

ORIGINAL ARTICLE

Interferon-beta-1b-induced short- and long-term signatures of treatment activity in multiple sclerosis

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Interferon beta (IFN β) reduces disease burden in relapsing-remitting multiple sclerosis (MS) patients. In this study, IFN β -1b-treated MS patient gene expression profiles and biological knowledgebases were integrated to study IFN β 's pleiotropic mechanisms of action. Genes involved in immune regulation, mitochondrial fatty acid metabolism and antioxidant activity were discovered. Plausible mediators of neuronal preservation included NRF2, downregulation of OLA1, an antioxidant suppressor, and the antioxidant gene ND6, implicated in optic neuropathy and MS-like lesions. Network analysis highlighted IKBKE, which likely has a role in both viral response and energy metabolism. A comparative analysis of therapy-naive MS- and IFN β -associated gene expression suggests an IFN β insufficiency in MS. We observed more gene expression changes in long-term treatment than during acute dosing. These distinct short- and long-term effects were driven by different transcription factors. Multi-gene biomarker signatures of IFN β treatment effects were developed and subsequently confirmed in independent IFN β -1b-treated MS studies, but not in glatiramer acetate-treated patients.

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INTRODUCTION

Multiple sclerosis (MS) is an autoimmune-mediated progressive neurodegenerative disease that results in permanent disability and a significant decrease in quality of life.^{1,2} Current standard of care MS drugs provide similar clinical benefit but vary considerably in side effects, dosing, route of administration and mechanism of action.³ There is an increasing need to provide guidance in treatment options to MS patients and their physicians. Treatment monitoring using biomarkers could provide assertive management of disease progression.^{4–7}

Interferon beta (IFN β) is a highly pleiotropic member of a family of cytokines that regulate innate and adaptive immune responses.^{8,9} IFN β suppresses activation of T and B cells, prevents translocation of immune cells from the periphery into the central nervous system, inhibits astrocyte and microglial cell differentiation, and is potentially neuroprotective.^{10–13} The mechanism of action of IFN β in MS remains uncertain but elucidating effects underlying treatment could provide guidance in treatment choices.

Previously, we have characterized¹⁴ and confirmed¹⁵ differential gene expression in short-term and long-term IFN β therapy.^{16,17} Gene expression microarrays were employed here to discover biological information relevant to the mechanism of action of IFN β -1b. Analysis built on an approach previously applied to study IFN β -1b effects in patients¹⁴ and was applied to a broader set of genes measured in mononuclear cells obtained from MS patients and healthy individuals using the Affymetrix HuEx1.0ST ('all exon') platform. As multi-gene effects are expected to be more stable across microarray platforms than single-gene effects,¹⁸ multi-gene IFN-biomarker signatures were characterized and subsequently confirmed in independently sampled, previously published patient data.

Gene expression microarray analysis presents a challenge to make sense of high-dimensional data comprised of measurements on tens of thousands of genes. We approached this challenge by bringing together genes into pre-defined biological contexts. Three biological themes were integrated into this study to infer protein effects upstream, downstream and interacting with the measured genes. First, for transcription factor binding, sets of genes that were annotated as bound by a specific transcription factor implicated events upstream of gene expression and facilitated pre-expression inference. Second, gene sets defined by canonical pathways and functions provided post-expression inference for biological effects downstream of drug-mediated gene expression changes. Finally, gene sets encoding the interaction neighbors of a given protein used gene expression to implicate proteins potentially influenced by a changing molecular environment. This protein network analysis supports peri-expression inference, or guilt-by-association. In this study, specific connections among these realms of knowledge and MS scientific literature were highlighted and permitted development of multi-gene IFN β -bioactivity signatures that were evaluated across larger groups of independently sampled patients.

MATERIALS AND METHODS

Study population

Fifty-two human subjects, including healthy individuals and clinically defined MS patients diagnosed according to McDonald criteria,¹⁹ were recruited to the study (Table 1). The study was approved by the Internal Review Board at the University of Chicago, and all patients gave written informed consent.

Seven patients were treated with IFN β -1b, and their expanded disability status scale (EDSS)²⁰ scores had a mean of 4.4 and ranged from 2.5 to 5.5.

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To obviate effects of disease activity on IFN responses, treated clinically 'stable' patients were free of exacerbations for at least 10 months before and 4 months after blood collection, and none were in the progressive phase of MS. No patients had received corticosteroids or immunosuppressants for 1 year before or during the study. Six of the seven patients had received high-dose, high-frequency Betaseron IFN β -1b therapy for > 3 years. Three patients had received prior treatment with other IFN β therapies (that is, IFN β -1a). Patients had received IFN β for an average of 6.9 (0.5–10.5) years and had discontinued therapy for at least 64 h before beginning this study. For this kinetic study, Betaseron IFN β -1b (8 million international units (IU), specific activity 2.3×10^7 IU mg $^{-1}$) was self-administered subcutaneously under a physician's supervision. Blood was drawn by the physician just before administration of drug ($t=0$) and at precisely controlled times at 4, 18 and 42 h post-administration.

