



**Original Article**

**Prognostic Value of Brain and Acute Leukemia Cytoplasmic Gene Expression in Egyptian Children with Acute Myeloid Leukemia**

Adel A. Hagag<sup>1</sup> and Amal Ezzat Abd El-Lateef<sup>2</sup>

Pediatrics<sup>1</sup> and Clinical Pathology<sup>2</sup> Departments, Faculty of Medicine, Tanta University, Egypt

**Abstract. Background:** Acute myeloid leukemia (AML) accounts for 25%-35% of acute leukemia in children. BAALC gene (Brain and Acute Leukemia Cytoplasmic gene) is a recently identified gene on chromosome 8q22.3 that has prognostic significance in AML. The aim of this work was to study the impact of BAALC gene expression on prognosis of AML in Egyptian children. **Patients and methods:** This study was conducted on 40 Egyptian children with newly diagnosed AML who were subjected to full history taking, clinical examination and laboratory investigations including: complete blood count, LDH, bone marrow aspiration, cytochemistry, immunophenotyping and assessment of BAALC Gene by real time PCR in bone marrow aspirate mononuclear cells before the start of chemotherapy.

**Results:** Positive BAALC gene expression was found in 24 cases (60%) and negative expression in 16 cases (40%). Positive BAALC gene expression group includes 14 males and 10 females with mean age at presentation of 8.35±2.63 while negative BAALC gene expression includes 10 males and 6 females with mean age at presentation of 7.74±3.23 with no statistically significant differences between patients with positive and negative BAALC gene expression regarding age, sex and clinical presentations at time of diagnosis including pallor, purpura, splenomegaly, hepatomegaly and lymphadenopathy and laboratory investigations including WBCs and platelets counts, hemoglobin and LDH levels, and peripheral blood and bone marrow blast cell counts. There was significant association between positive BAALC gene expression and M1 and M2 compared with negative BAALC gene expression which is significantly associated with M4. There were statistically significant differences in disease outcome between positive and negative BAALC gene expression groups with higher rate of relapse and death and lower rate of complete remission and disease free survival in positive BAALC gene expression group compared with negative BAALC gene expression group. (p = 0.017).

**Conclusion and Recommendation:** BAALC expression is an important bad prognostic factor in AML patients with normal karyotype and therefore we recommend its incorporation into novel risk-adapted therapeutic strategies to improve the currently disappointing cure rate of patients with AML.

**Citation:** Hagag A.A., El-Lateef A.E. Prognostic Value of Brain and Acute Leukemia Cytoplasmic Gene Expression in Egyptian Children with Acute Myeloid Leukemia. *Mediterr J Hematol Infect Dis* 2015, 7(1): e2015033, DOI: <http://dx.doi.org/10.4084/MJHID.2015.033>

**Published:** April 20, 2015

**Received:** March 4, 2015

**Accepted:** April 6, 2015

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Correspondence to: Dr. Adel A Hagag. Pediatrics Department, Faculty of Medicine, Tanta University, Egypt, E-mail: adelhagag20@yahoo.com

**Introduction.** Acute myeloid leukemia (AML) is a clonal malignant disease of the bone marrow in which hematopoietic progenitor cells are arrested at an early stage of development due to acquired genetic

alterations that lead to failure of differentiation and to over proliferation.<sup>1</sup> AML accounts for 25%-35% of the acute leukemia in children.<sup>2</sup>

Over the past decades, remarkable progress has been made in the treatment and the understanding of the molecular pathogenesis of acute myeloid leukemia. At present, up to 65% of pediatric AML patients experience long-term survival, owing to a more effective use of anti-leukaemic therapy, improvements in supportive care and better risk stratification<sup>(1)</sup>.

It is important to identify prognostic markers that predict patient's outcome more precisely, thereby allowing the development of molecular risk-adapted treatment strategies that may improve the clinical outcome. By the use of molecular genetics techniques, such as reverse transcriptase polymerase chain reaction (RT-PCR), global gene expression profiling and/or direct sequencing, several recurring molecular alterations of prognostic significance have been identified in patients with cytogenetically normal AML (CN-AML).<sup>3</sup>

BAALC gene (Brain and Acute Leukemia Cytoplasmic gene), is located on chromosome 8q22.3.<sup>4</sup> Expression of BAALC was found mainly in neuroectoderm-derived tissues and hematopoietic precursor cells. In hematopoietic cells, BAALC expression was restricted to the compartment of progenitor cells, whereas no expression was detected in mature bone marrow or circulating white blood cells.<sup>5</sup>

High BAALC gene expression was found in a subset of patients with AML, acute lymphoblastic leukemia (ALL), and blast phase of chronic myeloid leukemia (CML), whereas no BAALC expression could be detected in patients with chronic-phase CML or chronic lymphocytic leukemia (CLL)<sup>(4)</sup>. Additionally, high BAALC gene expression occurs in glioblastoma, melanoma, and childhood gastrointestinal stroma tumors, suggesting an oncogenic role for BAALC gene. However, the mechanisms underlying the deregulated expression are unknown.<sup>6</sup>

The prognostic significance of BAALC gene was first shown in CN-AML with association with significant higher refractoriness to induction treatment, lower rates of complete remission (CR), poor overall survival (OS) and disease free survival (DFS) for patients with high BAALC expression independent of other prognostic molecular markers with a gene expression signature consistent with less differentiated AML blasts.<sup>6-8</sup>

**Aim of the Work.** The aim of this work was to evaluate the prognostic value of BAALC gene expression in Egyptian children with acute myeloid leukemia.

