <i>MVIH</i> could inhibit the secretion of PGK1, and also promote the angiogenesis of tumors.
		<i>HOTTIP/</i> <i>HOXA13</i>	<ol style="list-style-type: none"> (1) <i>HOTTIP</i> has a possible as an early predictive marker of HCC.

TABLE 1 (Continued)

	Function	Genes	Result
			(2) Combined clinicopathological with <i>HOTTIP/HOXA13</i> expression data, which revealed that <i>HOTTIP/HOXA13</i> upregulation expression is related to metastasis of HCC patients and poor prognosis.
LncRNAs can inhibit the progression of HCC	Apoptosis of liver cancer cells	<i>MEG3</i>	(1) Increasing <i>MEG3</i> RNA could inhibit reduce the growth of HCC cells and induce cell apoptosis. (2) <i>Meg3</i> also plays an important role in other diseases such as ischemia-reperfusion injury and inflammation. (3) It can also directly bind to RNA-binding proteins, affecting the function of tumor cells.
	Tumor inhibitory	<i>LET</i>	(1) It was considered to have tumor suppressor activity. (2) The research on lncRNA <i>LET</i> has been reported in gastric cancer, bladder cancer, breast cancer, and so on. It mainly plays its tumor inhibitory function by competing to bind miRNA.
	Inhibit the growth and metastasis of HCC	<i>Dreh</i>	(1) It could inhibit HCC cells proliferation and metastasis. (2) <i>Dreh</i> could potentially interact with intermediate filament protein, inhibit its expression, and could also prevent HCC cells migration by changing the structure and morphology of cell cytoskeletons. (3) <i>Dreh's</i> function mainly focuses on liver cancer and glucose transport, moreover, there are few reports on related signaling pathways.
		<i>T-UCRs</i>	(1) If <i>TUC338</i> was knocked down, the anchorage-dependent, as well as anchorage-independent growth of the HCC cells, would be inhibited. This suggested that <i>T-UCR</i> played significantly role in HCC cells, also provided new ideas for the therapy of HCC targeting lncRNA. (2) <i>T-UCR300a</i> also could block the malignant pathological manifestations of the invasive performance of tumor cells.
	Apoptosis of liver cancer cells	<i>uc002mbe.2</i>	(1) <i>uc002mbe.2</i> played regulatory role in inducing apoptosis of the HCC cells induced by Trichostatin A. (2) <i>uc002mbe.2</i> knockdown could reduce apoptosis induced by TSA and promote the proliferation of cancer HCC cells.

we describe several lncRNAs that have been extensively studied.

3.1.3.1 | *MALAT1*

The *MALAT1* gene is located on chromosome 11q13 and is over 8000 nt in length. *MALAT1* regulates gene expression and influences posttranscriptional modifications of the primary transcripts. *MALAT1* is conserved among all species, indicating the importance of its functions. *MALAT1* was found to be upregulated in multiple tumor types [4]. Reduced expression of *MALAT1* in HCC cells inhibited the invasion and metastasis and promoted the apoptosis of HCC cells. *MALAT1* may thus be a potential biomarker for predicting the metastasis of HCC and a therapeutic target. *MALAT1* was also shown to be an

independent prediction index for the recurrence of HCC. Increased expression of *MALAT1* was identified in HCC cell lines and HCC tissues. Patients with highly expressed *MALAT1* in HCC cells, even following liver transplantation, still face the significant possibility of recurrence [62].

MALAT1 was found to be related with many malignant characteristics of tumors. Inhibition of *MALAT1* expression could reverse malignant characteristics of tumor cells, such as invasiveness, metastasis, and proliferation. Silencing of *MALAT1* by shRNA blocked cell growth and invasion activity of many malignant tumors through a mechanism involving the regulation of multiple genes, including *Caspase-3*, *Caspase-8*, and *Bax* genes [63]. Inhibition of *MALAT1* prevented tumor cell proliferation and

invasion in HCC and bladder cancer, and promoted tumor cell apoptosis [64].

3.1.3.2 | *HOTAIR*

HOTAIR is highly expressed in both primary and metastatic tumor tissues in breast carcinoma [65] and HCC [66]. The expression of *HOTAIR* in liver cancer tissue is significantly higher than that in adjacent tissues, and its expression level significantly correlated with tumor differentiation degree, tumor size, and TNM staging. Patients with distant metastasis, intravascular invasion, or advanced disease have significantly higher expression levels of *HOTAIR*, indicating that high expression of *HOTAIR* may play a role in promoting liver cancer proliferation, migration, and invasion. Furthermore, tumors with *HOTAIR* upregulation were associated with low survival rates [67] and a high recurrence rate [65]. Inhibition of *HOTAIR* in HCC cells decreased MMP9 production and vascular endothelial growth factor [68], which are critical for cell migration. These studies demonstrated that *HOTAIR* is related to the invasion and metastasis of HCC cells. Decreasing its expression may be a potential treatment strategy for HCC.

3.1.4 | LncRNAs associated with HBV infection

HEIH is a highly and specifically expressed lncRNA in HCC tissues that was discovered by Sun et al. in a study on the lncRNA expression spectrums in HCC in para-carcinoma tissues of hepatitis B virus-infected patients using lncRNA chip analysis [6]. Kaplan-Meier analysis determined that *HEIH* expression level may be a biomarker for predicting survival and its expression highly correlated with cancer recurrence. A functional study indicated that *lncRNA-HEIH* regulates the cell cycle and recruits PRC2 by interacting with EZH2 and affecting its transcriptional regulation function, thereby inhibiting the expression of downstream target genes. These findings indicate that *lncRNA-HEIH* plays an important regulatory role in hepatocarcinogenesis.