Forty-five treatment-naïve subjects were studied. Ten relapsing-remitting MS (RRMS) patients had blood drawn during a clinical exacerbation (active RRMS), and 11 RRMS patients were free of exacerbations 6 months before and after sample draw and showed no EDSS progression for 1 year (stable RRMS). Seven treatment-naïve patients had primary-progressive MS, and eight had secondary-progressive MS. Nine healthy individuals were included. Table 1 summarizes patient information.

RNA isolation and gene chip analysis

Heparinized blood (35 ml) was collected by venipuncture and peripheral blood mononuclear cells were immediately isolated using Ficoll–Paque Plus density gradients (Amersham Biosciences, NJ, USA). RNA was isolated using an RNeasy Midi Kit (Qiagen, Santa Clara, CA, USA). Quality and quantity of the RNA were determined using a Bioanalyzer 2100 (Agilent Instruments, Foster City, CA, USA). Samples having a 28S/18S ratio below 1.5 were excluded from this study. RNA was hybridized to GeneChip HuEx1.0ST arrays (Affymetrix, Santa Clara, CA, USA) containing approximately 1.4 million probe sets recognizing over 1 million exon clusters.

Comparative gene expression analysis using microarrays in a two other data sets

Gene Expression Omnibus (GEO, <http://www.ncbi.nlm.nih.gov/geo/>) record GSE16214 provided data from the Comprehensive Longitudinal Investigation of MS at the Brigham and Women's Hospital (CLIMB study), which included untreated ($n=82$), IFN β -treated ($n=94$), and glatiramer-acetate-treated ($n=64$) (Copaxone) patients. Timing of dosing relative to phlebotomy was not captured in the public CLIMB data. GEO record GSE24427 by Goertsches provided independent data from IFN β -1b-treated patients.²¹ We inferred that patients in the Goertsches study were sampled 48 h after administration.

Probe set (3' arrays) and transcript (HuEx1.0ST arrays) expression summarization was performed using the robust multichip analysis (RMA)²²

Table 1. Patient characteristics

	N	Gender	Mean age \pm s.d. (years)	Mean EDSS \pm s.d.
Control	9	4 F, 5 M	40.1 \pm 13.9	NA (0)
Stable RRMS	11	7 F, 4 M	38.2 \pm 13.8	2.3 \pm 1.48
Active RRMS	10	6 F, 4 M	43.4 \pm 11.5	4.72 \pm 1.3
PPMS	7	1 F, 6 M	49.9 \pm 13.6	5.43 \pm 1.79
SPMS	8	5 F, 3 M	46.5 \pm 7.1	5.91 \pm 1.56
IFN β treated	7	6 F, 1 M	42.6 \pm 14.3	4.4 \pm 1.0

Abbreviations: EDSS, expanded disability status scale; F, female; IFN β , interferon beta; M, male; MS, multiple sclerosis; NA, not applicable; PPMS, primary-progressive MS; RRMS, relapsing-remitting MS; SPMS, secondary-progressive MS.

implementation in Affymetrix Power Tools version 1.12.0. Probe set and transcript annotations used Affymetrix's NetAffx Release 30 (November 2009).

Knowledgebases

Gene sets by transcription factor binding were obtained from MSigDB (<http://www.broadinstitute.org/gsea/msigdb/index.jsp>), specifically the c3 TFT gene sets, and by searching the Biomolecular Object Network Databank (<http://bond.unleashedinformatics.com/>) for DNA–protein interactions. Pathway Commons²³ provided canonical pathways and a network of interacting proteins. Genes in these annotations were identified by Entrez Gene identifiers.

Data handling

Data handling was performed using Perl and PostgreSQL. Statistical analysis was performed in R.²⁴

Short- and long-term differential gene expression

The coincident extreme ranks in numerical observations method¹⁴ used a gene set analysis approach to investigate IFN β -1b short-term effects (hours post-administration), long-term effects (comparing samples from patients treated for years to those from therapy-naïve MS patients), and MS disease gene expression (therapy-naïve MS compared with healthy individuals). To facilitate a large number of gene set tests, a nested coincident extreme ranks in numerical observation testing routine replaced the Monte Carlo approach in Yamaguchi *et al.*¹⁴ Statistical testing including false discovery rate analysis details are provided as Supplementary Methods.

An integrated, cross-microarray platform database enabled flexible querying of microarray results to select genes regulated by IFN β -1b and genes annotated with specific functions or pathways. Selection of 3' array probe sets, with the intent to capture the effects measured on the