**Subjects and Method.** Approval for this study was granted by the ethical committee of Tanta University research center and written consent was obtained from the parents of all children involved in this study. The

study participants included 40 Egyptian children with newly diagnosed AML being followed up under the Oncology Unit of the Pediatric Department in the period from March 2012 to December 2014 including 24 males and 16 females with their ages ranging from 3-16 years with a mean age value of  $9.8 \pm 5.8$  years. All patients were subjected to follow up for 2 years to evaluate their prognosis.

*Inclusion criteria.* Children with cytogenetically normal AML.

All patients were subjected to the following:

- ❖ Full history taking.
- ❖ Thorough clinical examination with special attention to fever, pallor, purpura, hepatomegaly, splenomegaly, and lymphadenopathy.
- ❖ Laboratory investigations

*Specimen collection and handling.* Three ml venous blood were collected under complete aseptic technique. They were delivered into 2 tubes: 1 ml blood into a tube containing EDTA for complete blood count and 2 ml blood into the plain tube for assessment of Lactate dehydrogenase levels. Two ml of bone marrow aspirate were drawn into a sterile tube containing EDTA for mononuclear cell separation for polymerase chain reaction (PCR).

Laboratory investigations include the following:

- ◆ Complete blood count.
- ◆ Lactate dehydrogenase (LDH)
- ◆ Bone marrow aspiration with cytochemical examination and immunophenotyping.
- ◆ Cytogenetic analysis

Cytogenetic analyzes of bone marrow or peripheral blood were performed. Metaphase chromosomes were banded by G-banding technique and Karyotyped according to the International System for Human Cytogenetic Nomenclature. A minimum of 20 metaphases was required to be examined for any patient to be classified as having a normal cytogenetic study<sup>(9)</sup>.

◆ Assessment of BAALC Gene by real-time PCR.

Mononuclear cells were separated from the samples by centrifugation on Density gradient medium<sup>(10)</sup>. RNA was isolated using an RNA easy Mini Kit, and the concentration of extracted RNA was evaluated by spectrophotometry (SPEC)<sup>(11)</sup>. DNA amplification was done by real-time PCR using Gene Amp 5700 Sequence Detection System. Real-time PCR was used for the detection of BAALC gene<sup>(12)</sup>.

BAALC mRNA expression was normalized simultaneously analyzing the glucose phosphate isomerase (GPI) gene. The relative BAALC expression was determined using the comparative cycle threshold method. BAALC were amplified using 1  $\mu$ L cDNA, 1 $\times$  master mix (IQ Mix; BioRad, Munich, Germany).

- Glucose phosphate isomerase (GPI) forward primer  
5'-TCTTCGATGCCAACAAGGAC-3
- Glucose phosphate isomerase (GPI) reverse primer  
5'-GCATCACGTCCTCCGTCAC-3
- Glucose phosphate isomerase (GPI) probe  
-<sup>3</sup>HEX-TTCAGCTTGACCCTCAACACCAAC-TAMRA-<sup>5</sup>
- BAALC Gene forward primer:  
5'-GCCCTCTGACCCAAACAG-3';
- BAALC Gene reverse primer:  
5'-CTTTTGCAGGCATTCTCTTAGCA-3';
- BALCC Gene probe:  
5'-FAMCTCTTTTAGCCTCTGTGGT-3';

Reactions were performed using real-time PCR 7000 sequence detection system (Applied Biosystems, Foster City, CA, USA) <sup>(12)</sup>. A value of 0.166 for BAALC gene was attributed as cut off value, so BAALC gene expression was considered positive if BAALC gene expression is above 0.166. On the contrary, BAALC gene expression was considered negative if BAALC gene expression is below 0.166. Follow up of patients was carried out clinically and by blast count in BM on day 28 after two courses of induction therapy according to AML-protocol (MRC10). <sup>(13, 14)</sup>.

#### Course 1 (ADE)

Aracytin 100 mg/m<sup>2</sup> IV bolus every 12 hours on days 1-10 (20 doses), Daunorubicin 50 mg/m<sup>2</sup> IV on days 1, 3, 5, Etoposide 100 mg/m<sup>2</sup> (1 hour IV infusion) on days 1-5 and age adjusted intrathecal Aracytin at time of diagnostic lumbar puncture.

