

ORIGINAL ARTICLE

Loss of the Birt–Hogg–Dubé tumor suppressor results in apoptotic resistance due to aberrant TGF β -mediated transcription

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Birt–Hogg–Dubé (BHD) syndrome is an inherited cancer susceptibility disease characterized by skin and kidney tumors, as well as cystic lung disease, which results from loss-of-function mutations in the *BHD* gene. *BHD* is also inactivated in a significant fraction of patients with sporadic renal cancers and idiopathic cystic lung disease, and little is known about its mode of action. To investigate the molecular and cellular basis of BHD tumor suppressor activity, we generated mutant *Bhd* mice and embryonic stem cell lines. BHD-deficient cells exhibited defects in cell-intrinsic apoptosis that correlated with reduced expression of the BH3-only protein Bim, which was similarly observed in all human and murine BHD-related tumors examined. We further demonstrate that Bim deficiency in *Bhd*^{-/-} cells is not a consequence of elevated mTOR or ERK activity, but results instead from reduced *Bim* transcription associated with a general loss of TGF β -mediated transcription and chromatin modifications. In aggregate, this work identifies a specific tumor suppressive mechanism for BHD in regulating TGF β -dependent transcription and apoptosis, which has implications for the development of targeted therapies.

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Introduction

Birt–Hogg–Dubé (BHD) syndrome is a rare inherited cancer susceptibility disease characterized by benign hair follicle tumors and lung cysts in a majority of patients, and renal cell carcinoma (RCC) in approximately one third of diagnosed BHD cases (Schmidt *et al.*, 2005). Genetic analyses of families affected by BHD syndrome previously identified germline mutations in the *BHD* gene on chromosome 17p11.2, most of which were predicted to prematurely truncate the encoded protein (Nickerson *et al.*, 2002; Toro *et al.*,

2008). Mutational analyses of BHD-related tumors from patients, as well as those from mouse and rat models of BHD syndrome, identified inactivating ‘second hits’ in the remaining wild-type *BHD* allele, formally establishing BHD as a tumor suppressor (Okimoto *et al.*, 2004; Vocke *et al.*, 2005; Hasumi *et al.*, 2009). *BHD* inactivation has also been described in a subset of von Hippel–Lindau-independent RCC syndromes, in approximately one third of sporadic renal cancers, and in 60% of idiopathic cystic lung disease cases (Khoo *et al.*, 2003; Gunji *et al.*, 2007; Woodward *et al.*, 2008).

While the genetic basis of BHD syndrome is well understood, the cellular and molecular mechanisms involving the protein encoded by *BHD*, also called Folliculin, remain unclear. Dissecting BHD's molecular functions is particularly challenging since the *BHD* gene product bears no sequence or functional homology to any known protein. Studies in *Schizosaccharomyces pombe* have suggested a role for the putative yeast BHD ortholog in amino-acid homeostasis, while siRNA-mediated knockdown of *Drosophila* BHD in the fly has implicated it in male germline stem cell maintenance through Stat and/or BMP signaling (Singh *et al.*, 2006; van Slegtenhorst *et al.*, 2007). Finally, mammalian cell culture analyses and mouse models have linked BHD and mammalian target of rapamycin (mTOR) signaling, potentially mediated through an interaction with its binding partner, called Folliculin-interacting protein, and AMP-activated kinase (Baba *et al.*, 2006). These studies demonstrate that BHD loss can paradoxically result in either stimulation or inhibition of mTOR depending on the system examined (Baba *et al.*, 2006, 2009; Hudon *et al.*, 2008, 2010).

To investigate the cellular and molecular mechanisms by which BHD suppresses tumorigenesis, we established *Bhd*^{-/-} embryonic stem (ES) cell lines, since the early lethality of *Bhd*^{-/-} embryos precluded the development of other cellular models. We observed that the major cellular consequence of BHD loss is resistance to apoptosis caused by decreased Bim expression. This phenomenon was independent of mTOR and ERK hyperactivation and was instead associated with a general loss of TGF β -dependent chromatin modifications and transcription. These data present new insight into how BHD functions as a tumor suppressor and

suggest novel targeted therapeutics for BHD syndrome, sporadic RCC, and cystic lung disease.

Results

Homozygous mutant Bhd embryos die during early embryogenesis

To define the basic molecular functions of BHD, we initially attempted to establish mouse embryonic fibroblasts from a gene-trap mouse model. In order to fully characterize this mutant *Bhd* allele (denoted as *m*), we mapped the insertional position of the trap cassette to intron 8 of the murine *Bhd* gene, and established Southern and PCR-based genotyping assays to

discriminate wild type from the *m* allele (Figures 1a–c). Using multiple quantitative real-time (qRT)–PCR probes, we observed a 40–50% reduction in *Bhd* mRNA expression in *Bhd*^{+/*m*} ES cells compared with *Bhd*^{+/*+*} controls by qRT–PCR (Figure 1d), and confirmed expression of the BHD–βgeo fusion product by staining the ES cells with X-Gal (Figure 1e).

Bhd^{+/*m*} intercrosses yielded no viable *Bhd*^{*m/m*} embryos after e7.5; although we were unable to genotype e6.5 embryos, we observed that ~25% of these were severely abnormal or partially resorbed upon dissection, exhibiting grossly atrophied and disorganized egg cylinders, distorted visceral endoderm and lack of cavitation (Figures 1f and g). This phenotype is consistent with a recently reported embryonic lethal phenotype for an independent mutant *Bhd* allele (Hasumi *et al.*, 2009).