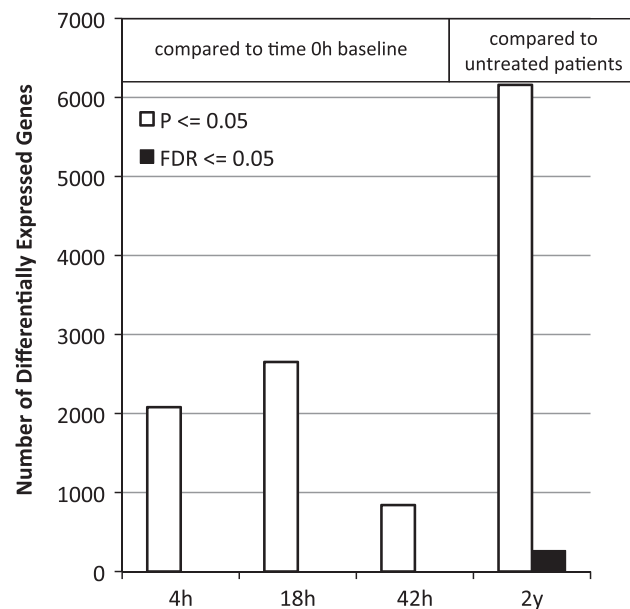


Figure 1. Differentially expressed genes at the $P \leq 0.05$ cutoff are roughly threefold more abundant in the long-term comparison than in the short-term comparisons. No probe sets exhibit a false discovery rate (FDR) of 0.05 or less in the short-term comparisons. In all, 264 genes have at least one probe set that exhibits a t -test FDR of 0.05 in the long-term comparison (black bar). In all, 715 probe sets map to these 264 genes. The FDR analysis tells us that, even considering the large number of tests (312 023) for transcript differential expression, there is an excess of probe sets yielding small P -values, such that we expect only 5% to be false discoveries. Thus, there is confidence that the vast majority of the 264 genes are regulated by long-term interferon beta (IFN β)-1b treatment. h, hours; y, years.

HuEx1.0ST array, were completed before running tests on the published Goertsches²¹ data (see Supplementary Methods).

Signatures were selected based on statistical evidence derived from short- and long-term IFN-β-induced gene expression, and grouped according to treatment-affected biological functions and MS-relevant biology described in published literature. Supplementary Information provides in-depth descriptions of signature definition criteria, as well as the lists of the selected genes and probe sets (Supplementary Tables 1–10).

Signature testing was performed using Global Test,²⁵ a multiple regression method that used only the gene expression results within a set of transcript measurements, and thus explicitly ignored expression levels estimated from the entire microarray. Global Test does not specifically model multiple transcript measurements per gene, so only one transcript or probe set measurement per gene was permitted in any signature.

Coincident extreme ranks in numerical observation was used to identify the most important differential expression effects of IFNβ-1b in the research sample profiles. Global Test was used to evaluate whether signatures were significantly regulated in particular studies.

RESULTS

IFNβ-1b treatment: differential expression and timescales

Differential gene expression in MS patient peripheral blood mononuclear cells at pre-dose levels was compared with 4, 18 and 43 h post-IFNβ-1b ('short-term' comparisons). Pharmacokinetics of differential gene expression after a single dose of IFNβ-1b using HuEx1.0ST arrays were similar to those previously observed.¹⁵ Classic acute IFN response genes were among the