#### Course 2 (ADE)

Aracytin 100 mg/m<sup>2</sup> IV bolus every 12 hours on days 1-8 (16 doses), Daunorubicin 50 mg/m<sup>2</sup> IV daily on days 1, 3, 5, Etoposide 100 mg/m<sup>2</sup> daily (1 hour IV infusion) on days 1-5 and age-adjusted intrathecal Aracytin on day 1 of course 2.

#### Course 3 (ACE)

Aracytin 1gm/m<sup>2</sup>/dose IV every 12 hours on days 1-5 (10 doses) and Etoposide 150 mg/m<sup>2</sup>/dose daily (1 hour IV infusion) on days 1-5 with age-adjusted intrathecal Aracytin on day 1 of course 3.

#### Course 4 (MIDAC)

Mitoxantrone 10 mg/m<sup>2</sup> IV daily (short infusion) days 1-5, Aracytin 1gm/m<sup>2</sup> /12hours (2 hours IV infusion) on days 1-3 (6 doses) and age-adjusted intrathecal Aracytin on day 1 of course 4.

#### Course 5 (CLASP)

Aracytin 3gm/m<sup>2</sup> IV every 12 hours on days 1, 2, 8 and 9 (8 doses), L-asparaginase 6,000 IU/m<sup>2</sup> IM on days 2 and 9 (3 hours after completion of Aracytin).

Age-adjusted intrathecal chemotherapy with Aracytin: 20 mg for age less than 1 year; 30 mg for age of 1- 2 years; 50 mg for age of 2 - 3 years and 60 mg for age of 3 year or older.

**Statistical Analysis.** The patient's data were collected and statistically analyzed using SPSS software statistical computer package version 12. All Data were expressed as in terms of mean values  $\pm$  SD. The difference between two means was statistically analyzed using the student (t) test. A chi-square test ( $X^2$ ) and Fischer exact test was used as a test of significance. The log-rank test was used to assess survival. Significance was adopted at  $p \leq 0.05$ .

**Results.** Positive BAALC gene expression was found in 24 cases (60%) and negative expression in 16 cases (40%).

Positive BAALC gene expression group (n=24) includes 14 males and ten females with a mean age at presentation of 8.35 $\pm$ 2.63. Pallor was found in 20 cases, purpura in 22 cases, splenomegaly in 19 cases, hepatomegaly in 16 cases and lymphadenopathy in 8 cases. The negative BAALC gene expression group (n=16) includes ten males and six females with a mean age at presentation of 7.74 $\pm$ 3.23. In this group pallor was found in 13 cases, purpura in 14 cases, splenomegaly in 13 cases, hepatomegaly in 11 cases and lymphadenopathy in 6 cases. There were no statistically significant differences between patients with positive and negative BAALC gene expression regarding age, sex and clinical presentations at the time of diagnosis. (**Table 1**).

There were no significant differences between positive and negative BAALC gene expression groups regarding WBCs and platelets counts, hemoglobin, and LDH levels, and peripheral blood and bone marrow blast cell counts. (The mean WBCs count was 75.51 $\pm$ 51.31 in the positive BAALC gene group versus and 76.44 $\pm$ 46.67 in the negative BAALC gene group, with a p-value of 0.845. The mean platelets count was 69.86 $\pm$ 33.15 in the positive BAALC gene expression group versus 77.13 $\pm$ 48.07 in the negative BAALC gene group, with a p-value of 0.712. The mean hemoglobin level was 8.13 $\pm$ 1.57 in the positive BAALC gene group versus 7.86 $\pm$ 2.37 in the negative BAALC gene group, with a p-value of 0.632. The mean LDH level was 1312.73 $\pm$  695.63 in the positive BAALC gene group versus 1299.57 $\pm$ 315.47 in the negative BAALC gene expression group, with a p-value of 0.082. The mean peripheral blood blast cells in the positive BAALC gene expression group was 63  $\pm$ 20.05 versus 57.35 $\pm$ 28.48 in the negative BAALC gene expression group, with a p-value of 0.416. The mean bone marrow blast cells in the positive BAALC gene expression group was 78.21 $\pm$ 9.83 versus 75.39 $\pm$ 21.79 in the negative BAALC gene expression group with p-value of 0.096) (**Table 1**).

There was a significant association between positive BAALC gene expression and M1 and M2 subtypes compared with negative BAALC gene expression significantly associated with the M4 subtype. In fact, of 24 cases with positive BAALC gene expression; 9

were M1, 13 were M2, and 3 M4 whereas of 16 patients with negative BAALC gene expression; 2 were M1, 4 M2, and 9 M4. The presence of positive and negative BAALC were statistically different in the FAB subtypes) (Table 1).