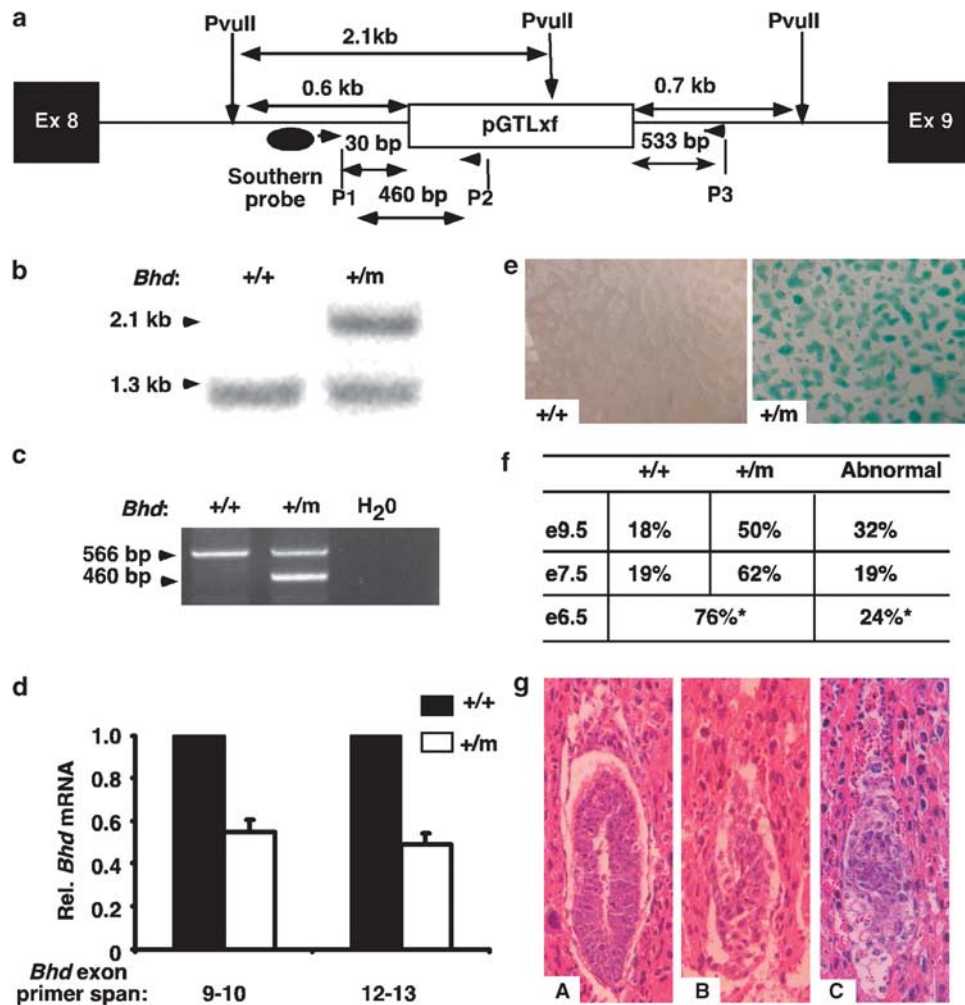


Figure 1 *Bhd*^{*m/m*} embryos are early embryonic lethal. (a) Schematic depicting integration site of Bay Genomics *pG2Lxf* gene-trap vector in murine *Bhd* intron 8, location of probe and restriction sites for Southern blot genotyping and location of primers (P1, P2, and P3) for PCR-based genotyping. (b) Southern blot of *PvuII*-digested genomic DNA and (c) PCR-based genotypes of *Bhd*^{+/*+*} or *Bhd*^{+/*m*} ES cells to discriminate wild-type from mutant alleles. ‘*m*’ specifically designates the gene-trapped mutant *Bhd* allele from Bay Genomics throughout the manuscript. (d) Relative *Bhd* mRNA levels in *Bhd*^{+/*+*} versus *Bhd*^{+/*m*} ES cells assayed by qRT–PCR using primers spanning exons just downstream of the trap cassette (exons 9–10) or across the terminal exons of the *Bhd* gene (exons 12–13), ruling out the presence of aberrant transcripts generated from cryptic transcriptional start sites downstream of the trap cassette. (e) X-Gal staining of *Bhd*^{+/*+*} and *Bhd*^{+/*m*} ES cells showing *Bhd*^{+/*m*} cells express the *Bhd*–βgalactosidase fusion product. (f) Table of progeny obtained from *Bhd*^{+/*-*} intercrosses showing no *Bhd*^{-/-} embryos could be recovered after e6.5. (g) 200 × images of H&E-stained cross-sections of paraffin-embedded e6.5 embryos from *Bhd*^{+/*m*} intercrosses. (A) Exemplifies normal morphology while (B) and (C) demonstrate gross abnormalities, probable *Bhd*^{*m/m*} genotypes.

Loss of BHD does not affect proliferation, but results in decreased cellular size and resistance to cell-intrinsic apoptosis

Since the early lethality of the *Bhd*^{mi/mi} embryos precluded the development of differentiated cell types, we generated an ES cell model for *in vitro* studies. To do this, we used an independent mutant *Bhd* allele (denoted as '-') with a gene trap in *Bhd* intron 1, which ablated expression of the entire *Bhd* open reading frame. The *Bhd*^{+/-} ES cells were characterized in the same manner as the *Bhd*^{+/+} cells to validate efficacy of the trap cassette (Supplementary Figures 1a–e).

Bhd^{-/-} ES cells were generated from the parental *Bhd*^{+/-} cells as shown by PCR, and western blot confirmed complete loss of BHD protein expression (Figures 2a and b). To verify BHD loss did not affect the undifferentiated state of *Bhd*^{-/-} ES cells, we analyzed protein levels of Oct-4, Nanog, Sox2 and phospho-Stat3, and alkaline phosphatase activity in *Bhd*^{+/+}, *Bhd*^{+/-}, and *Bhd*^{-/-} cells (Supplementary Figures 2a–c). We observed no difference in all parameters of ES cell pluripotency, allowing for valid comparison in *in vitro* studies across genotypes.

To gain insight into BHD's tumor suppressive properties, we assessed *Bhd*^{+/+}, *Bhd*^{+/-}, and *Bhd*^{-/-} ES cell lines for proliferation, cell growth, and apoptotic responses. *Bhd*^{-/-} ES cells did not proliferate significantly faster at days 1–3 after plating, but confluent cultures analyzed at day 4 were found to contain 15–20% more cells compared with *Bhd*^{+/+} and *Bhd*^{+/-} counterparts (Figure 2c). Intriguingly, this difference was attributed to a 15–20% decrease in cellular volume in *Bhd*^{-/-} cells (Figure 2d).

Finally, we evaluated the response of *Bhd*^{-/-} cells to cell-intrinsic apoptosis by starving cells of serum, glucose, or amino acids for 24 h. *Bhd*^{-/-} cells clearly demonstrated a resistance to all three apoptosis-inducing stresses as revealed by light microscopy, decreased accumulation of sub-G1 populations, and reduced Caspase-3 and Parp cleavage (Figures 2e–h). This effect was specific to the intrinsic apoptotic pathway, as *Bhd*^{-/-} cells showed no differential response to the death-receptor ligand Tumor necrosis factor α (Figure 2g). Thus, BHD deficiency does not confer an obvious increase in cellular proliferation or growth, but does reduce cell-intrinsic apoptosis, thereby identifying a plausible cellular role for BHD as a tumor suppressor.

Bim expression is lost in *Bhd*^{-/-} cells and BHD-related tumors and contributes to apoptotic resistance

The cell-intrinsic apoptotic resistance of *Bhd*^{-/-} cells could either be due to increased intracellular nutrient stores, an effect previously observed in a mutant yeast model of BHD, or result from a defect in the cell-intrinsic death machinery (van Slegtenhorst *et al.*, 2007). To assess the former possibility, we measured levels of 13 different intracellular amino acids by high performance liquid chromatography and observed no differences between *Bhd*^{+/+} and *Bhd*^{-/-} ES cells, suggesting that nutrient homeostasis was not playing a role in the apoptotic-resistance phenotype (Figure 3a).

To determine if the cell-intrinsic death machinery might be impaired in *Bhd*^{-/-} ES cells, we examined protein levels of several Bcl2 family members and found many pro-apoptotic BH3-only proteins to be misregulated in *Bhd*^{-/-} cells. Both extra-long (EL) and long (L) isoforms of Bim, as well as Puma α and Bad were significantly downregulated in *Bhd*^{-/-} ES cells, while Bid and BMF levels were upregulated (Figure 2b). Bik, Noxa, and Puma β , as well as the multi-domain Bcl2 family members BclXL and Bak were unchanged. Similar results were obtained using three independent BHD-deficient ES clones homozygous for the *Bhd*^{mi} allele (Supplementary Figures 3a and b). We focused primarily on Bim regulation in subsequent experiments, given its dominant role among BH3-only proteins in mediating the cell-intrinsic death response and frequent loss in RCC (Kim *et al.*, 2006; Zantl *et al.*, 2007; Guo *et al.*, 2009).

To test whether the apoptotic resistance of *Bhd*^{-/-} cells was Bim dependent, we restored either *BimEL* or *Bhd* expression by retroviral transduction in *Bhd*^{-/-} cells. Restoration of *BimEL* or *Bhd* expression, or treatment of cells with the BH3-only chemical mimetic ABT-737, rescued the apoptotic response of *Bhd*^{-/-} ES cells to amino-acid deprivation, as assessed by FACs analysis (Figure 3c, upper panel), and Parp and Caspase-3 cleavage (Figure 3c, lower panel). Importantly, *Bhd* reconstitution was sufficient to restore Bim protein levels. Interestingly, autophagy inhibitors 3-methyladenine and chloroquine rescued the apoptotic response in *Bhd*^{-/-} cells (Figure 3c), similar to apoptosis-deficient *Bax*^{-/-}/*Bak*^{-/-} double knockout cells (Lum *et al.*, 2005).

To determine whether loss of BHD results in Bim deficiency *in vivo*, we performed immunohistochemistry on BHD-related tumors from both human BHD patients and aged *Bhd*^{+/+} mice. 3/3 human-derived tumors (one fibrofolliculoma, one chromophobe-clear cell hybrid RCC, and one chromophobe RCC) and 5/5 solid murine renal tumors (four hybrid oncocyctic and one papillary mass projecting from a cyst), exhibited absent or significantly reduced Bim expression, confirming that loss of Bim expression is a common event in BHD-related tumors of diverse histologies (Figure 3d).