	compared to time 0h baseline			compared to untreated patients	
	4h	18h	43h	long-term	CLIMB
IRF (STTTCRNTTT_V\$IRF_Q6)	3.E-09	6.E-11	1.E+00	9.E-01	2.E-03
ISRE (V\$ISRE_Q1)	1.E-08	4.E-10	1.E+00	8.E-01	2.E-03
IRF (V\$IRF_Q6)	3.E-08	1.E-11	1.E+00	6.E-01	2.E-03
ICSBP (V\$ICSBP_Q6)	7.E-06	4.E-07	1.E+00	6.E-01	2.E-03
IRF2 (V\$IRF2_Q1)	2.E-05	3.E-03	1.E+00	7.E-01	3.E-01
ETS2 (RYTTCCTG_V\$ETS2_B)	6.E-04	2.E-08	1.E+00	7.E-01	5.E-01
PEA3 (V\$PEA3_Q6)	6.E-04	3.E-02	1.E+00	6.E-01	5.E-01
AML (RACCACAR_V\$AML_Q6)	2.E-03	2.E-01	1.E+00	1.E+00	7.E-01
NFAT (TGGA_A_V\$NFAT_Q4_Q1)	2.E-03	7.E-03	1.E+00	8.E-01	4.E-01
IRF7 (V\$IRF7_Q1)	1.E-03	2.E-04	1.E+00	4.E-01	2.E-04
ELF1 (V\$ELF1_Q6)	5.E-03	1.E-07	1.E+00	9.E-01	3.E-01
NFKB (V\$NFKB_C)	4.E-03	1.E-01	1.E+00	5.E-01	7.E-01
IRF1 (V\$IRF1_Q1)	5.E-03	2.E-01	1.E+00	7.E-01	3.E-01
STAT (V\$STAT_Q1)	1.E-02	2.E-01	1.E+00	9.E-01	5.E-01
NFKB (GGGNNTTCC_V\$NFKB_Q6_Q1)	1.E-02	2.E-06	1.E+00	5.E-01	1.E-01
PU1 (RGAGGAARY_V\$PU1_Q6)	3.E-02	4.E-04	1.E+00	5.E-01	8.E-01
GABP (MGGAAGTG_V\$GABP_B)	4.E-02	7.E-03	1.E+00	3.E-01	3.E-01
AP1 (TGANTCA_V\$AP1_C)	2.E-02	6.E-03	1.E+00	1.E-01	6.E-01
(GGGYGTGNY_UNKNOWN)	6.E-02	2.E-03	1.E+00	4.E-01	3.E-01
TEL2 (V\$TEL2_Q6)	6.E-02	1.E-03	1.E+00	1.E-01	5.E-01
NRF2 (V\$NRF2_Q1)	3.E-01	3.E-03	1.E+00	3.E-01	4.E-01
USF (V\$USF_C)	5.E-01	7.E-03	1.E+00	4.E-01	5.E-01
AP4 (CAGCTG_V\$AP4_Q5)	3.E-01	2.E-03	1.E+00	2.E-01	1.E+00
AP2 (V\$AP2_Q3)	2.E-01	5.E-03	1.E+00	1.E-01	8.E-01
NFE2 (V\$NFE2_Q1)	2.E-01	2.E-04	1.E+00	1.E-01	2.E-01
CREL (V\$CREL_Q1)	3.E-01	7.E-03	1.E+00	1.E-01	4.E-01
MYC (CAGCTG_V\$MYC_Q2)	1.E-01	7.E-03	1.E+00	4.E-02	7.E-02
AP1 (V\$AP1_Q6_Q1)	3.E-01	9.E-03	1.E+00	6.E-02	9.E-01
CETS1P54 (V\$CETS1P54_Q1)	3.E-01	3.E-03	1.E+00	3.E-02	4.E-01
AP1 (V\$AP1_Q6)	6.E-01	6.E-03	1.E+00	3.E-02	7.E-01
ATF (V\$ATF_Q1)	2.E-01	2.E-01	1.E+00	1.E-02	6.E-01
TCF11MAFG (TGANNYRGCA_V\$TCF11MAFG_Q1)	6.E-01	7.E-04	1.E+00	2.E-02	5.E-01
E12 (CAGGTG_V\$E12_Q6)	5.E-01	7.E-03	1.E+00	2.E-02	9.E-01
MAZ (GGGAGGRR_V\$MAZ_Q6)	3.E-01	2.E-04	1.E+00	9.E-03	9.E-01
SP1 (V\$SP1_Q4_Q1)	5.E-01	6.E-01	1.E+00	9.E-03	9.E-01
ELK1 (V\$ELK1_Q2)	6.E-01	1.E-01	1.E+00	9.E-03	3.E-01
TFIII (V\$TFIII_Q6)	3.E-01	9.E-02	1.E+00	4.E-03	1.E+00
NFE2 (TGASTMAGC_V\$NFE2_Q1)	5.E-01	1.E-02	1.E+00	6.E-03	8.E-01
RBL2	1.E+00	1.E+00	1.E+00	1.E-02	4.E-03
E2F4	9.E-01	1.E+00	1.E+00	9.E-03	1.E-03
E2F1	1.E+00	3.E-01	1.E+00	9.E-03	7.E-02
(GKCGCNNNNNTGAYG_UNKNOWN)	6.E-01	9.E-01	1.E+00	4.E-03	3.E-01
E4F1 (GTGACGY_V\$E4F1_Q6)	6.E-01	1.E-01	1.E+00	1.E-03	6.E-02
ERR1 (TGACCTY_V\$ERR1_Q2)	6.E-01	3.E-01	1.E+00	1.E-03	9.E-01
SREBP1 (TCANNTGAY_V\$SREBP1_Q1)	2.E-01	2.E-06	1.E+00	3.E-04	4.E-01
CREB (V\$CREB_Q2)	9.E-01	6.E-01	1.E+00	3.E-04	2.E-01
ELK1 (SCGGAAGY_V\$ELK1_Q2)	3.E-01	3.E-03	1.E+00	6.E-06	1.E-01
SP1 (GGGCGGR_V\$SP1_Q6)	2.E-01	2.E-04	1.E+00	6.E-08	7.E-02
MAX	2.E-01	7.E-03	1.E+00	6.E-08	1.E-03
MYC	4.E-01	3.E-04	1.E+00	4.E-08	1.E-03
TAF1	1.E-01	2.E-04	1.E+00	1.E-13	7.E-12
HNF4A	4.E-01	2.E-04	1.E+00	8.E-20	5.E-07

Figure 2. Gene sets characterized by transcription factor binding (rows) show excessive differential expression in sample comparisons (columns) as indicated by false discovery rate (FDR) coincident extreme ranks in numerical observation (CERNO) values. The most significant differentially expressed gene sets (dark orange) are distinct between short-term and long-term comparisons. Independent data from the Comprehensive Longitudinal Investigation of MS at the Brigham and Women's Hospital (CLIMB) study (Affymetrix 3' array HG-U133 Plus 2.0) also show evidence of both short- and long-term treatment effects, which corroborates the multi-gene analysis of the HuEx1.0ST data.

