Contents lists available at ScienceDirect



Journal of Clinical & Translational Endocrinology

journal homepage: www.elsevier.com/locate/jcte

Original research

Prevalence of hypovitaminosis D, and its association with hypoadiponectinemia and hyperfollistatinemia, in Saudi women with naïve polycystic ovary syndrome



Osama Adnan Kensara

Department of Clinical Nutrition, Faculty of Applied Medical Sciences, Umm Al-Qura University, PO Box 7607, Holy Makkah, Saudi Arabia

ARTICLE INFO	A B S T R A C T
<i>Keywords:</i> Polycystic ovary syndrome Saudi women Serum 25-hydroxyvitamin D Adiponectin Follistatin	Aims: The association between vitamin D and polycystic ovary syndrome (PCOS) is an active area of growing research. However, data in Saudi Arabia are scarce. This study aimed to define serum 25-hydroxyvitamin D (25(OH)D) levels among Saudi women with naïve PCOS, and to investigate the associations of their 25(OH)D status with their serum adiponectin and follistatin levels, along with indices of insulin resistance and hormonal deteriorations. <i>Methods:</i> In this case-control observational study, 63 women with PCOS and 65 age-and body mass index (BMI)-matched control women were assessed. PCOS was diagnosed based on the revised criteria of Rotterdam. Fasting serum levels of 25(OH)D, adiponectin, follistatin, insulin, follicle-stimulating hormone (FSH), luteinizing hormone (LH), total testosterone (TT), androgen (Δ_4 -androstenedione), estradiol, progesterone, along with fasting plasma glucose (FPG), homeostasis model assessment-insulin resistance (HOMA-IR) index and lipid profile were measured in both groups. <i>Results:</i> The prevalence of hypovitaminosis D (serum 25(OH)D < 30 ng/ml) was higher in PCOS group than control group (77.8% vs. 12.3%). Serum adiponectin and FSH concentrations were significantly lower, while serum follistatin, LH, TT, Δ_4 -androstenedione and insulin levels, as well as FPG and HOMA-IR, FPG, LH, testosterone, and Δ_4 -androstenedione levels. <i>Conclusion:</i> Hypovitaminosis D, coexisted and correlated with hypoadiponectinemia and hyperfollistatinemia, is being an alarming risk factor in Saudi women with PCOS. Further investigational and explanatory studies in large size samples are warranted to realize these findings and to improve both diagnostic and treatment tools in Saudi women with PCOS.

Introduction

Polycystic ovary syndrome (PCOS) is a multifactorial, heterogeneous, endocrine-metabolic disorder that commonly affects women at their reproductive age. PCOS is a complex syndrome characterized by chronic oligo-or anovulation, menstrual irregularities, hyperandrogenism, infertility, and polycystic ovarian morphologic features [1–3]. Excess luteinizing hormone (LH) and low follicle stimulatinghormone (FSH) are also common, and approximately 60%–80% of all PCOS cases are more vulnerable to develop insulin resistance (IR) and compensatory hyperinsulinemia, which exacerbates ovarian androgen production and ovulation dysfunction in PCOS patients [1–3].

During the last decade, vitamin D has gained and sustained immense interest in the fields of biomedical and human health research [4,5] including those related to female reproductive-metabolic disorders [6]. At that respect, while the extent of this association remains inconsistent and requires further investigations, there is an increasing evidence that hyovitaminosis D (vitamin D insufficiency/deficiency) is associated with an increased prevalence of PCOS [7–9]. It has been suggested that vitamin D depletion plays a potential role in increasing IR and metabolic abnormalities and in disrupting ovarian folliculogenesis and hormonal secretion in PCOS patients [7–12], and it is therefore essential to screen vitamin D status among all the PCOS patients [13]. Furthermore, some recent interventional trials demonstrated that appropriate supplementation of vitamin D in vitamin Ddeficient-PCOS women had resulted in enhancing their insulin sensitivity and improving their deteriorated sex-hormones and metabolic profiles, re-enforcing the importance of vitamin D status in the etiology

https://doi.org/10.1016/j.jcte.2018.04.001

2214-6237/ © 2018 The Author. Published by Elsevier Inc. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/BY-NC-ND/4.0/).

E-mail address: o.kensara@gmail.com.

Received 30 January 2018; Received in revised form 12 April 2018; Accepted 13 April 2018

and management of PCOS [14-17].