3.1.5 | LncRNA associated with liver cancer prognosis

3.1.5.1 | *MVIH*

Yuan et al. [69] reported that the level of *MVIH* (NCBI N0. AK094613) in HCC tissues was significantly higher than that in peri-carcinoma tissues and *MVIH* is involved

in the angiogenesis of tumors. The authors performed a microarray analysis of tumor tissues and paired paracancerous tissues of 40 patients with HCC related to hepatitis B. Clinical research data of 215 HCC patients revealed that high expression of *MVIH* was closely related to microvascular invasion, intrahepatic metastasis, and poor prognosis. *MVIH* was found to inhibit the secretion of PGK1 and promote tumor angiogenesis.

3.1.5.2 | *HOTTIP/HOXA13*

The lncRNA *HOXA* transcript at the distal tip (*HOTTIP*) is located in adjacent with *HOXA13* [70]. *HOTTIP*, similar to lncRNA *Xist* and *HOTAIR*, plays a key role in gene expression regulation by influencing chromatin modification [71]. *HOTTIP* and *HOXA13* are upregulated in HCC and are involved in hepatocarcinogenesis [72]. Upregulation of *HOTTIP* was also observed in nontumor liver diseases (such as cirrhosis and HCV-infected cirrhosis), which suggests that *HOTTIP* imbalance may be the early step of *HOXA13* leading to hepatocarcinogenesis [72]. Therefore, *HOTTIP* may be an early predictive marker of HCC. Upregulation of *HOTTIP/HOXA13* expression was found to be related to metastasis and poor prognosis of HCC.

3.2 | LncRNAs that inhibit the progression of HCC

In contrast to the lncRNAs described above, many lncRNAs function as tumor suppressors in HCC through their activities in promoting apoptosis and inhibiting metastasis and proliferation of tumor cells. For example, both lncRNA *CASC2* [73] and *RUNX1-IT1* [74] promote the apoptosis of HCC cells. Below we discuss several important lncRNAs that can inhibit HCC.

3.2.1 | *Maternally expression gene 3 (MEG3)*

MEG3 is expressed in various normal tissues and was found to exhibit a tumor suppression function. The *MEG3* lncRNA gene is a single-copy imprinted gene that contains 10 exons. As a result of various splicing patterns, 12 isoforms are expressed, which display three secondary domains [75, 76].

MEG3 is widely studied, and its expression has been reported in various tumors. *MEG3* was found to interact with cAMP and *p53* [77, 78]. The effect of *MEG3* on *p53* activation depends on the secondary structure of *MEG3* [78]. *MEG3* expression is controlled by epigenetic modifications, and abnormal methylation of CpGs in *MEG3* was observed in a variety of cancer types [79–82]. Compared with normal liver cells, HCC cells show

downregulation of *MEG3* by 210-fold. Increasing *MEG3* RNA inhibited the growth of HCC cells and induced cell apoptosis [82]. Furthermore, the authors found that *miR-29* promotes *MEG3* expression. Compared with wild-type control mice, *miR-29a/b1* knock-out mice showed a downregulation of *MEG3* in liver tissues [82].

MEG3 plays an important role in diseases such as ischemia-reperfusion injury and inflammation. Its mechanism of action is relatively complex, and it regulates cell function by its action as a ceRNA [83]. *MEG3* also directly binds to RNA binding proteins, affecting the function of tumor cells. The role of *MEG3* in hepatocarcinogenesis and development is still under study.

3.2.2 | *LncRNA-LET*

LncRNA LET expression has been reported in gastric cancer, bladder cancer, breast cancer, and HCC. It mainly exerts a tumor inhibitory function through its activity as a ceRNA [84–87].

Yang et al. [88] found that *lncRNA-LET* (NCBI number AK055007) was downregulated in HBV-related HCC. Further study showed that *lncRNA-LET* exhibited reduced expression in HCC, colorectal, and squamous cell lung cancers. Furthermore, downregulated *lncRNA-LET* promoted HCC metastasis. Thus, *lncRNA-LET* was considered to have tumor suppressor activity. Hypoxia-induced histone deacetylase 3 inhibits *lncRNA-LET* by reducing histone acetylation of the *lncRNA-LET* promoter [88]. *lncRNA-LET* downregulation stabilizes nuclear factor 90, which leads to hypoxia-induced invasion of cancer cells.

3.2.3 | *Dreh*

Huang et al. [89] examined the alterations of lncRNA expression induced by *HBx* and found that *Dreh* inhibited the growth and metastasis of HCC. The authors found that *HBx* transgenic mice have a specific profile of liver lncRNAs compared with wild-type mice and identified *lncRNA-Dreh* as a lncRNA downregulated by *HBx*. Functional experiments showed that *Dreh* inhibits HCC cell proliferation and metastasis. *Dreh* may also interact with intermediate filament protein and inhibit its expression, and it prevents HCC cell migration by changing the structure and morphology of cell cytoskeletons. In HCC patients with high expression of *Dreh*, the recurrence rate was low and the prognosis was good. While research on *Dreh* function mainly focuses on liver cancer and glucose transport, some studies have investigated the related signaling pathways.