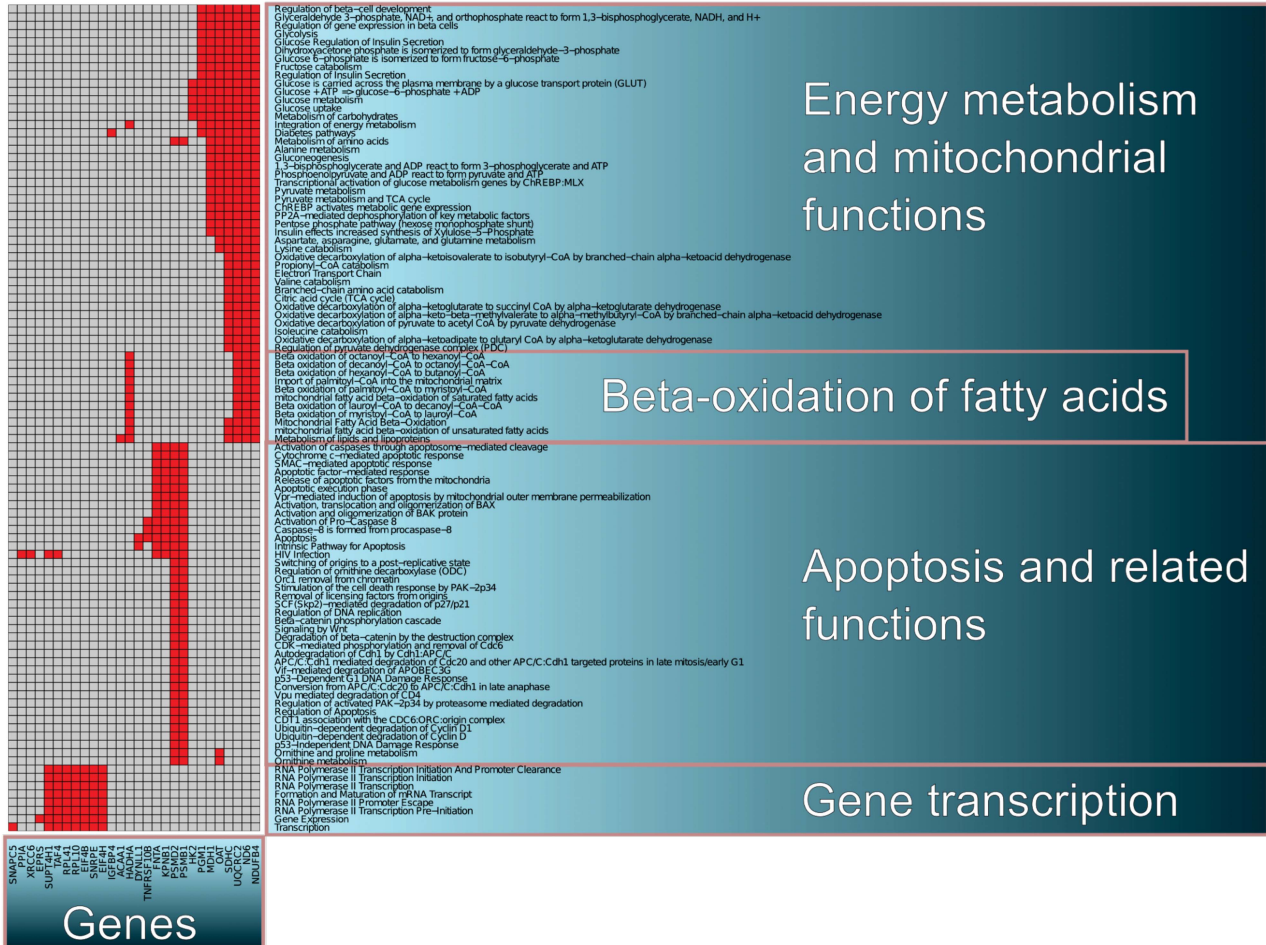


Figure 3. The 100 most significantly changed functional pathways, as detected with coincident extreme ranks in numerical observation (CERNO), are shown. In all, 28 of the 264 long-term interferon beta (IFNβ)-1b-regulated genes map to these 100 pathways. Membership of the genes in the pathways is indicated by a red square. The membership matrix was ordered using hierarchical clustering, revealing a pattern that includes a cluster of functions and genes related to beta-oxidation of fatty acids.

most differentially expressed (OAS1/2/3, MX1/MxA, CXCL10/IP-10 and others).

Short-term changes were contrasted with long-term treatment changes. ‘Long-term’ changes were defined by the difference between gene expression in therapy-naïve patients and patients after an average of 6.9 years of IFNβ-1b therapy. The long-term comparison revealed a larger number of differentially expressed genes than did short-term comparisons, whether differential expression was measured by a nominal *P*-value cutoff or by false discovery rate (FDR) analysis (Figure 1). Although the range of expression ratios was similar for short- and long-term effects, more significant up and downregulation was observed in the long term. Moreover, for most transcripts, long-term treatment reversed the effects of short-term dosing, with larger changes in magnitude in the long term (Supplementary Figure 1).

