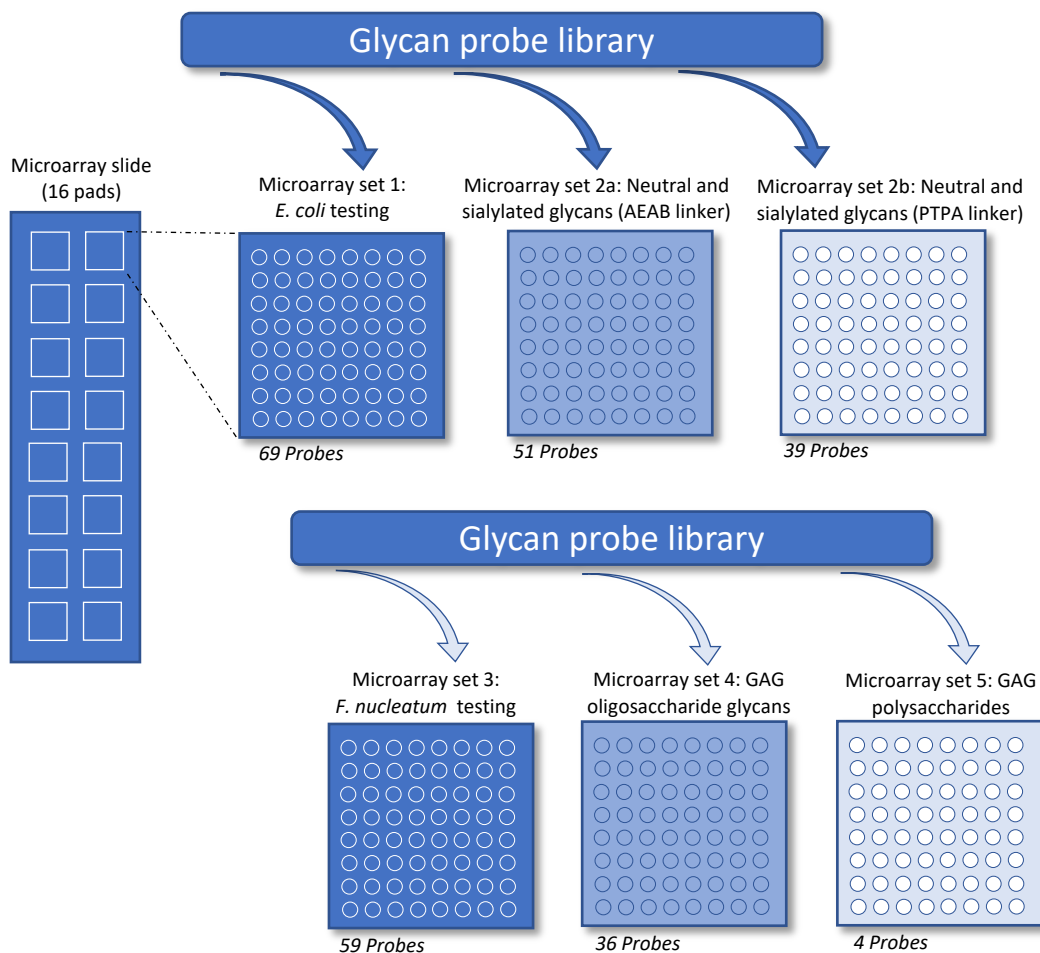
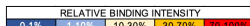




Supplementary Figure 1: Map of glycan microarray sets used in the study. Diagram representing the six glycan microarray sets used for analysis of whole bacteria binding to sequence-defined glycans. Microarray set 1 with a subset of 69 probes was used to investigate binding with four *E. coli* strains and for comparison of live and fixed *E. coli* C600 binding. Microarray set 3 was used for glycan binding comparison analysis of live and fixed *F. nucleatum* 23726 cultures. Microarray set 2a and 2b with a total of 90 neutral and sialylated glycans and Microarray set 4 with 36 GAG oligosaccharide probes were used to test the whole bacterial collection of 22 strains (Supplementary data 4) . Microarray set 5 with four GAG polysaccharide probes was used to test selected bacteria (Fig. 5B-C).