In accordance with vitamin D, there is also a durable hypothesis that the dysregulated circulating levels of adipocytokines; the pleiotropic bioactive products secreted from the adipose tissue and other body cells, are involved in the development of PCOS [18,19]. Among these adipokines, due to its optimistic insulin-sensitizing, anti-inflammatory, and vasculoprotective properties, adiponectin has attracted much research interest among these adipokines in both PCOS [19] and diabetic research [20]. At that respect, low circulating adiponectin concentration has been reported in both lean and obese women with PCOS [21–23], and suggested as a predictor for PCOS patients to develop type II diabetes [24]. Oppositely, PCOS is usually linked with aberrant increases in circulating follistatin; another adipocytokine with a distinctive infertility role among female population through inhibiting FSH production and ovarian folliculogenesis [25,26].

Notably, studies related to vitamin D and PCOS are scanty in Saudi Arabia. In addition, less is known whether there is any relationship between vitamin D, adiponectin, and follistatin among Saudi population. Hence, the present study was designed to measure the serum levels of 25(OH)D in Saudi women with untreated PCOS and compare their values with those of age-and BMI matched non-PCOS controls and also; to elucidate is there any association of 25(OH)D levels of PCOS patients with their serum adiponectin or follistatin concentrations, and/or with their indices of metabolic and hormonal deteriorations.

Subjects and methods

Subjects

Sixty-three Saudi women with naïve PCOS (case group; mean age: 31.6 \pm 6.4 years, and mean body mass index (BMI): 22.2 \pm 2.6 kg/ m²), along with 65 age- and BMI matched non-PCOS control Saudi women with regular menstrual cycles (control group), who met the inclusion and exclusion criteria listed below, were enrolled and evaluated in this case-control observational study. All participants were outpatients referred to the Gynecology and Endocrinology Clinic of Obstetrics & Gynecology Department, Heraa' General Hospital, The Western Region of Saudi Arabia. The enrollment process of study subjects was carried out at both the local media and the hospital outpatient's clinical units by using IRB-approved advertisement materials and notifications, including both electronic and paper advertisements. The advertisement called for young Saudi women with history of irregular menses, infertility, ovary cysts, excess hair, etc., and for healthy volunteers Saudi women who had regular ovulatory cycles and came to the clinic for annual check-up or their partners had male fertility problems. Our plan was to enroll a 1:1 ratio between eligible PCOS and healthy control women matching by age and BMI. Based on available worldwide clinical data examined the prevalence of hypovitaminosis D in women with PCOS [7–11], sample size of 50 women per group was determined regard 0.05 and 0.2 of α and β error, respectively, and a power of 80%. Herein, 63 women in case group and 65 women in control group were enrolled considering any dropouts from study participation. Diagnosis of PCOS was made as per the revised Rotterdam PCOS consensus criteria [27], which referred as the presence of at least two of the following three standard criteria: chronic oligo/amenorrhea and/or chronic anovulation (i.e., having menstruation at longer than 35 day intervals, or \leq 9 cycles/year), clinical and/or biochemical signs of hyperandrogenism (i.e., hirsutism and/or excess serum androgens levels), and polycystic ovaries morphological features detected by ultrasound (i.e., having ovarian volume of over 10 cm³ and/or presence of more than 12 ovarian follicles of 2-9 mm in length) [27]. Exclusion criteria of the case group included women with known causes of anovulation, infertility, and hyperandrogenemia; including congenital adrenal hyperplasia, androgen-secreting tumors, Cushing's syndrome, and hyperprolactinemia, or any disease that could possibly affect reproductive physiology. Women with diabetes, thyroid dysfunction,

kidney or liver disease, or pregnancy were also excluded. All enrolled eligible PCOS and control women were not smokers or alcohol or drug abusers, and none of them had received any vitamin D supplement, contraceptives or any other drugs or hormonal treatment known to affect vitamin D metabolism or interfere with metabolic variables and hypothalamic–pituitary–gonadal axis for at least 3 months prior to the study. In addition, according to their ultrasound findings, none of the healthy women in the control group had evidence of polycystic ovaries. Institutional Ethical approval and written informed consent of all participants were obtained before the initiation of the study. The study was conducted according to the principals of the Declaration of Helsinki.

Blood sampling and laboratory parameters measurement

After 12 h fasting, two peripheral blood samples were obtained from each participant. Blood samples of menstruating women were taken during the mid-follicular phase of their menstrual cycle, while those of women with amenorrhea were taken randomly. The first blood sample was collected into a tube contained EDTA-anticoagulant and immediately used to estimate fasting plasma glucose (FPG) level, while the second one was collected into a plain tube without anticoagulant, centrifuged, and aliquots of its corresponding serum were immediately stored at -80 °C until analyzed for serum levels of 25(OH)D, adiponectin, follistatin, insulin, FSH, LH, total testosterone, androgen (Δ_4 androstenedione), estradiol, progesterone, and lipid profile parameters. In addition, insulin resistance (IR) was determined using the homeostasis model assessment of insulin resistance (HOMA-IR) index calculated as FPG (in mg/dL) multiplied by fasting insulin (in µU/mL) divided by 405, and a HOMA-IR value of 2.5 or above was considered as insulin resistant [28].