Bim is transcriptionally downregulated in *Bhd*^{-/-} cells independent of mTORC1, mTORC2, ERK, or GCN2-eIF2 α pathway aberrations

Bim is regulated transcriptionally by several well-characterized signal transduction pathways, translationally by miRNAs, and on the level of protein stability (Dijkers *et al.*, 2000; Dehan *et al.*, 2009; Su *et al.*, 2009). We initially assessed *Bim* mRNA and found it to be decreased by ~80% in *Bhd*^{-/-} cells compared with *Bhd*^{+/+} and *Bhd*^{+/-} cells (Figure 4a). To investigate a possible additional role for enhanced proteolytic degradation of Bim, we treated *Bhd*^{-/-} cells with the proteasome inhibitor MG-132, but observed no increase in Bim protein (Supplementary Figure 4a) (Dehan *et al.*, 2009). We also examined levels of *mir-19* and *mir-92*,

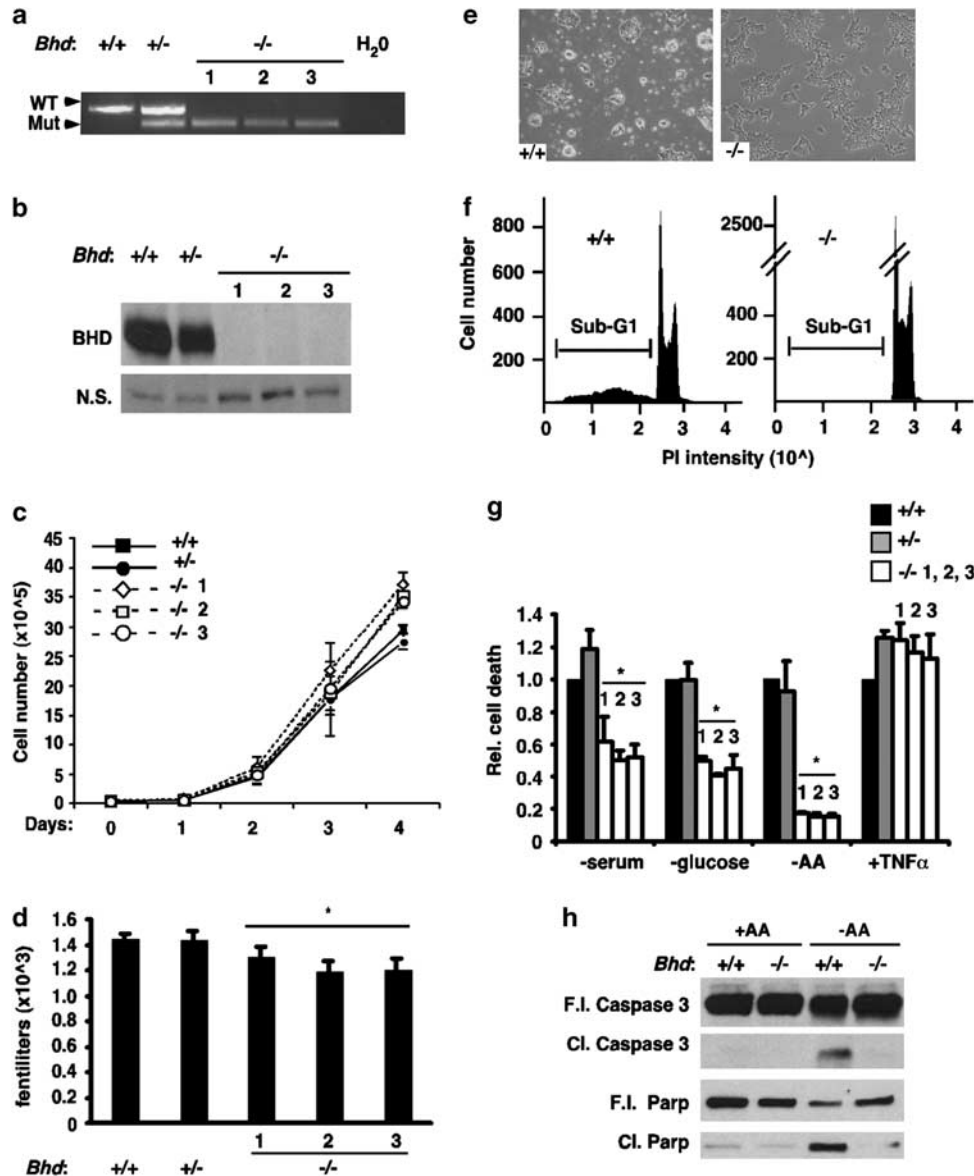


Figure 2 *Bhd*^{-/-} ES cells do not have a proliferative or growth advantage, but are resistant to apoptosis. (a) PCR genotypes verifying loss of wild-type *Bhd* allele in three independent ES clones, designated as ‘1’, ‘2’, and ‘3,’ after step-up selection. WT = wild-type, Mut = mutant alleles. (b) Western blot demonstrating loss of BHD protein expression in three independent *Bhd*^{-/-} ES clones with a non-specific (NS) band shown as a loading control. (c) Growth curve showing *Bhd*^{-/-} ES cells proliferate at similar rates to *Bhd*^{+/+} and *Bhd*^{+/-} counterparts, but have increased saturation density at day 4. (d) Coulter counter size measurements showing *Bhd*^{-/-} cells are significantly smaller than *Bhd*^{+/+} and *Bhd*^{+/-} ES cells (**P* = 0.000623). *Bhd*^{-/-} ES cells are resistant to stress-induced apoptosis as shown by (e) brightfield images of *Bhd*^{+/+} and *Bhd*^{-/-} ES cells starved of amino acids for 1 day, or (g) FACS analysis of cells containing sub-G₁ DNA content after 1 day serum, glucose and amino-acid (AA) deprivation compared with *Bhd*^{+/+} and *Bhd*^{+/-} cells (**P* < 0.0000345), but not by Tumor necrosis factor α treatment. (f) Representative FACS plot showing *Bhd*^{-/-} ES cells have significantly less sub-G₁ DNA content following 1 day of amino-acid starvation. (h) Western blot of Caspase 3 and Parp cleavage with and without amino-acid starvation in *Bhd*^{+/+} and *Bhd*^{-/-} ES cells. All bar graphs represent averages of three independent experiments with error bars showing standard error of the mean (s.e.m.).

miRNAs previously shown to regulate *Bim* translation in ES cells (Su *et al.*, 2009), but detected no differences between *Bhd*^{-/-} and control cells (Supplementary Figure 4b).

To determine which transcriptional networks and upstream signaling pathways were involved in *Bim* misregulation, we investigated the mTOR and ERK pathways that have previously been linked to *Bim*

regulation (Dijkers *et al.*, 2000; Dehan *et al.*, 2009), and are hyperactivated in late-stage cysts and tumors derived from a mutant *Bhd* mouse model (Baba *et al.*, 2008). Western blot analysis of *Bhd*^{+/+}, *Bhd*^{+/-}, and *Bhd*^{-/-} ES cells revealed that *Bhd*^{-/-} ES cells exhibited hyperphosphorylation of S6K1 and 4E-BP1 (indicative of increased mTOR complex 1 (mTORC1) activity), hyperphosphorylation of Akt and the FoxO family of