Concanavalin A



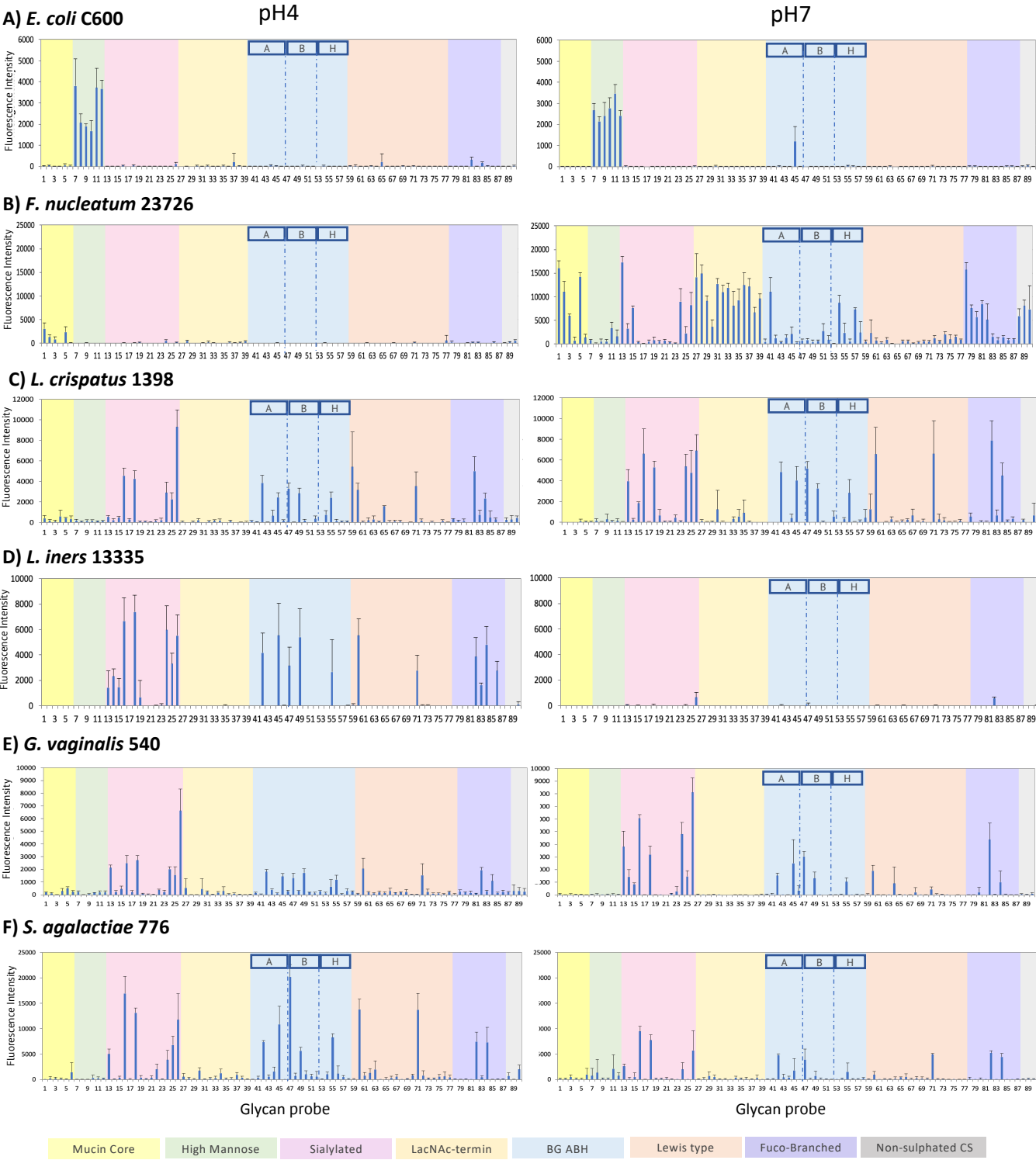
<i>E. coli</i> C600 Assay_1				<i>E. coli</i> C600 Assay_2			<i>E. coli</i> C600 Assay_3			
Probe Name	MFI	SD	RANK	MFI	SD	RANK	MFI	SD	RANK	RANK 3
Man5-GN2-AEAB	4681	1348	71	13832	1139	100	10470	5608	100	90
Man5-GN2-Gly	6428	719	97	6887	923	50	9963	2444	95	81
Man6-GN2-AEAB	5266	1558	80	7139	3976	52	8754	2795	84	72
Man8-GN2-AEAB	5707	2618	86	12414	1159	90	3812	1780	36	71
Man9-GN2-AEAB	5702	1405	86	8750	4240	63	4309	2244	41	64
Man6-GN2-Gly	6619	1811	100	6130	197	44	4058	1384	39	61
Man7D1GN2-AEAB	5992	2520	91	6742	760	49	4527	736	43	61
Man7D3GN2-AEAB	2850	1325	43	3167	3778	23	1639	715	16	27
NA2-Gly	336	668	5	2738	1870	20	826	291	8	11

<i>E. coli</i> 789 Assay_1				<i>E. coli</i> 789 Assay_2			<i>E. coli</i> 789 Assay_3			
Probe Name	MFI	SD	RANK	MFI	SD	RANK	MFI	SD	RANK	RANK 3
Man6-GN2-AEAB	7814	3442	85	7992	1573	100	6078	2983	71	86
Man5-GN2-Gly	7913	2849	87	7822	518	98	4937	1914	58	81
Man7D1GN2-AEAB	5494	562	60	6146	436	77	8530	1747	100	79
Man6-GN2-Gly	6572	3218	72	5631	258	70	6943	1881	81	75
Man9-GN2-AEAB	9146	3722	100	5691	2823	71	3057	996	36	69
Man5-GN2-AEAB	6521	3099	71	7422	900	93	2541	372	30	65
Man8-GN2-AEAB	3384	2320	37	6602	698	83	2953	1071	35	51
Man7D3GN2-AEAB	1614	1038	18	4430	1105	55	5132	1270	60	44
NA2-Gly	786	848	9	2567	840	32	2546	427	30	24