There was a statistically significant difference in disease outcome between positive and negative BAALC gene expression groups with higher rate of relapse and death and lower rate of complete remission and disease free survival in positive BAALC gene

expression group compared with negative BAALC gene expression group. ( $p = 0.017$ ). Of 24 patients positive; 10 achieved complete remission, 8 died and 6 suffered from relapse, while of 16 patients negative; 12 achieved complete remission, 1 suffered from relapse and 3 died with a median disease-free survival in the positive group of 6.03 months compared with 21.63 months in negative group. (Table 2, 3 and Figure 1, 2).

**Table 1.** Comparison between patients with positive and negative BAALC gene expression regarding clinical and laboratory data.

Parameters		Positive BAALC gene expression group (n=24)		Negative BAALC gene expression group (n=16)		t test or X <sup>2</sup>	P- value
		N	%	N	%		
Sex	Males	14	35%	10	25%	0.260	0.610
	Females	10	25%	6	15%		
Age (years)	Mean ± SD	8.35±2.63		7.74±3.23		0.536	0.336
Pallor	Present	20	83.3%	13	81.25%	0.20	0.654
Purpura	Present	22	91.6%	14	87.5%	2.53	0.112
Lymphadenopathy	Present	8	33.33%	6	37.5%	0.361	0.547
Hepatomegaly	Present	16	66.66%	11	68.75%	0.095	0.758
Splenomegaly	Present	19	79.16%	13	81.25	0.49	0.481
Hb (gm/dl)	Mean ±SD	8.13±1.57		7.86±2.37		0.96	0.632
Platelets (x10 <sup>3</sup> /mm <sup>3</sup> )	Mean ±SD	69.86±33.15		77.13±48.07		1.85	0.712
TLC (x10 <sup>3</sup> /mm <sup>3</sup> )	Mean ±SD	75.51±51.31		76.44±46.67		0.93	0.845
Blast cells (%) in peripheral blood	Mean ±SD	63 ±20.05		57.35±28.48		0.833	0.416
BM blast cells (%)	Mean ±SD	78.21±9.83		75.39±21.79		0.924	0.096
LDH (U/L)	Mean ±SD	1312.73± 695.63		1299.57±315.47		0.86	0.082
FAB classification							
M1 (11 cases)		9	37.5%	2	12.5%	3.510	0.04*
M2 (17 cases)		13	54.16%	4	25%	4.331	0.02*
M4 (12 cases)		3	12.5%	9	56.25%	3.359	0.031*

\* Significant ( $p < 0.05$ ). SD=Standard deviation. TLC= Total leucocytic count. BM=bone marrow. LDH=lactate dehydrogenase.

**Table 2.** Outcome of studied patients in relation to BAALC gene expression.

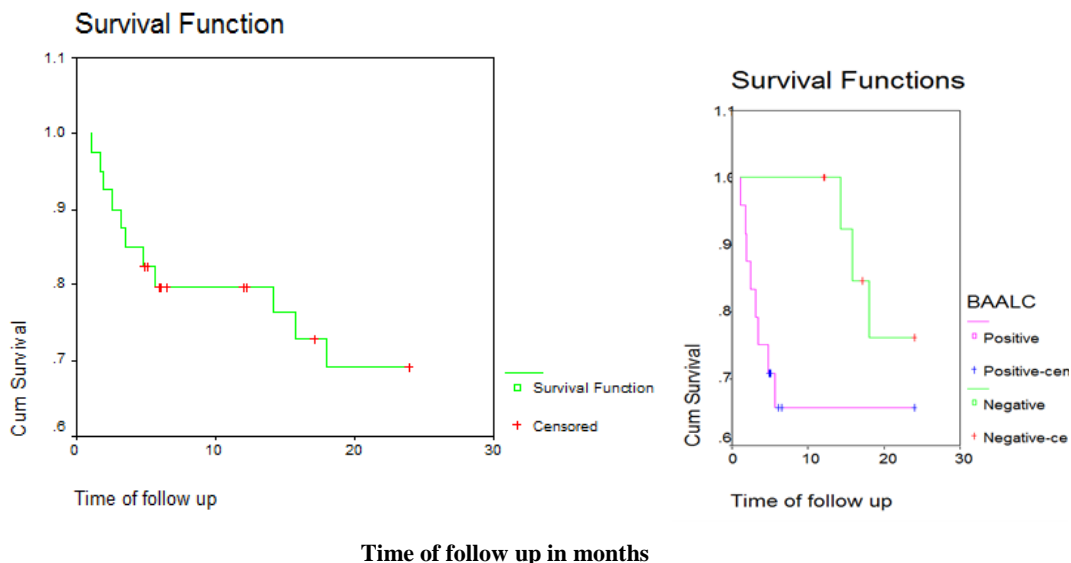
BAALC gene expression	Patients outcome (No=40)		
	Death (11 cases)	Remission (22 cases)	Relapse (7 cases)
Positive BAALC gene expression group (No=24) (60%)	8 (20)	10 (25)	6 (15)
Negative BAALC expression group (No=16) (40%)	3 (7.5)	12 (30)	1 (2.5)
Total Number = 40 (100%)	11 (27.5)	22 (55)	7 (17.5)
Chi-square	X <sup>2</sup>	5.421	
	P-value	0.017*	

\*Significant.