3.2.4 | *T-UCR338*

Transcribed ultra-conserved regions (T-UCRs), a class of lncRNAs, are transcribed from ultra-conserved regions [12]. *UCRs* are DNA noncoding genomic segments of at least 200 bp in length that are completely conserved across humans, mice, and rats. A total of 481 *UCRs* have been identified, some of which overlap with coding exons; more than half of them are noncoding genes [90]. Approximately 68% of *UCRs* are transcribed, constituting a new category of ncRNAs, the *T-UCRs* [91]. *T-UCR* expressions are altered in human tumorigenesis, such as leukemia, neuroblastoma, colorectal cancer, and HCC [91–93]. *T-UCRs* are aberrantly expressed in malignant hepatocytes. *T-UCR 338* is a *T-UCR* with a length of 590 nt. Knockdown of *TUC338* led to inhibition of anchorage-dependent and anchorage-independent growth of HCC cells. This suggested that *T-UCR* plays a significant role in regulating the growth of HCC cells [93]. Other studies have shown that reducing other *T-UCRs*, such as *T-UCR300a*, could block the invasion of HCC cells [94]. These findings may lead to the development of new therapeutic strategies for HCC.

3.2.5 | *uc002mbe.2*

The lncRNA *uc002mbe.2* is downregulated in liver cancer, and its downregulation inhibits the apoptosis of tumor cells. Furthermore, *uc002mbe.2* mediates trichostatin-induced apoptosis of liver cancer cells [95, 96]. Chen et al. [97] found that the interaction between *uc002mbe.2* and *hnRNPA2B1* can mediate AKT deactivation and p21 induction is related, thereby participating in the cytostatic effect of trichostatin in HCC cells. The global expression of lncRNAs and coding genes was analyzed with the Human lncRNA Array V2.0 after 24 h treatment of liver cancer cells with Trichostatin A. Among the differentially expressed lncRNAs, *uc002mbe.2* change the most. Knockdown of *uc002mbe.2* reduced the apoptosis induced by TSA and promoted the proliferation of HCC cells. Moreover, *uc002mbe.2* was significantly deregulated in HCC cell lines and tissues compared with normal human hepatocytes and adjacent noncancerous tissues. The function of *uc002mbe.2* in other tumors has not been reported yet.

4 | FUTURE PROSPECTS

The functions of lncRNA are complex and diverse. In cancer, lncRNAs can function as oncogenic factors that promote tumor occurrence and development or tumor suppressors that inhibit cancer growth. Some lncRNAs

promote tumor distant metastasis and are associated with poor prognosis while some lncRNAs are associated with improved prognosis. LncRNAs may serve as tumor molecular markers or therapeutic targets, providing new strategies for the diagnosis and treatment of tumors. More research on the relationship between lncRNAs and tumor development is required.

LncRNAs form very large and complex post-transcriptional and pre-protein translation regulatory networks. The number of lncRNAs is large, and the number of lncRNAs in human cells can reach up to tens of thousands. However, the lncRNAs that have been currently studied only account for a few of the total lncRNAs, and research on the function of lncRNA in tumors, including in HCC, remains in the initial stage. With further investigations on the molecular mechanism of HCC, new lncRNAs related to HCC will be continuously discovered, and the regulation mechanisms will be further revealed. These findings will help provide insights to aid in early diagnosis of HCC, establish molecular targeted therapies, and improve the survival of HCC patients.

AUTHOR CONTRIBUTIONS

Jingli Du: Conceptualization (equal); methodology (equal); resources (equal); writing—original draft (equal). **Yue Su:** Data curation (equal); resources (equal). **Jianzhi Gao:** Investigation (equal); supervision (equal). **Yanhong Tai:** Formal analysis (equal); Writing—review and editing (equal).

ACKNOWLEDGMENTS

None.

CONFLICT OF INTEREST STATEMENT

The authors declare no conflict of interest.

DATA AVAILABILITY STATEMENT

Data are openly available in a public repository that issues data sets with DOIs.

ETHICS STATEMENT

Not Applicable.

INFORMED CONSENT

Not Applicable.