In the present study, serum levels of 25(OH)D; which is the most widely accepted biomarker for monitoring the overall vitamin D status [29,30], were quantitatively measured by commercially available enzyme-linked immunosorbent assay (ELISA) kit (IDS, Boldon, UK), following the manufacturer's instructions. Based on the The Endocrine Society Clinical Practice Guidelines, interpretation of vitamin D status among the participants was as follow: 25(OH)D levels of 21-29 ng/mL and $\leq 20 \text{ ng/mL}$ were considered as cases of vitamin D insufficiency and deficiency, respectively, while 25(OH)D of more than 30 ng/mL was regarded as a normal level [29,30]. Commercially available ELISA Kits were also used to determine the serum levels of adiponectin (Mediagnost, Uppsala, Germany), insulin (Monobind, Uppsala, US), and follistatin (R&D Systems, Inc., MN, USA) and following their corresponding manufacturers' recommendations. As reported by the manufactures, the intra- and inter-assay coefficients of variation (CVs) for serum 25(OH)D, adiponectin, follistatin and insulin were 5.4, 1.5%, 2.3%, 5.5% and 5.5, 2%, 8%, 5.8%, respectively, while the sensitivity of the assays for 25(OH)D, adiponectin, follistatin and insulin were 2.6 ng/mL, 0.6 ng/mL, 89 pg/mL and 2 μ U/mL, respectively. All ELISA assays of 25(OH)D, adiponectin, and follistatin were performed in duplicate on a fully automated ELISA system (DYNEX, Technologies MRX II, VA, USA). The remaining biochemical and hormonal assays, which routinely carried out at Gynecology and Endocrinology Clinic Lab, were performed according to their standard protocols and procedures as described elsewhere.

Statistical analysis

Data are expressed as mean \pm standard deviation (SD) for quantitative variables, while those for qualitative variables are presented as percentages or numbers. Analyses were performed using the statistical software version 17.0 for windows (SPSS Inc., Chicago, IL, USA). Comparison of mean variables between the two groups was estimated by Student's *t*-test, and χ^2 test was used for frequency analysis. The associations between serum 25(OH)D levels and other measured variables in PCOS group were performed by the Pearson's correlation test. A

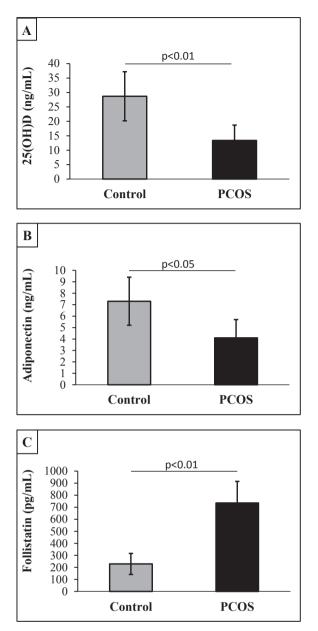


Fig. 1. Baseline levels of fasting serum 25-hydroxy-vitamin D (25(OH)D), adiponectin, and follistatin of study's participants. Sixty-three Saudi women with naïve polycystic ovary syndrome (PCOS group) along with 65 age- and body mass index (BMI) matched non-PCOS Saudi women (Control group), were enrolled and studied. After 12 h fasting, peripheral blood samples were obtained, and aliquots of their corresponding serum were used to measure the fasting serum levels of 25(OH)D, adiponectin and follistatin. Data are presented as mean ± SD. *P* < 0.05 and *P* < 0.01 are significant and highly significant differences *vs* normal control group.

p-value < 0.05 was considered as statistically significant.

Results

Following inclusion and exclusion criteria, 128 Saudi women, of whom 63 had PCOS (case group) and 65 non-PCOS women served as control group, were assessed in this study. As shown in Fig. 1A, PCOS group had significantly lower serum level of 25(OH)D compared with control group (13.4 ± 5.3 ng/mL vs 28.7 ± 8.5 ng/mL; p < 0.01). Similarly, serum concentration of adiponectin was significantly lower (p < 0.05) in women with PCOS compared with control women (Fig. 1B). However, serum follistatin level was significantly higher

Table 1

Distribution of Vitamin D status among study's participants. Sixty-three Saudi women with naïve polycystic ovary syndrome (PCOS group), along with 65 age and body mass index (BMI) matched non-PCOS Saudi women (Control group), were enrolled and studied. After 12 h fasting, peripheral blood samples were obtained, and aliquots of their corresponding serum were used to measure the fasting serum levels of 25(OH)D. Based on the measured value of serum 25(OH) D, interpretations of vitamin D status among all subjects of both groups were as follow: 25(OH)D levels of 21–29 ng/mL and \leq 20 ng/mL were considered as cases of vitamin D insufficiency and deficiency, respectively, while 25(OH)D of more than 30 ng/mL was regarded as a normal level.