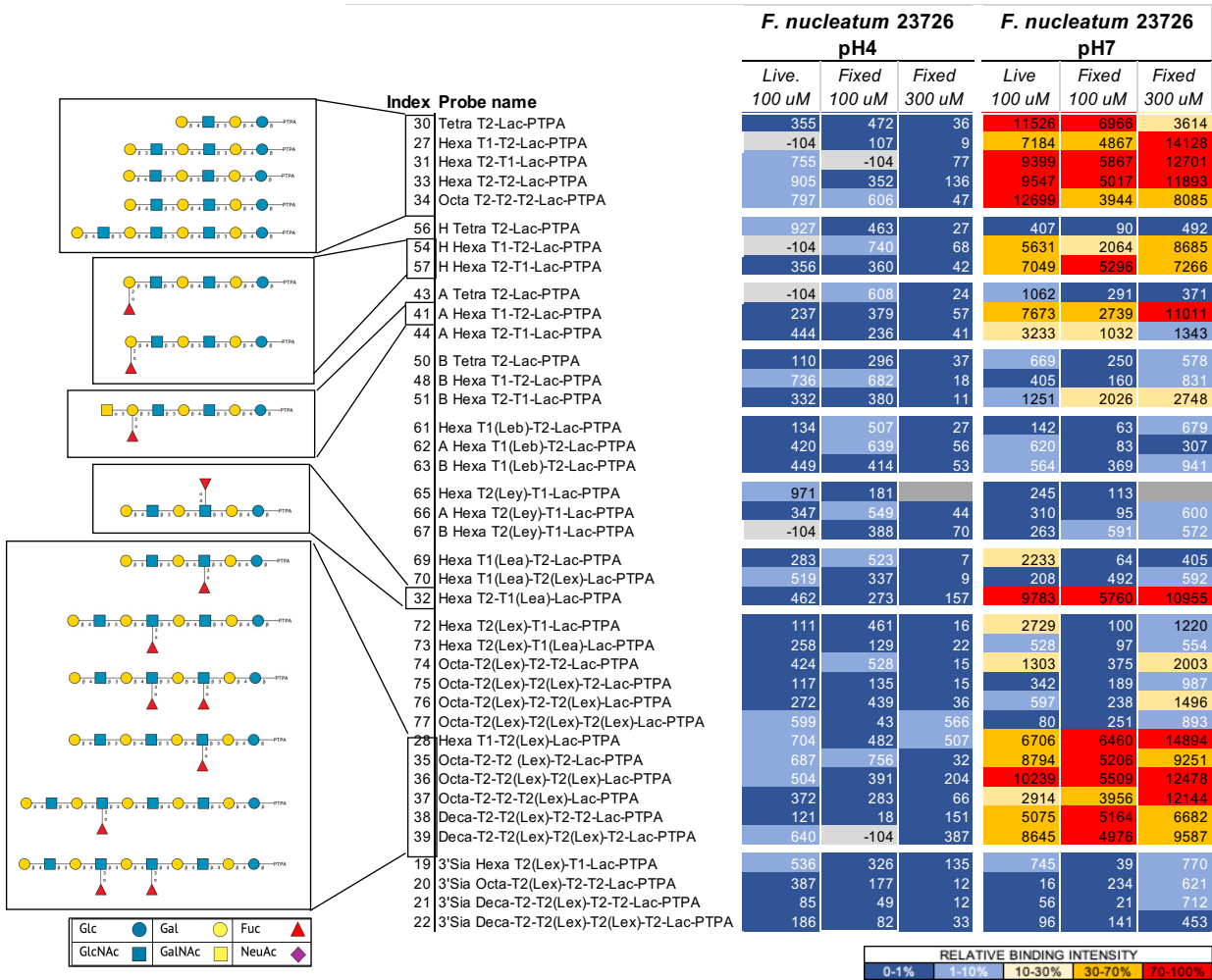
<i>E. coli</i> 901 Assay_1				<i>E. coli</i> 901 Assay_2			<i>E. coli</i> 901 Assay_3			
Probe Name	MFI	SD	RANK	MFI	SD	RANK	MFI	SD	RANK	RANK 3
Man5-GN2-AEAB	7881	1986	72	10084	988	100	14757	10894	100	91
Man5-GN2-Gly	10947	1683	100	8159	350	81	12068	4072	82	88
Man6-GN2-AEAB	3762	1761	34	7225	1099	72	8799	3348	60	55
Man7D1GN2-AEAB	3966	994	36	8380	1582	83	6600	2147	45	55
Man6-GN2-Gly	5814	523	53	7147	252	71	5643	617	38	54
Man8-GN2-AEAB	2445	957	22	5554	216	55	3740	773	25	34
Man9-GN2-AEAB	2349	795	21	5333	955	53	3429	2490	23	33
Man7D3GN2-AEAB	1885	506	17	5114	555	51	1534	850	10	26
NA2-Gly	20	24	0	1509	149	15	401	107	3	6

RELATIVE BINDING INTENSITY			
0-1%	10-30%	30-70%	70-100%
0.1%	10.3%	30.7%	70.1%

Supplementary Figure 2. (A) Top: Microarray images of fluorescently labelled *E. coli* C600 live or fixed cells binding to glycans on Microarray set 1 (Supplementary Figure 1). Bottom: Quantified microarray data presented as heatmaps of relative binding intensities for each condition separately. Dark blue (0-1 %); Light blue (1-10 %); Yellow (10-30 %); Orange (30-70%); Red (70-100%). 100%, the maximum binding score observed for a given condition. Concanavalin A was used as a control for binding to oligo and high mannose N-glycans. (B) Table with mean fluorescence intensities (MFI), standard deviation of four replicates on the microarray (SD), RANK (or % of maximum intensity) and average rank of three independent binding assays (RANK3) of *E. coli* C600, 789 and 901 to the glycan Microarray set 1. The RANK for individual assays is calculated as $100 \times \text{Mean fluorescence intensity of probe} / \text{Max mean fluorescence intensity in range of probes}$. Average rank of three assays (RANK3) is shown and coloured as in A. Source data are provided in Source Data file 3.



Supplementary Figure 3: Vaginal bacteria binding to neutral and sialylated glycans on sequence-defined covalent microarrays. Bar charts showing the binding fluorescence intensities to neutral and sialylated glycans at pH4 and pH7 of fixed cultures of A) *E. coli* C600 and B) *F. nucleatum* 23726 and live cultures of C) *L. crispatus* 1398, D) *L. iners* 13335, E) *G. vaginalis* 540 and F) *S. agalactiae* 776. Results are shown as mean fluorescence intensities of quadruplicate spots on high density (probes printed at 300 μ M) Microarray set 2a and 2b (Supplementary Figure 1). Error bars represent SD of quadruplicate spots for each glycan probe (n=1). Glycan categories: Yellow: Mucin core related; Green: High Mannose related; Pink: Sialylated; Orange: LacNAc terminating; Blue: Blood group A, B, H glycans; Peach: Lewis blood group related; Purple: Fucosylated branched glycans from human milk; Grey: Non-sulphated chondroitin sulphate. The glycan sequences and structures corresponding to glycans 1-90 are included in Supplementary data 1. Glycan probe with index number #65 didn't pass quality control on Microarray set 2b. Source data are provided in Source Data file 4.