ORCID

Jingli Du  <http://orcid.org/0000-0002-4852-5864>

REFERENCES

- Fattahi N, Rezaei N, Asadi-Lari M, Yousefi M, Madadi Z, Gohari K, et al. Hepatocellular carcinoma incidence at national and provincial levels in Iran from 2000 to 2016: a meta-regression analysis. *PLoS One*. 2021;16(1):e0245468. <https://doi.org/10.1371/journal.pone.0245468>
- Peng WX, Koirala P, Mo YY. LncRNA-mediated regulation of cell signaling in cancer. *Oncogene*. 2017;36(41):5661–7. <https://doi.org/10.1038/onc.2017.184>
- Bhat AA, Younes SN, Raza SS, Zarif L, Nisar S, Ahmed I, et al. Role of non-coding RNA networks in leukemia progression, metastasis and drug resistance. *Mol Cancer*. 2020;19(1):57. <https://doi.org/10.1186/s12943-020-01175-9>
- Tang X, Qiao X, Chen C, Liu Y, Zhu J, Liu J. Regulation mechanism of long noncoding RNAs in colon cancer development and progression. *Yonsei Med J*. 2019;60(4):319–25. <https://doi.org/10.3349/ymj.2019.60.4.319>
- Hua JT, Chen S, He HH. Landscape of noncoding RNA in prostate cancer. *TIG*. 2019;35(11):840–51. <https://doi.org/10.1016/j.tig.2019.08.004>
- Yang F, Zhang L, Huo XS, Yuan JH, Xu D, Yuan SX et al. Long noncoding RNA high expression in hepatocellular carcinoma facilitates tumor growth through enhancer of zeste homolog 2 in humans. *Hepatology*. 2011;54(5):1679–89. <https://doi.org/10.1002/hep.24563>
- Wang J, Zhu S, Meng N, He Y, Lu R, Yan GR. ncRNA-encoded peptides or proteins and cancer. *Mol Ther*. 2019;27(10):1718–25. <https://doi.org/10.1016/j.ymthe.2019.09.001>
- Goodall GJ, Wickramasinghe VO. RNA in cancer. *Nat Rev Cancer*. 2021;21(1):22–36. <https://doi.org/10.1038/s41568-020-00306-0>
- Wu T, Du Y. LncRNAs: from basic research to medical application. *Int J Biol Sci*. 2017;13(3):295–307. <https://doi.org/10.7150/ijbs.16968>
- Pardini B, Calin G. MicroRNAs and long non-coding RNAs and their hormone-like activities in cancer. *Cancers*. 2019;11(3):378. <https://doi.org/10.3390/cancers11030378>
- Ratti M, Lampis A, Ghidini M, Salati M, Mirchev MB, Valeri N, et al. MicroRNAs (miRNAs) and long non-coding RNAs (lncRNAs) as new tools for cancer therapy: first steps from bench to bedside. *Target Oncol*. 2020;15(3):261–78. <https://doi.org/10.1007/s11523-020-00717-x>
- Kopp F. Molecular functions and biological roles of long non-coding RNAs in human physiology and disease. *J Gene Med*. 2019;21(8):e3104. <https://doi.org/10.1002/jgm.3104>
- Ali T, Grote P. Beyond the RNA-dependent function of LncRNA genes. *eLife*. 2020;9:e60583. <https://doi.org/10.7554/eLife.60583>
- Jarroux J, Morillon A, Pinskaya M. History, discovery, and classification of lncRNAs. *Adv Exp Med Biol*. 2017;1008:1–46. https://doi.org/10.1007/978-981-10-5203-3_1
- Weng W, Zhang Z, Huang W, Xu X, Wu B, Ye T, et al. Correction to: identification of a competing endogenous RNA network associated with prognosis of pancreatic adenocarcinoma. *Cancer Cell Int*. 2020;20:282. <https://doi.org/10.1186/s12935-020-01381-x>
- Zhou B, Yang H, Yang C, Bao Y, Yang S, Liu J, et al. Translation of noncoding RNAs and cancer. *Cancer Lett*. 2021;497:89–99. <https://doi.org/10.1016/j.canlet.2020.10.002>
- Yamamoto T, Saitoh N. Non-coding RNAs and chromatin domains. *Curr Opin Cell Biol*. 2019;58:26–33. <https://doi.org/10.1016/j.ceb.2018.12.005>
- Guo X, Plank-Bazinet J, Krivega I, Dale RK, Dean A. Embryonic erythropoiesis and hemoglobin switching require