Vitamin D status (serum 25(OH)D ng/mL)	Control subjects $(n = 65)$		PCOS patients $(n = 63)$		P-value
	No. of cases	%	No. of cases	%	
Adequate (> 30)	57	87.7	14	22.2	< 0.01
Insufficient (20-29)	6	9.2	16	25.4	< 0.01
Deficient (< 20)	2	3.1	33	52.4	< 0.001

(p < 0.01) in PCOS patients compared with non-PCOS controls (Fig. 1C). Women of both groups were sub-categorized based on their vitamin D status (i.e., their serum 25(OH)D levels), and the prevalence of hypovitaminosis D (< 30 ng/mL) was significantly (p < 0.01) higher in Saudi women with PCOS than in non-PCOS controls. As shown in Table 1, of the 63 eligible women with PCOS, only 14 cases (22.2%) had a sufficient level of vitamin D, while 49 cases (77.8%) had hypovitaminosis D distributed as follow: 6 cases (25.4%) had vitamin D insufficiency and 33 cases (52.4%) had vitamin deficiency. In contrary, among the eligible 65 non-PCOS controls, 57 women (87.7%) had adequate levels of vitamin D and only 8 women (12.3%) showed a status of hypovitaminosis D (6 women (9.2%) with vitamin D insufficiency and 2 women (3.1%) with vitamin D deficiency) (Table 1).

The characteristic metabolic and hormonal features of the studied PCOS and control groups are summarized in Table 2. The results showed that the levels of serum insulin, HOMA-IR and blood glucose, along with the serum levels of LH, testosterone and androgen (Δ_4 -androstenedione) hormones were significantly higher, while serum FSH concentrations were significantly lower, in the PCOS group (p < 0.05) compared with non-PCOS control group. Indices of dyslipidemia were higher in women of PCOS group than in the control women; however, this difference was not statistically significant (p > 0.05). Also, there were no significant differences regarding systolic and diastolic blood pressure, and the serum estradiol and progesterone levels between PCOS and control groups (all p not significant) (Table 2).

As demonstrated in Table 3, significant (p < 0.05) positive correlations were observed between the serum 25(OH)D levels of Saudi women with PCOS and their serum adiponectin and FSH levels. By contrary, serum 25(OH)D levels of these PCOS patients were negatively (p < 0.05) correlated with the serum levels of follistatin, LH, testosterone, Δ_4 -androstenedione, and insulin, as well as with FPG and HOMA-IR levels (Table 3). There were no significant correlations between serum 25(OH)D levels and other measured variables in PCOS group (Table 3).

Discussion

To date, the exact etiology and underlying pathogenic mechanisms of PCOS remain largely elusive. To this end, although its conclusion is still inconsistent and representing an active area of energetic research [31], impaired vitamin D status has been emphasized as an important risk factor in the development of PCOS and increased its metabolic and hormonal abnormalities [6–9]. Nevertheless, whether hypovitaminosis D has a causal association with PCOS among Saudi patients is not known. Data of the present study demonstrated that, besides indices of metabolic and hormonal deteriorations such as insulin resistance (IR), hyperinsulinemia, low circulating FSH, but excess LH testosterone, and

Table 2

Clinical, metabolic and hormonal characteristics of study's participants. Sixtythree Saudi women with naïve polycystic ovary syndrome (PCOS group), along with 65 age- and body mass index (BMI) matched non-PCOS Saudi women (Control group), were enrolled and studied. After 12 h fasting, two peripheral blood samples were obtained from each participant: the 1st sample was collected into a tube contained EDTA-anticoagulant and immediately used to estimate fasting plasma glucose (FPG) level, while the 2nd sample was collected into a plain tube without anticoagulant, centrifuged, and aliquots of its corresponding serum were used to assess the fasting serum levels of insulin, total testosterone, Δ_4 -androstenedione (Δ_4 -A), follicle-stimulating hormone (FSH), luteinizing hormone (LH), estradiol, progesterone, and lipid profile parameters (TC, total cholesterol; LDL-C, low density lipoprotein-cholesterol; TG, triglyceride; VLDL-C, very low density lipoprotein-cholesterol; HDL-C, high density lipoprotein-cholesterol). In addition, insulin resistance (HOMA-IR) index.