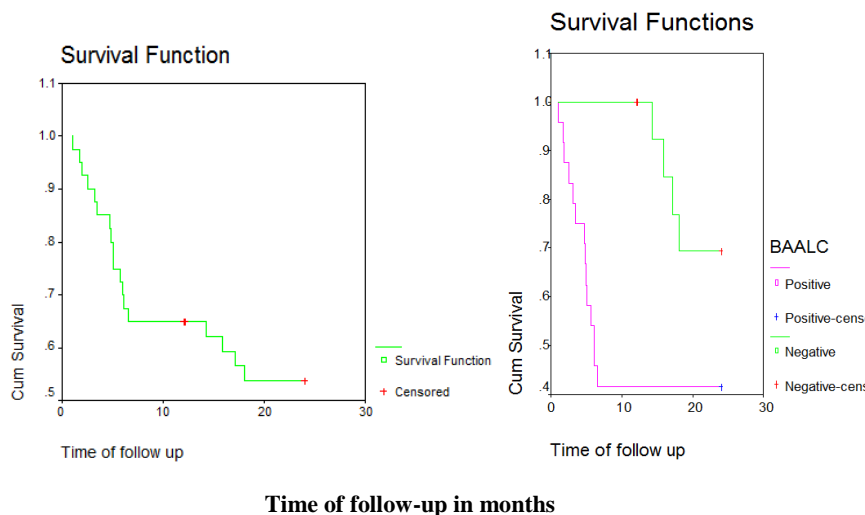
**Table 3.** Log Rank test of overall and disease-free survival.

	Overall survival			Log Rank	
	Median	SE	CI 95%	test value	P-value
Negative	22.110	0.970	(20.21-24.01 )	1.750	0.185
Positive	16.840	2.070	(12.79- 20.90)		
	Disease free survival			Log Rank	
	Median	SE	CI 95%	test value	P-value
Negative	21.630	1.010	( 19.650-23.610 )	5.450	0.020*
Positive	6.030	0.880	( 4.310-7.750 )		

\*Significant. SE=Standard error. CI= Confidence Interval.



**Figure 1.** Kaplan Meir curve showing overall survival (OAS) in positive and negative BAALC gene expression groups.



**Figure 2.** Kaplan Meir curve showing Disease Free Survival (DFS) in positive and negative BAALC gene expression groups.

**Discussion.** Acute myeloid leukemia is a clonal malignant disease of hematopoietic tissue that is characterized by the proliferation of abnormal myeloblast cells principally in marrow and impaired production of normal blood cells<sup>(15)</sup>. The prognosis of AML varies dramatically and is strongly influenced by a number of factors, including age, performance status, and cytogenetic and/or molecular alterations.<sup>16</sup>

In recent years, a major focus of molecular cancer research has been the analysis of genes that may be the cause of carcinogenesis (oncogenes).<sup>17</sup> Multiple chromosomal and gene rearrangements have been identified in AML, such as MLL, PML/RARA, DEK/CAN, and AML1/ETO<sup>18</sup> Chromosomal rearrangements involving the MLL gene at band 11q23 are the most common genetic alteration encountered in



infant acute myeloid leukemia. Reciprocal translocation represents the most frequent form of MLL rearrangement.<sup>19</sup> NRAS mutation varies considerably in patients with childhood AML and was found in about 15% of pediatric AML patients in some studies.<sup>17, 20</sup>

The present study was designed to use Real-time PCR analysis to study the prognostic value of BAALC gene expression in Egyptian children with AML.

In this study, positive BAALC gene expression was found in 24 cases (60%). This is in substantial accord with Yahya et al 2013,<sup>21</sup> who found high BAALC gene expression in 22 of 45 patients (48.9%) and Damiani et al 2013,<sup>22</sup> who found BAALC gene overexpression in 87/175 (50%) of their studied patients.

In the present study, there were no statistically significant differences between patients with positive and negative BAALC gene expression regarding age, sex and clinical presentation at time of diagnosis including pallor, purpura, hepatomegaly, splenomegaly, and lymphadenopathy. These results were in agreement with Yahya et al. 2013<sup>21</sup> who found no significant differences between positive and negative BAALC gene expression regarding clinical parameters of patients at the time of diagnosis.