- transcriptional repressor ETO2 to modulate chromatin organization. *Nucleic Acids Res.* 2020;48(18):10226–40. <https://doi.org/10.1093/nar/gkaa736>
19. Ribeiro de Almeida C, Hendriks RW, Stadhouders R. Dynamic control of long-range genomic interactions at the immunoglobulin κ light-chain locus. *Adv Immunol.* 2015;128:183–271. <https://doi.org/10.1016/bs.ai.2015.07.004>
 20. Trendel J, Schwarzl T, Horos R, Prakash A, Bateman A, Hentze MW, et al. The human RNA-binding proteome and its dynamics during translational arrest. *Cell.* 2019;176(1–2):391–403. <https://doi.org/10.1016/j.cell.2018.11.004>
 21. Raut SK, Khullar M. The big entity of new RNA world: long non-coding RNAs in microvascular complications of diabetes. *Front Endocrinol.* 2018;9:300. <https://doi.org/10.3389/fendo.2018.00300>
 22. Barman P, Reddy D, Bhaumik SR. Mechanisms of antisense transcription initiation with implications in gene expression, genomic integrity and disease pathogenesis. *noncoding. RNA.* 2019;5(1):11. <https://doi.org/10.3390/ncrna5010011>
 23. Sachani SS, Landschoot LS, Zhang L, White CR, MacDonald WA, Golding MC, et al. Nucleoporin 107, 62 and 153 mediate Kcnq1ot1 imprinted domain regulation in extraembryonic endoderm stem cells. *Nat Commun.* 2018;9(1):2795. <https://doi.org/10.1038/s41467-018-05208-2>
 24. Llères D, Imaizumi Y, Feil R. Exploring chromatin structural roles of non-coding RNAs at imprinted domains. *Biochem Soc Trans.* 2021;49(4):1867–79. <https://doi.org/10.1042/BST20210758>
 25. Sanli I, Feil R. Chromatin mechanisms in the developmental control of imprinted gene expression. *Int J Biochem Cell Biol.* 2015;67:139–47. <https://doi.org/10.1016/j.biocel.2015.04.004>
 26. Pinter SF. A tale of two cities: how xist and its partners localize to and silence the bicompartamental X. *Semin Cell Dev Biol.* 2016;56:19–34. <https://doi.org/10.1016/j.semcdb.2016.03.023>
 27. Yu W, Gius D, Onyango P, Muldoon-Jacobs K, Karp J, Feinberg AP, et al. Epigenetic silencing of tumour suppressor gene *p15* by its antisense RNA. *Nature.* 2008;451(7175):202–6. <https://doi.org/10.1038/nature06468>
 28. Sanchez Calle A, Kawamura Y, Yamamoto Y, Takeshita F, Ochiya T. Emerging roles of long non-coding RNA in cancer. *Cancer Sci.* 2018;109(7):2093–100. <https://doi.org/10.1111/cas.13642>
 29. Quan M, Chen J, Zhang D. Exploring the secrets of long noncoding RNAs. *Int J Mol Sci.* 2015;16(3):5467–96. <https://doi.org/10.3390/ijms16035467>
 30. Ouyang J, Zhong Y, Zhang Y, Yang L, Wu P, Hou X, et al. Long non-coding RNAs are involved in alternative splicing and promote cancer progression. *Br J Cancer.* 2022;126(8):1113–24. <https://doi.org/10.1038/s41416-021-01600-w>
 31. Statello L, Guo CJ, Chen LL, Huarte M. Gene regulation by long non-coding RNAs and its biological functions. *Nat Rev Mol Cell Biol.* 2021;22(2):96–118. <https://doi.org/10.1038/s41580-020-00315-9>
 32. Zeng T, Ni H, Yu Y, Zhang M, Wu M, Wang Q, et al. BACE1-AS prevents BACE1 mRNA degradation through the sequestration of BACE1-targeting miRNAs. *J Chem Neuroanat.* 2019;98:87–96. <https://doi.org/10.1016/j.jchemneu.2019.04.001>
 33. Fischer JW, Busa VF, Shao Y, Leung AKL. Structure-mediated RNA decay by UPF1 and G3BP1. *Mol Cell.* 2020;78(1):70–84. <https://doi.org/10.1016/j.molcel.2020.01.021>