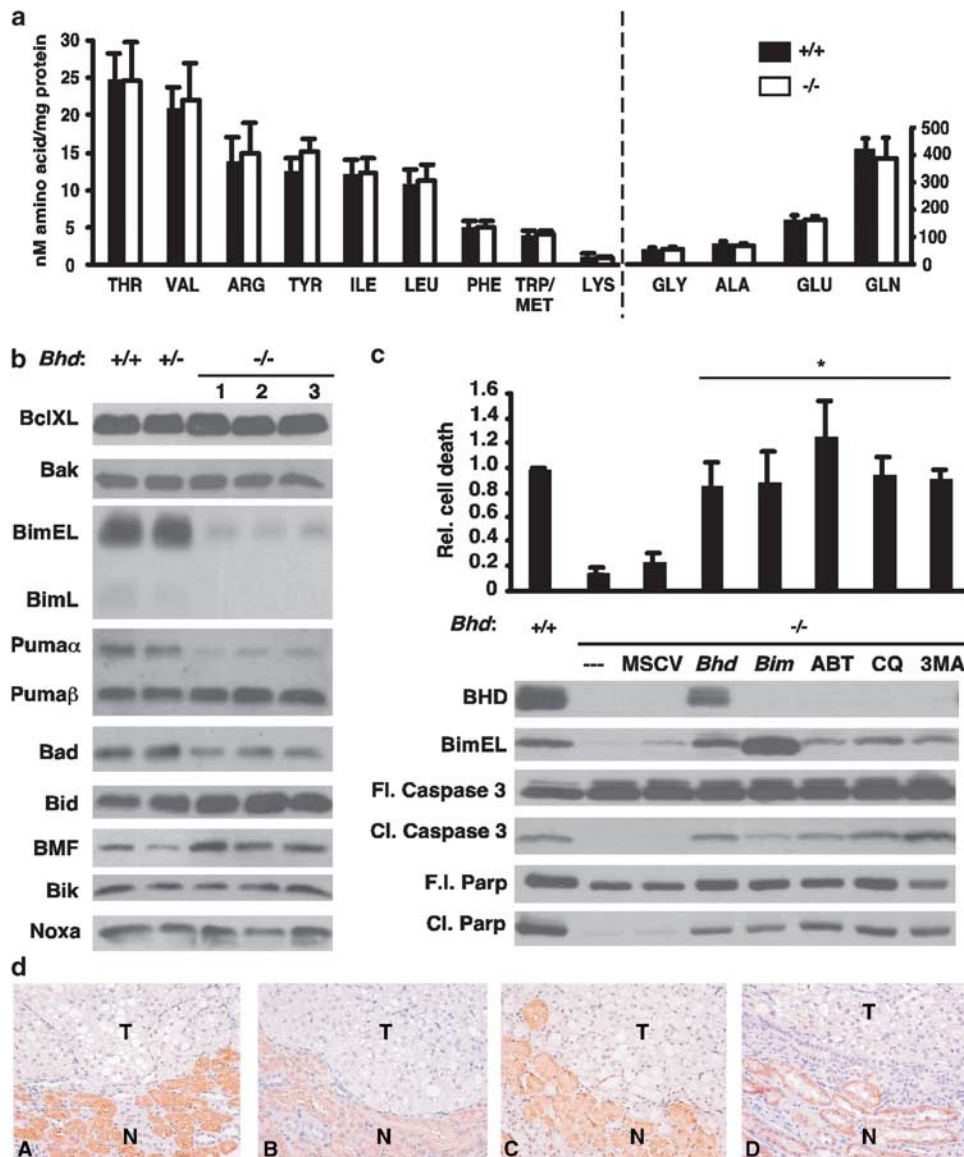


Figure 3 Death resistance of *Bhd*^{-/-} cells is not due to increased intracellular amino-acid levels, but instead results from decreased Bim protein levels. **(a)** Intracellular amino-acid levels normalized to total protein in *Bhd*^{+/+} versus *Bhd*^{-/-} ES cells as assessed by HPLC. Data represent average of three independent experiments and error bars reflect standard deviation. **(b)** Western blot showing *Bhd*^{-/-} ES cells have decreased levels of several BH3-only proteins, most notably Bim. **(c)** Quantification of sub-G₁ populations by flow cytometry in upper panel shows restoration of *Bhd* and *BimEL* expression, or treatment with ABT-737 (ABT), chloroquine (CQ) or 3-methyladenine (3MA) rescues the death-resistance phenotype of *Bhd*^{-/-} ES cells in response to 2-day amino-acid starvation compared with untreated or vector only (MSCV) infected controls. Bar graph represents three independent experiments and error bars show s.e.m. Lower panel is a western blot verifying re-expression of BHD and Bim, and correlate changes in Caspase-3 and Parp cleavage. **(d)** 200 \times images of paraffin-embedded sections of solid renal tumors (labeled T) from *Bhd*^{+/*m*} mice 18–21 months in age (A–C) or a human patient (D) exhibiting loss of Bim expression compared with normal adjacent tissue (labeled N). (A–C) Hybrid clear cell-oncocytic lesions from aged *Bhd*^{+/*m*} mice and (D) is a human chromophobe RCC.

proteins (indicative of mTOR complex 2 (mTORC2) hyperactivation) and hyperactivation of the MEK–ERK–p90RSK cascade (Figure 4b).

To determine if these pathways were playing a role in decreased *Bim* levels in *Bhd*^{-/-} cells, we treated cells with inhibitors of each at doses that reduced signaling in *Bhd*^{-/-} cells to levels observed in *Bhd*^{+/+} cells. We found that pharmacologic inhibition of PI3K-dependent mTORC2, MEK, or mTORC1 activities, either singly or

in combination, was insufficient to restore Bim protein levels in *Bhd*^{-/-} cells (Figure 4c). Treatment of *Bhd*^{-/-} cells with the same panel of inhibitors was similarly unable to rescue the death-resistance phenotype as assessed by sub-G₁ population quantification (Figure 4d, upper panel), or Caspase and Parp cleavage following 2-day amino-acid starvation (Figure 4d, lower panel).

Bim expression is also known to be regulated by Chop, a transcription factor activated downstream of