Variable	Control group $(n = 65)$	PCOS group $(n = 63)$	P-value
Age (years)	30.4 ± 5.2	31.6 ± 6.4	NS
BMI (kg/m ²)	21.4 ± 3.1	22.2 ± 2.6	NS
FPG (mg/dL)	83.5 ± 6.4	91.3 ± 7.5	< 0.05
Insulin (µU/mL)	8.2 ± 2.2	13.1 ± 3.6	< 0.05
HOMA-IR (n)	1.7 ± 0.7	2.9 ± 1.4	< 0.05
Total Testosterone (ng/dl)	46.5 ± 19.3	71.7 ± 23.2	< 0.05
Δ_4 -A (ng/mL)	1.1 ± 0.4	2.6 ± 0.8	< 0.05
FSH (mIU/mL)	8.7 ± 3.4	4.9 ± 1.2	< 0.05
LH (mIU/mL)	4.4 ± 1.6	8.8 ± 3.4	< 0.05
Estradiol (pg/mL)	45.5 ± 14.4	39.2 ± 11.2	NS
Progesterone (ng/mL)	3.1 ± 1.2	2.8 ± 0.9	NS
TC (mg/dL)	161.6 ± 24.5	176.4 ± 27.3	NS
LDL-C (mg/dL)	76.9 ± 11.7	84.1 ± 13.2	NS
TG (mg/dL)	115.3 ± 28.4	121.3 ± 31.2	NS
VLDL (mg/dL)	23.2 ± 5.7	24.3 ± 6.3	NS
HDL-C (mg/dL)	47.6 ± 8.5	44.7 ± 8.1	NS
Systolic blood pressure (mmHg)	118.5 ± 3.5	119.0 ± 9.0	NS
Diastolic blood pressure (mmHg)	$77.0~\pm~5.5$	79.5 ± 8.0	NS

Data are presented as mean \pm SD. Values are significant at P < 0.05; NS, not significant.

Table 3

Relations of serum 25(OH)D concentrations with serum adiponectin and follistatin levels, and with metabolic and hormonal indexes of PCOS group.

25(OH)D

Parameter	r	p-value
Adiponectin	0.328	< 0.05
Follistatin	-0.364	< 0.05
Insulin	-0.273	< 0.05
FPG	-0.288	< 0.05
HOMA-IR	-0.295	< 0.05
TC	-0.123	NS
LDL-C	-0.082	NS
TG	-0.102	NS
VLDL-C	-0.055	NS
HDL-C	0.103	NS
Total Testosterone	-0.283	< 0.05
Δ_4 -A	-0.277	< 0.05
FSH	0.292	< 0.05
LH	-0.312	< 0.05
Estradiol	0.086	NS
Progesterone	0.077	NS

Values are significant at P < 0.05; NS, not significant.

Abbreviations: FPG, fasting plasma glucose; HOMA-IR, homeostasis model assessment index of insulin resistance; TC, total cholesterol; LDL-C, low density lipoprotein-cholesterol; TG, triglyceride; VLDL-C, very low density lipoproteincholesterol; HDL-C, high density lipoprotein-cholesterol; Δ_4 -A, Δ_4 -androstenedione; FSH, follicle-stimulating hormone; LH, luteinizing hormone. androgen levels [1–3], Saudi women with PCOS had evidence of significant hypovitaminosis D status as indicated by their remarkably decreased serum 25(OH)D levels as compared to healthy controls. Most importantly, serum 25(OH)D levels of these PCOS patients were correlated inversely with IR indices (HOMA-IR and FPG levels) and serum levels of LH, testosterone and androgen, and positively with FSH levels. These findings may coherently support the sustained assumption that hypovitaminosis D is an alarming contributor in prevalence of PCOS and its metabolic and hormonal complications [10–13], and support the raised necessity to monitor PCOS patients for their vitamin D status [13].

Indeed, the mechanisms behind the possible association between vitamin D and pathophysiology of PCOS are not vet covered well: nevertheless, the regulating effects of vitamin D on insulin signaling and sensitivity and expression of its receptors in both classical and nonclassical insulin-sensitive tissues, including ovaries [10-13], and vitamin D is a predictor of IR in PCOS [7], should not be neglected. In addition to that, vitamin D receptors (VDR) are existed in the ovarian, fallopian and endometrial cells, and vitamin D may be involved in ovarian physiology and reserve through regulating the activities of genes involved in follicular development, steroidogenesis and androgen production [6], [32]. Low circulating 25(OH)D [6,32], as well as IR and its compensatory hyperinsulinemia [33,34], may lead to overproduction of both LH and androgen with consequent aggravation of ovulatory dysfunction in PCOS. Taken together, the aforementioned observations and suggestions can in turn explain, at least in part, the correlations that were reported here between the serum 25(OH)D levels of PCOS patients and their IR indices, serum LH, testosterone and FSH levels. In support, optimized regimens of vitamin D-based therapy in vitamin D-deficient-PCOS women have lately shown to improve their insulin sensitivity and normalize their elevated circulating androgen and other deregulated hormones [14-17].