Pathways and network analysis of long-term IFNβ-1b effects in patients

Three gene set analyses were computed to infer protein effects upstream, downstream and interacting with the measured long-term gene expression changes. The inferred upstream effects of transcription factors clearly showed a contrast between short-term and long-term transcriptional changes (Figure 2). The multi-gene analysis approach revealed distinct short-term and long-term gene expression patterns. The independently sampled CLIMB

study data, which comprises patient samples drawn at various times after beginning IFNβ therapy, shows a mixture of the patterns.

Downstream effects revealed a large number of canonical pathways influenced by long-term IFNβ-1b treatment (224 pathways at FDR < 0.05 of 953 tested). To relate biological pathways to single-gene changes, 28 of the 264 long-term differentially expressed genes were identified as members of the top-100 differentially expressed pathways. A cluster analysis of these 28 genes was used to group the observed pathway effects (Figure 3). Three major effects of IFNβ-1b long-term treatment appeared within the top-100 differentially expressed pathways. These effects included gene transcription, apoptosis and metabolism. Metabolism pathways included mitochondrial beta-oxidation of fatty acids, depicted as the intersection of a set of annotated pathways and a distinct pattern of gene membership. Canonical pathways spotlighted mitochondrial function and energy metabolism, both of which have an important role in maintaining neuronal integrity²⁶ as well as immune cell function.

Peri-expression analysis of protein interaction neighborhoods revealed additional regulatory proteins, not necessarily ones in named pathways (Figure 4). Disrupted levels of gene transcripts were significantly associated with neighbors of particular proteins. As larger signature sets provided more power to detect smaller effects on test ranks, we employed a plotting technique to determine *P*-values associated with a given protein neighbor that

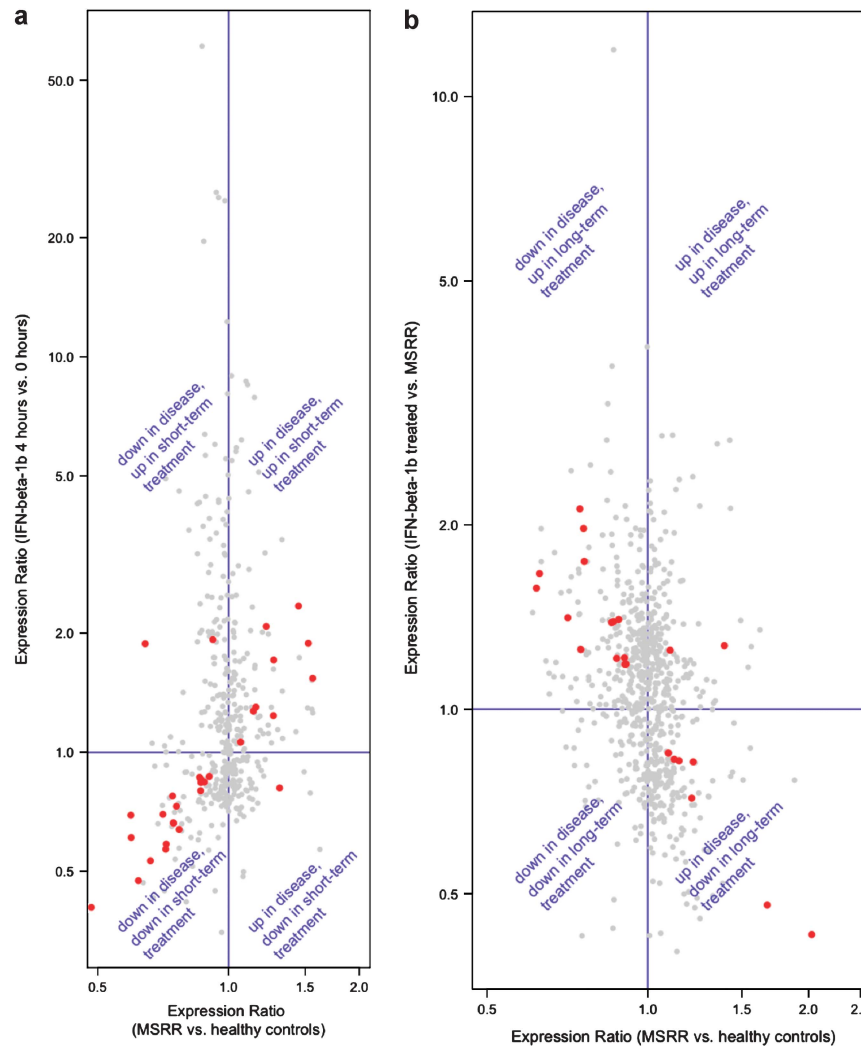


Figure 5. Long-term, but not short-term, interferon beta (IFN β) effects counter-regulate disease-related changes. **(a)** Short-term changes: genes exhibiting the most significant changes (t -test $P \leq 0.002$) observed 4-h after injection tend to be either unchanged in (stable) disease or change in the same direction as characteristic of multiple sclerosis (MS) patients, which is especially true of those probe sets more significantly changed in disease ($P \leq 0.05$, red points show positive slope). **(b)** Long-term changes: expression ratios in the long-term-treated patients versus therapy-naïve patients compared with the ratios found in therapy-naïve patients relative to healthy controls. The long-term-regulated gene expression for these differentially expressed probe sets (false discovery rate (FDR) ≤ 0.05 based on t -test) tends to reverse the expression changes that characterize disease, especially when focusing on the probe sets exhibiting more significant changes in disease ($P \leq 0.05$, red points show negative slope).