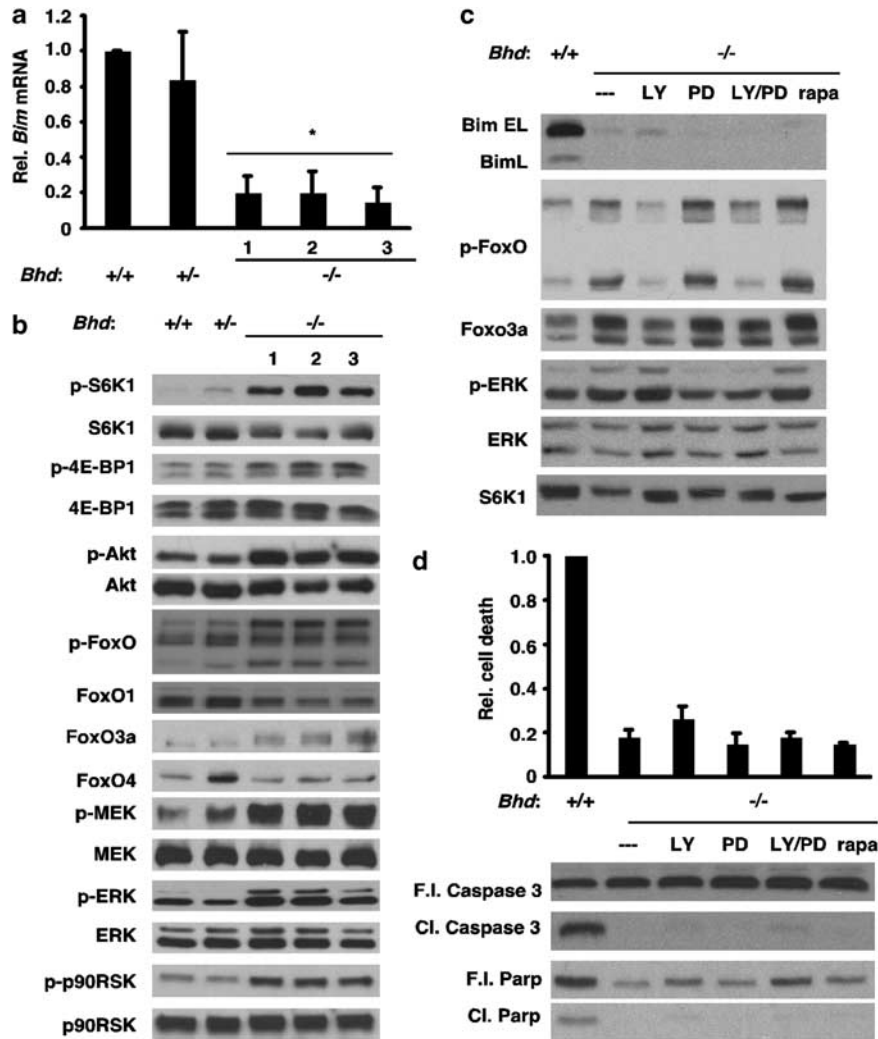


Figure 4 *Bim* is transcriptionally downregulated in *Bhd*^{-/-} cells independent of mTORC1, mTORC2, or ERK hyperactivation. (a) qRT-PCR analysis showing *Bim* mRNA is reduced in *Bhd*^{-/-} ES cells ($*P = 1.44 \times 10^{-7}$). (b) Western blot demonstrating that mTORC1 (indicated by p-S6K1 [T389] and p-4E-BP1 [T70]), mTORC2 (indicated by p-Akt [S473] and p-FoxO [FoxO1 [Thr24]/FoxO3a [Thr32]/FoxO4 [Thr28]) and ERK (indicated by p-MEK [S217/271], p-ERK [T202/Y204], and p-p90RSK [T359/S363]) signaling pathways are hyperactivated in *Bhd*^{-/-} ES cells. (c) Inhibition of hyperactivated mTORC2 by 5 μM LY294002 (LY) or ERK by 10 μM PD98059 (PD) or a combination of both, or inhibition of mTORC1 by 20 nM rapamycin (rapa) for 24 h does not restore *Bim* protein expression levels in *Bhd*^{-/-} cells by western blot. Drug efficacy was shown by decreased p-FOXO levels with LY treatment and decreased p-ERK levels with PD treatment in *Bhd*^{-/-} cells. Decreased mobility shifts in S6K demonstrate effectiveness of both LY and rapa treatments. Doses were chosen based on their ability to bring activation levels of signaling components in *Bhd*^{-/-} cells to those observed in *Bhd*^{+/+} cells. (d) *Bhd*^{-/-} ES cells were pre-treated for 6 h with the same panel of inhibitors as (c), then starved of amino acids for 2 days with inhibitors re-added after 1 day of starvation to maintain concentrations. Death was assessed by quantification of sub-G₁ populations by flow cytometry (upper panel, $*P < 0.0345$) and Caspase and Parp cleavage by western blot (lower panel). All bar graphs in this figure represent averages of three independent experiments with error bars reflecting s.e.m.

eukaryotic initiation factor α (eIF2α) kinases as part of the unfolded protein response and as an adaptation to nutrient withdrawal (Puthalakath *et al.*, 2007). Interestingly, *Bhd*^{-/-} cells exhibited decreased basal and amino-acid deprivation-induced levels of p-eIF2α levels, but not ER stress-induced p-eIF2α after thapsigargin treatment (Supplementary Figures 5a–c). However, induction of the downstream effectors ATF4 and Chop was only slightly attenuated in response to amino-acid deprivation (Supplementary Figure 5b). More importantly, reintroduction of Chop into *Bhd*^{-/-} cells failed to

rescue *Bim* expression (Supplementary Figure 5d). Collectively, these data suggest that the downregulation of *Bim* and apoptotic resistance observed in *Bhd*^{-/-} cells are not due to signaling aberrations in the mTORC1, mTORC2, ERK, and eIF2α signaling pathways.

Bhd^{-/-} ES cells exhibit many phenotypic and molecular defects characteristic of TGFβ-pathway mutants
We next considered the TGFβ pathway, since many reports had previously shown it regulated *Bim*

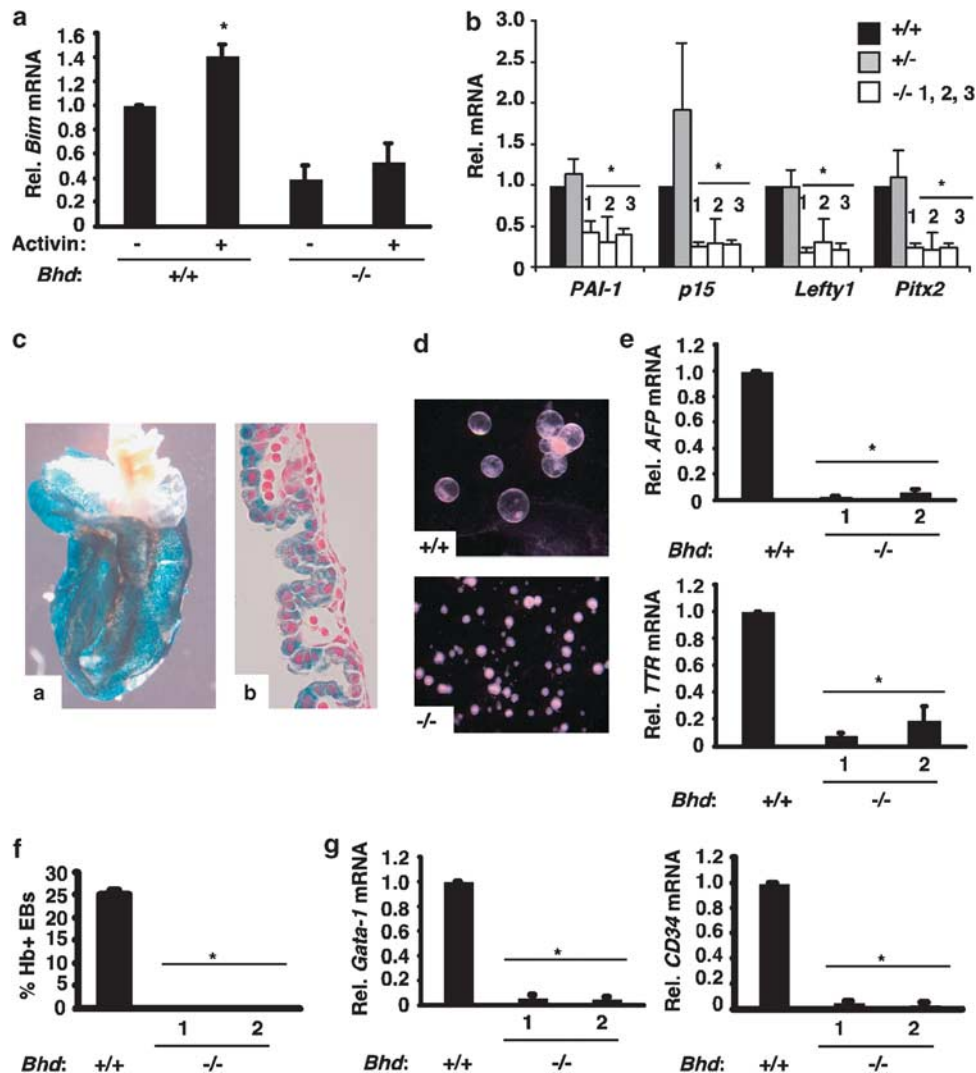


Figure 5 *Bhd*^{-/-} ES cells exhibit phenotypes and transcriptional defects characteristic of TGF β -signaling components. (a) *Bhd*^{+/+} and *Bhd*^{-/-} ES cells were cultured for 24 h in N2B27 media, then stimulated with 10 ng/ml of Activin. *Bim* mRNA levels were significantly induced in *Bhd*^{+/+} cells after stimulation, but not in *Bhd*^{-/-} ES cells ($*P=0.00864$). (b) qRT-PCR analysis showing canonical TGF β target genes *PAI-1*, *p15*, *Lefty1*, and *Pitx2* are significantly downregulated in multiple *Bhd*^{-/-} ES cell clones labeled 1, 2 and 3 ($*P<0.04$). (c) X-gal stained 70 \times image of (a) whole mount or 200 \times image of (b) sectioned yolk sac taken from a e10.5 *Bhd*^{+/m} embryo exhibiting high *Bhd* expression in visceral endoderm of the yolk sac. (d) 200 \times image of day 12 EBs showing *Bhd*^{+/+} EBs form expanded cystic structures reminiscent of yolk sacs while *Bhd*^{-/-} EBs fail to do so. (e) qRT-PCR analysis on RNA from EBs, showing two independent *Bhd*^{-/-} EBs (clones 1 and 2) fail to express maximal levels of mature yolk sac markers α -feto-protein (*Afp*) and *Trithioredoxin* (*Ttr*). Averages represent maximal expression levels for each mRNA, which occurred between days 6 and 9 of EB development ($*P<1.0 \times 10^{-5}$ for both mRNAs). (f) Quantification of benzediene-positive EBs that express hemoglobin (Hb) at day 10 in methylcellulose cultures demonstrating *Bhd*^{-/-} EBs fail to form erythroid lineages ($*P=2.32 \times 10^{-6}$). Data represent the average of three independent experiments with error bars showing standard deviation. (g) qRT-PCR analysis of *Gata-1* and *CD34* mRNA expression in day 10 EBs in methylcellulose cultures, which is significantly reduced in *Bhd*^{-/-} EBs ($P<4.49 \times 10^{-6}$).