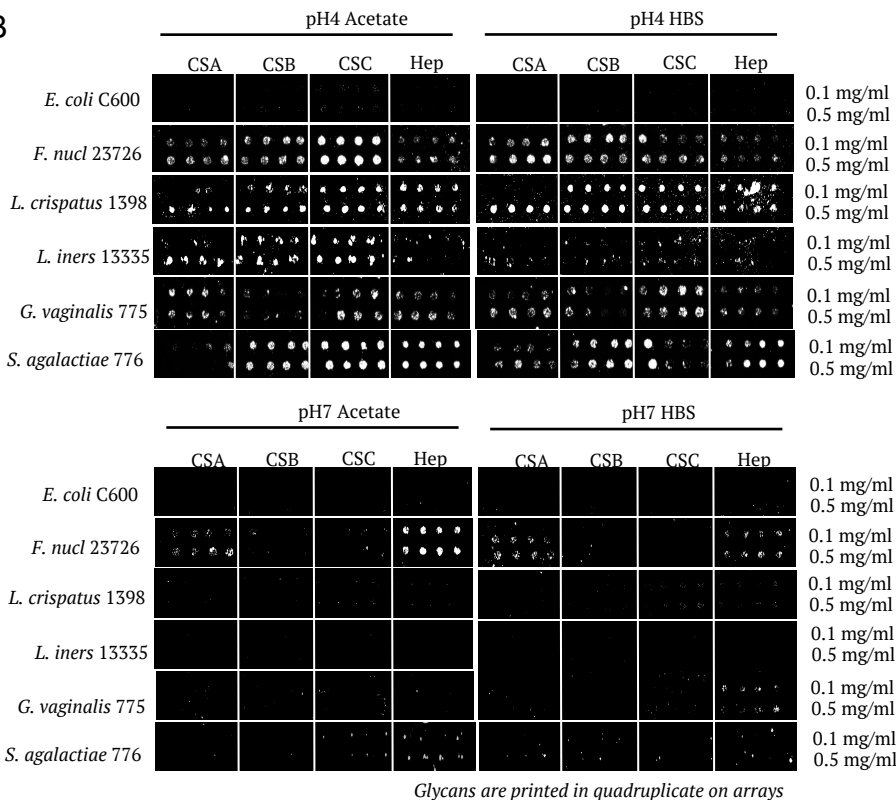
Supplementary Figure 4: Comparison of glycan binding by fixed or live cultures *F. nucleatum* 23726. Fluorescently labelled *F. nucleatum* 23726 live or fixed cultures were tested on Microarray set 3 (probes printed at 100 μM) and Microarray set 2b (300 μM) (Supplementary Figure 1). Heatmaps showing the mean fluorescence intensities of quadruplicates on microarrays at pH4 and pH7 are shown. Colour code: Dark blue (0-1 %); Light blue (1-10 %); Yellow (10-30 %); Orange (30-70%); Red (70-100%). Grey cells were flagged when they did not pass quality control in that microarray set (Probe index # 65 on microarray set 2b) or (-104) for artifact/background on slide. 100%, the maximum binding score observed for a given condition. Glycan structures corresponding to bound glycan probes are shown. Source data are provided in Source Data file 5.

A

Probe Name	<i>L. crispatus</i> 1398				<i>L. iners</i> 13335				<i>G. vaginalis</i> 775				<i>S. agalactiae</i> 776				<i>E. coli</i> C600*				<i>F. nucleatum</i> 23726			
	pH4		pH7		pH4		pH7		pH4		pH7		pH4		pH7		pH4		pH7		pH4		pH7	
	Acetate	HBS	Acetate	HBS	Acetate	HBS	Acetate	HBS	Acetate	HBS	Acetate	HBS	Acetate	HBS	Acetate	HBS	Acetate	HBS	Acetate	HBS	Acetate	HBS	Acetate	HBS
CSA-DP6-AEAB	1148	1308	254	7	647	4311	16	102	8482	11027	11	2	10301	9546	8	15	499	303	7	4	4510	4137	15	7
CSA-DP10(3S)-AEAB	1361	2626	174	3	1353	5499	9	5	1610	9773	5	10	9004	10718	46	2	330	108	7	3	4991	8950	565	1004
CSA-DP10(5S)-AEAB	3090	5781	10	13	5773	12335	14	1	1395	12606	11	1	7475	21719	0	25	584	215	0	0	9482	14803	3	227
CSA-DP14-AEAB	658	617	30	6	5455	6467	4	4	748	13658	8	26	11225	17680	89	19	535	166	5	3	7912	8122	229	281
CSB-DP6(2S)-AEAB	548	835	11	5	312	989	9	6	3844	9655	12	4	6048	3148	6	3	327	92	4	2	3533	3945	178	33
CSB-DP6(3S)-AEAB	1476	2777	177	14	2114	5153	27	17	4193	6972	100	11	11735	16888	33	6	646	154	7	3	7888	9345	28	661
CSB-DP10(4S)-AEAB	2133	4210	6	5	5726	7677	1	1	495	11388	0	0	5889	16452	0	0	452	127	1	0	8202	11365	16	574
CSB-DP10(5S)-AEAB	395	334	63	15	983	1455	11	7	578	8620	20	7	6689	5595	24	3	508	258	7	3	3764	4582	269	268
CSB-DP14-AEAB	2992	3880	178	72	8178	12452	19	13	10392	20159	19	18	22513	23099	148	75	544	179	8	4	10247	8719	268	68
CSC-DP6-AEAB	2093	3644	57	6	3231	3439	10	9	3878	11002	563	16	8795	18436	9	3	674	120	8	2	7117	6010	12	8
CSC-DP10-AEAB	3336	6427	102	25	13419	10922	10	6	1773	14140	16	2	11163	22609	127	2	674	212	2	2	8941	13495	16	215
CSC-DP14-AEAB	601	962	16	13	1840	5340	35	11	2250	13781	37	12	9418	24353	122	224	719	171	8	3	4674	7704	217	168
Heparin-DP6-AEAB	737	1298	77	24	1002	1913	21	13	7266	13510	16	7	14891	17047	9	5	583	178	9	5	2267	3198	98	16
Heparin-DP10-AEAB	4309	5656	669	540	10325	6930	178	73	11323	12098	1849	81	23800	27118	118	10	1216	434	51	6	8703	13719	55	976
Heparin-DP14-AEAB	5497	8887	1106	686	17504	12002	363	72	2748	7515	566	22	18340	26822	170	10	1938	823	6	4	13958	16540	180	541