Disturbed secretions of adipokines have also been accentuated in the pathophysiology of PCOS and its metabolic and reproductive dysfunctions [18,19]. As demonstrated here, Saudi women with PCOS showed lower adiponectin, but higher follistatin, concentrations in their serum compared to control group, and their 25(OH)D levels were correlated positively with adiponectin and negatively with follistatin levels. In constancy with these findings, aberrant hyperfollistatinemia is common in PCOS, in despite of their BMI, and may be involved in induction of ovarian testosterone production and in arresting FSH-ovarian granulosa cell axis and ovarian folliculogenesis [25,26,35,36]. Furthermore, could vitamin D directly reduce the synthesis of follistatin has been previously documented by Woeckel et al. [37] on human osteoblast cells; implying the explanation of the negative association between serum levels of 25(OH)D and follistatin that were observed here in PCOS patients. Concerning the current findings related to the lower adiponectin levels in PCOS group, other researchers have also reported the same findings in both obese and non-obese PCOS patients [22-24], including data of a meta-analysis of more than 3500 females of various ages [21]. In addition, circulating adiponectin level was found to be more reduced at lower levels of serum 25(OH)D in women with PCOS [38], and vitamin D supplementation improved serum adiponectin levels in women with PCOS [17]. However, other researchers found no significant difference in the circulating adiponectin concentrations between PCOS and non-PCOS groups [34], and such discrepancy could be related to the ethnic and demographic variations among the studied populations [34]. Toward this latter suggestion, positive correlations of systemic vitamin D status with circulating adiponectin, and hypoadiponectinemia is a link between hypovitaminosis D and IR, have been detected among diabetic Saudi patients [39] and also among African American patients with cardiovascular disease [40].

Strengths and limitations

To the best of our knowledge, this study is the first to investigate the

possible association/correlation between hypovitaminosis D to the pathology of PCOS among Saudi women- and highlighted the significant prevalence of vitamin D deficiency and low adiponectin and follistatin levels in their serum. It also suggests the necessity of screening Saudi women with PCOS for their serum 25(OH)D levels. In despite of this, the present study is an observational one with included small sample size and therefore, causation cannot be claimed. Coherently, correction of vitamin D level via a next step clinical trial with vitamin D supplement therapy in Saudi women with vitamin D-deficiency- and PCOS could provide more knowledge and explore a causal relationship of impaired vitamin D status in etio-pathogenesis of PCOS among Saudi women. It has also been reported that vitamin D binding protein (VDBP) levels decrease in PCOS, which may partly account for the reduced 25(OH)D levels and therefore estimation of VDBP with 25(OH)D in a further future study would be more informative. In addition, BMI seems to be an important variable in PCOS, hence the relationship of vitamin D deficiency with increased IR, infertility and hyperandrogenism have been reported to be more intense in obese-PCOS than lean-PCOS patients. However, this variable was not covered here since the BMI of PCOS and control groups were similar and it, in turn, requires further studies with large size samples and variable BMI to be clarified.

Conclusions

Overall, data of the current study indicate, for the first time, that hypovitaminosis D was highly prevalent among the investigated women with naïve PCOS, and significantly coexisted and correlated with hyperadiponectinemia, hyperfollistatinemia, IR and altered sex-hormonal parameters of these PCOS patients. However, in view of using convenience sampling, the present observations cannot be generalized to Saudi population and further studies in large size samples and at different Saudi locations are warranted to realize these findings and to improve the diagnostic and treatment strategies of Saudi women with PCOS.

Conflict of interest

The author declares that there is no conflict of interests.

Funding

This study was funded by King Abdul Aziz City for Science and Technology (KACST) the Kingdom of Saudi Arabia, Award Number (AT-34-86).

Acknowledgments

The author would like to thank Dr. El-Shemi AG, Dr. Refaat B and Dr. Azzeh F (Depts. of Laboratory Medicine and Clinical Nutrition, Faculty of Applied Medical Sciences, Umm Al-Qura University), and Dr. Yasser G (Dept. of Gynecology & Obstetrics, Batterjee Medical College, KSA) for their active support during the study.