In this work, there were no significant differences between positive and negative BAALC gene expression regarding WBCs and platelets counts, hemoglobin, and LDH levels, and blast cell counts in the peripheral blood and bone marrow. This is in agreement with Elsharnouby et al. 2010<sup>23</sup> who found no significant differences between positive and negative BAALC gene expression regarding WBCs and platelets counts, hemoglobin, and LDH levels, blast cell counts, both in the peripheral blood and bone marrow. But not in agreement with Baldus et al. 2003<sup>24</sup> who found the association between positive BAALC gene expression and significantly higher WBCs and blast cell counts in the peripheral blood and bone marrow.

Variation between the results of this study and the previous studies may be explained by different age and

number of studied patients, different localities, different presentations of leukemia, different duration of studies and different duration of follow-up.

In the current study, there was a significant association between BAALC gene expression and certain FAB subtypes with predominant positive BAALC gene expression in M1 and M2 and predominant negative BAALC gene expression in M4. These data are in agreement with Bienz et al 2005<sup>26</sup> and Elsharnouby et al 2010<sup>(23)</sup> who both found high BAALC expression more frequently in M1, M2 and less in M4, M5, and Yanaihara et al 2006<sup>25</sup> who stated that high BAALC gene expression is more often present in M1.

In our study there were statistically significant differences in disease outcome between positive and negative BAALC gene expression groups with higher rate of relapse and death and lower rate of complete remission and disease free survival in positive BAALC gene expression group compared with negative BAALC gene expression group. This is in agreement with Nibourel et al 2010<sup>27</sup> who stated that BAALC gene expression was found to be an independent negative prognostic factor in CN-AML, Yahya et al 2013<sup>21</sup> who found that high BAALC expression had significantly lower incidence of CR, higher mortality rate, significantly shorter DFS, and inferior overall survival, Weber et al 2014<sup>28</sup> who revealed an independent adverse prognostic impact of high BAALC expression on overall survival and event-free survival and Damiani et al 2013<sup>22</sup> who found that overexpression of BAALC gene confers poor prognosis in cytogenetically normal AML patients, with negative impact on CR achievement, overall survival but have no influence on relapse probability.

**Conclusion and Recommendation.** BAALC expression is an important bad prognostic factor in AML patients with normal karyotype, and therefore we recommend its incorporation into novel risk-adapted therapeutic strategies to improve the currently disappointing cure rate of patients with AML.

## References:

1. Arber DA, Brunning RD, Orazi A, Le Beau M. Acute myeloid leukemia with myelodysplasia-related changes. In: Swerdlow SH, Campo E, Harris NL (eds.), WHO Classification of Tumors of Hematopoietic and Lymphoid Tissues. (4th edition). 2008b: 124-126.
2. Juliusson G, Lazarevic VL, Horstedt AS, Hagberg O, Höglund M, Swedish Acute Leukemia Registry Group. Acute myeloid leukemia in the real world: Why population-based registries are needed. *Blood* 2012 Apr 26; 119(17):3890-9. doi: 10.1182/blood-2011-12-379008. Epub 2012 Mar 1. <http://dx.doi.org/10.1182/blood-2011-12-379008>
3. Mrozek K, Marcucci G, Paschka P, Whitman SP, Bloomfield CD. Clinical relevance of mutations and gene-expression changes in adult acute myeloid leukemia with normal cytogenetics: Are we ready for a prognostically prioritized molecular classification? *Blood* 2007; 109:431-448. <http://dx.doi.org/10.1182/blood-2006-06-001149> PMID:16960150 PMCid:PMC1785102
4. Tanner SM, Austin JL, Leone G, Rush LJ, Plass C, Heinonen K, Mrózek K, Sill H, Knuutila S, Kolitz JE, Archer KJ, Caligiuri MA, Bloomfield CD, de La Chapelle. BAALC, the human member of a novel mammalian neuroectoderm gene lineage, is implicated in hematopoiesis and acute leukemia. *Proc Natl Acad Sci USA* 2001 Nov 20; 98(24):13901-6. Epub 2001 Nov 13. <http://dx.doi.org/10.1073/pnas.241525498> PMID:11707601 PMCid:PMC61139
5. Ilencíková D, Sykora J, Mikulaon Z, Repiská V. Identification of molecular markers in children with acute myeloid leukemia. *Clin Oncol*. 2012; 25(1):26-35.
6. Eisfeld AK, Marcucci G, Liyanarachchi S, Döhner K, Schwind S, Maharry K, Leffel B, Döhner H, Radmacher MD, Bloomfield CD, Tanner SM, de la Chapelle A. Heritable polymorphism predisposes to high BAALC expression in acute myeloid leukemia. *Proc Natl Acad Sci USA* 2012 Apr 24; 109(17):6668-73. Epub 2012 Apr 9. <http://dx.doi.org/10.1073/pnas.1203756109>
7. Baldus CD, Martus P, Burmeister T, Schwartz S, Gökbueget N, Bloomfield CD, Hoelzer D, Thiel E, Hofmann WK. Low ERG and