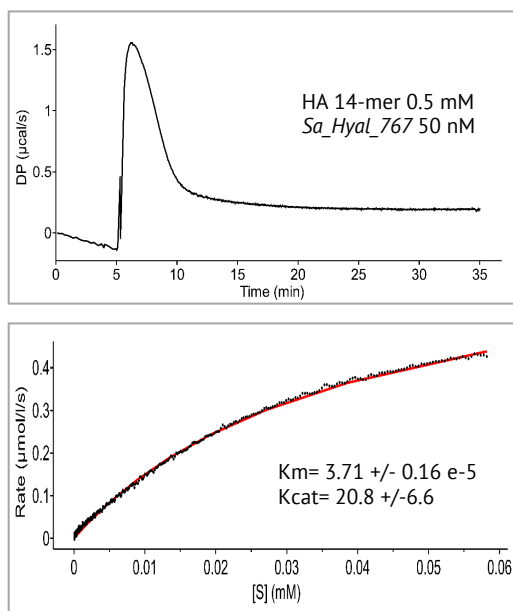
RELATIVE BINDING INTENSITY				
0-10%	10-25%	25-50%	50-70%	70-100%

B

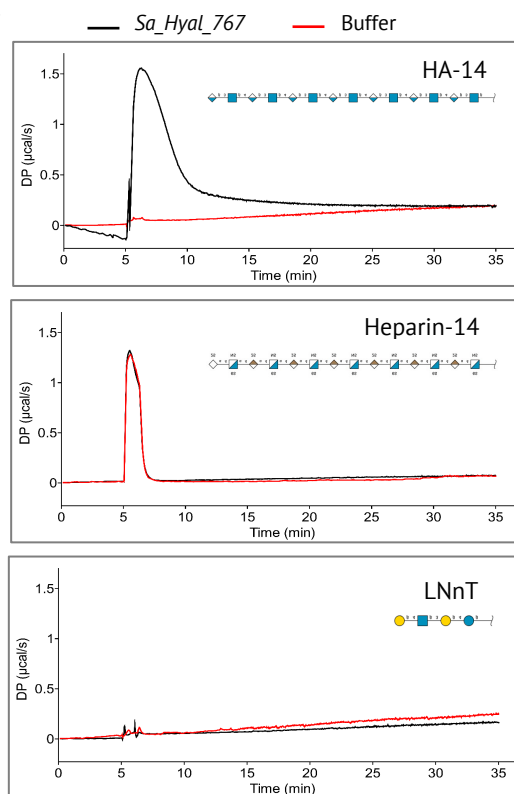


Supplementary Figure 5. Glycosaminoglycan binding by bacteria of the vaginal microbiota in acetate and HBS buffers is pH dependent. (A) Heatmap showing the relative binding intensities to CS and Heparin oligosaccharides in acetate or HBS buffer and at pH4 and pH7 of live fluorescently labelled strains of *E. coli*, *F. nucleatum*, *L. crispatus*, *L. iners*, *G. vaginalis* and *S. agalactiae* on Microarray set 4 (Supplementary Figure 1) coloured as follows: Dark blue (0-10 %); Light blue (10-25 %); Yellow (25-50 %); Orange (50-70%); Red (70-100%). 100%, the maximum binding score observed for a given strain and condition. *Due to the low binding signals of *E. coli* C600 to GAGs in this experiment, a different colour palette has been applied for the heatmap: Dark blue (0-1 %); Light blue (1-10 %); Yellow (10-30 %); Orange (30-70%); Red (70-100%). Source data are provided in Source Data file 6. (B) Glycan microarray images of fluorescently labelled *E. coli* C600, *F. nucleatum* 23726, *L. crispatus* 1398, *L. iners* 13335, *G. vaginalis* 775 and *S. agalactiae* 776 binding to GAG polysaccharides in acetate or HBS buffer at pH4 or pH7. Assay is representative of two independent experiments.