DISCUSSION

IFN β -bioactivity markers have been proposed,^{6,30–33} yet validation in independent studies and links to treatment outcomes that are relevant to clinical endpoints such as disability, exacerbation rate, cognition loss, brain atrophy and EDSS remain limited. A small number of studies have suggested that certain IFN β -bioactivity markers, such as transient, neutralizing antibody effects, may predict magnetic resonance imaging outcomes.³⁴ However, treatment response markers associated with MS-relevant regulation of biological functions are lacking.^{1,2,11,12,35} Treatment outcome biomarkers from differential gene expression studies in IFN β -treated MS patients have been a challenge to confirm.⁶ Mechanistically themed gene expression patterns that are confirmed across studies should better represent treatment-mediated changes. To identify such biomarkers, we integrated biological knowledgebases with the effects of IFN β -1b treatment in a well-controlled differential gene expression study^{14,15} and defined 10 multi-gene signatures to measure specific treatment effects. Statistical power

compared with single-gene models is increased using this approach, which allowed for rigorous confirmatory hypothesis testing in independent IFN β -treated MS patient data (Table 2).

The incorporation of biological themes relevant in MS therapy into these confirmed signatures, in combination with other measures (such as relapse rate, number of Gd-enhancing lesions, presence of neutralizing antibodies), can lead to mechanistically specific biomarkers able to facilitate treatment decisions. In addition, multi-gene RNA signatures measured before treatment provide an advantage over genome-wide association study biomarkers because the latter yield odds ratios often too low for predictive engineering.^{36,37} An obvious advantage to RNA signatures measured during treatment is that these characterize actual treatment-mediated changes in patients.

The finding that long-term IFN β -1b treatment reversed particular MS-related gene expression abnormalities (Figure 5), together with the observation that short-term treatment effects are similar to some MS-related changes, raises the possibility that

Table 2. Independent confirmation of IFN β -1b gene expression signatures

	Goertsches				CLIMB
	2 d vs 0 m	1 m vs 0 m	12 m vs 0 m	24 m vs 0 m	GA vs untreated
ST-4h+18h	4.33E-06	2.51E-15	1.17E-05	4.88E-08	5.10E-01
ST-4h	2.46E-03	2.52E-12	7.00E-05	7.92E-06	1.27E-01
ST-18h	8.50E-02	3.18E-09	3.18E-02	8.80E-07	3.37E-01
LT	1.16E-03	2.57E-10	2.43E-05	9.39E-03	1.08E-01
LT2	2.26E-02	3.44E-05	3.88E-04	5.72E-02	9.96E-02
MT-OX	5.10E-03	3.54E-04	1.56E-02	6.21E-01	5.02E-02
NRF2	1.47E-01	8.94E-03	3.51E-01	5.90E-01	2.89E-01
BBB	1.41E-06	4.03E-11	4.91E-06	2.59E-05	8.68E-01
IKE	4.17E-02	1.67E-05	1.12E-01	4.86E-02	3.16E-03
DR	2.60E-02	6.96E-07	2.78E-02	5.53E-02	7.47E-02

Abbreviations: BBB, blood-brain barrier; CLIMB, Comprehensive Longitudinal Investigation of MS at the Brigham and Women's Hospital; DR, disease reversal; GA, glatiramer acetate; IFN β , interferon beta; IKE, nuclear factor kappa-B kinase subunit epsilon related; LT, long-term; m, months; MT-OX, mitochondrial; NRF2, nuclear factor erythroid 2-related factor; ST, short-term.

Values are globaltest *P*-values for multi-gene signatures. Values in bold are individually significant at $P \leq 0.05$.

these MS patients exhibited an IFN β deficiency³⁸ that was corrected over time by repeated IFN β -1b dosing. Long-term IFN β -1b treatment effects were clearly distinct from short-term effects. Short-term effects influenced immune regulation and antiviral responses as expected.³⁵ Long-term treatment instead influenced metabolic functions, mitochondrial energetics and specific anti-oxidant mechanisms, which can preserve neuronal homeostasis.³⁹ Fatigue and cognitive function are associated with decreased brain metabolism in MS patients.^{40,41} This study presents multi-gene evidence for IFN β -1b regulation of mitochondrial function, in particular metabolic processes that control the balance of fatty acids and their derivative biomolecules. Mitochondrial dysfunction and neuroprotection in MS is extensively reviewed in Mao and Reddy³⁹ and Guzmán and Blázquez.⁴² IFN β -1b effects on the central nervous system could be mediated by carnitine regulation, as we observed dysregulation of the specific function 'import of palmitoyl-CoA into the mitochondrial matrix' (Figure 3). If MS brain exhibits defects in mitochondrial function, IFN β -1b effects on extrahepatic ketone body catabolism/formation may influence alternative energy sources for neurons.