 34. Guay C, Abdulkarim B, Tan JY, Dubuis G, Rütli S, Laybutt DR, et al. Loss-of-function of the long non-coding RNA A830019P07Rik in mice does not affect insulin expression and secretion. *Sci Rep.* 2020;10(1):6413. <https://doi.org/10.1038/s41598-020-62969-x>
 35. Cooper D, Carter G, Li P, Patel R, Watson J, Patel N. Long non-coding RNA NEAT1 associates with SRp40 to temporally regulate PPAR γ 2 splicing during adipogenesis in 3T3-L1 cells. *Genes.* 2014;5(4):1050–63. <https://doi.org/10.3390/genes5041050>
 36. Yu AM, Choi YH, Tu MJ. RNA drugs and RNA targets for small molecules: principles, progress, and challenges. *Pharmacol Rev.* 2020;72(4):862–98. <https://doi.org/10.1124/pr.120.019554>
 37. Li Y, Egranov SD, Yang L, Lin C. Molecular mechanisms of long noncoding RNAs-mediated cancer metastasis. *Genes Chromosom Cancer.* 2019;58(4):200–7. <https://doi.org/10.1002/gcc.22691>
 38. Sheng L, Ye L, Zhang D, Cawthorn WP, Xu B. New insights into the long non-coding RNA SRA: physiological functions and mechanisms of action. *Front Med.* 2018;5:244. <https://doi.org/10.3389/fmed.2018.00244>
 39. Peckham E, Scheurer M, Danysh H, Lubega J, Langlois P, Lupo P. Residential radon exposure and incidence of childhood lymphoma in Texas, 1995–2011. *Int J Environ Res Public Health.* 2015;12(10):12110–26. <https://doi.org/10.3390/ijerph121012110>
 40. Pan Y, Qin T, Feng L, Yu Z. Expression profile of altered long non-coding RNAs in patients with HBV-associated hepatocellular carcinoma. *J Huazhong Univ Sci Technol.* 2013;33(1):96–101. <https://doi.org/10.1007/s11596-013-1078-y>
 41. Xu K, Xia P, Gongye X, Zhang X, Ma S, Chen Z, et al. A novel lncRNA RP11-386G11.10 reprograms lipid metabolism to promote hepatocellular carcinoma progression. *Mol Metab.* 2022;63:101540. <https://doi.org/10.1016/j.molmet.2022.101540>
 42. Liu F, Tian T, Zhang Z, Xie S, Yang J, Zhu L, et al. Long non-coding RNA SNHG6 couples cholesterol sensing with mTORC1 activation in hepatocellular carcinoma. *Nat Metab.* 2022;4(8):1022–40. <https://doi.org/10.1038/s42255-022-00616-7>
 43. Peng JY, Cai DK, Zeng RL, Zhang CY, Li GC, Chen SF, et al. Upregulation of Superenhancer-driven lncRNA FASRL by USF1 promotes de novo fatty acid biosynthesis to exacerbate hepatocellular carcinoma. *Adv Sci (Weinh).* 2022;10(1):e2204711. <https://doi.org/10.1002/adv.202204711>
 44. Wang L, Li B, Bo X, Yi X, Xiao X, Zheng Q. Hypoxia-induced lncRNA DACT3-AS1 upregulates PKM2 to promote metastasis in hepatocellular carcinoma through the HDAC2/FOXA3 pathway. *Exp Mol Med.* 2022;54(6):848–60. <https://doi.org/10.1038/s12276-022-00767-3>
 45. Shaker O, Mahfouz H, Salama A, Medhat E. Long non-coding HULC and miRNA-372 as diagnostic biomarkers in hepatocellular carcinoma. *Rep Biochem Mol Biol.* 2020;9(2):230–40. <https://doi.org/10.29252/rbmb.9.2.230>
 46. Ruan L, Huang L, Zhao L, Wang Q, Pan X, Zhang A, et al. The interaction of lncRNA-HEIH and lncRNA-HULC with HBXIP in hepatitis B patients. *Gastroenterol Res Pract.* 2018;2018:1–6. <https://doi.org/10.1155/2018/9187316>
 47. Klec C, Gutschner T, Panzitt K, Pichler M. Involvement of long non-coding RNA HULC (highly up-regulated in liver