A



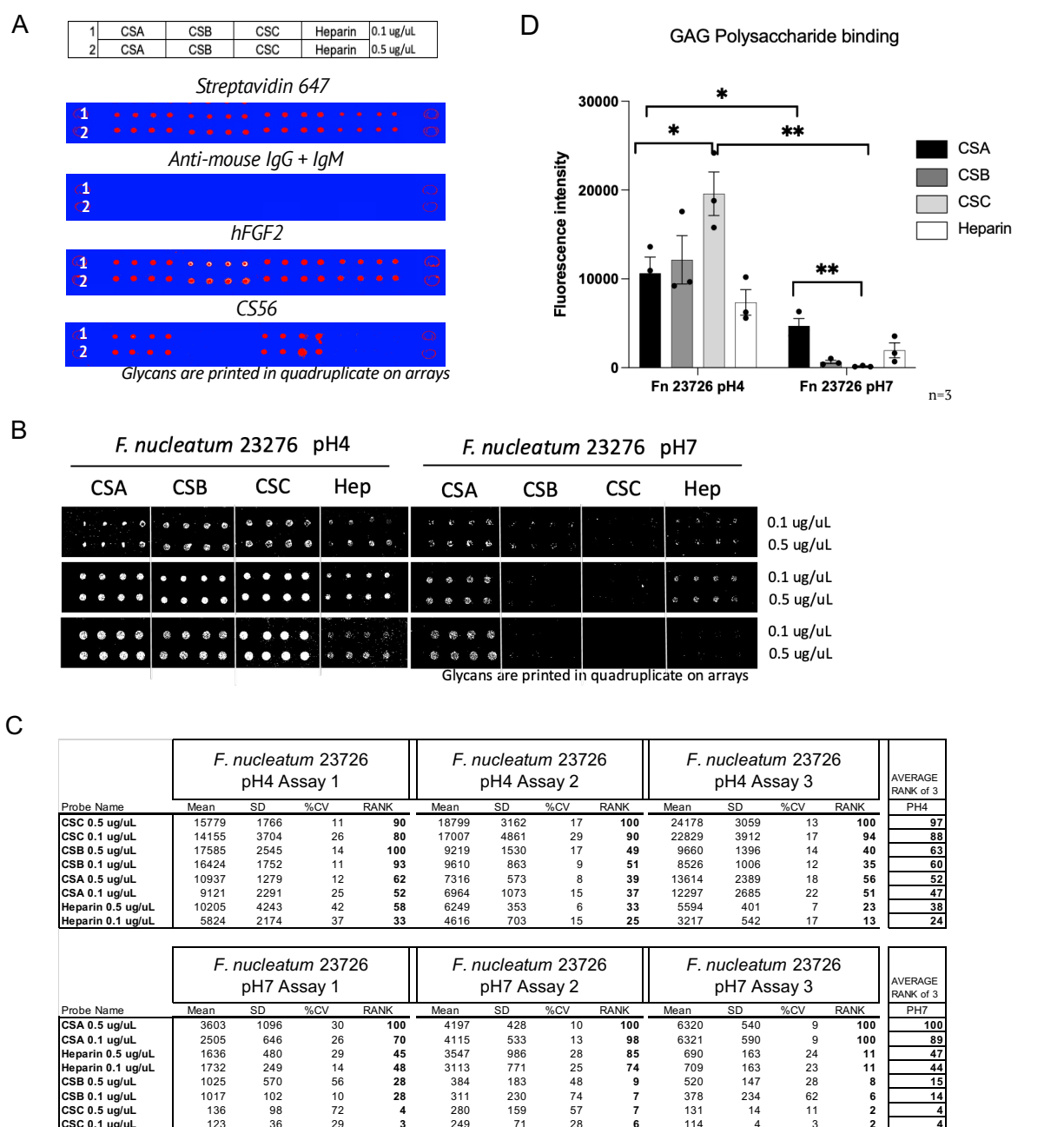
C



B

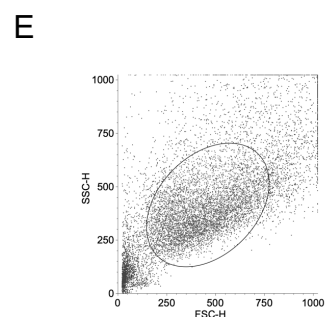
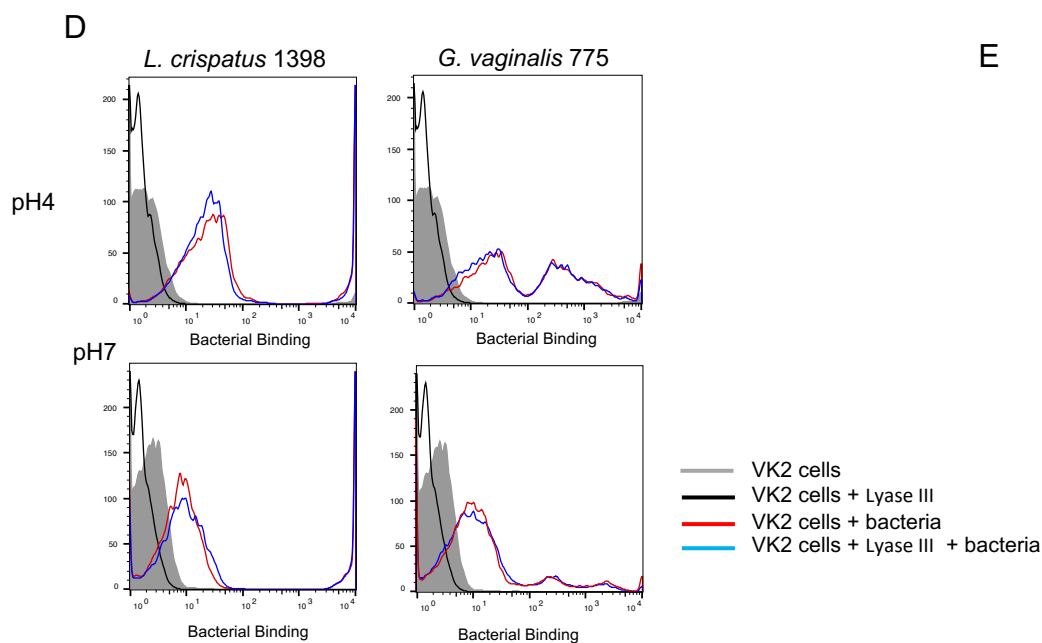
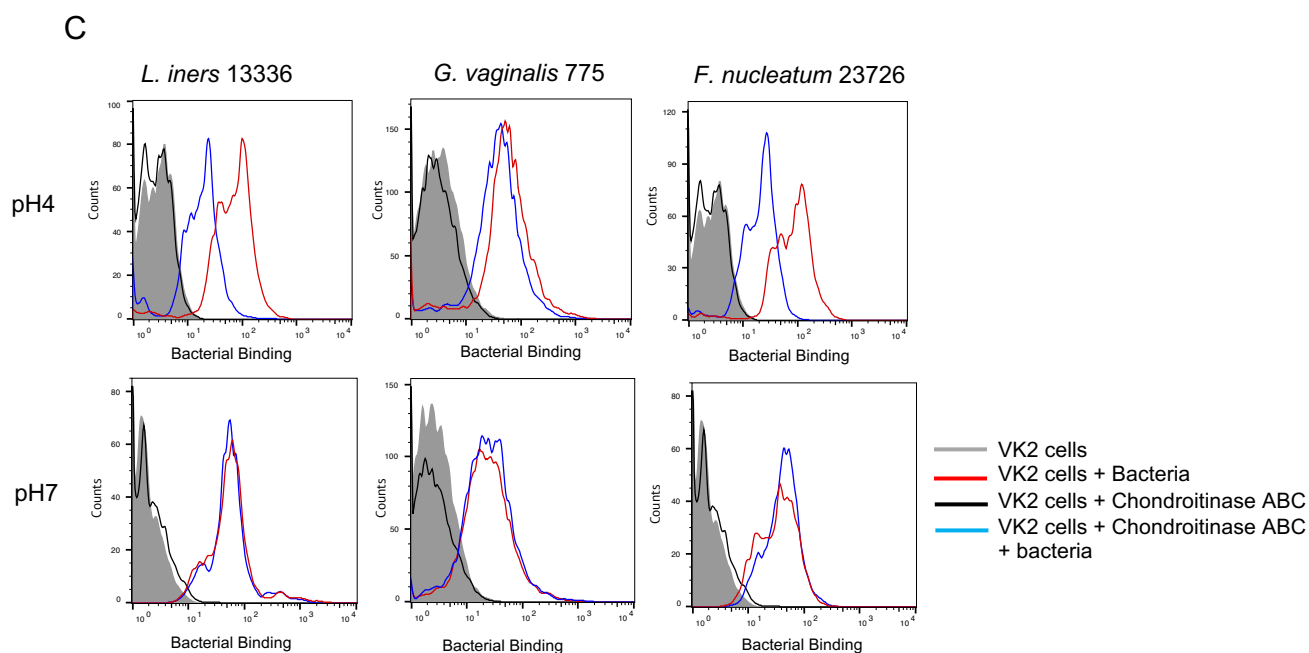
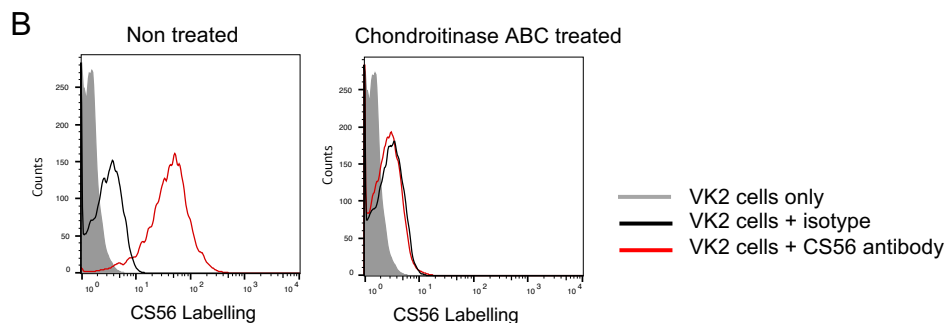
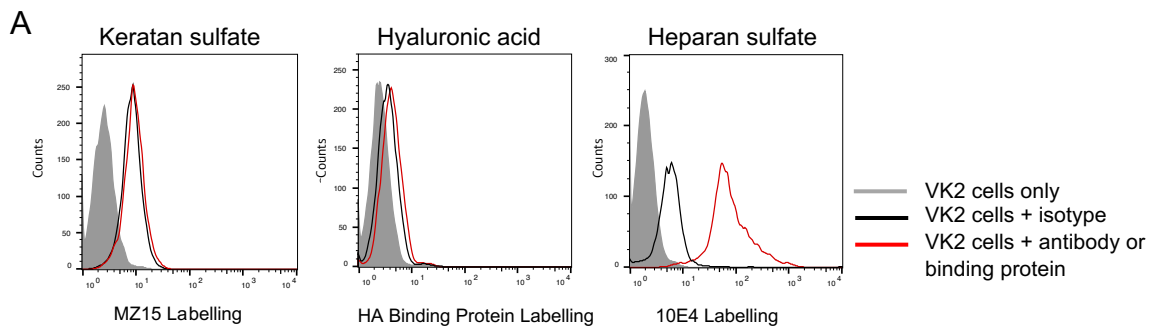
SAMPLE	T ^e (°C)	[Enzyme] (M)	[Substrate] (M)	ΔH (kcal/mol)	k _{cat} (1/s)	K _M (M)	Red. Chi-Sqr. ($\mu\text{mol/l/s}$) ²
HA-14 pH 7.5	25.1	N/A	5.00E-04	N/A	N/A	N/A	N/A
<i>Sa_Hyal_767</i> + HA14 pH7.5_001	25.1	5.00E-08	5.00E-04	16	12	3.59E-05	2.70E-05
<i>Sa_Hyal_767</i> + HA14 pH7.5_002	25.1	5.00E-08	5.00E-04	18.9	17.5	3.84E-05	2.60E-05
<i>Sa_Hyal_767</i> + HA14 pH7.5_003	25.1	5.00E-08	5.00E-04	14.1	29.4	3.55E-05	5.90E-05
<i>Sa_Hyal_767</i> + HA14 pH7.5_004	25.1	5.00E-08	5.00E-04	18.3	24.5	3.86E-05	4.60E-05