MS relapses, magnetic resonance imaging activity, cognition and even survival^{33,43} are influenced by IFN β treatment. However, no convincing association is observed between relapse reduction and long-term disease progression,⁴⁴ which suggests there exist different mechanisms for neurodegenerative disease progression. Susceptibility of neurons to degeneration through mechanisms other than acute inflammation warrants consideration. Many energy metabolism genes are influenced by IFN β and are likely to be important in MS pathology.³⁹ As in Parkinson's disease and amyotrophic lateral sclerosis (ALS), energy metabolism and cytoprotective genes are important in chronic degenerative MS, and likely mediate beneficial effects of treatment.

This study's results suggest NADH dehydrogenase 6 (ND6), GPRC5C, OLA1 and NRF2 could potentially mediate long-term antioxidant effects of treatment. IFN β -1b-treated patients exhibited significantly higher expression of the mitochondrially encoded ND6 gene than treatment-naïve patients. Wild-type mt-ND6 maintains levels of reactive oxygen species,⁴⁵ potentially reducing oxidative damage. Mutated mt-ND6 is causative of Leber hereditary optic neuropathy,⁴⁶ a disease with MS-like white matter lesions.⁴⁷ This suggests a mechanism by which IFN β -1b treatment-modified mitochondria-mediated oxidative-stress responses could influence oligodendroglia in MS. GPRC5C was itself somewhat increased by IFN β -1b treatment (23% increase in long-term comparison, $P = 0.015$) as well as defining a neighborhood significantly regulated by treatment. This G protein-coupled

receptor family protein has been suggested to mediate retinoic acid effects on G protein signal transduction cascade,⁴⁸ but is otherwise little studied. OLA1 is an NRF2-independent, transcription-independent regulator of oxidative stress.⁴⁹ OLA1, identified in the long-term treatment comparison, was a contributing neighbor of GPRC5C and was significantly decreased by treatment. The observation was confirmed in the CLIMB study data ($P = 0.002$). This particular oxidative-stress-reducing effect of IFN β -1b has not been previously identified. In addition, our observed treatment effects related to NRF2, a major mediator of oxidative control provides a neuroprotective mechanism that requires further investigation.⁵⁰

IKK ϵ (encoded by IKBKE) is a link between the protein network analysis, in which IKK ϵ was the hub of the most significantly regulated network neighborhood, and the canonical pathway analysis, in which mitochondrial energy metabolism was highlighted. IKK ϵ regulates energy balance.⁵¹ A role for IKK ϵ in long-term IFN β treatment is consistent with the anti-inflammatory synergy between IKK ϵ and IFN β *in vivo*.⁵² IFN β induces rapid phosphorylation and activation of IKK ϵ .⁵³ IKK ϵ is critical to IFN β 's role in response to viral infection⁵³ as it integrates important signals from pattern recognition receptors that detect viral nucleic acids,⁵⁴ particularly notable here in light of the association of viruses and exacerbations in MS.

All 10 RNA signatures developed in this study were significantly regulated in the Goertsches study data,²¹ indicating that both long-term and short-term effects were captured in the Goertsches patient profiles and that signatures of IFN β -1b-controlled biological functions were confirmed. The temporal relationship between IFN-induced differential gene expression and physiological response can now be defined across different studies. The 10 signatures described provide a family of IFN-bioactivity markers that target distinct molecular mechanisms of action that can be studied further, especially in regard to patient response and clinical or magnetic resonance imaging outcomes.

Treatment of MS patients with GA reduces disease burden and relapses.⁵⁵ However, as with IFN β treatment, a cohort of patients appears to be non-responsive to treatment. With the sole exception of the IKE signature that characterized IKK ϵ -related expression changes, signatures derived from IFN β treatment effects were not changed by GA treatment, confirming that the signatures developed in this study measure IFN β -1b-specific treatment effects. Monitoring the activity of these signatures should now serve as a foundation to predict effective combination drug use and provide guidance in selecting treatment options. In addition, the IKBKE signature may point to an

important common treatment mechanism and warrants further investigation.

Large multi-gene expression patterns reflect biological effects observable across different gene expression platforms and different MS patient treatment studies. More focused multi-gene signatures permitted confirmation of diverse IFN β -1b effects in independent study data. Analysis revealed distinct long-term and short-term effects of IFN β -1b treatment that actually contrasted in direction of change. Protein interaction network analysis strongly underscored a key role for IKK ϵ in long-term IFN β -1b treatment effects. Long-term IFN β -1b treatment affected numerous pathways, including oxidative control and mitochondrial mechanisms important in maintaining normal brain homeostasis.

CONFLICT OF INTEREST

The efforts of HS and KDY were compensated through a contract funded by Bayer HealthCare Pharmaceuticals, EC and VK were employed by Bayer HealthCare Pharmaceuticals, and ATR has consulted for Bayer, Biogen, Novartis, Serono and Teva Pharmaceuticals.

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