

Supporting Information

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Macrophage-Disguised Manganese Dioxide Nanoparticles for Neuroprotection by Reducing Oxidative Stress and Modulating Inflammatory Microenvironment in Acute Ischemic Stroke

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Figure S1. UV-vis absorption spectra of KMnO₄ solution, MnO₂, FTY-loaded MnO₂,

and Ma@(MnO₂+FTY).



Figure S2. PXRD pattern of MnO₂ nanospheres.



Figure S3. EDS analysis of MnO_2 nanospheres.



Figure S4. SEM images of MnO₂ nanospheres.



Figure S5. Nitrogen sorption isotherms of MnO₂ nanospheres.



Figure S6. The pore distribution of MnO₂ nanospheres.



Figure S7. Optimal prescription screening for the ratio of cell membrane (quantified by protein content) to MnO_2 +FTY by monitoring the change of zeta-potential values. Data are reported as means ± SD, n = 3.



Figure S8. The NTA result of size distribution and the corresponding concentration of $Ma@(MnO_2+FTY)$ nanoparticles.



Figure S9. Membrane surficial proteins in macrophage membrane (1), macrophage membrane vesicles (2) and $Ma@(MnO_2+FTY)$ nanoparticles (3), analyzed with western-blots.



Immunostaining CD11b (extracellular)

No primary stain

Figure S10. Representative TEM images of immunostaining which revealed the right-out-side CD11b (extracellular domain) on the Ma@(MnO₂+FTY) nanoparticles.



(Scale bar, 200 nm).

Figure S11. (a) Representative images of MnO_2 and $Ma@MnO_2$ in H_2O , saline and PBS over 72 h. The size and PDI of $Ma@MnO_2$ (b) and $Ma@(MnO_2+FTY)$ (c) in PBS over 3 days. Results are presented as means \pm SD, n = 3).



Figure S12. The production of O_2 after adding different concentration of MnO_2 into H_2O_2 (100 mM) solution for 5 min. The concentrations of MnO_2 from left to right are 100, 50, 25, 12.5, 6.25 and 0 µg mL⁻¹, respectively.



Figure S13. Representative TEM images of Ma@(MnO₂+FTY) after incubation in PBS 6.0 with 100 μ M H₂O₂ at 37 °C for 30 min.



Figure S7. (a) T1-weighted MR images of Ma@(MnO₂+FTY) after incubation in PBS 6.0 and 7.4, all with 100 μ M H₂O₂. (b) T1 relaxation rate (1/T1) raised linearly with the concentration of Mn²⁺ in Ma@(MnO₂+FTY) nanoparticles processed with 100 μ M H₂O₂ in PBS 6.0, r1 were 54.30 mm⁻¹ s⁻¹ and 3.745 mm⁻¹ s⁻¹ for Ma@(MnO₂+FTY) at pH 6.0 and 7.4, respectively.



Figure S15. The biocompatibility investigation of Ma@MnO₂ on SH-SY5Y cells, cells were incubated with Ma@MnO₂ in different concentration for 1 day. Results are presented as means \pm SD, n = 5, *P < 0.05.



Figure S16. Representative images of the H₂D-CFDA fluorescence in SH-SY5Y cells treated with OGD/R or different concentration of Ma@MnO₂ (Scale bar, 100 μ m).



Figure S17. The generation of O₂ in SH-SY5Y cells treated with OGD/R, cells in 96-well plates were incubated with Ma@MnO₂ of different concentration for 30 min, and the fluorescence intensity at 620 nm was analyzed with a microplate reader. Data are reported as means \pm SD, n = 6, ***P* < 0.01.



Figure S18. The safety of Ma@MnO₂, Ma@(MnO₂+FTY), and FTY on BV2 cells, cells were incubated with drugs in different concentration for 24 hours. Results are presented as means \pm SD, n = 5, ***P* < 0.01.



Figure S19. The internalization mechanism of Ma@(MnO₂+FTY) in BV2 cells. a) The representative flow cytometry analysis results of the fluorescence intensity in BV2 cells with different treatment. b) The semi-quantitative results of the flow cytometry analysis. Data are presented as means \pm SD, n = 3, ***P* < 0.01, ****P* < 0.001.



Figure S20. (a) Representative images of cellular uptake in BV2 cells after incubated with Ma@MnO₂ or Ma@(MnO₂+FTY) for 0.5 h, 1 h, (Scale bar, 100 μ m). (b) The flow cytometry results of cellular uptake, treated with above nanoparticles for different time.



Figure S21. Representative images of the H_2D -CFDA fluorescence in BV2 cells treated with OGD/R or different formulations (Scale bar, 100 μ m).



Figure S22. (a) Representative immunofluorescence images of primary microglia stained with CD11b (Scale bar, 100 μ m). (b) The flow cytometry results of primary microglia stained with CD11b, confirming cell purity greater than 97%.



Figure S23. The bio-distribution of Ma@(MnO₂+FTY) in main organs of tMCAO/R

rats. The ID % of RM@(MnO₂+FTY) nanoparticles were applied as a control. Data are presented as means \pm SD, n = 3, ***P* < 0.01.



Figure S24. T1-weighted MR images of tMCAO/R rat after injection with $Ma@(MnO_2+FTY)$ for 4 h.



Figure S25. Treated with Ma@(MnO₂+FTY) promoted the polarization of activated microglia from M1 to M2 in ischemic brain. a) Representative images of microglia

phenotype polarization analyzed with flow cytometry. b) The gating strategy of $CD45^{int}CD11b^+$ microglia. c) The semi-quantitative results of the flow cytometry analysis. Data are reported as means \pm SD, n = 3, ****P* < 0.001.



Figure S26. Representative images of the expression of p-P65 in microglia in ischemic hemisphere (Scale bar, $100 \ \mu m$).



Figure S27. Expression of p-P65 in the ischemic hemisphere of MCAO rats after

treated with different drugs.



Figure S28. The neurological assessment of tMCAO/R rats treated with different formulations. Data are presented as means \pm SD, n = 3, **P* < 0.01, ***P* < 0.01.



Figure S29. Representative images of hemolysis experiment with different formulations. Data are presented as means \pm SD, n = 3.



Figure S30. The level of AST and ALT in rats after treated with different formulations. Results are reported as means \pm SD, n = 3, *P < 0.05.



Figure S31. The H&E staining images of main organs with full view.



Figure S32. Representative images of main organs with H&E staining after the tMCAO/R rats were treated with different formulations (Scale bar, 250 μ m).

Table S1. Pharmacokinetic parameters of free FTY and Ma@(MnO₂+FTY) nanoparticles at a dose of 1.5 mg Kg⁻¹. Data are presented as means \pm SD, n = 3.

Formulation	AUC _{0-∞}	MRT	t _{1/2}
	[mg/L×h]	[h]	[h]
Free FTY	2.285 ± 0.274	7.676 ± 0.437	14.597 ± 1.623
Ma@(MnO ₂ +FTY)	7.803 ± 1.841	16.680 ± 0.662	30.208 ± 2.804