- BAALC expression identifies a new subgroup of adult acute T Lymphoblastic leukemia with a highly favorable outcome. *J Clin Oncol*. 2007 Aug 20; 25(24):3739-45. Epub 2007 Jul 23. <http://dx.doi.org/10.1200/JCO.2007.11.5253> PMID:17646667
8. Santamaría C, Chillón MC, García-Sanz R, Pérez C, Caballero MD, Mateos MV, Ramos F, de Coca AG, Alonso JM, Giraldo P, Bernal T, Queizán JA, Rodríguez JN, Puig N, Balanzategui A, Sarasquete ME, Alcoceba M, Díaz-Mediavilla J, San Miguel J, González M. BAALC is an important predictor of refractoriness to chemotherapy and poor survival in intermediate-risk acute myeloid leukemia. *Ann Hematol*. 2010 May; 89(5):453-8. Epub 2009 Nov 27. <http://dx.doi.org/10.1007/s00277-009-0864-x>
  9. Byrd JC, Mrózek K, Dodge RK, Carroll AJ, Edwards CG, Arthur DC, Pettenati MJ, Patil SR, Rao KW, Watson MS, Koduru PR, Moore JO, Stone RM, Mayer RJ, Feldman EJ, Davey FR, Schiffer CA, Larson RA, Bloomfield CD; Cancer and Leukemia Group B (CALGB 8461). Pretreatment cytogenetic abnormalities are predictive of induction success, cumulative incidence of relapse, and overall survival in adult patients with de novo acute myeloid leukemia: Results from Cancer and Leukemia Group B (CALGB 8461). *Blood* 2002 Dec 15; 100(13):4325-36. Epub 2002 Aug 1. <http://dx.doi.org/10.1182/blood-2002-03-0772> PMID:12393746
  10. Boyam A. Isolation of mononuclear cells and granulocytes from human blood. Isolation of mononuclear cells by one centrifugation and of granulocytes by combining centrifugation and sedimentation at 1g. *Scand J Clin Lab Invest Suppl*. 1968; 97:77-89
  11. Guo X, Shi P, Chen F, Zha J, Liu B, Li R, Dong H, Zheng H, Xu B. Low MDR1 and BAALC expression identifies a new subgroup of intermediate cytogenetic risk acute myeloid leukemia with a favorable outcome. *Blood Cells Mol Dis*. 2014 Sep; 53(3):144-8. Epub 2014 May 20. <http://dx.doi.org/10.1016/j.bcmd.2014.05.001>
  12. Baladus A., Thiede C, Soucek S. BAALC expression and FLT-3 internal tandem duplication mutations in acute myeloid leukemia patients with normal cytogenetic: Prognostic implications. *J Clin Oncol*. 2006; 24:790-7. <http://dx.doi.org/10.1200/JCO.2005.01.6253> PMID:16418499
  13. Hann IM, Webb DK, Gibson BE, Harrison CJ. MRC trials in childhood acute myeloid leukaemia. *Ann Hematol*. 2004; 83 (Suppl 1): S108-12. PMID:15124698
  14. Gibson BE, Webb DK, Howman AJ, De Graaf SS, Harrison CJ, Wheatley K; United Kingdom Childhood Leukaemia Working Group and the Dutch Childhood Oncology Group. Results of a randomized trial in children with Acute Myeloid Leukemia: medical research council AML12 trial. *Br J Haematol*. 2011 Nov; 155(3):366-76. Epub 2011 Sep 9. <http://dx.doi.org/10.1111/j.1365-2141.2011.08851.x>
  15. Ishikawa Y, Kiyoi H, Naoe T. Prevalence and clinical characteristics of N-terminally truncated WT1 expression in acute myeloid leukemia. *Leuk Res*. 2011 May; 35(5):685-8. Epub 2011 Jan 21. <http://dx.doi.org/10.1016/j.leukres.2011.01.002>
  16. Döhner H, Estey EH, Amadori S, Appelbaum FR, Büchner T, Burnett AK, Dombret H, Fenaux P, Grimwade D, Larson RA, Lo-Coco F, Naoe T, Niederwieser D, Ossenkoppele GJ, Sanz MA, Sierra J, Tallman MS, Löwenberg B, Bloomfield CD, European Leukemia Net. Diagnosis and management of AML in adults: Recommendation from an international expert panel, on behalf of the European leukemia net. *Blood* 2010 Jan 21; 115(3):453-474. Epub 2009 Oct 30. <http://dx.doi.org/10.1182/blood-2009-07-235358>
  17. Aly RM, El-Sharnoby MR, Hagag AA. Prognostic Significance of NRAS Gene Mutations in Children with Acute Myelogenous leukemia. *Mediterr J Hematol Infect Dis*. 2011; 3(1):e2011055. <http://dx.doi.org/10.4084/mjihid.2011.055>