- cancer) in pathogenesis and implications for therapeutic intervention. *Expert Opin Ther Targets*. 2019;23(3):177–86. <https://doi.org/10.1080/14728222.2019.1570499>
48. Liu Y, Pan S, Liu L, Zhai X, Liu J, Wen J, et al. A genetic variant in long non-coding RNA HULC contributes to risk of HBV-related hepatocellular carcinoma in a Chinese population. *PLoS One*. 2012;7(4):e35145. <https://doi.org/10.1371/journal.pone.0035145>
 49. Wang C, Ke S, Li M, Lin C, Liu X, Pan Q. Downregulation of lncRNA GAS5 promotes liver cancer proliferation and drug resistance by decreasing PTEN expression. *Mol Genet Genomics*. 2020;295(1):251–60. <https://doi.org/10.1007/s00438-019-01620-5>
 50. Chen Y, Xiang D, Zhao X, Chu X. Upregulation of lncRNA NIFK-AS1 in hepatocellular carcinoma by m(6A) methylation promotes disease progression and sorafenib resistance. *Hum Cell*. 2021;34(6):1800–11. <https://doi.org/10.1007/s13577-021-00587-z>
 51. Han S, Han B, Li Z, Sun D. Retracted article: downregulation of long noncoding RNA CRNDE suppresses drug resistance of liver cancer cells by increasing microRNA-33a expression and decreasing HMGA2 expression. *Cell Cycle*. 2019;18(19):2524–37. <https://doi.org/10.1080/15384101.2019.1652035>
 52. Wang H, Huo X, Yang XR, He J, Cheng L, Wang N, et al. STAT3-mediated upregulation of lncRNA HOXD-AS1 as a ceRNA facilitates liver cancer metastasis by regulating SOX4. *Mol Cancer*. 2017;16(1):136. <https://doi.org/10.1186/s12943-017-0680-1>
 53. Castle JC, Armour CD, Löwer M, Haynor D, Biery M, Bouzek H, et al. Digital genome-wide ncRNA expression, including snoRNAs, across 11 human tissues using polyA-neutral amplification. *PLoS One*. 2010;5(7):e11779. <https://doi.org/10.1371/journal.pone.0011779>
 54. Olivero CE, Dimitrova N. Identification and characterization of functional long noncoding RNAs in cancer. *FASEB J*. 2020;34(12):15630–46. <https://doi.org/10.1096/fj.202001951R>
 55. Tietze L, Kessler SM. The good, the bad, the question-H19 in hepatocellular carcinoma. *Cancers*. 2020;12(5):1261. <https://doi.org/10.3390/cancers12051261>
 56. Ardelit MA, Pachmayr J. The long non-coding RNA H19—a new player in hepatocellular carcinoma. *Cell Stress*. 2017;1(1):4–6. <https://doi.org/10.15698/cst2017.10.102>
 57. Tsang WP, Kwok TT. Riboregulator H19 induction of MDR1-associated drug resistance in human hepatocellular carcinoma cells. *Oncogene*. 2007;26(33):4877–81. <https://doi.org/10.1038/sj.onc.1210266>
 58. Ye Y, Shen A, Liu A. Long non-coding RNA H19 and cancer: a competing endogenous RNA. *Bull Cancer*. 2019;106(12):1152–9. <https://doi.org/10.1016/j.bulcan.2019.08.011>
 59. Raveh E, Matouk IJ, Gilon M, Hochberg A. The H19 long non-coding RNA in cancer initiation, progression and metastasis—a proposed unifying theory. *Mol Cancer*. 2015;14:184. <https://doi.org/10.1186/s12943-015-0458-2>
 60. Cai Y, Lyu T, Li H, Liu C, Xie K, Xu L, et al. lncRNA CEBPA-DT promotes liver cancer metastasis through DDR2/ β -catenin activation via interacting with hnRNPC. *J Exp Clin Cancer Res*. 2022;41(1):335. <https://doi.org/10.1186/s13046-022-02544-6>
 61. Xia A, Yuan W, Wang Q, Xu J, Gu Y, Zhang L, et al. The cancer-testis lncRNA lnc-CTHCC promotes hepatocellular carcinogenesis by binding hnRNP K and activating YAP1 transcription. *Nat Cancer*. 2022;3(2):203–18. <https://doi.org/10.1038/s43018-021-00315-4>
 62. Lai M, Yang Z, Zhou L, Zhu Q, Xie H, Zhang F, et al. Long non-coding RNA MALAT-1 overexpression predicts tumor recurrence of hepatocellular carcinoma after liver transplantation. *Med Oncol*. 2012;29(3):1810–6. <https://doi.org/10.1007/s12032-011-0004-z>
 63. Hu J, Sun Z, Hu K, Tang M, Sun S, Fang Y, et al. Overexpression of Hsa-miR-23b-3p suppresses proliferation, migration, invasion and epithelial-mesenchymal transition of human cervical cancer CasKi cells. *Xi Bao Yu Fen Zi Mian Yi Xue Za Zhi*. 2020;36(11):983–9. (Chinese)
 64. Han Y, Liu Y, Nie L, Gui Y, Cai Z. Inducing cell proliferation inhibition, apoptosis, and motility reduction by silencing long noncoding ribonucleic acid metastasis-associated lung adenocarcinoma transcript 1 in urothelial carcinoma of the bladder. *Urology*. 2013;81(1):209.e1–7. <https://doi.org/10.1016/j.urology.2012.08.044>
 65. Huang H, Li L, Wen K. Interactions between long non-coding RNAs and RNA-binding proteins in cancer (review). *Oncol Rep*. 2021;46(6):256. <https://doi.org/10.3892/or.2021.8207>
 66. Zhong DN, Luo YH, Mo WJ, Zhang X, Tan Z, Zhao N, et al. High expression of long non-coding HOTAIR correlated with hepatocarcinogenesis and metastasis. *Mol Med Rep*. 2018;17(1):1148–56.
 67. Jiang D, Xu L, Ni J, Zhang J, Cai M, Shen L. Functional polymorphisms in lncRNA HOTAIR contribute to susceptibility of pancreatic cancer. *Cancer Cell Int*. 2019;19:47. <https://doi.org/10.1186/s12935-019-0761-x>
 68. Geng Y, Xie S, Li Q, Ma J, Wang G. Large intervening non-coding RNA HOTAIR is associated with hepatocellular carcinoma progression. *J Int Med Res*. 2011;39(6):2119–28. <https://doi.org/10.1177/147323001103900608>
 69. Yuan SX, Yang F, Yang Y, Tao QF, Zhang J, Huang G, et al. Long noncoding RNA associated with microvascular invasion in hepatocellular carcinoma promotes angiogenesis and serves as a predictor for hepatocellular carcinoma patients' poor recurrence-free survival after hepatectomy. *Hepatology*. 2012;56(6):2231–41. <https://doi.org/10.1002/hep.25895>
 70. Murillo-Maldonado JM, Riesgo-Escovar JR. The various and shared roles of lncRNAs during development. *Dev Dyn*. 2019;248(11):1059–69. <https://doi.org/10.1002/dvdy.108>
 71. Dykes IM, Emanuelli C. Transcriptional and post-transcriptional gene regulation by long non-coding RNA. *Genomics Insights*. 2017;15(3):177–86. <https://doi.org/10.1016/j.gpb.2016.12.005>
 72. Quagliata L, Matter MS, Piscuoglio S, Arabi L, Ruiz C, Procino A, et al. Long noncoding RNA HOTTIP/HOXA13 expression is associated with disease progression and predicts outcome in hepatocellular carcinoma patients. *Hepatology*. 2014;59(3):911–23. <https://doi.org/10.1002/hep.26740>
 73. Fan JC, Zeng F, Le YG, Xin L. lncRNA CASC2 inhibited the viability and induced the apoptosis of hepatocellular carcinoma cells through regulating miR-24-3p. *JCB*. 2018;119(8):6391–7. <https://doi.org/10.1002/jcb.26479>
 74. Yan PH, Wang L, Chen H, Yu FQ, Guo L, Liu Y, et al. lncRNA RUNX1-IT1 inhibits proliferation and promotes apoptosis of hepatocellular carcinoma by regulating MAPK pathways. *Eur Rev Med Pharmacol Sci*. 2019;23(19):8287–94. https://doi.org/10.26355/eurrev_201910_19139