References

- Bachelot A. Polycystic ovarian syndrome: clinical and biological diagnosis. Ann Biol Clin (Paris) 2016;74:661–7.
- [2] McCartney CR, Marshall JC. Clinical practice. Polycystic ovary syndrome. N Engl J Med 2016;375:54–64.
- [3] Macut D, Bjekić-Macut J, Rahelić D, Doknić M. Insulin and the polycystic ovary syndrome. Diabetes Res Clin Pract 2017;130:163–70.
- [4] Papadimitriou DT. The big vitamin D mistake. J Prev Med Public Health 2017;50(4):278–81.
- [5] Kimball SM, Emery JCH, Lewanczuk RZ. Effect of a vitamin and mineral supplementation on glycemic status: results from a community-based program. J Clin Transl Endocrinol 2017;10:28–35.
- [6] Shahrokhi SZ, Ghaffari F, Kazerouni F. Role of vitamin D in female reproduction.

Clin Chim Acta 2016;455:33-8.

- [7] Patra SK, Nasrat H, Goswami B, Jain A. Vitamin D as a predictor of insulin resistance in polycystic ovarian syndrome. Diabetes Metab Syndr 2012;6:146–9.
- [8] He C, Lin Z, Robb SW, Ezeamama AE. Serum vitamin D levels and polycystic ovary syndrome: a systematic review and meta-analysis. Nutrients 2015;7:4555–77.
- [9] Bacopoulou F, Kolias E, Efthymiou V, Antonopoulos CN, Charmandari E. Vitamin D predictors in polycystic ovary syndrome: a meta-analysis. Eur J Clin Invest 2017;47(10):746–55.
- [10] Wehr E, Pilz S, Schweighofer N, Giuliani A, Kopera D, Pieber TR, et al. Association of hypovitaminosis D with metabolic disturbances in polycystic ovary syndrome. Eur J Endocrinol 2009;161:575–8.
- [11] Li HW, Brereton RE, Anderson RA, Wallace AM, Ho CK. Vitamin D deficiency is common and associated with metabolic risk factors in patients with polycystic ovary syndrome. Metabolism 2011;60:1475–81.
- [12] Thomson RL, Spedding S, Buckley JD. Vitamin D in the aetiology and management of polycystic ovary syndrome. Clin Endocrinol 2012;77:343–50.
- [13] Kumar A, Barki S, Raghav V, Chaturvedi A, Kumar KVSH. Correlation of vitamin D with metabolic parameters in polycystic ovarian syndrome. J Family Med Prim Care 2017;6:115–9.
- [14] Jamilian M, Foroozanfard F, Rahmani E, Talebi M, Bahmani F, Asemi Z. Effect of two different doses of vitamin d supplementation on metabolic profiles of insulinresistant patients with polycystic ovary syndrome. Nutrients 2017;9(12). pii: E1280.
- [15] Foroozanfard F, Talebi M, Samimi M, Mehrabi S, Badehnoosh B, Jamilian M, et al. Effect of two different doses of vitamin D supplementation on metabolic profiles of insulin-resistant patients with polycystic ovary syndrome: a randomized, doubleblind, Placebo-controlled trial. Horm Metab Res 2017;49(8):612–7.
- [16] Karadağ C, Yoldemir T, Yavuz DG. Effects of vitamin D supplementation on insulin sensitivity and androgen levels in vitamin-D-deficient polycystic ovary syndrome patients. J Obstet Gynaecol Res 2017;44(2):270–7.
- [17] Seyyed Abootorabi M, Ayremlou P, Behroozi-Lak T, Nourisaeidlou S. The effect of vitamin D supplementation on insulin resistance, visceral fat and adiponectin in vitamin D deficient women with polycystic ovary syndrome: a randomized placebocontrolled trial. Gynecol Endocrinol 2017:1–6. doi: 10.1080/09513590.2017. 1418311. [Epub ahead of print].
- [18] Behboudi-Gandevani S, Ramezani Tehrani F, Bidhendi Yarandi R, Noroozzadeh M, Hedayati M, Azizi F. The association between polycystic ovary syndrome, obesity, and the serum concentration of adipokines. J Endocrinol Invest 2017;40(8):859–66.
- [19] Polak K, Czyzyk A, Simoncini T, Meczekalski B. New markers of insulin resistance in polycystic ovary syndrome. J Endocrinol Invest 2017;40:1–8.
- [20] Achari AE, Jain SK. Adiponectin, a therapeutic target for obesity, diabetes, and endothelial dysfunction. Int J Mol Sci 2017:18(6). pii: E1321.
- [21] Lee BC, Lee J. Cellular and molecular players in adipose tissue inflammation in the development of obesity-induced insulin resistance. Biochim Biophys Acta 2014;1842(3):446–62.
- [22] Sharifi F, Hajihosseini R, Mazloomi S, Amirmogaddami H, Nazem H. Decreased adiponectin levels in polycystic ovary syndrome, independent of body mass index. Metab Syndr Relat Disord 2010;8(1):47–52.
- [23] Yildiz Y, Ozaksit G, Serdar Unlu B, Ozgu E, Energin H, Kaba M, et al. Serum adiponectin level and clinical, metabolic, and hormonal markers in patients with polycystic ovary syndrome. Int J Fertil Steril 2014;7:331–6.
- [24] Sepilian V, Nagamani M. Adiponectin levels in women with polycystic ovary syndrome and severe insulin resistance. J Soc Gynecol Investig 2005;12(2):129–34.
- [25] Norman RJ, Milner CR, Groome NP, Robertson DM. Circulating follistatin concentrations are higher and activin concentrations are lower in polycystic ovarian syndrome. Hum Reprod 2001;16(4):668–72.
- [26] Chen MJ, Chen HF, Chen SU, Ho HN, Yang YS, Yang WS. The relationship between follistatin and chronic low-grade inflammation in women with polycystic ovary syndrome. Fertil 2009;92(6):2041–4.
- [27] Rotterdam ESHRE/ASRM-Sponsored PCOS consensus workshop group. Revised 2003 consensus on diagnostic criteria and long-term health risks related to polycystic ovary syndrome (PCOS). Hum Reprod 2004;19:41–7.
- [28] Keshavarz MA, Moradi S, Emami Z, Rohani F. Association between serum 25(OH) vitamin D and metabolic disturbances in polycystic ovary syndrome. Neth J Med 2017;75(5):190–5.
- [29] Holick MF, Binkley NC, Bischoff-Ferrari HA, Gordon CM, Hanley DA, Heaney RP, et al. Evaluation, treatment, and prevention of vitamin D deficiency: an Endocrine Society clinical practice guideline. J Clin Endocrinol Metab 2011;96(7):1911–30.
- [30] Smith TJ, Lanham-New SA, Hart KH. Vitamin D in adolescents: are current recommendations enough? J Steroid Biochem Mol Biol 2017;173:265–72.
- [31] Krul-Poel YH, Snackey C, Louwers Y, Lips P, Lambalk CB, Laven JS, et al. The role of vitamin D in metabolic disturbances in polycystic ovary syndrome: a systematic review. Eur J Endocrinol 2013;169(6):853–65.
- [32] Bakhshalizadeh S, Amidi F, Alleyassin A, Soleimani M, Shirazi R, Shabani Nashtaei M. Modulation of steroidogenesis by vitamin D3 in granulosa cells of the mouse model of polycystic ovarian syndrome. Syst Biol Reprod Med 2017;63(3):150–61.
- [33] Kauffman RP, Baker VM, DiMarino P, Castracane VD. Hyperinsulinemia and circulating dehydroepiandrosterone sulfate in white and Mexican American women with polycystic ovary syndrome. Fertil Steril 2006;85:1010–6.
- [34] Spritzer PM, Lecke SD, Satler F, Morsch DM. Adipose tissue dysfunction, adipokines, and low-grade chronic inflammation in polycystic ovary syndrome. Reproduction 2015;149(5):R219–27.
- [35] Eldar-Geva T, Spitz IM, Groome NP, Margalioth EJ, Homburg R. Follistatin and activin A serum concentrations in obese and non-obese patients with polycystic ovary syndrome. Hum Reprod 2001;16(12):2552–6.
- [36] Teede H, Ng S, Hedger M, Moran L. Follistatin and activins in polycystic ovary