Supplementary Figure 6: ITC shows that HA is a substrate of *S. agalactiae* 767 Hyaluronidase. (A) Representative thermal profile of a single injection experiment (top) and fitting to a Michaelis Menten curve (bottom) of calorimetry data generated following incubation of HA 14-mer with hyaluronidase from *S. agalactiae* 767 (*Sa_Hyal_767*) in the ITC cell. Black line (raw data), red line (fitted curve). (B) Table with K_m and K_{cat} values inferred from four independent experiments. (C) Single injection isotherms generated following incubation of HA 14-mer, Heparin 14-mer and LNnT glycans with either buffer only (red line) or *Sa_Hyal_767* (black line) in the ITC cell.

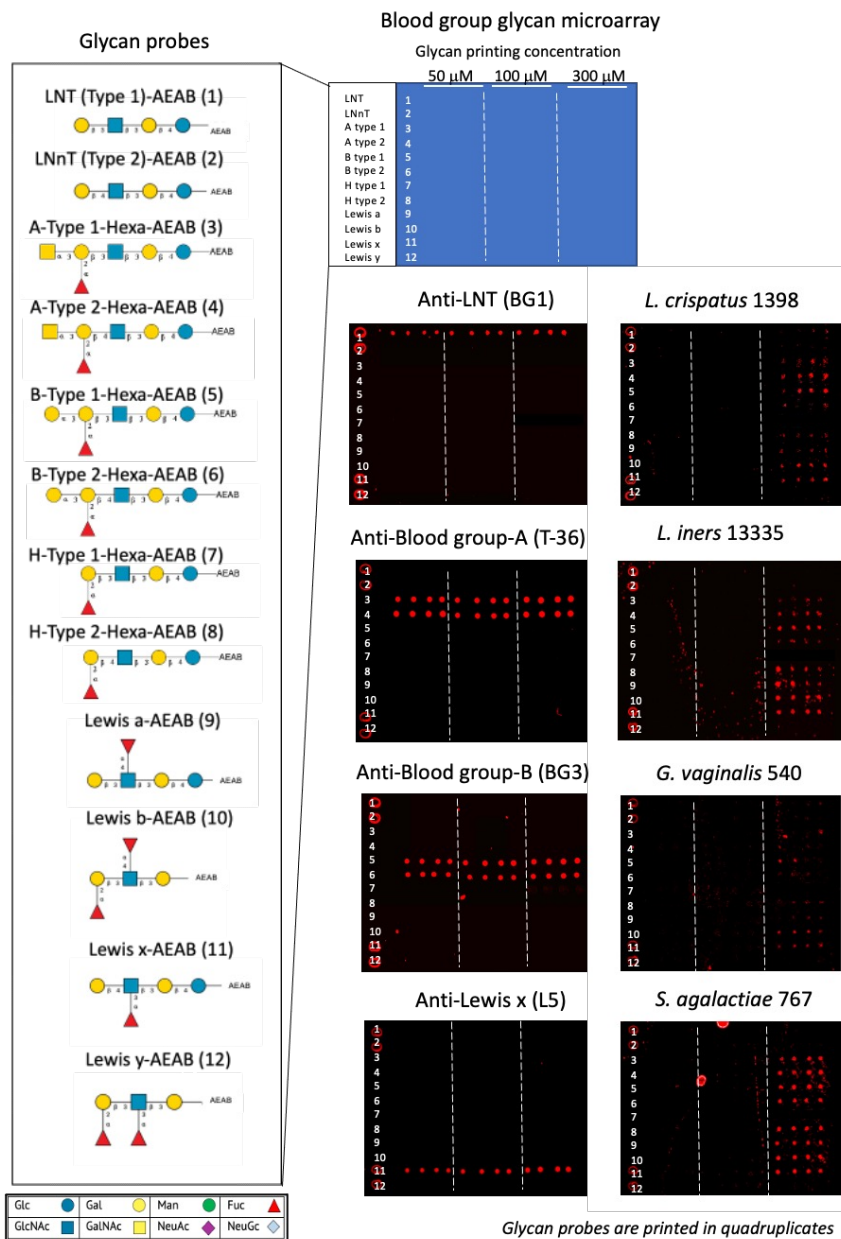


Supplementary Figure 7: Chondroitin sulphate and heparin polysaccharides are bound by *F. nucleatum* 23276.

(A) Top: Layout of the GAG polysaccharide Microarray set 5 containing biotinylated chondroitin sulphate A, B, C and Heparin probes printed in quadruplicate from 0.1 $\mu\text{g}/\mu\text{L}$ and 0.5 $\mu\text{g}/\mu\text{L}$ solutions. Bottom: Microarray images of polysaccharide binding by Streptavidin-Alexa647 or the indicated antibodies. (B) Microarray images from three independent experiments of fixed fluorescently labelled *F. nucleatum* 23276 binding at pH4 or pH7 to GAG polysaccharides. (C) Table with the analysis of the images from B showing the background subtracted mean fluorescence intensities (Mean), SD, Coefficient of variance (%CV), and RANK (or % of MAX that is calculated as $100 \times \text{Mean for probe}/\text{Max Mean in range of probes}$). The average rank of three independent experiments is shown. (D) Bar charts show the average fluorescence intensities of *F. nucleatum* ATCC 23276 binding to GAG polysaccharides on glycan microarrays at pH4 and pH7. The average of the background-subtracted mean fluorescence intensities of bacterial binding from three independent experiments with quadruplicate spots printed using 0.5 $\mu\text{g}/\mu\text{L}$ of polysaccharide on the microarray are shown. Error bars represent SD of $n=3$. * $p<0.05$, ** $p<0.01$ Unpaired t-test. Source data are provided in Source Data File 7.



Supplementary Figure 8: Analysis of the expression of chondroitin sulphate, keratan sulphate, heparan sulphate and hyaluronic acid on vaginal epithelial VK2 cells. (A) VK2 cells were stained with the anti-keratan sulphate antibody MZ15, anti-heparan sulphate 10E4 antibody or HA binding protein and analysed by flow cytometry. (B) VK2 cells treated or not with chondroitinase were stained with the anti- CSA/CSC antibody CS56 and analysed by flow cytometry. (C) Flow cytometry-based analysis of live fluorescently labeled bacteria binding to VK2 cells treated or not with chondroitinase. (D) Flow cytometry-based analysis of fluorescently labeled bacteria binding to VK2 cells treated or not with Lyase III representative of three biological independent experiments. (E) Gating strategy applied to select the intact VK2 cells. Intact VK2 cells were gated based on their FFS and SSC.



Supplementary Figure 9: Glycan density requirements for detecting binding of vaginal commensals and pathobionts to blood group glycans. Design of the blood group glycan microarray. The names of the 12 glycan probes and the corresponding glycan structures are shown. Three different glycan printing concentrations were used: 50, 100 and 300 μ M. Microarray images of selected anti-carbohydrate antibodies and live fluorescently labelled bacteria binding are shown.

Met His tag

ATG GGCAGCAGC CATCATCATCATCATCAGCAGCGGCCTGGTGCCGCGCGGCAGCCATATGGCTAGCATGAC
 TGTTGGACAGCAAATGGGTGCGGATCCATGGGTGACTTCAAGGAAAAGATCATCGACAAGAAAATTGATAAGA
 AAAGCCAGTGGACCAACCTGTACGGCGCGAAGGACTGGAACACCTATATCGATCAGACCAAAAGCGTTAACAAG
 AGCCCGATCATTCAACGTACCGAGCAGGGTCAAGTGAGCCTGAGCAGCGATAAGGGTTTTCTGGCGCGGTGAC
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 GAAGGTTGGCAACGATTATCAAAACGTGTACTATCAGCCGCAACCATGACCAAAACCGACCAACTGGCGATCT
 AA

Supplementary Figure 10: Nucleotide sequence used for the expression of *SaHyal_767*. The sequence specified encoding the *SaHyal_767* hyaluronate lyase from *Streptococcus agalactiae* strain 767 was codon optimized and cloned in the plasmid pET28a (+) for expression in *E. coli*. The recombinant full-length *SaHyal_767* enzyme includes the appended carbohydrate binding module (CBM70), and the catalytic module in which Tyr578 was mutated to Phe (highlighted in pink). The Y578F mutant was designed to reduce catalytic activity to perform glycan binding assays with the full-length enzyme (Supplementary Figure 6).