75. Hamilton S, de Cabo R, Bernier M. Maternally expressed gene 3 in metabolic programming. *Biochim Biophys Acta Gene Regul Mech.* 2020;1863(4):194396. <https://doi.org/10.1016/j.bbagr.2019.06.007>
76. Zhou Y, Zhang X, Klibanski A. MEG3 noncoding RNA: a tumor suppressor. *J Mol Endocrinol.* 2012;48(3):R45–53. <https://doi.org/10.1530/JME-12-0008>
77. Modali SD, Parekh VI, Kebebew E, Agarwal SK. Epigenetic regulation of the lncRNA MEG3 and its target c-MET in pancreatic neuroendocrine tumors. *Mol Endocrinol.* 2015;29(2):224–37. <https://doi.org/10.1210/me.2014-1304>
78. Li J, Bian EB, He XJ, Ma CC, Zong G, Wang HL, et al. Epigenetic repression of long non-coding RNA MEG3 mediated by DNMT1 represses the p53 pathway in gliomas. *Int J Oncol.* 2016;48(2):723–33. <https://doi.org/10.3892/ijo.2015.3285>
79. Gao Y, Huang P, Zhang J. Hypermethylation of MEG3 promoter correlates with inactivation of MEG3 and poor prognosis in patients with retinoblastoma. *J Transl Med.* 2017;15(1):268. <https://doi.org/10.1186/s12967-017-1372-8>
80. Gejman R, Batista DL, Zhong Y, Zhou Y, Zhang X, Swearingen B, et al. Selective loss of MEG3 expression and intergenic differentially methylated region hypermethylation in the MEG3/DLK1 locus in human clinically nonfunctioning pituitary adenomas. *J Clin Endocrinol Metab.* 2008;93(10):4119–25. <https://doi.org/10.1210/jc.2007-2633>
81. Nardi V, Hasserjian RP. Genetic testing in acute myeloid leukemia and myelodysplastic syndromes. *Surgical Pathology Clinics.* 2016;9(1):143–63. <https://doi.org/10.1016/j.path.2015.10.004>
82. Li Y, Ren M, Zhao Y, Lu X, Wang M, Hu J, et al. MicroRNA-26a inhibits proliferation and metastasis of human hepatocellular carcinoma by regulating DNMT3B-MEG3 axis. *Oncol Rep.* 2017;37(6):3527–35. <https://doi.org/10.3892/or.2017.5579>
83. Sun HJ, Zhang FF, Xiao Q, Xu J, Zhu LJ. lncRNA MEG3, acting as a ceRNA, modulates RPE differentiation through the miR-7-5p/Pax6 axis. *Biochem Genet.* 2021;59(6):1617–30. <https://doi.org/10.1007/s10528-021-10072-9>
84. Zhou CX, Wang X, Yang N, Xue SK, Li WC, Xie PP. lncRNA LET function as a tumor suppressor in breast cancer development. *Eur Rev Med Pharmacol Sci.* 2018;22(18):6002–7. https://doi.org/10.26355/eurrev_201809_15935
85. Liang JH, Xu QD, Gu SG. lncRNA RSU1P2-microRNA let-7a-testis-expressed protein 10 axis modulates tumorigenesis and cancer stem cell-like properties in liver cancer. *Bioengineered.* 2022;13(2):4285–300. <https://doi.org/10.1080/21655979.2022.2031394>
86. Retraction: lncRNA DANCR promotes migration and invasion through suppression of lncRNA-LET in gastric cancer cells. *Biosci Rep.* 2020;40(10):BSR-20171070_RET. https://doi.org/10.1042/BSR-20171070_RET
87. Zhuang J, Shen L, Yang L, Huang X, Lu Q, Cui Y, et al. TGFβ1 promotes gemcitabine resistance through regulating the lncRNA-LET/NF90/miR-145 signaling axis in bladder cancer. *Theranostics.* 2017;7(12):3053–67. <https://doi.org/10.7150/thno.19542>
88. Yang F, Huo X, Yuan S, Zhang L, Zhou W, Wang F, et al. Repression of the long noncoding RNA-LET by histone deacetylase 3 contributes to hypoxia-mediated metastasis. *Mol Cell.* 2013;49(6):1083–96. <https://doi.org/10.1016/j.molcel.2013.01.010>
89. Huang J, Guo Y, Zhao C, Yuan S, Wang Y, Tang G, et al. Hepatitis B virus X protein (HBx)-related long noncoding RNA (lncRNA) down-regulated expression by HBx (Dreh) inhibits hepatocellular carcinoma metastasis by targeting the intermediate filament protein vimentin. *Hepatology.* 2013;57(5):1882–92. <https://doi.org/10.1002/hep.26195>
90. Habic A, Mattick JS, Calin GA, Kreske R, Konc J, Kunej T. Genetic variations of ultraconserved elements in the human genome. *OMICS J Integr Biol.* 2019;23(11):549–59. <https://doi.org/10.1089/omi.2019.0156>
91. Calin GA, Liu C, Ferracin M, Hyslop T, Spizzo R, Sevignani C, et al. Ultraconserved regions encoding ncRNAs are altered in human leukemias and carcinomas. *Cancer Cell.* 2007;12(3):215–29. <https://doi.org/10.1016/j.ccr.2007.07.027>
92. Watters KM, Bryan K, Foley NH, Meehan M, Stallings RL. Expressional alterations in functional ultra-conserved non-coding RNAs in response to all-trans retinoic acid-induced differentiation in neuroblastoma cells. *BMC Cancer.* 2013;13:184. <https://doi.org/10.1186/1471-2407-13-184>
93. Bo C, Li N, Li X, Liang X, An Y. Long noncoding RNA uc.338 promotes cell proliferation through association with BMI1 in hepatocellular carcinoma. *Hum Cell.* 2016;29(4):141–7. <https://doi.org/10.1007/s13577-016-0140-z>
94. Wei C, Yang C, Wang S, Shi D, Zhang C, Lin X, et al. Crosstalk between cancer cells and tumor associated macrophages is required for mesenchymal circulating tumor cell-mediated colorectal cancer metastasis. *Mol Cancer.* 2019;18(1):64. <https://doi.org/10.1186/s12943-019-0976-4>
95. Yu J, Han J, Zhang J, Li G, Liu H, Cui X, et al. The long noncoding RNAs PVT1 and uc002mbe.2 in sera provide a new supplementary method for hepatocellular carcinoma diagnosis. *Medicine.* 2016;95(31):e4436. <https://doi.org/10.1097/MD.0000000000004436>
96. Yang H, Zhong Y, Xie H, Lai X, Xu M, Nie Y, et al. Induction of the liver cancer-down-regulated long noncoding RNA uc002mbe.2 mediates trichostatin-induced apoptosis of liver cancer cells. *Biochem Pharmacol.* 2013;85(12):1761–9. <https://doi.org/10.1016/j.bcp.2013.04.020>
97. Chen T, Gu C, Xue C, Yang T, Zhong Y, Liu S, et al. lncRNA-uc002mbe.2 interacting with hnRNPA2B1 mediates AKT deactivation and p21 up-regulation induced by trichostatin in liver cancer cells. *Front Pharmacol.* 2017;8:669. <https://doi.org/10.3389/fphar.2017.00669>

How to cite this article: Du J, Su Y, Gao J, Tai Y. The expression and function of long noncoding RNAs in hepatocellular carcinoma. *Cancer Innov.* 2023;2:488–499. <https://doi.org/10.1002/cai2.90>