syndrome: relationship to metabolic and hormonal markers. Metabolism 2013;62(10):1394–400.

- [37] Woeckel VJ, van der Eerden BC, Schreuders-Koedam M, Eijken M, Van Leeuwen JP. 1α,25-dihydroxyvitamin D3 stimulates activin A production to fine-tune osteoblastinduced mineralization. J Cell Physiol 2013;228:2167–74.
- [38] Mazloomi S, Sharifi F, Hajihosseini R, Kalantari S, Mazloomzadeh S. Association between hypoadiponectinemia and low serum concentrations of calcium and vitamin D in women with polycystic ovary syndrome. ISRN Endocrinol 2012;2012:949427.
- [39] Al-Daghri NM, Al-Attas OS, Alokail MS, Alkharfy KM, Al-Othman A, Draz HM, et al. Hypovitaminosis D associations with adverse metabolic parameters are accentuated in patients with Type 2 diabetes mellitus: a body mass index-independent role of adiponectin? J Endocrinol Invest 2013;36(1):1–6.
- [40] Khan RJ, Gebreab SY, Riestra P, Sims M, Gaye A, Xu R, et al. Associations between vitamin D and cardiovascular disease risk factors in African Americans are partly explained by circulating adipokines and C-Reactive Protein: the Jackson Heart Study. J Nutr 2016;146(12):2537–43.