transcription in diverse cell types (Willey *et al.*, 2003; Ramjaun *et al.*, 2007; Ramesh *et al.*, 2008; Yu *et al.*, 2008; Houde *et al.*, 2009). We first verified that the TGF β ligand family regulates *Bim* mRNA in ES cells by treating them with Activin, a TGF β family ligand to which ES cells are responsive. *Bim* mRNA was increased within 2 h of Activin treatment in *Bhd*^{+/+} ES cells, which did not occur in *Bhd*^{-/-} cells (Figure 5a). To ascertain whether this effect was specific to *Bim*, we analyzed the mRNA levels of several other BH3-only family members in response to Activin. Contrary to *Bim*, *Bid*, *BMF*, and *Bad* mRNAs were not induced by

Activin in wild-type cells, and actually appeared elevated in *Bhd*^{-/-} cells (Supplementary Figure 6a). The latter finding was consistent with increased Bid and BMF protein levels in *Bhd*^{-/-} cells, but appeared opposite to that of Bad protein levels (Figure 3b). While *Pumax* mRNA was induced by Activin, this was not BHD dependent as it occurred in both *Bhd*^{+/+} and *Bhd*^{-/-} cells (Supplementary Figure 6a). Taken together, these data identify *Bim* as a specific BH3-only mRNA regulated by TGF β in a BHD-dependent manner.

To determine if TGF β -mediated transcriptional defects were a general phenomenon in *Bhd*^{-/-} cells, we

examined the basal mRNA levels of several established TGF β -transcriptional targets involved in diverse biological processes. mRNA levels for the cell cycle regulator *p15*, pro-apoptotic plasminogen inhibitor activator type I (*PAI-1*) and *Lefty1* and *Pitx2*, which have developmental roles were all lower in *Bhd*^{-/-} cells compared with *Bhd*^{+/+} cells (Figure 5b). Other canonical TGF β mRNA targets such as *SnoN*, *p21*, *Colla2*, and *Lefty2* were similarly downregulated in *Bhd*^{-/-} cells (Supplementary Figure 6b). Other TGF β targets previously implicated in cell cycle regulation and apoptosis were unaffected in *Bhd*^{-/-} cells, including *Gadd45b* and *p57*, while the death-associated protein kinase was actually upregulated (Supplementary Figure 6c). Overall, these data support a positive role for BHD in TGF β -mediated transcription that governs biological processes implicated in cancer such as apoptosis, cell cycle, and differentiation.

TGF β ligands and receptors are required for normal yolk sac vasculogenesis and embryonic hematopoiesis *in vivo*, and in ES cell-derived cystic embryoid body (EB) cultures *in vitro* (Dickson *et al.*, 1995; Oshima *et al.*, 1996; Goumans *et al.*, 1999; Larsson *et al.*, 2001). Interestingly, strong BHD expression was observed in yolk sac visceral endoderm, as revealed by X-Gal staining of E10.5 embryos heterozygous for the *Bhd*^{+/+} allele (Figure 5c). To investigate whether BHD is required for yolk sac development, we generated day 12 EBs from *Bhd*^{+/+} and *Bhd*^{-/-} ES cells. *Bhd*^{-/-} ES cells failed to form expanded cystic EBs, which are reminiscent of mature yolk sacs *in vivo* (Figure 5d), or induce mature yolk sac markers α -fetoprotein (*Afp*) and *Trithioredoxin* (*Ttr*) (Doetschman *et al.*, 1985) (Figure 5e). To further evaluate the role of BHD in embryonic hematopoietic stem cell development, we generated *Bhd*^{+/+} and *Bhd*^{-/-} EBs in methylcellulose cultures, which promotes the differentiation of hematopoietic lineages. *Bhd*^{-/-} EBs failed to form hemoglobinized colonies at day 10 in culture (Figure 5f; Supplementary Figure 6b) and exhibited severely reduced mRNA levels of the erythroid gene *Gata-1* and *CD34* (a marker of both hematopoietic and endothelial lineages) (Figure 5g). Interestingly, we observed a delayed induction of the mesodermal marker *Brachyury*, a super-induction of *Fgf-5* and attenuated maintenance of *Hnf4* at later stages (Supplementary Figure 7). However, these minor effects are unlikely to explain the dramatic hematopoietic and yolk sac defects of *Bhd*^{-/-} EBs, as maximal induction of these target genes was still robust in *Bhd*^{-/-} EBs, increasing by orders of magnitude.

Bhd^{-/-} cells exhibit hypo-acetylation of TGF β target gene promoters, resulting in their death-resistance phenotype
Because BHD deficiency resulted in molecular and phenotypic defects similar to TGF β receptor and ligand mutants, we next investigated whether the transcriptional defects occurred at the level of receptor-mediated phosphorylation and nuclear translocation of Smad-transcriptional regulators. Therefore, we examined

nuclear levels of phosphorylated and total receptor-regulated Smad2 and the common partner Smad4, which form a complex and translocate to the nucleus upon cellular engagement of TGF β ligands (Zhang *et al.*, 1996). Western blot analysis did not reveal any differences in basal or Activin-induced nuclear accumulation of phospho-Smad2, total Smad2, or total Smad4 in *Bhd*^{+/+} versus *Bhd*^{-/-} ES cells (Figures 6a and b).

Since Smads have previously been shown to activate transcription via acetylation of Histone H3 at target promoters, we performed chromatin immunoprecipitations for acetylated Histone H3 at the well-characterized *Lefty1* promoter (Ross *et al.*, 2006). Whereas *Bhd*^{+/+} ES cells demonstrated a significant increase in acetyl-H3 levels at the *Lefty1* promoter following Activin treatment, *Bhd*^{-/-} cells exhibited no response (Figure 6c). A similar effect was observed at the promoter of *PAI-1*, which has been implicated in the apoptotic response (Kortlever *et al.*, 2006; Lademann and Romer, 2008) (Supplementary Figure 8a). This difference was not due to a global reduction of acetyl-H3 levels, which appeared unaffected based on western blot assays (Figure 6d). To determine whether restoration of acetylated Histone levels rescues *Bhd*^{-/-} cell defects, *Bhd*^{-/-} cells were treated with the histone deacetylase (HDAC) inhibitor Trichostatin A (TSA), which restored mRNA levels of multiple TGF β target genes (*PAI-1*, *p15*, and *Bim*), *Bim* protein levels and cell death (Figures 6e and f). TSA appeared to have variable effects on other misregulated BH3-only proteins in *Bhd*^{-/-} cells under nutrient deprivation (Supplementary Figure 8b). First, nutrient deprivation itself changed protein levels of some BH3-only proteins in *Bhd*^{-/-} cells, with BMF actually decreasing and Bad equilibrating to wild-type levels, in contrast to nutrient replete conditions (Figure 3a). TSA appeared to have no effect on Puma α and Bid levels abundance under nutrient limitation, while it slightly elevated Bad and BMF levels. It seems unlikely, however, that Bad and BMF induction can explain the effects of TSA, given the ability of *Bim* re-expression alone to completely reverse the phenotype of *Bhd*^{-/-} cells. In summary, BHD loss appears to contribute to apoptotic resistance and TGF β -dependent transcriptional defects via aberrations in chromatin modifications at target gene promoters including *Bim* (Figure 7).