  18. Mrózek K, Heinonen K, Bloomfield CD. Clinical importance of cytogenetics in acute myeloid leukemia. *Best Pract Res Clin Haematol*. 2001; 14:19-47. <http://dx.doi.org/10.1053/beha.2000.0114> PMID:11355922
  19. Launay E, Henry C, Meyer C, Chappé C, Taque S, Boulland ML, Ben Abdelali R, Dugay F, Marschalek R, Bastard C, Fest T, Gandemer V, Belaud-Rotureau MA. MLL-SEPT5 fusion transcript in infant acute myeloid leukemia with t(11;22)(q23;q11). *Leuk Lymphoma* 2014 Mar; 55(3):662-7. <http://dx.doi.org/10.3109/10428194.2013.809528>
  20. Liang DC, Shih LY, Fu JF, Li HY, Wang HI, Hung IJ, Yang CP, Jaing TH, Chen SH, Liu HC. K-ras mutations and N-ras mutations in childhood acute leukemias with or without mixed-lineage leukemia gene rearrangements. *Cancer* 2006; 106: 950-956. <http://dx.doi.org/10.1002/cncr.21687> PMID:16404744.
  21. Yahya RS, Sofan MA, Abdelmasseih HM, Saady N, Sharaf-Eldein MA. Prognostic implication of BAALC gene expression in adult acute myeloid leukemia *Clin Lab*. 2013; 59(5-6):621-8. PMID:23865362
  22. Damiani D, Tiribelli M, Franzoni A, Michelutti A, Fabbro D, Cavallin M, Toffoletti E, Simeone E, Fanin R, Damante G. BAALC overexpression retains its negative prognostic role across all cytogenetic risk groups in acute myeloid leukemia patients. *Am J Hematol*. 2013 Oct; 88(10):848-52. Epub 2013 Jul 23. <http://dx.doi.org/10.1002/ajh.23516>
  23. El-Sharnouby JA, Sayed Ahmed LM, Taha AM, Okasha K. Prognostic Significance of CEBPA Mutations and BAALC Expression in Acute Myeloid Leukemia Patients with Normal Karyotype. *Eur J Gen Med*. 2010; 7(1):17-28.
  24. Baldus CD, Tanner SM, Ruppert AS, Whitman SP, Archer KJ, Marcucci G, Caligiuri MA, Carroll AJ, Vardiman JW, Powell BL, Allen SL, Moore JO, Larson RA, Kolitz JE, de la Chapelle A, Bloomfield CD. BAALC expression predicts clinical outcome of de novo acute myeloid leukemia patients with normal cytogenetic: A Cancer and Leukemia Group B study. *Blood* 2003 Sep 1; 102(5):1613-8. Epub 2003 May 15. <http://dx.doi.org/10.1182/blood-2003-02-0359> PMID:12750167
  25. Yanaihara N, Caplen N, Bowman E, Seike M, Kumamoto K, Yi M, Stephens RM, Okamoto A, Yokota J, Tanaka T, Calin GA, Liu CG, Croce CM, Harris CC. Unique microRNA molecular profiles in lung cancer diagnosis and prognosis. *Cancer Cell* 2006 Mar; 9(3):189-98. <http://dx.doi.org/10.1016/j.ccr.2006.01.025> PMID:16530703
  26. Bienz M, Ludwig M, Leibundgut EO, Mueller BU, Ratschiller D, Solenthaler M, Fey MF, Pabst T. Risk assessment in patients with acute myeloid leukemia and a normal karyotype. *Clin Cancer Res*. 2005 Feb 15; 11(4):1416-24. <http://dx.doi.org/10.1158/1078-0432.CCR-04-1552> PMID:15746041
  27. Nibourel O, Kosmider O, Cheok M, Boissel N, Renneville A, Philippe N, Dombret H, Dreyfus F, Quesnel B, Geffroy S, Quentin S, Roche-Lestienne C, Cayuela JM, Roumier C, Fenaux P, Vainchenker W, Bernard OA, Soulier J, Fontenay M, Preudhomme C. . Incidence and prognostic value of TET2 alterations in de novo acute myeloid leukemia achieving complete remission. *Blood* 2010 Aug 19; 116(7):1132-5. Epub 2010 May 20. <http://dx.doi.org/10.1182/blood-2009-07-234484>
  28. S Weber, T Alpermann, F Dicker, S Jeromin, N Nadarajah, C Eder, A Fasanl, A Kohlmann, M Meggendorfer, C Haferlach, W Kern, T Haferlach and S Schnittger. BAALC expression: a suitable marker for prognostic risk stratification and detection of residual disease in cytogenetically normal acute myeloid leukemia. *Blood Cancer Journal* 2014 January 10; 4, e173; doi:10.1038/bcj.2013.71. <http://dx.doi.org/10.1038/bcj.2013.71>