Discussion

To date, the cellular and molecular mechanisms of tumor suppression by BHD have remained elusive. Uncovering BHD protein functions is particularly challenging, given the lack sequence homology to any known protein. In this study, we initially observed that the major cellular consequence of BHD loss is apoptotic resistance due to loss of *Bim* expression, a phenomenon that can be observed in BHD-related renal tumors. These findings prompted us to ask what signaling pathways and/or transcriptional networks were implicated

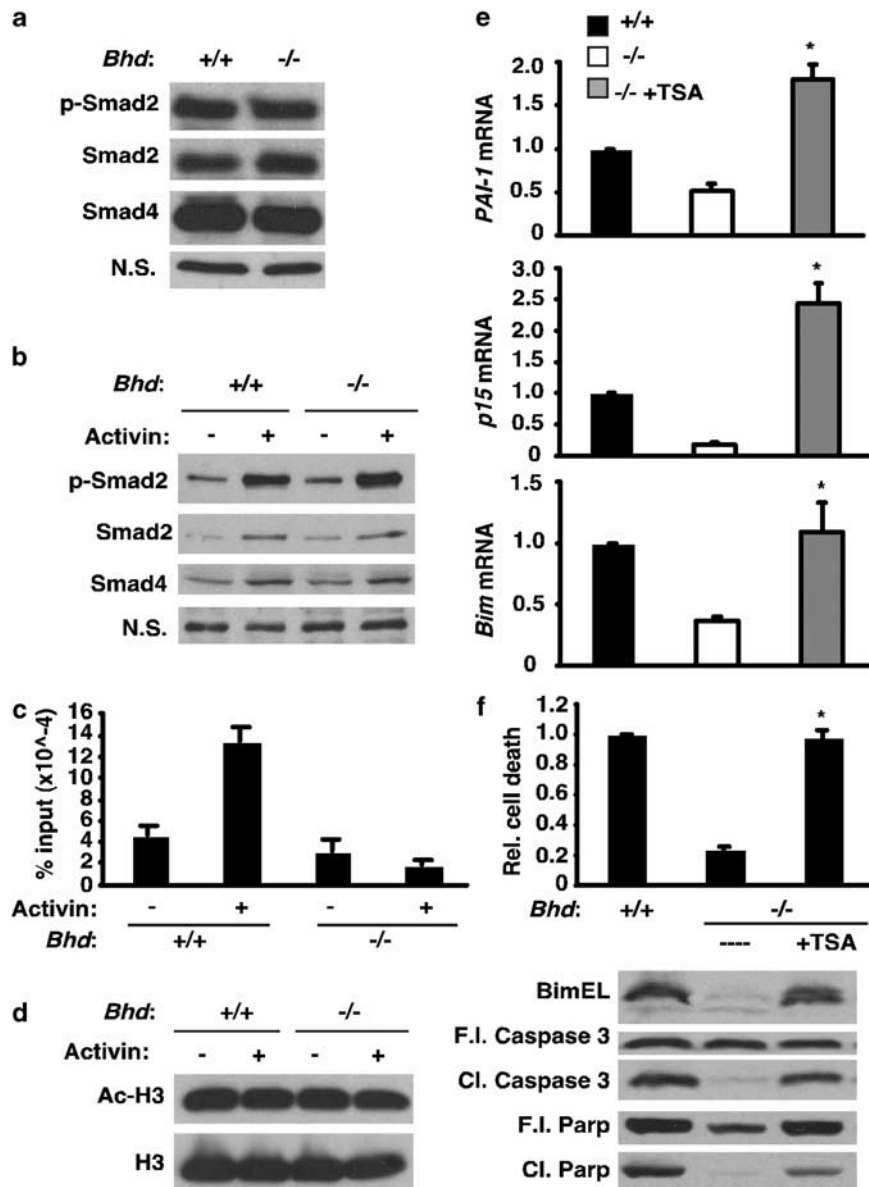


Figure 6 Loss of BHD results in HDAC-mediated silencing of TGF β -transcriptional targets. Western blot of nuclear extracts showing (a) basal or (b) Activin-induced levels of nuclear phospho-smad2, Smad2, and Smad4 are unaffected in *Bhd*^{-/-} ES cells. Non-specific (NS) band is shown as a loading control. (c) qRT-PCR using primers located in the *Lefty1* promoter on genomic DNA immunoprecipitated with acetyl-histone H3 antibody from ES cells stimulated with activin for 1 h following 24 h culture in N2B27. (d) Western blot showing total levels of acetyl-histone H3 are unaffected in *Bhd*^{+/+} versus *Bhd*^{-/-} cells with and without Activin treatment. (e) Treatment of *Bhd*^{-/-} cells with TSA restores mRNA levels of *PAI-1*, *p15*, and *Bim* as shown by qRT-PCR analysis of mRNA expression levels (**P* < 0.0362). (f) Treatment of *Bhd*^{-/-} ES cells with TSA rescues the *Bhd*^{-/-} cellular death-resistance phenotype and *Bim* expression as shown by analysis of sub-G₁ populations (above, **P* = 0.000145) and western blot for *Bim*, and Caspase and Parp cleavage (below).

in this phenotype. We initially examined the mTOR pathway, as multiple published reports have shown correlations between BHD loss and mTOR activity (Hartman *et al.*, 2009; Hasumi *et al.*, 2009; Hudon *et al.*, 2010). While we did observe aberrant mTOR signaling in *Bhd*^{-/-} cells, this was surprisingly unrelated to the apoptotic defects.

We turned our attention to the TGF β pathway, given its known role in regulating *Bim* transcription and tumor suppression. For instance, in humans, *Smad4* germline mutations are associated with juvenile

polyposis syndrome, which predisposes individuals to gastrointestinal cancers, and *TGF β RI*, *TGF β RII* and *activin receptor II* are mutated or downregulated in a broad spectrum of sporadic cancers (Levy and Hill, 2006). Epigenetic silencing of genes involved in TGF β signaling has also been specifically observed in RCCs (McRonald *et al.*, 2009). Genetic ablation of these signaling components in mice have confirmed the tumor suppressive function of this pathway in models of colon, mammary, and pancreatic cancer (Massague, 2008). Our data demonstrate that BHD loss results in yolk sac and

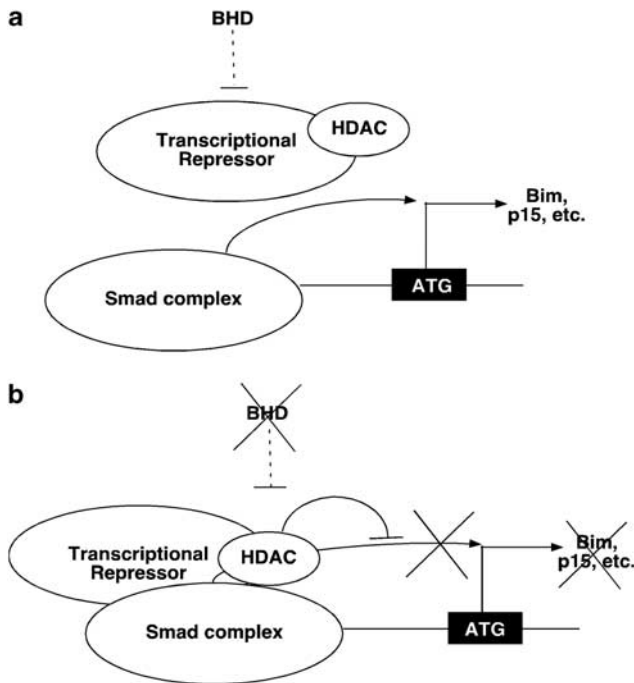


Figure 7 Model: Loss of BHD leads to increased HDAC-dependent repression of Smad-specific promoters. (a) In the presence of BHD, Smad-dependent transcription is active, allowing transcription of pro-apoptotic genes like *Bim* and anti-proliferative genes like *p15* to promote tumor suppression. (b) However, when BHD is lost, Smad-dependent transcription is repressed and apoptotic and anti-proliferative responses are lost, leading to tumor formation.

hematopoietic defects reminiscent of TGFβ receptor and ligand mutants. These phenotypes correlate with global aberrations in TGFβ-mediated transcription, which appear to occur downstream of nuclear accumulation of phosphorylated Smads. Our data indicate this defect is most likely due to HDAC-mediated effects on chromatin at Smad-dependent promoters, given the ability of TSA to reverse the biological and molecular defects of *Bhd*^{-/-} cells (Figure 7). Further work will be needed to elucidate the nature of these effects.

Previous studies have suggested a contradictory role for mTOR in BHD-related tumorigenesis, showing that BHD loss can result in either upregulation or down-regulation of mTOR signaling. These studies have been largely correlative and have not demonstrated a causative relationship between the signaling aberrations and biological outcomes (Baba *et al.*, 2006; Hartman *et al.*, 2009; Hasumi *et al.*, 2009; Hudon *et al.*, 2010). While we observed increased mTOR signaling in *Bhd*^{-/-} ES cells, as reported for late-stage kidney tumors taken from a BHD mouse model, this phenomenon appeared unrelated to the observed apoptotic resistance. We speculate that effects of BHD loss on mTOR signaling likely result from compensation and/or crosstalk with more proximal defects in TGFβ signaling.

Collectively, the data presented here establish a novel model whereby BHD exerts its major tumor suppressive effect through TGFβ-mediated transcriptional output to

regulate cell-intrinsic apoptosis. A recent publication has further supported this model, suggesting TGFβ-dependent transcriptional defects in a human BHD-derived tumor cell line (Hong *et al.*, 2010). This study demonstrated that BHD deficiency results in decreased levels of TGFβ pathway signaling components themselves including the ligands TGFβ-2 and Inhibin β A and Smad3. This contrasts with our data that demonstrate an effect of BHD downstream of signaling, on canonical Smad target gene promoters. Further, the report by Hong *et al.* suggested that TGFβ-2 stimulates cell growth and survival in the absence of BHD, while the TGFβ ligand Activin does the opposite. The latter is surprising in light of our findings, given the failure of BHD null ES cells to mount any transcriptional response to Activin treatment. This discrepancy likely derives from differential cell type, dosage and/or length of treatment, which has been well documented for TGFβ ligands (Massague, 2008). In any case, these molecular defects likely extend to other tumor-promoting effects such as cell-type specific proliferative advantages, due to p15 loss, and stromal alterations in BHD-related tumors *in vivo* (Massague, 2008). Although our *in vitro* study did not uncover a role for BHD in proliferation, this is likely due to the unusual cell cycle machinery of ES cells, which is refractory to cyclin-dependent kinase inhibitors (Savatier *et al.*, 1996; Faast *et al.*, 2004). It is probable that BHD loss and consequent defective TGFβ-mediated signaling *in vivo* results in a proliferative advantage. Collectively, these findings suggest several targeted therapeutics for the treatment of BHD-related tumors, as well as sporadic cases of cystic lung disease and RCC, including BH3-only mimetics, autophagy inhibitors, and HDAC inhibitors, which are all currently in clinical use for the treatment of other types of cancer.

Materials and methods

Sub-G₁ quantification

Cells were fixed and stained as previously described, then fluorescence was measured by flow cytometry (Riccardi and Nicoletti, 2006). At least 10 000 cells were counted for each experiment and data were analyzed using FlowJo software.

X-Gal staining of tissues and embryos

X-gal staining was carried out as previously described for both cells and whole mount tissues fixed in 4% paraformaldehyde according to the Sanger Institute gene-trap protocol. X-gal stained yolk sac sections came from tissue snap-frozen in OCT.

Alkaline phosphatase staining

Cells were stained using the Alkaline Phosphatase Detection Kit from Millipore (Bellerica, MA, USA) according to the manufacturer's instructions.

RNA isolation and qRT

RNA was extracted using Qiagen (Valencia, CA, USA) RNeasy columns with DNAase treatment and cDNA was written off ~1 μg of RNA per reaction using the ABI (Foster

City, CA, USA) one-step RT-PCR Master Mix. qRT-PCR reactions were run in triplicate on an ABI 7900HT machine for each experiment, utilizing either SYBR green or Taqman primer probe sets as noted in Supplementary Materials. All target mRNA levels were normalized to β -actin or *18s* expression levels. All qRT-PCR data reflect average mRNA levels from three independent RNA extractions and reverse transcription reactions with error bars showing standard error of the mean.

Protein extraction and western blotting

Whole-cell lysates were prepared using a buffer containing 120 mM NaCl, 1 mM EDTA, 10 mM pyrophosphate, 10 mM glycerophosphate, 50 mM NaF, 1% Triton, and freshly added protease inhibitor cocktail tablets from Roche (Basel, Switzerland) and 2 mM orthovanadate. Nuclear extracts were prepared using the Active Motif Nuclear Extraction kit, with 500 mM NaCl added after nuclease digestion followed by sonication of nuclear extracts. Lysates were run on polyacrylamide gels, transferred to nitrocellulose, blotted using standard protocols, and visualized on film using HRP-conjugated secondary antibodies and ECL reagent.

Cell size measurements

Cell size was measured using a Z2 Coulter particle size analyzer. At least 10 000 cells were measured for each experiment.

Characterization of gene-trapped ES cells and mice

Bhd^{+/*m*} ES cells were obtained from Bay Genomics and *Bhd*^{+/*-*} ES cells were obtained from the German Gene Trap Consortium, with isogenic parental *Bhd*^{+/*+*} lines being obtained from each respective source. Gene-trap insertions were mapped by first PCR amplifying the boundary of the fusion transcript using primers within the 3' end of the trap cassette and within the sequence tag, on cDNA generated from heterozygous ES cells. The precise genomic location of the cassette was mapped by direct PCR amplification of the respective intronic-trap cassette boundary using primers in the exon just upstream from the insertion and within the 5' end of the trap cassette. Southern blots were carried out on either *Pvu*II or *Bam*HI-digested DNA on *Bhd*^{+/*m*} or *Bhd*^{+/*-*} ES cell, respectively, with 5' end-directed probes being generated by PCR amplification from genomic DNA and subsequent TOPO cloning. *Bhd*^{+/*m*} mice were generated and intercrossed and embryo collection was performed as previously described (Covello *et al.*, 2006; Hartman *et al.*, 2009).

Proliferation assay

In all, 5×10^4 cells were plated in a gelatinized 12-well plate and counted using the Countess automatic counter from Invitrogen (Carlsbad, CA, USA) each day after plating.

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Immunohistochemistry

Paraffin-embedded tissue was immunostained using a rabbit polyclonal anti-Bim antibody from Novus following antigen retrieval in sodium citrate solution and developed using diaminobenzidine.

Embryoid bodies

For suspension cultures, 8×10^6 ES cells were plated in 10 cm bacterial dishes in DMEM containing 10% FBS and β -mercaptoethanol. Media was changed every other day, with EBs being split 1:2 on day 3. Methylcellulose cultures and benzidine staining were performed as previously described (Cooper *et al.*, 1974; Adelman *et al.*, 1999).

Northern blotting

Northern blots for miRNAs were done as previously described (Gruber *et al.*, 2009).

Chromatin immunoprecipitation

In all, 1×10^6 ES cells were fixed for 10 min in 1% formaldehyde for 10 min at room temperature. Crosslinking was quenched with 125 mM glycine for 5 min, then cells were lysed and sonicated to shear DNA for 3 min with on/off pulses using a Fisher Sonic Dismembrator Model 500 at 20% power. Chromatin immunoprecipitation was carried using the Imgenex QuikChIP kit according to the manufacturer's instructions using a pan acetyl-H3 antibody from Millipore. Eluted protein/DNA complexes were purified by phenol/chloroform extraction and ethanol precipitation.

Statistics

All *P*-values were generated using a two-tailed Student's *t*-test.

Conflict of interest

The authors declare no conflict of interest.

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